

## Supplemental material

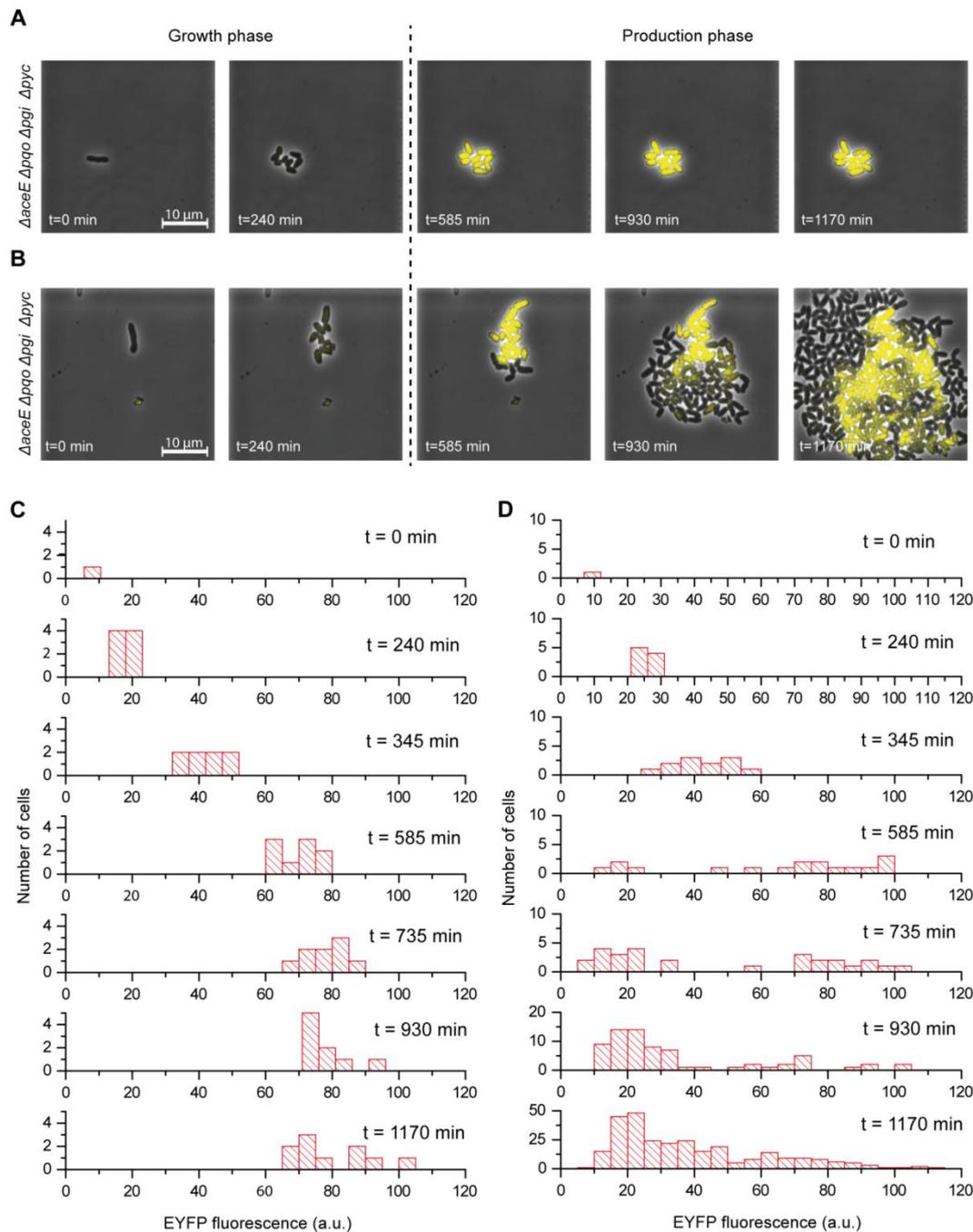
# Application of a genetically encoded biosensor for live cell imaging of L-valine production in pyruvate dehydrogenase complex-deficient *Corynebacterium glutamicum* strains

Nuriye Mustafi<sup>1§</sup>, Alexander Grünberger<sup>1§</sup>, Regina Mahr<sup>1</sup>, Stefan Helfrich<sup>1</sup>, Katharina Nöh<sup>1</sup>, Bastian Blombach<sup>2</sup>, Dietrich Kohlheyer<sup>1</sup>, and Julia Frunzke<sup>1\*</sup>

