



Opinion

Measles Immune Suppression: Functional Impairment or Numbers Game?

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Introduction

Measles remains a significant cause of childhood morbidity and mortality. Hallmark of the disease is a generalized immune suppression that can last for several weeks to months after resolution of measles virus (MV) infection [1–3], resulting in increased susceptibility to opportunistic infections [4–7]. At the same time, measles is associated with immune activation and induces strong MV-specific immune responses that confer lifelong immunity [8]. This contradiction is known as the “measles paradox”. Although measles-associated immune suppression has been a subject of study since the beginning of the 20th century [9], the importance of possible underlying mechanisms remains disputed.

The Immune System as “Viral Friend”

From the perspective of MV, cells of the immune system are both friend and foe. MV efficiently replicates in cells of the immune system, especially during the initial stages of the infection [10,11]. However, the virus preferentially infects specific subsets of lymphocytes and dendritic cells (DCs). The relative susceptibility of these cells to MV infection is governed by their expression level of the cellular receptor CD150 [11–14]. Memory T-lymphocytes, which express CD150, are preferentially infected [13,14]. In secondary and tertiary lymphoid tissues, the virus also replicates to high levels in follicular and marginal zone B-lymphocytes [10,11,13]. DCs can also be infected by MV [11,15–17] and may serve as initial target cells [18,19].

The Immune System as “Viral Foe”

In the majority of cases MV infection is self-limiting and induces strong virus-specific cellular and humoral immune responses resulting in lifelong immunity [20]. Virus neutralizing antibodies are an important correlate of protection against MV infection, but cytotoxic T-lymphocytes are

crucial for clearance of infected cells [21–23]. Resolution of MV infection is associated with increased lymphoproliferation [8,24] and enlargement of lymph nodes [13]. Thus, the immune system efficiently restricts MV replication and clears MV-infected cells.

Mechanisms of Measles Immune Suppression

Measles is associated with lymphopenia [25] and extensive depletion of lymphocytes from lymphoid tissues [13,26,27]. However, lymphocyte numbers return to normal within a week after clinical symptoms of measles have disappeared, while measles immune suppression extends for several weeks to months. Therefore, immune cell depletion was initially dismissed as a mechanism for measles immune suppression [3]. Alternative mechanisms have been proposed to explain measles-associated immune suppression, as summarized in Table 1. However, the relevance of these observations to enhanced susceptibility to opportunistic infections *in vivo* remains unclear.

Is Suppressed Lymphoproliferation Important?

Reduced responsiveness of peripheral blood mononuclear cells to stimulation with mitogens *in vitro* has been considered an important mechanism underlying measles-associated immune suppression.

Although the observations in these studies are not disputed, we find it difficult to reconcile this *in vitro* observation with the observed immune activation *in vivo*. Measles results in dramatic expansion of MV-specific lymphocytes followed by resolution of viremia and lymphopenia [8,25,28]. We recently demonstrated extensive lymphoproliferation in lymphoid tissues early after MV infection *in vivo* [13]. Thus, there is no evidence of suppressed lymphoproliferative responses, at least towards MV, *in vivo*. Rather, we believe that alterations in the composition of the peripheral lymphocyte populations before and after measles may explain these *in vitro* observations [13].

Do Dendritic Dells Play a Crucial Role?

DC subsets have been shown susceptible to MV infection *in vitro* [15–17] and in nonhuman primates *in vivo* [11,19]. Therefore, it is likely that infection, depletion, or functional modulation of DCs contributes to measles-associated immune suppression. Nevertheless, antigen presentation does not seem to be impaired *in vivo*, as strong MV-specific immune responses develop during the acute stage of the disease.

Measles Damages the Respiratory Epithelium

Whereas MV targets CD150 to infect lymphoid and myeloid cells, the virus uses poliovirus receptor like 4 (also known as

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Table 1. Reported mechanisms of measles immune suppression.

