Table S6.PCR reaction conditions

PCR reaction conditions				
Enzyme	System		Procedure	Samples
KOD HotStart DNA Polymerase	10×PCRbuffer	5ul	95℃ 2min	clone sequencing
	dNTP（2mM）	5ul	95℃ 20s	
	Primer forward(10uM)	1.5ul	Tm 10s	
	Primer reverse(10uM)	1.5ul	70℃ 20s	
	Enzyme	1ul	70℃ 1min	
	$MgSO_4$（25mM）	3ul	4℃ forever	
	Template DNA	100-200ng		
	ddH_2O			
	total	50ul	38cycles	
Platinum® PCR SuperMix	SuperMix	40ul	94℃ 2min	clone sequencing
	Primer forward(10uM)	1.5ul	94℃ 15s	
	Primer reverse(10uM)	1.5ul	Tm 30s	
	Template DNA	100-200ng	68℃ 1min	
			68℃ 5min	
			4℃ forever	
	total	45ul	38cycles	
TaKaRa rTaq DNA Polymerase	10×PCRbuffer	4ul	95℃ 3min	Whole genome sequencing
	dNTP（10mM）	0.5ul	95℃ 40s	
	Primer forward(10uM)	0.8ul	Tm 40s	
	Primer reverse(10uM)	0.8ul	72℃ 40s	
	Enzyme	0.4ul	72℃ 1min	
	Template DNA	100-200ng	4℃ forever	
	ddH_2O			
	total	40ul	38cycles	
HotStarTaq Master Mix Kit	10×PCRbuffer	4ul	95℃ 15min	Whole genome sequencing ,E7 ,E6,L1(SPF/GP6+) amplification
	dNTP（10mM）	0.6ul	94℃ 40s	
	Primer forward(10uM)	1.6ul	Tm 40s	
	Primer reverse(10uM)	1.6ul	72℃ 30s	
	Enzyme	0.2ul	72℃ 1min	
	$MgCL_2$（25mM）	0.8ul	4℃ forever	
	Template DNA	50ng		
	ddH_2O			
	total	40ul	38cycles	
Trans Taq HiFi DNA	10×PCRbuffer	4ul	95℃ 3min	Whole genome sequencing
	dNTP（10mM）	0.5ul	95℃ 40s	

Polymerase	Primer forward(10uM)	0.8ul	Tm 40s	
	Primer reverse(10uM)	0.8ul	72℃ 40s	
	Enzyme	0.3ul	72℃ 1min	
	Template DNA	100-200ng	4℃ forever	
	ddH_2O			
	total	40ul	38cycles	
2XPCR TaqMix	2XPCR TaqMix	10ul	94℃ 2min	PCR during Clone sequencing
	Primer forward(10uM)	1ul	94℃ 30s	
	Primer reverse(10uM)	1ul	Tm 30s	
	Template DNA	100-200ng	72℃ 75s	
	ddH_2O		72℃ 5min	
			4℃ forever	
	total	20ul	38cycles	