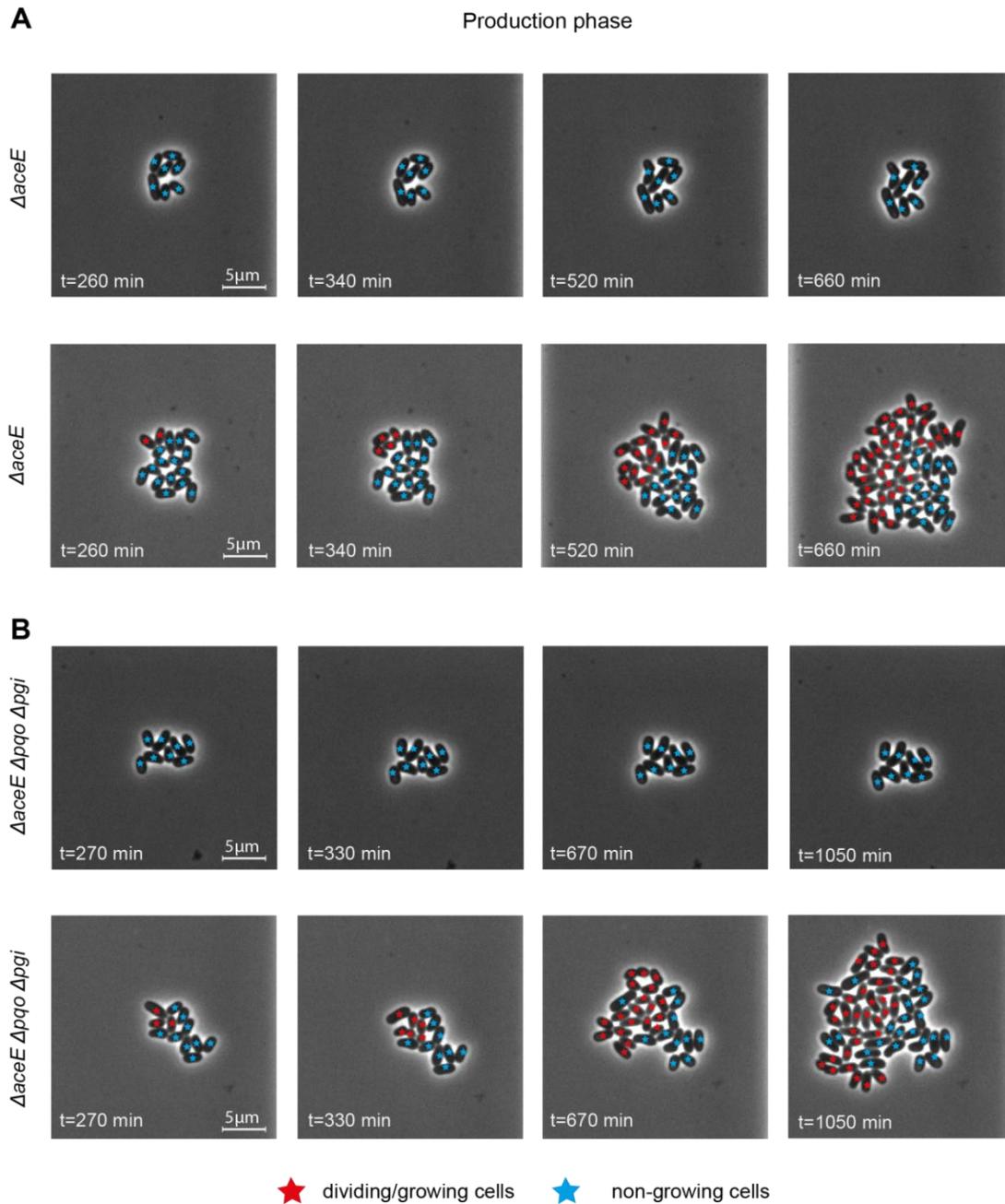
<sup>1</sup>IBG-1: Biotechnology, Forschungszentrum Jülich GmbH (Research Centre Jülich), 52425 Jülich, Germany

<sup>2</sup>Institute of Biochemical Engineering, University of Stuttgart, 70569 Stuttgart, Germany

