

S3 Table. Plasmids used in this study

Plasmid Name	Stock Name	Description	Origin, marker	Reference
pET15b	pET15b	Overexpression vector for protein purification	pBR322, Amp ^R	Novagen
pET15b- <i>luxT-6xHis</i>	pME127	<i>luxT-6xHis</i> overexpression vector for protein purification, cloned in pET15b	pBR322, Amp ^R	This study
pRE112	pRE112	allelic exchange vector harboring <i>sacB</i> as a counter-selectable marker	R6K γ , Cam ^R	[1]
pRE112- Δ <i>luxT</i>	pME12	<i>V. harveyi luxT</i> deletion construct in pRE112	R6K γ , Cam ^R	This study
pRE112- <i>aphA-3xFLAG</i>	pME146	<i>V. harveyi aphA-3xFLAG</i> allele exchange construct in pRE112	R6K γ , Cam ^R	This study
pRE112- <i>3xFLAG-luxR</i>	pME147	<i>V. harveyi 3xFLAG-luxR</i> allele exchange construct in pRE112	R6K γ , Cam ^R	This study
pFED343	pFED343	P _{tac} overexpression vector	P15A, Cam ^R	[2]
P _{qrr1} - <i>mRuby3</i>	pME98	<i>V. harveyi qrr1-mRuby3</i> transcriptional reporter in pFED343 (excluding the P _{tac} promoter)	P15A, Cam ^R	This study
P _{luxO} - <i>mRuby3</i>	pME96	<i>V. harveyi luxO-mRuby3</i> transcriptional reporter in pFED343 (excluding the P _{tac} promoter)	P15A, Cam ^R	This study
P _{qrr2} - <i>mRuby3</i>	pME100	<i>V. harveyi qrr2-mRuby3</i> transcriptional reporter in pFED343 (excluding the P _{tac} promoter)	P15A, Cam ^R	This study
P _{qrr3} - <i>mRuby3</i>	pME102	<i>V. harveyi qrr3-mRuby3</i> transcriptional reporter in pFED343 (excluding the P _{tac} promoter)	P15A, Cam ^R	This study
P _{qrr4} - <i>mRuby3</i>	pME103	<i>V. harveyi qrr4-mRuby3</i> transcriptional reporter in pFED343 (excluding the P _{tac} promoter)	P15A, Cam ^R	This study
P _{qrr5} - <i>mRuby3</i>	pME105	<i>V. harveyi qrr5-mRuby3</i> transcriptional reporter in pFED343 (excluding the P _{tac} promoter)	P15A, Cam ^R	This study
p <i>luxCDABE</i>	pBB1	<i>V. harveyi luxCDABE</i> cloned in pLAFR (expressed from its native promoter)	oriV Tet ^R	[3]
p <i>luxR</i>	pME125	<i>V. harveyi luxR</i> overexpression vector, cloned in pFED343	P15A, Cam ^R	This study
pKP8-35	pKP8-35	P _{BAD} overexpression vector	pBR322, Amp ^R	[4]

