

Table S3 mRNA leader regions that coIP with Hfq

Name	Ligand	Peak Length	Peak Coordinates	RPKM Ratio ^a
<i>ilvB</i>	tRNA	76	2897048-2897123	317
<i>yybP</i>	Unknown	101	4169826-4169926	211
<i>mgtE</i>	Magnesium	203	1395622-1395824	130
<i>rplT</i>	Protein	177	2953417-2953593	115
<i>tyrS</i>	tRNA	219	3037940-3038158	88.6
<i>xpt</i>	Purine	135	2320065-2320199	68.3
<i>glpF</i>	Protein	96	1002367-1002462	56.1
<i>ydaO</i>	Unknown	195	486090-486284	41.5
<i>thiU</i>	TPP	172	1391680-1391851	38.7
<i>rplJ</i>	Protein	100	119850-119949	37.6
<i>ykoY</i>	Unknown	130	1410625-1410754	36.9
<i>ptsG</i>	Protein	96	1456978-1457073	31.8
<i>bglP</i>	Protein	121	4035698-4035818	31.6
<i>trpE</i>	Protein	108	2377504-2377611	30.3
<i>rpsD</i>	Protein	118	3035565-3035682	30.1
<i>thrS</i>	tRNA	233	2961244-2961476	28.3
<i>thiT</i>	TPP	123	3179111-3179233	28.1
<i>yitJ</i>	SAM	203	1180645-1180847	27.2
<i>pyrG^b</i>	Protein	76	3812400-3812475	26.7
<i>trpS</i>	tRNA	223	1219144-1219366	24.1
<i>queC</i>	Purine	82	1439273-1439354	18.6
<i>lysP</i>	Lysine	229	3421133-3421361	18.4
<i>ktrA</i>	Unknown	172	3188194-3188365	18.3
<i>fmnP</i>	FMN	199	2410680-2410878	12.3
<i>yxjG</i>	SAM	123	3999167-3999289	12.2
<i>yxjA</i>	Purine	149	4005557-4005705	11.6
<i>tenA</i>	TPP	140	1242260-1242399	11.2
<i>metQ</i>	SAM	166	3364355-3364520	11.1
<i>thiC</i>	TPP	165	955648-955812	9.92
<i>lysC</i>	Lysine	171	2910880-2911050	5.44
<i>pbuG</i>	Purine	131	694450-694580	4.56
<i>ileS</i>	tRNA	197	1613087-1613283	4.14
<i>gcvT</i>	Glycine	172	2549415-2549586	3.82
<i>ribD</i>	FMN	132	2431468-2431599	3.08
<i>purE</i>	Purine	140	698396-698535	2.93
<i>yoaD</i>	SAM	125	2025141-2025265	2.79
<i>cspC^b</i>	Protein	45	559567-559611	1.91

^a The expression of each peak was quantified in reads per kilobase per million mapped reads, or ‘RPKM’ (Mortazavi et al., 2008). The ratio of these values for the Hfq^{FLAG} and mock control samples was taken as an indicator of Hfq-mediated enrichment.

^b Almost all of the Hfq CoIP peaks shown in this table directly correspond to the aptamer portions of the cis-acting regulatory RNAs. There are two exceptions – *pyrG* and *cspC*. *pyrG* does not have a discrete aptamer domain according to current literature. Instead, it regulates downstream gene expression through reiterative incorporation of nontemplated G residues under conditions of pyrimidine

limitation, which in turn affects formation of a downstream intrinsic terminator. The Hfq CoIP peak for *pyrG* exactly corresponds to the entire regulatory RNA, from the start of transcription to the transcription terminator element. Therefore, despite the absence of a recognizable aptamer domain, the ligand-sensing portion of the *pyrG* regulatory RNA still corresponds to the Hfq CoIP peak. Expression of *cspB* and *cspC* genes is auto-regulated through a transcription attenuation mechanism (Graumann and Marahiel, 1999). The aptamer portion of these leader regions has not been fully identified; however, the *cspC* Hfq CoIP peak covers the region between the transcription start site and the downstream transcription terminator, and is therefore likely to encompass the putative CspC binding site.