Progesterone II

Progesterone

12145383 122

100

English

Intended use

Immunoassay for the in vitro quantitative determination of progesterone in human serum and plasma.

The electrochemiluminescence immunoassay “ECLIA” is intended for use on Elecsys and cobas e immunoassay analyzers.

Summary

The gestagen progesterone is a steroid hormone having a molecular-weight of 314.5 daltons. Progesterone is mainly formed in the cells of the corpus luteum and during pregnancy in the placenta.

The progesterone concentration correlates with the development and regression of the corpus luteum. Whereas progesterone is barely detectable in the follicular phase of the female cycle, a rise in the progesterone level is observed one day prior to ovulation. Increased progesterone synthesis occurs during the luteal phase. In the second half of the cycle progesterone is excreted in urine as the main degradation product of progesterone.

Progesterone brings about the conversion of the uterine mucosa into a tissue rich in glands (secretion phase), in order to prepare for the intrauterine implantation of the fertilized ovum. During pregnancy, progesterone inhibits the contraction of the myometrium. In the mammary gland, progesterone (together with estrogens) promotes the proliferation and secretion disposition of the alveoli.

The determination of progesterone is utilized in fertility diagnosis for the detection of ovulation and assessment of the luteal phase.

Test principle

Competition principle. Total duration of assay: 18 minutes.

- 1st incubation: 30 µL sample - in the presence of a biotinylated monoclonal progesterone-specific antibody and a progesterone derivative labeled with ruthenium complex - are incubated with Danazol to release progesterone. Progesterone from the sample competes with the labeled progesterone derivative for the antibody binding sites.

- 2nd incubation: After addition of streptavidin-coated microparticles, the complex becomes bound to the solid phase via interaction of biotin and streptavidin. The amount of the labeled progesterone derivative bound to the solid phase is inversely proportional to the progesterone content of the sample.

- The reaction mixture is aspirated into the measuring cell where the microparticles are magnetically captured onto the surface of the electrode. Unbound substances are then removed with ProCell/ProCell M. Application of a voltage to the electrode then induces chemiluminescent emission which is measured by a photomultiplier.

- Results are determined via a calibration curve which is instrument-specifically generated by 2-point calibration and a master curve provided via the reagent bar code.

Reagents - working solutions

The reagent rackpack is labeled as PROG II.

M Streptavidin-coated microparticles (transparent cap), 1 bottle, 6.5 mL; Streptavidin-coated microparticles 0.72 mg/mL; preservative.

R1 Anti-progesterone-Ab-biotin (gray cap), 1 bottle, 10 mL; Biotinylated monoclonal anti-progesterone antibody (mouse) 0.15 mg/L, phosphate buffer 25 mmol/L, pH 7.0; preservative.

R2 Progesterone-peptide–Ru(bpy)³⁺ (black cap), 1 bottle, 8 mL:

Progesterone (of vegetable origin) coupled to a synthetic peptide labeled with ruthenium complex, 10 ng/mL; phosphate buffer 25 mmol/L, pH 7.0; preservative.

Precautions and warnings

For in vitro diagnostic use. Exercise the normal precautions required for handling all laboratory reagents.

Disposal of all waste material should be in accordance with local guidelines.

Safety data sheet available for professional user on request.

Avoid foam formation in all reagents and sample types (specimens, calibrators and controls).

Reagent handling

The reagents in the kit have been assembled into a ready-for-use unit that cannot be separated.

All information required for correct operation is read in from the respective reagent barcodes.

Storage and stability

Store at 2-8 °C.

Do not freeze.

Store the Elecsys reagent kit upright in order to ensure complete availability of the microparticles during automatic mixing prior to use.

Stability:

- unopened at 2-8 °C up to the stated expiration date
- after opening at 2-8 °C 12 weeks
- on the analyzers 8 weeks

Specimen collection and preparation

Only the specimens listed below were tested and found acceptable.

- Serum collected using standard sampling tubes or tubes containing separating gel.

- Na-, Li-heparin, K₃-EDTA, sodium citrate and sodium fluoride/potassium oxalate plasma. When sodium citrate is used, the results must be corrected by + 10 %.

Cohesion: Recovery within 90-110 % of serum value or slope 0.9-1.1 + intercept within ± 2x analytical sensitivity (LDL) + coefficient of correlation > 0.95.

Stable for 5 days at 2-8 °C, 6 months at -20 °C. Freeze only once.

Stability of serum obtained with separating tubes: 48 hours at 2-8 °C (note the data provided by the tube manufacturer).