Functional Impairment	
<i>Mechanism</i>	<i>References</i>
Suppression of lymphocyte proliferation	[41–45]
Altered cytokine profiles	[42,43,46–50]
Lymphoproliferation defect due to MV-infected DC	[15,17,51]
Immune modulation mediated by viral proteins	[44,52–56]
Modulation of cell membrane components	[57,58]
Inhibition of hematopoiesis	[59,60]
Depletion	
<i>Mechanism</i>	<i>References</i>
Lymphocyte infection & depletion	[11,13,14,46,61–64]
Bystander lymphocyte apoptosis	[65–67]
DC infection & depletion	[11,16,68,69]
T-cell apoptosis after interaction with MV-infected DC	[16,70]

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nectin-4) as an alternative cellular receptor to infect epithelial cells [29–31]. Whilst infection of epithelial cells contributes to

viral transmission [32], MV also causes extensive epithelial damage in the respiratory tract [33,34]. This epithelial injury

may provide an opportunity for respiratory bacteria to adhere, replicate, and invade with increased efficiency [35].

Morbillivirus infections

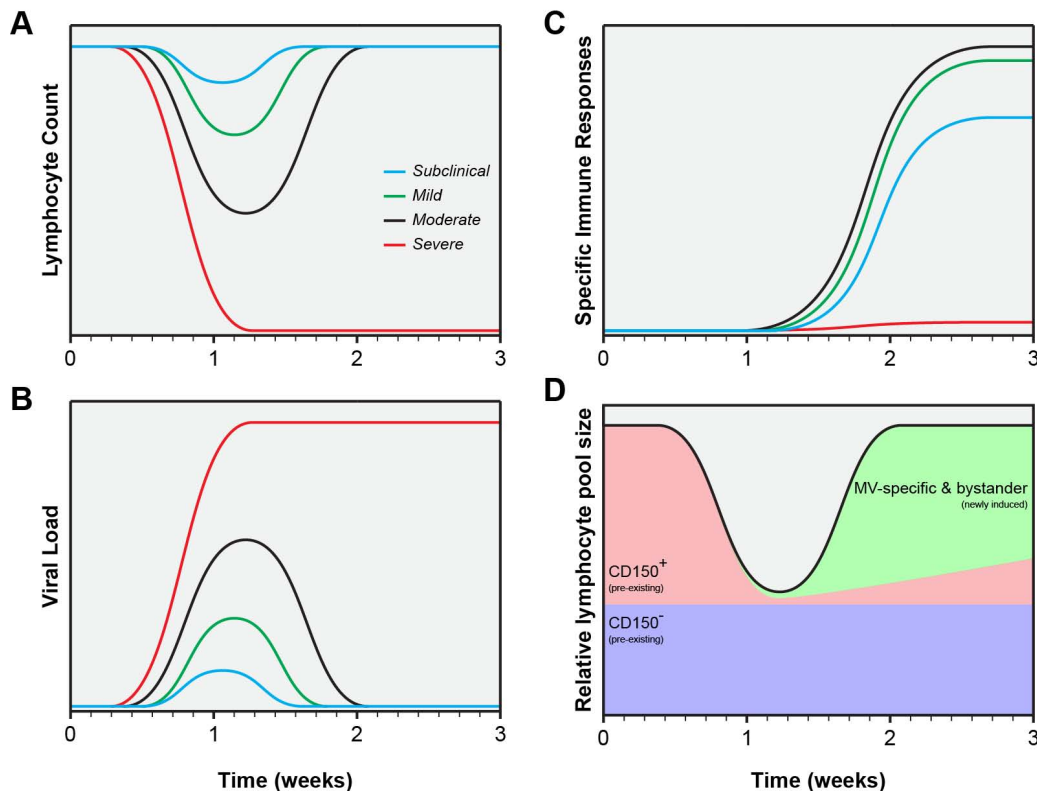


Fig. 1. Schematic representation of the measles paradox. Different levels of lymphopenia (A), systemic virus loads (B), and virus-specific immune responses (C) after subclinical (blue), mild (green), moderate (black), or severe (red) morbillivirus infections. Panel D shows a model for immune suppression caused by moderate morbillivirus infection as shown in panels A, B, and C (adapted from [13]).
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Attenuated, Mild, Moderate, or Severe Morbillivirus Infections

MV infections display a large variability in clinical severity, ranging from vaccination with attenuated viruses, via subclinical or mild infections, to severe disease. Closely related animal morbilliviruses may even overwhelm the immune system, resulting in functional paralysis and virtual absence of virus-specific immune responses [36–40]. This variation is also reflected in a wide range of levels of lymphopenia, viremia, and specific immune responses (Fig. 1A–C) [13]. Natural MV infection of the naive host will normally follow the pattern of either a mild or moderate infection as displayed in Fig. 1. Whereas mild measles results in limited depletion of pre-existing CD150⁺ memory lymphocytes, moderate measles is associated with infection and subsequent depletion of a large fraction of those lymphocytes (Fig. 1D). Whether this depletion is mediated by necrosis, apoptosis, pyroptosis, or cytotoxic T-cells remains

to be determined, but the effect is always the same: to a varying degree, measles erases immunological memory.

