## **Software Minimum Requirements**

- Fiji or ImageJ version 1.440 or newer
- Image5D (pre-installed with Fiji)
- Java 1.6 or newer
- 32 or 64 bit Windows, OSX or Linux.
- 2Gb RAM

## **Segmentation User Interface**

Select an Experiment Select an experiment Select an experiment to load or provide the name of a new experiment to create. Select the directory which will hold the experiments     /Users/mike/phd/temp/data/     Create New Experiment Experiment Name     Load Experiment	the 'data' dire name or type	der which will hold t ectory below. Either s in the name of a new he directory structure	select an e v experim	experiment
expt1				
expt2				
(Cancel) (Load)				
🚞 data 🔹 🕨	expt1 🕨	📄 info.xml	ļ	E10f01_ch1.tif
	expt2 🕨	💼 pictures	► I	E10f01_ch2.tif
		i rois	Þ	
		🚞 tables	⊳	

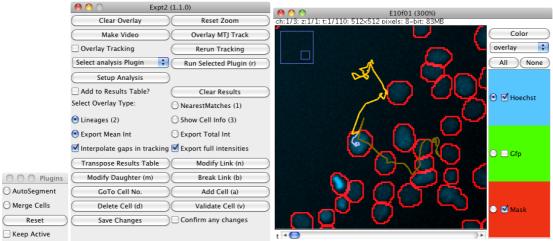
Experiment directory structure. The info.xml file describes the experiment. The 'rois' and 'tables' directories are filled when the experiment is analysed.

		0 0		Edi	t Experiment		
🔴 🔿 🔹 Edit Segmentation		Remove	Channel Name		Image Stack	Store?	Segment? 👗
Segmentation Method:	Seeded Growth						
Edit Segmentation	Edit Settings	-	Hoechst		E10f01_ch1.zip		
Edit Channels	Preview	-	GFP		E10f01_ch2.zip		
Preview Speed	1	-	Segmented Mask		E10f01_mask.zip		
☑ Run Tracking (	Edit Tracking						
Run Segmentation	Display						
							Ŧ
(	Quit		Add	)	OK Cancel		

Control	Function
Edit Experiment Window:	
Add	Adds a new image channel. Images must be uncompressed TIFF stacks.
[-]	Remove the image channel
Store	Store intensity information for this channel
Segment	Use this channel in the segmentation algorithm
Edit Segmentation Windo	w:
Segmentation Method	Select the segmentation method.
Edit Segmentation	Edit method-specific segmentation parameters.
Edit Settings	Experiment-specific segmentation parameters including maximum & minimum object size and background subtraction.

Edit Channels	Opens the 'Edit Experiment' window.
Preview	Displays a window previewing the segmentation.
Edit Tracking	Edit the tracking parameters
Run Segmentation	Run the segmentation (and tracking if required)
Display	Call the Tracking Viewer

## **Tracking View User Interface**



Tracking viewer running as an ImageJ plug-in, displaying the trajectory of a tracked cell. The plugin selector window allows rapid switching between data analysis or editing plugins.

Control	Function		
Control Panel Window:			
Clear Overlay	Remove any overlaid highlighting from the window.		
Reset Zoom	Reset to 100% magnification.		
Make Video	Create a image time-series complete with highlighted cells and tracks.		
Load MTJ Track	Load and overlay an MTrackJ trajectory.		
Overlay Tracking	When a tracked cell is selected, draw the full trajectory from current timepoint onwards.		
Rerun Tracking	Allows tracking to be run with a different method or parameters.		
Select Analysis Plugin	If any additional analysis plugins have been installed, they can be selected from here. When a cell is clicked in the viewer window, the plugin will be called for that cell.		
Do Analysis	Call the selected analysis plugin for all cells in the experiment.		
Setup Analysis	Edit any settings (if needed) for the analysis plugin. Also used to display a brief description of the plugin.		
Add to Results Table	When a cell is selected, add fluorescence intensity data to an ImageJ Results Table		
Clear Results	Clear any currently open Results Tables.		
Overlay Types: the next 3 buttons control the information displayed or added to a results table when a cell is highlighted in the window.			

1) Nearest Matches	Overlay the Movement Scores for trajectories leaving the highlighted cell.			
2) Lineages	Display tracking, including daughter cells.			
3) Cell Info	Display fluorescence information for current timepoint.			
Export Total/Mean Int	Selects between exporting the integrated or mean cell intensity to the results table.			
Interpolate Gaps	Interpolates the intensity if tracking skips a frame.			
Export Full Intensities	ON: Exports the full lineage intensity for daughter cells OFF: Only export intensities from division onwards for daughter cells.			
between two cells	ttons modify tracking based on the previous two highlighted cells. To alter the link (in adjacent frames), click on one cell then move to the next frame and click on nally select one of the following buttons.			
Modify Link	Add the two selected cells as a trajectory			
Modify Daughter	Add a trajectory, marking it as a cell division. If the parent cell already has a tracked 'next cell', that is marked as the other daughter cell.			
Break Link	Remove the tracking between the two selected cells.			
Go To CellID	Select a cell based on its ID number			
Save Changes	Save any segmentation or tracking changes.			
Add Cell	Expects a 'closed' ROI (such as oval or polygon). Calculates the cell features and adds a new cell.			
Delete Cell	Deletes the currently selected cell.			
Validate Cell	Marks the currently selected trajectory as 'validated'. The validated cells are drawn with filled-in masks for easy identification.			
Confirm Changes	Prompts the user before any actions which change the cell data.			
Cell Image Wind	0W			
The tick-boxes sho	ow or hide the following image channels.			
Time t	The primary fluorescence channel			
Mask	Segmentation outlines			
GFP	Any additional fluorescence channels.			
Plugins Window				
This window lists plugins are curre	all available 'Analysis Plugins' and allows easy selection between them. Two ntly available.			
AutoSegment	After selecting this and clicking on a cell in the image window, the plugin will attempt to calculate the outline of the cell and add it to the experiment data. A dialog box appears allowing the segmentation parameters to be adjusted.			
Merge Cells	Requires an ROI to be drawn in the image window. When 'Run Selected Plugin' is clicked, any cells within the ROI will be merged together.			
Reset	Un-selects the current selected plugin.			
Keep Active	If un-selected, the current plugin will be used for the next click in the cell image window. Selecting this will allow the plugin to be run several times on different cells.			

## **Benchmarking Software**

The segmentation benchmarking consists of an ImageJ macro which receives the segmentation mask and compares the cell positions with a manually-produced 'gold standard'. The tracking benchmark software is a plugin for the LineageTracker application which requires a manually validated cell lineage. This software will be available to download from the LineageTracker website<sup>1</sup>.

<sup>&</sup>lt;sup>1</sup> <u>http://go.warwick.ac.uk/lineagetracker</u>