The sample types listed were tested with a selection of sample collection tubes that were commercially available at the time of testing, i.e. not all available tubes of all manufacturers were tested. Sample collection systems from various manufacturers may contain differing materials which could affect the test results in some cases. When processing samples in primary tubes (sample collection systems), follow the instructions of the tube manufacturer.

Centrifuge samples containing precipitates before performing the assay. Do not use heat-inactivated samples.

Do not use samples and controls stabilized with azide.

Ensure the samples, calibrators and controls are at 20-25 °C prior to measurement.

Due to possible evaporation effects, samples, calibrators and controls on the analyzers should be analyzed/measured within 2 hours.
Progestørone II

Materials provided
See “Reagents – working solutions” section for reagents.

Materials required (but not provided):
- 12145391122, Progestørone II CalSet, for 4 x 1 mL
- 11731416190, PreciControl Universal, for 2 x 3 mL each of PreciControl Universal 1 and 2
- 03028542122, Diluent Estradiol/Progestørone, 2 x 22 mL sample
- General laboratory equipment
- Elecsys 2010, MODULAR ANALYTICS E170 or cobas e analyzer
- Accessories for Elecsys 2010 and cobas e 411 analyzers:
  - 11662988122, ProCell, 6 x 380 mL system buffer
  - 11662970122, CleanCell, 6 x 380 mL measuring cell cleaning solution
  - 11930346122, Elecsys SysWash, 1 x 500 mL water wash additive
  - 11933159001, Adapter for SysClean
  - 11706802001, Elecsys 2010 AssayCup, 60 x 60 reaction vessels
  - 11706799001, Elecsys 2010 AssayTip, 30 x 120 pipette tips
- Accessories for MODULAR ANALYTICS E170, cobas e 601 and cobas e 682 analyzers:
  - 048600340190, ProCell M, 2 x 2 L system buffer
  - 04860293190, CleanCell M, 2 x 2 L measuring cell cleaning solution
  - 03023141001, PC/CC-Cups, 12 cups to prewarm ProCell M and CleanCell M before use
  - 03005712190, ProbeWash M, 12 x 70 mL cleaning solution for run initialization and rinsing during reagent change
  - 12102137001, AssayTip/AssayCup Combimagazine M, 48 magazines x 84 reaction vessels or pipette tips, waste bags
  - 03023150001, WasteLiner, waste bags
  - 03027651001, SysClean Adapter M
- Accessories for all analyzers:
  - 11298500316, Elecsys SysClean, 5 x 100 mL system cleaning solution

Assay
For optimum performance of the assay follow the directions given in this document for the analyzer concerned. Refer to the appropriate operator’s manual for analyzer-specific assay instructions.

Resuspension of the microparticles takes place automatically prior to use. Read in the test-specific parameters via the reagent barcode. If in exceptional cases the barcode cannot be read, enter the 15-digit sequence of numbers.

Bring the cooled reagents to approx. 20 °C and place on the reagent disk (20 °C) of the analyzer. Avoid foam formation. The system automatically regulates the temperature of the reagents and the opening/closing of the bottles.

Calibration
Traceability: This method has been standardized via ID-GC/MS ("Isotope Dilution Gas Chromatography Mass Spectrometry").

Every Elecsys reagent set has a barcoded label containing specific information for calibration of the particular reagent lot. The predefined master curve is adapted to the analyzer using the relevant CalSet.

Calibration frequency: Calibration must be performed once per reagent lot using fresh reagent (i.e. not more than 24 hours since the reagent kit was registered on the analyzer). Renewed calibration is recommended as follows:
- after 1 month (28 days) when using the same reagent lot
- after 7 days when using the same reagent kit on the analyzer
- as required: e.g. quality control findings outside the defined limits

Quality control
For quality control, use PreciControl Universal.

In addition, other suitable control material can be used. Controls for the various concentration ranges should be run individually at least once every 24 hours when the test is in use, once per reagent kit, and following each calibration.

The control intervals and limits should be adapted to each laboratory’s individual requirements. Values obtained shall fall within the defined limits. Each laboratory should establish corrective measures to be taken if values fall outside the defined limits.

Follow the applicable government regulations and local guidelines for quality control.

Calculation
The analyzer automatically calculates the analyte concentration of each sample (either in nmol/L, ng/mL or μg/L).

Conversion factors: \[ \text{nmol/L} \times 0.314 = \text{ng/mL (μg/L)} \]
\[ \text{ng/mL} \times 3.18 = \text{nmol/L} \]

Limitations - interference
The assay is unaffected by icterus (bilirubin disatrate < 923 μmol/L or < 54 mg/dL), hemolysis (≥ 0.6%, ≤ 1.0 g/dL), lpramia (Intralipid < 720 mg/dL) and biotin (< 82 μmol/L or < 20 mg/mL).

