



S2 Figure: Surface plasmon resonance (SPR) analysis of PA-receptor binding affinity. As indicated in an earlier report (14), the ultraslow dissociation kinetics made accurate measurement of the binding affinity between receptor and PA difficult using SPR. Here, using the TF-tagged receptors, TF-R318(WT) and TF-R318(C255/279A), we were able to measure the binding affinity between the TF-tagged receptors and PA₈₃. Briefly, PA₈₃ was covalently coupled to a CM5 chip in a Biacore 100 system at 25°C. 2 μM of the protein was immobilized in sodium acetate pH 4.0 buffer at 5 μl min⁻¹ until the baseline reached <4,000 response units and then blocked with ethanolamine. Purified proteins of TF-R318(WT) (**A**) or TF-R318(C255/279A) (**B**) were diluted into HBS buffer (10mM Hepes pH 7.4, 150mM NaCl, 1 mM MgCl₂) and serial injections were made at a constant flow rate of 10 μl min⁻¹. Concentrations of receptor proteins ranged from 38nM to 5 μM. The CM5 chip was regenerated using 0.5 M NaCO₃ (pH 10.5) with a 30 μl pulse, resulting in <1% loss of base line per injection. Kinetic and affinity studies were done on a 1:1 fit model and steady state, respectively.

The binding kinetics and affinity are shown in **S1 Table** below:

Binding Kinetics and Affinity between TF-R318 receptors and PA₈₃

	k_a (1/Ms)	k_d (1/s)	K_D (M)	Affinity K_D (M)
TF-R318(WT): PA ₈₃	1222	0.001678	1.37E-06	5.91E-06
TF-R318(C255/279A):PA ₈₃	8075	7.71E-04	9.55E-08	9.24E-07

As indicated in **S1 Table**, compared to TF-R318(WT), TF-R318(C255/279A) has a slightly higher affinity to PA₈₃, indicating that deletion of the disulfide bond C255-C279 does not reduce the binding affinity to PA₈₃.