Figure S7

**A**

<table>
<thead>
<tr>
<th>PPI:</th>
<th>-</th>
<th>-</th>
<th>+</th>
</tr>
</thead>
<tbody>
<tr>
<td>CIP:</td>
<td>-</td>
<td>+</td>
<td>+</td>
</tr>
</tbody>
</table>

**PPI:**

- Immunoblotting
- Rubisco
- P-LIP5
- LIP5

**Phos-tag Immunoblotting**

**B**

<table>
<thead>
<tr>
<th>PPI:</th>
<th>-</th>
<th>-</th>
<th>+</th>
</tr>
</thead>
<tbody>
<tr>
<td>CIP:</td>
<td>-</td>
<td>+</td>
<td>+</td>
</tr>
</tbody>
</table>

**PPI:**

- Immunoblotting
- Rubisco
- P-LIP5
- LIP5

**Phos-tag Immunoblotting**
**Figure S7.** Dephosphorylation of *in vivo* Phosphorylated LIP5 Proteins.

Protein extracts were isolated from transgenic *NtMEK2\textsuperscript{DD}/myc-LIP5\textsuperscript{WT}* at 24 hours after DEX treatment (A) or *lip5-1/myc-LIP5\textsuperscript{WT}* (B) plants at 24 hpi of *PstDC3000*. The protein extracts was treated at 37°C for 45 minutes with calf intestinal alkaline phosphatase (CIP) in the absence or presence of a phosphatase inhibitor cocktail (10 mM NaF, 7 mM β-glycerophosphate and 5 mM Na-pyrophosphate). Reactions without CIP and phosphatase inhibitors (-) were used as control. The protein extracts were subsequently separated on the regular SDS-PAGE and Phos-tag gels for immunoblot analysis using an anti-myc monoclonal antibody. Rubisco staining of the regular SDS-PAGE gel was used for assessing equal protein loading.