Criterio: Recovery within ± 10 % of initial value.

Samples should not be taken from patients receiving therapy with high biotin doses (i.e. > 5 mg/day) until at least 8 hours following the last biotin administration.

No interference was observed from rheumatoid factors up to a concentration of 2000 IU/mL.

In vitro tests were performed on 18 commonly used pharmaceuticals. Of these, only phenylbutazone at therapeutic dosage levels showed interference with the assay (progestørone values depressed).

In rare cases, interference due to extremely high titers of antibodies to analyte-specific antibodies, streptavidin or ruthenium can occur. These effects are minimized by suitable test design.

For diagnostic purposes, the results should always be assessed in conjunction with the patient’s medical history, clinical examination and other findings.

Limits and ranges

Measuring range
0.095-191 nmol/L or 0.030-60.0 ng/mL (defined by the lower detection limit and the maximum of the master curve). Values below the lower detection limit are reported as < 0.095 nmol/L or < 0.030 ng/mL. Values above the measuring range are reported as > 191 nmol/L or > 60.0 ng/mL.

Lower limits of measurement
Lower detection limit: 0.095 nmol/L (0.030 ng/mL)

The lower detection limit represents the lowest measurable analyte level that can be distinguished from zero. It is calculated as the value lying two standard deviations above that of the lowest standard (master calibrator, standard 1 + 2 SD, repeatability study, n = 21).

Dilution
Samples with progesterone concentrations above the measuring range can be diluted with Diluent Estradiol/Progestørone or a suitable human serum with a low analyte concentraion. The recommended dilution is 1:10. The concentration of the diluted sample must be > 6 nmol/L (> 2 ng/mL).

After dilution, multiply the result by the dilution factor.

Depending on the biological variance of the diluted patient sample and the human serum matrix used for production of Diluent Estradiol/Progestørone, lower recovery of diluted samples may be observed.

Expected values
Studies with the Elecsys Progestørone II assay have revealed the following progesterone values:
Obtained:

Controls

Results

Representative Specific ranges.

Elecsys Progesterone II Multicenter Evaluation, 5/99

Each laboratory should investigate the transferability of the expected values to its own patient population and if necessary determine its own reference ranges.

Specific performance data

Representative performance data on the analyzers are given below. Results obtained in individual laboratories may differ.

Precision

Precision was determined using Elecsys reagents, pooled human sera and controls in a modified protocol (EP5-A) of the CLSI (Clinical and Laboratory Standards Institute): 6 times daily for 10 days (n = 60); repeatability on MODULAR ANALYTICS E170 analyzer, n = 21. The following results were obtained:

Elecsys Progesterone II Multicenter Evaluation, 5/99

<table>
<thead>
<tr>
<th>Test subjects</th>
<th>N</th>
<th>Percentiles</th>
<th>50th</th>
<th>5-95th</th>
<th>50th</th>
<th>5-95th</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td></td>
<td>nmol/L</td>
<td>ng/mL</td>
<td>nmol/L</td>
<td>ng/mL</td>
</tr>
<tr>
<td>Men</td>
<td>33</td>
<td></td>
<td>1.8</td>
<td>0.7-4.3</td>
<td>0.6</td>
<td>0.2-1.4</td>
</tr>
<tr>
<td>Women</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>• Follicular phase</td>
<td>192</td>
<td></td>
<td>2.1</td>
<td>0.6-4.7</td>
<td>0.7</td>
<td>0.2-1.5</td>
</tr>
<tr>
<td>• Ovulation phase</td>
<td>13</td>
<td></td>
<td>3.9</td>
<td>2.4-9.4</td>
<td>1.2</td>
<td>0.8-3.0</td>
</tr>
<tr>
<td>• Luteal phase</td>
<td>158</td>
<td></td>
<td>36</td>
<td>5.3-86</td>
<td>11</td>
<td>1.7-27</td>
</tr>
<tr>
<td>• Postmenopause</td>
<td>89</td>
<td></td>
<td>1.0</td>
<td>0.3-2.5</td>
<td>0.3</td>
<td>0.1-0.8</td>
</tr>
</tbody>
</table>