Future Directions: Studies in Naturally Infected Measles Patients

To improve our understanding of measles immune suppression, a transition from *in vitro* to *in vivo* studies is required. Two aspects are of crucial importance: viral tropism and depletion of immune cell subsets. We feel that it is important to characterize the phenotype of MV-infected cells during the prodromal phase of natural measles, with special emphasis on infection of DCs and memory lymphocytes. Furthermore, to address depletion of immune cell subsets, paired blood samples from children before and after measles will be required. Staining of immune cells specific for previously encountered pathogens, rather than functional assays, will allow us to

distinguish between subset depletion and functional paralysis.

Conclusions

Experimental MV infections in animal models have demonstrated that percentages of infected lymphocyte subsets are higher than previously thought, especially in secondary and tertiary lymphoid tissues [11,13]. We believe that measles immune suppression mainly results from depletion of immune cell subsets, which is masked by the rapid proliferation of MV-specific and bystander lymphocytes (Fig. 1D). This model is fully compatible with the measles paradox. Clinical studies are required to test our hypothesis that measles immune suppression is mainly a numbers game.

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References

- Schneider-Schaulies S, Schneider-Schaulies J (2009) Measles virus-induced immunosuppression. *Curr Top Microbiol Immunol* 330: 243–269.
- Hahm B (2009) Hostile communication of measles virus with host innate immunity and dendritic cells. *Curr Top Microbiol Immunol* 330: 271–287.
- Griffin DE (2010) Measles virus-induced suppression of immune responses. *Immunol Rev* 236: 176–189.
- Beckford AP, Kaschula RO, Stephen C (1985) Factors associated with fatal cases of measles. A retrospective autopsy study. *S Afr Med J* 68: 858–863.
- Akramuzzaman SM, Cutts FT, Wheeler JG, Hossain MJ (2000) Increased childhood morbidity after measles is short-term in urban Bangladesh. *Am J Epidemiol* 151: 723–735.
- Van den Hof S, Conyn-van Spaendonck MAE, van Steenberghe JE (2002) Measles epidemic in the Netherlands, 1999–2000. *J Infect Dis* 186:1483–1486.
- Shanks GD, Lee SE, Howard A, Brundage JF (2011) Extreme mortality after first introduction of measles virus to the polynesian island of Rotuma, 1911. *Am J Epidemiol* 173: 1211–1222.
- Griffin DE, Ward BJ, Jauregui E, Johnson RT, Vaisberg A (1989) Immune activation in measles. *N Engl J Med* 320: 1667–1672.
- Von Pirquet CE (1908) Das Verhalten der kutanen Tuberkulin-reaktion während der Maseren. *Dtsch Med Wochenschr* 34: 1297–1300.
- McChesney MB, Miller CJ, Rota PA, Zhu Y, Antipa L, et al. (1997) Experimental measles I. Pathogenesis in the normal and the immunized host. *Virology* 233: 74–84.
- De Swart RL, Ludlow M, De Witte L, Yanagi Y, Van Amerongen G, et al. (2007) Predominant infection of CD150⁺ lymphocytes and dendritic cells during measles virus infection of macaques. *PLoS Pathog* 3: e178.
- Tatsuo H, Ono N, Tanaka K, Yanagi Y (2000) SLAM (CDw150) is a cellular receptor for measles virus. *Nature* 406: 893–897.
- De Vries RD, McQuaid S, Van Amerongen G, Yüksel S, Verburgh RJ, et al. (2012) Measles immune suppression: lessons from the macaque model. *PLoS Pathog* 8: e1002885.
- Conrack C, Grivel JC, Devaux P, Margolis L, Cattaneo R (2007) Measles virus vaccine attenuation: suboptimal infection of lymphatic tissue and tropism alteration. *J Infect Dis* 196: 541–549.
- Grosjean I, Caux C, Bella C, Berger I, Wild F, et al. (1997) Measles virus infects human dendritic cells and blocks their allostimulatory properties for CD4⁺ T cells. *J Exp Med* 186: 801–812.