<i>pluxT</i>	pME109	<i>V. harveyi luxT</i> overexpression vector, cloned in pKP8-35	pBR322, Amp ^R	This study
<i>pRE112-</i> <i>ΔVIBHAR_RS03920</i>	pME64	<i>V. harveyi VIBHAR_RS03920</i> deletion construct in pRE112	R6K γ , Cam ^R	This study
<i>pRE112-luxO D61E</i> <i>Δqrr1</i>	pME148	<i>V. harveyi luxO D61E Δqrr1</i> allele exchange construct in pRE112	R6K γ , Cam ^R	This study
<i>pluxT</i>	pME69	<i>V. harveyi luxT</i> overexpression vector, cloned in pFED343	P15A, Cam ^R	This study
<i>P₁₁₇₈₅-lux</i>	pME188	<i>VIBHAR_RS11785-luxCDABE</i> transcriptional reporter in pFED343 (excluding the <i>P_{tac}</i> promoter)	P15A, Cam ^R	This study
<i>P₁₁₆₂₀-lux</i>	pME189	<i>VIBHAR_RS11620-luxCDABE</i> transcriptional reporter in pFED343 (excluding the <i>P_{tac}</i> promoter)	P15A, Cam ^R	This study
<i>P₁₆₉₈₀-lux</i>	pME190	<i>VIBHAR_RS16980-luxCDABE</i> transcriptional reporter in pFED343 (excluding the <i>P_{tac}</i> promoter)	P15A, Cam ^R	This study
<i>P₂₅₆₇₀-lux</i>	pME191	<i>VIBHAR_RS25670-luxCDABE</i> transcriptional reporter in pFED343 (excluding the <i>P_{tac}</i> promoter)	P15A, Cam ^R	This study
<i>pXB300</i>	pXB300	<i>P_{tetA}</i> overexpression vector	pBR322, Amp ^R	[5]
<i>ptetA-Kan</i>	pME149	<i>P_{tetA}</i> overexpression vector (Amp ^R replaced with Kan ^R in pXB300)	pBR322, Kan ^R	This study
<i>P_{tetA-11785}-</i> <i>'mVenus</i>	pME150	<i>VIBHAR_RS11785</i> translational <i>mVenus</i> reporter, expressed from the <i>tetA</i> promoter	pBR322, Kan ^R	This study
<i>P_{tetA-11620}-</i> <i>'mVenus</i>	pME151	<i>VIBHAR_RS11620</i> translational <i>mVenus</i> reporter, expressed from the <i>tetA</i> promoter	pBR322, Kan ^R	This study
<i>P_{tetA-16980}-</i> <i>'mVenus</i>	pME152	<i>VIBHAR_RS16980</i> translational <i>mVenus</i> reporter, expressed from the <i>tetA</i> promoter	pBR322, Kan ^R	This study
<i>P_{tetA-25670}-</i> <i>'mVenus</i>	pME153	<i>VIBHAR_RS25670</i> translational <i>mVenus</i> reporter, expressed from the <i>tetA</i> promoter	pBR322, Kan ^R	This study
<i>pqrr1</i>	pME154	<i>V. harveyi qrr1</i> overexpression vector, cloned in pFED343	P15A, Cam ^R	This study
<i>pRE112-ΔluxT (V.</i> <i>cholerae)</i>	pME112	<i>V. cholerae luxT</i> deletion construct in pRE112	R6K γ , Cam ^R	This study
<i>P_{qrr1}-luxCDABE</i>	pBK1001	<i>qrr1-luxCDABE</i> promoter fusion	Cam ^R	[6]
<i>pRE112-ΔswrT (V.</i> <i>parahaemolyticus)</i>	pME155	<i>V. parahaemolyticus swrT</i> deletion construct in pRE112	R6K γ , Cam ^R	This study

pRE112- <i>luxO</i> D61E (<i>V.</i> <i>parahaemolyticus</i>)	pME156	<i>V. parahaemolyticus luxO</i> D61E allele exchange construct in pRE112	R6K γ , Cam ^R	This study
P _{qrr1} - <i>mRuby3</i> (<i>V.</i> <i>parahaemolyticus</i>)	pME157	<i>V. parahaemolyticus qrr1-mRuby3</i> transcriptional reporter in pFED343 (excluding the P _{tac} promoter)	P15A, Cam ^R	This study
pRE112- <i>luxO</i> D55E (<i>A. fischeri</i>)	pME158	<i>A. fischeri luxO</i> D55E allele exchange construct in pRE112	R6K γ , Cam ^R	This study
P _{qrr1} - <i>mRuby3</i> (<i>A.</i> <i>fischeri</i>)	pME159	<i>A. fischeri qrr1-mRuby3</i> transcriptional reporter in pFED343 (excluding the P _{tac} promoter)	P15A, Cam ^R	This study
pRE112- <i>luxO</i> D55E Δ <i>qrr1</i> (<i>A. fischeri</i>)	pME160	<i>A. fischeri luxO</i> D55E Δ <i>qrr1</i> allele exchange construct in pRE112	P15A, Cam ^R	This study

Table S3 References

1. Edwards RA, Keller LH, Schifferli DM. Improved allelic exchange vectors and their use to analyze 987P fimbria gene expression. *Gene*. 1998 Jan 30;207(2):149–57.
2. Swem LR, Swem DL, Wingreen NS, Bassler BL. Deducing receptor signaling parameters from in vivo analysis: LuxN/AI-1 quorum sensing in *Vibrio harveyi*. *Cell*. 2008 Aug 8;134(3):461–73.
3. Miller MB, Skorupski K, Lenz DH, Taylor RK, Bassler BL. Parallel quorum sensing systems converge to regulate virulence in *Vibrio cholerae*. *Cell*. 2002 Aug 9;110(3):303–14.
4. Papenfort K, Pfeiffer V, Mika F, Lucchini S, Hinton JCD, Vogel J. SigmaE-dependent small RNAs of *Salmonella* respond to membrane stress by accelerating global *omp* mRNA decay. *Mol Microbiol*. 2006 Dec;62(6):1674–88.
5. Bina XR, Wong EA, Bina TF, Bina JE. Construction of a tetracycline inducible expression vector and characterization of its use in *Vibrio cholerae*. *Plasmid*. 2014 Nov;76:87–94.
6. Svenningsen SL, Waters CM, Bassler BL. A negative feedback loop involving small RNAs accelerates *Vibrio cholerae*'s transition out of quorum-sensing mode. *Genes Dev*. 2008 Jan 15;22(2):226–38.