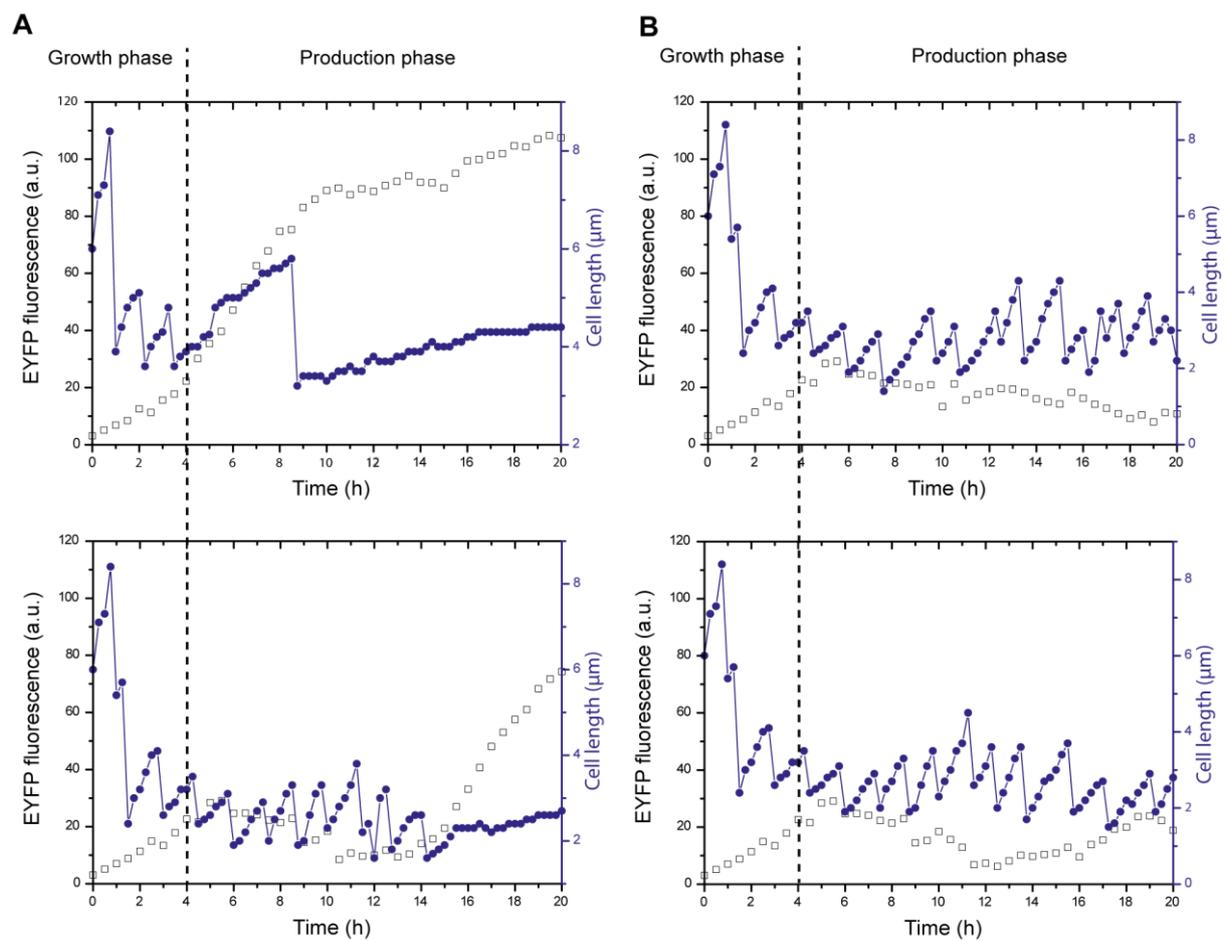
<sup>§</sup> Both authors contributed equally to this work.



**Figure S1. Phenotypic heterogeneity of the  $\Delta aceE \Delta pqo \Delta pgi \Delta pyc$  sensor strain upon switch from growth to production phase.** (A) Microcolony showing transition to producing cells or (B) a mixture of growing and producing cells after medium switch (initiated after 240 min). In approximately 50% of the recorded colonies one or several single cells continued growth after medium switch. (C, D) Fluorescence histograms depicting single cell fluorescence to selected times during growth (0-240 min) and production phase (0-1200 min) of the microcolonies shown in A (C) and B (D). Cultivation was performed in CGXII minimal medium containing 154 mM acetate, 222 mM glucose and 0.5% BHI during growth phase or 222 mM glucose and 0.5% BHI during production phase, respectively.



**Figure S2. Phenotypic heterogeneity of  $\Delta aceE$  and  $\Delta aceE \Delta pqo \Delta pgi$  upon switch from growth to production phase.** (A)  $\Delta aceE$  microcolonies where all cells stopped growth (blue stars) upon transition to the production phase (upper row) or a mixture of growing (red stars) and non-growing cells (lower row) after initiation of the production phase. In approximately 50% of the recorded colonies one or several single cells continued growth after medium switch (initiated after 250 min). (C)  $\Delta aceE \Delta pqo \Delta pgi$  microcolonies. In the upper row, all cells stopped growth whereas in the lower row a microcolony is shown where some cells continued growth after initiation of the production phase. In approximately 50% of the recorded colonies one or several single cells continued growth after medium switch (initiated after 250 min). These findings confirm that the phenotypic split shown in Figure 5 is not due to the presence of the Lrp-sensor. Cultivation was performed in CGXII minimal medium containing 154 mM acetate, 222 mM glucose and 0.5% BHI during growth phase or 222 mM glucose and 0.5% BHI during production phase, respectively.



**Figure S3. Single cell traces of the  $\Delta aceE \Delta pqo \Delta pgi \Delta pyc$  sensor strain upon switch from growth to production phase. (A)** Single cell traces showing the switch from growth (cell length=blue line) to production (fluorescence=squares) after several cell divisions during production phase ( $t=8.5$  h,  $t=15.0$  h). **(B)** Single cell traces showing no switch from growth to production. Single cell traces are taken from the cultivation of  $\Delta aceE \Delta pqo \Delta pgi \Delta pyc$  sensor strain shown in Figure S1.