- Fugier-Vivier I, Servet-Delprat C, Rivallier P, Risoan MC, Liu YJ, et al. (1997) Measles virus suppresses cell-mediated immunity by interfering with the survival and functions of dendritic and T cells. *J Exp Med* 186: 813–823.
- Schnorr JJ, Xanthakos S, Keikavoussi P, Kampgen E, Ter Meulen V, et al. (1997) Induction of maturation of human blood dendritic cell precursors by measles virus is associated with immunosuppression. *Proc Natl Acad Sci U S A* 94: 5326–5331.
- Ferreira CS, Frenze M, Leonard VH, Welstead GG, Richardson CD, et al. (2010) Measles virus infection of alveolar macrophages and dendritic cells precedes spread to lymphatic organs in transgenic mice expressing human signaling lymphocytic activation molecule (SLAM, CD150). *J Virol* 84: 3033–3042.
- Lemon K, De Vries RD, Mesman AW, McQuaid S, Van Amerongen G, et al. (2011) Early target cells of measles virus after aerosol infection of non-human primates. *PLoS Pathog* 7: e1001263.
- Moss WJ, Griffin DE (2012) Measles. *Lancet* 379: 153–164.
- Van Binnendijk RS, Poelen MCM, Kuijpers KC, Osterhaus ADME, UytdedeHaag FGCM (1990) The predominance of CD8⁺ T cells after infection with measles virus suggests a role for CD8⁺ class I MHC-restricted cytotoxic T lymphocytes (CTL) in recovery from measles. *J Immunol* 144: 2394–2399.
- Permar SR, Klumpp SA, Mansfield KG, Kim WK, Gorgone DA, et al. (2003) Role of CD8⁺ lymphocytes in control and clearance of measles virus infection of rhesus monkeys. *J Virol* 77: 4396–4400.
- De Vries RD, Yüksel S, Osterhaus ADME, De Swart RL (2010) Specific CD8⁺ T-lymphocytes control dissemination of measles virus. *Eur J Immunol* 40: 388–395.
- Mongkolsapaya J, Jaye A, Callan MFC, Magnusen AF, McMichael AJ, et al. (1999) Antigen-specific expansion of cytotoxic T lymphocytes in acute measles virus infection. *J Virol* 73: 67–71.
- Ryon JJ, Moss WJ, Monze M, Griffin DE (2002) Functional and phenotypic changes in circulating lymphocytes from hospitalized Zambian children with measles. *Clin Diagn Lab Immunol* 9: 994–1003.
- Finkeldey W (1931) Über Riesenzellbefunde in den Gaumenmandeln, zugleich ein Beitrag zur Histopathologie der Mandelveränderungen im Maserinkubationsstadium. *Virchows Arch* 281: 323–329.
- Warthin AS (1931) Occurrence of numerous large giant cells in the tonsils and pharyngeal mucosa in the prodromal stage of measles. *Arch Pathol* 11: 864–874.
- Lisse I, Samb B, Whittle H, Jensen H, Soumare M, et al. (1998) Acute and long-term changes in T-lymphocyte subsets in response to clinical and subclinical measles. A community study from rural Senegal. *Scand J Infect Dis* 30: 17–21.
- Noyce RS, Bondre DG, Ha MN, Lin LT, Sisson G, et al. (2011) Tumor cell marker PVRL4 (nectin 4) is an epithelial cell receptor for measles virus. *PLoS Pathog* 7: e1002240.
- Mühlebach MD, Mateo M, Sinn PL, Pruffer S, Uhlig KM, et al. (2011) Adherens junction protein nectin-4 is the epithelial receptor for measles virus. *Nature* 480: 530–533.
- Pratakpriya W, Seki F, Otsuki N, Sakai K, Fukuhara H, et al. (2012) Nectin4 is an epithelial cell receptor for canine distemper virus and involved in the neurovirulence. *J Virol* 86: 10207–10210.
- Racaniello V (2011) An exit strategy for measles virus. *Science* 334: 1650–1651.
- Ludlow M, Lemon K, De Vries RD, McQuaid S, Millar E, et al. (2013) Measles virus infection of epithelial cells in the macaque upper respiratory tract is mediated by sub-epithelial immune cells. *J Virol* 87: 4033–4042.
- Ludlow M, De Vries RD, Lemon K, McQuaid S, Millar E, et al. (2013) Infection of lymphoid tissues in the macaque upper respiratory tract