MODULAR ANALYTICS E170, cobas e 601 and cobas e 602 analyzers

<table>
<thead>
<tr>
<th>Sample</th>
<th>Mean</th>
<th>SD</th>
<th>CV</th>
</tr>
</thead>
<tbody>
<tr>
<td>Human serum 1</td>
<td>4.99</td>
<td>1.57</td>
<td>0.04</td>
</tr>
<tr>
<td>Human serum 2</td>
<td>38.2</td>
<td>12.0</td>
<td>0.17</td>
</tr>
<tr>
<td>Human serum 3</td>
<td>96.0</td>
<td>30.2</td>
<td>0.76</td>
</tr>
<tr>
<td>PreciControl U1</td>
<td>28.1</td>
<td>8.83</td>
<td>0.19</td>
</tr>
<tr>
<td>PreciControl U2</td>
<td>66.1</td>
<td>20.8</td>
<td>0.35</td>
</tr>
</tbody>
</table>

b) U = Universal

Elecsys 2010 and cobas e 411 analyzers

<table>
<thead>
<tr>
<th>Sample</th>
<th>Mean</th>
<th>SD</th>
<th>CV</th>
</tr>
</thead>
<tbody>
<tr>
<td>Human serum 1</td>
<td>4.99</td>
<td>1.57</td>
<td>0.09</td>
</tr>
<tr>
<td>Human serum 2</td>
<td>38.2</td>
<td>12.0</td>
<td>0.53</td>
</tr>
<tr>
<td>Human serum 3</td>
<td>96.0</td>
<td>30.2</td>
<td>1.54</td>
</tr>
<tr>
<td>PreciControl U1</td>
<td>28.1</td>
<td>8.83</td>
<td>0.38</td>
</tr>
<tr>
<td>PreciControl U2</td>
<td>66.1</td>
<td>20.8</td>
<td>0.77</td>
</tr>
</tbody>
</table>

Method comparison

A comparison of the Elecsys Progesterone II assay (y) with the Elecsys Progesterone assay (x) using clinical samples gave the following correlations (ng/mL):

Number of samples measured: 88

Passing/Bablok

\[ y = 0.91x + 0.29 \]

Linear regression

\[ y = 0.90x + 0.50 \]

\[ r = 0.998 \]

The sample concentrations were between approximately 0.6 and 140 nmol/L (approximately 0.2 and 44 ng/mL).

Analytical specificity

For the antibody derivative used, the following cross-reactivities were found (in %):

Androstenediol 0.002
Androstenedione 0.136
Corticosterone 0.687
Cortisol 0.005
Danazol 0.002
DHEA-S 0.009
D(-)-Norgestrel 0.008
Estradiol 0.009
Ethisterone 0.002
Ethinolindiol acetate n.d."\v
Medroxyprogesterone 0.812
Norethindrone 0.010
Norethindrone acetate n.d.
Testosterone 0.020
4-Pregn-11β-17α-diol-3,20-dione 0.338
11-Desoxycorticosterone 0.296
11-Desoxycorticisol 0.392
5-α-Dihydrotestosterone 0.040
5-β-Dihydroprogesterone 20.7
5α-Pregnen-3β-ol-20-one 0.858
5α-Pregnan-3α-ol-20-one 0.211
6α-Methyl-17α-hydroxy-progesterone acetate 0.257
6α-Methylprednisolone n.d.
17α-Hydroxyprognenolon 0.018
17α-Hydroxyprogesterone 1.30
20α-Hydroxy-4-pregnen-3-one 0.016
c) n.d. = not detectable

Functional sensitivity

0.48 nmol/L (0.15 ng/mL)
The functional sensitivity is the lowest analyte concentration that can be reproducibly measured with an intermediate precision CV of < 20 %.

References

For further information, please refer to the appropriate operator’s manual for the analyzer concerned, the respective application sheets, the product information and the Method Sheets of all necessary components (if available in your country).

A point (period/stop) is always used in this Method Sheet as the decimal separator to mark the border between the integral and the fractional parts of a decimal numeral. Separators for thousands are not used.

Symbols
Roche Diagnostics uses the following symbols and signs in addition to those listed in the ISO 15223-1 standard.

<table>
<thead>
<tr>
<th>CONTENT</th>
<th>SYSTEM</th>
<th>REAGENT</th>
<th>CALIBRATOR</th>
</tr>
</thead>
<tbody>
<tr>
<td>Contents of kit</td>
<td>Analyzers/Instruments on which reagents can be used</td>
<td>Reagent</td>
<td>Calibrator</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Volume after reconstitution or mixing</td>
<td></td>
</tr>
</tbody>
</table>

COBAS, COBAS E, ELECSYS, MODULAR and PRECICONTROL are trademarks of Roche. INTRALIPID is a trademark of Fresenius Kabi AB.

All other product names and trademarks are the property of their respective owners.

Significant additions or changes are indicated by a change bar in the margin.

© 2013, Roche Diagnostics