- contributes to the emergence of transmissible measles virus. *J Gen Virol* 94: 1933–1944.
35. Vareille M, Kieninger E, Edwards MR, Regamey N (2011) The airway epithelium: soldier in the fight against respiratory viruses. *Clin Microbiol Rev* 24: 210–229.
 36. Beineke A, Puff C, Sechusen F, Baumgärtner W (2009) Pathogenesis and immunopathology of systemic and nervous canine distemper. *Vet Immunol Immunopathol* 127: 1–18.
 37. Nguyen DT, Ludlow M, Van Amerongen G, De Vries RD, Yüksel S, et al. (2012) Evaluation of synthetic infection-enhancing lipopeptides as adjuvants for a live-attenuated canine distemper virus vaccine administered intra-nasally to ferrets. *Vaccine* 30: 5073–5080.
 38. McCullough B, Krakowka S, Koestner A (1974) Experimental canine distemper virus-induced lymphoid depletion. *Am J Pathol* 74: 155–170.
 39. Von Messling V, Springfield C, Devaux P, Cattaneo R (2003) A ferret model of canine distemper virus virulence and immunosuppression. *J Virol* 77: 12579–12591.
 40. Von Messling V, Milosevic D, Cattaneo R (2004) Tropism illuminated: lymphocyte-based pathways blazed by lethal morbillivirus through the host immune system. *Proc Natl Acad Sci U S A* 101: 14216–14421.
 41. Hirsch RL, Griffin DE, Johnson RT, Cooper SJ, Lindo de Soriano I, et al. (1984) Cellular immune responses during complicated and uncomplicated measles virus infections of man. *Clin Immunol Immunopathol* 31: 1–12.
 42. Ward BJ, Johnson RT, Vaisberg A, Jauregui E, Griffin DE (1991) Cytokine production *in vitro* and the lymphoproliferative defect of natural measles virus infection. *Clin Immunol Immunopathol* 61: 236–248.
 43. Griffin DE, Ward BJ (1993) Differential CD4 T cell activation in measles. *J Infect Dis* 168: 275–281.
 44. Schlender J, Schnorr JJ, Spielhofer P, Cathomen T, Cattaneo R, et al. (1996) Interaction of measles virus glycoproteins with the surface of uninfected peripheral blood lymphocytes induces immunosuppression *in vitro*. *Proc Natl Acad Sci U S A* 93: 13194–13199.
 45. Schnorr JJ, Seufert M, Schlender J, Borst J, Johnston ICD, et al. (1997) Cell cycle arrest rather than apoptosis is associated with measles virus contact-mediated immunosuppression *in vitro*. *J Gen Virol* 78: 3217–3226.
 46. Moussallem TM, Guedes F, Fernandes ER, Pagliari C, Lancellotti CLP, et al. (2007) Lung involvement in childhood measles: severe immune dysfunction revealed by quantitative immunohistochemistry. *Hum Pathol* 38: 1239–1247.
 47. Polack FP, Hoffman SJ, Moss WJ, Griffin DE (2002) Altered synthesis of interleukin-12 and type 1 and type 2 cytokines in rhesus macaques during measles and atypical measles. *J Infect Dis* 185: 13–19.
 48. Atabani SF, Byrnes AA, Jaye A, Kidd IM, Magnusen AF, et al. (2001) Natural measles causes prolonged suppression of interleukin-12 production. *J Infect Dis* 184: 1–9.
 49. Hoffman SJ, Polack FP, Hauer DA, Griffin DE (2003) Measles virus infection of rhesus macaques affects neutrophil expression of IL-12 and IL-10. *Viral Immunol* 16: 369–379.
 50. Karp CL, Wysocka M, Wahl LM, Ahearn JM, Cuomo PJ, et al. (1996) Mechanism of suppression of cell-mediated immunity by measles virus. *Science* 273: 228–231.
 51. Steineur MP, Grosjean I, Bella C, Kaiserlian D (1998) Langerhans cells are susceptible to measles virus infection and actively suppress T cell proliferation. *Eur J Dermatol* 8: 413–420.
 52. Niewiesk S, Eisenhuth I, Fooks A, Clegg JC, Schnorr JJ, et al. (1997) Measles virus-induced immune suppression in the cotton rat (*Sigmodon hispidus*) model depends on viral glycoproteins. *J Virol* 71: 7214–7219.
 53. Avota E, Avots A, Niewiesk S, Kane LP, Bommhardt U, et al. (2001) Disruption of Akt kinase activation is important for immunosuppression induced by measles virus. *Nat Med* 7: 725–731.
 54. Marie JC, Saltel F, Escola JM, Jurdic P, Wild TF, et al. (2004) Cell surface delivery of the measles virus nucleoprotein: a viral strategy to induce immunosuppression. *J Virol* 78: 11952–11961.
 55. Kerdiles YM, Cherif B, Marie JC, Tremillon N, Blanquier B, et al. (2006) Immunomodulatory properties of morbillivirus nucleoproteins. *Viral Immunol* 19: 324–334.
 56. Kerdiles YM, Sellin CI, Druelle J, Horvat B (2006) Immunosuppression caused by measles virus: role of viral proteins. *Rev Med Virol* 16: 49–63.
 57. Gassert E, Avota E, Harms H, Krohne G, Gulbins E, et al. (2009) Induction of membrane ceramides: a novel strategy to interfere with T lymphocyte cytoskeletal reorganization in viral immunosuppression. *PLoS Pathog* 5:e1000623.
 58. Avota E, Schneider-Schaulies S (2014) The role of sphingomyelin breakdown in measles virus immunomodulation. *Cell Physiol Biochem* 34: 20–26.
 59. Manchester M, Smith KA, Eto DS, Perkin HB, Torbett BE (2002) Targeting and hematopoietic suppression of human CD34⁺ cells by measles virus. *J Virol* 76: 6636–6642.
 60. Boussaad I, Varagnolo L, Hornich V, Rieger L, Krockenberger M, et al. (2011) Wild-type measles virus interferes with short-term engraftment of human CD34⁺ hematopoietic progenitor cells. *J Virol* 85: 7710–7718.
 61. Addae MM, Komada Y, Taniguchi K, Kamiya T, Osei-Kwasi M, et al. (1998) Surface marker patterns of T cells and expression of interleukin-2 receptor in measles infection. *Acta Paediatr Jpn* 40: 7–13.
 62. Sullivan JL, Barry DW, Lucas SJ, Albrecht P (1975) Measles infection of human mononuclear cells. I. Acute infection of peripheral blood lymphocytes and monocytes. *J Exp Med* 142: 773–784.
 63. Huddlestone JR, Lampert PW, Oldstone MBA (1980) Virus-lymphocyte interactions: infection of Tg and Tm subsets by measles virus. *Clin Immunol Immunopathol* 15: 502–509.
 64. Ito M, Watanabe M, Ihara T, Kamiya H, Sakurai M (1997) Measles virus induces apoptotic cell death in lymphocytes activated with phorbol 12-myristate 13-acetate (PMA) plus calcium ionophore. *Clin Exp Immunol* 108: 266–271.
 65. Okada H, Kobune F, Sato TA, Kohama T, Takeuchi Y, et al. (2000) Extensive lymphopenia due to apoptosis of uninfected lymphocytes in acute measles patients. *Arch Virol* 145: 905–920.
 66. Pignata C, Fiore M, de Filippo S, Cavalcanti M, Gaetaniello L, et al. (1998) Apoptosis as a mechanism of peripheral blood mononuclear cell death after measles and varicella-zoster virus infections in children. *Pediatr Res* 43: 77–83.
 67. Vuorinen T, Peri P, Vainionpää R (2003) Measles virus induces apoptosis in uninfected bystander T cells and leads to granzyme B and caspase activation in peripheral blood mononuclear cell cultures. *Eur J Clin Invest* 33: 434–442.
 68. Servet-Delprat C, Vidalain PO, Azocar O, Le Deist F, Fischer A, et al. (2000) Consequences of Fas-mediated human dendritic cell apoptosis induced by measles virus. *J Virol* 74: 4387–4393.
 69. Servet-Delprat C, Vidalain PO, Bausinger H, Manie S, Le Deist F, et al. (2000) Measles virus induces abnormal differentiation of CD40 ligand-activated human dendritic cells. *J Immunol* 164: 1753–1760.
 70. Vidalain PO, Azocar O, Lamouille B, Astier A, Rabourdin-Combe C, et al. (2000) Measles virus induces functional TRAIL production by human dendritic cells. *J Virol* 74: 556–559.