

RESEARCH ARTICLE

Life on the margin: Rainwater tanks facilitate overwintering of the dengue vector, *Aedes aegypti*, in a sub-tropical climate

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Abstract

A key determinant of insect persistence in marginal habitats is the ability to tolerate environmental extremes such as temperature. *Aedes aegypti* is highly invasive and little is known about the physiological sensitivity of the species to fluctuating temperature regimes at the lower critical threshold for development. A temperature that may limit the establishment and persistence of the species in sub-optimal regions. Daily winter temperatures were measured in common Australian larval habitats, replicated in environmental chambers and used to investigate the effect of fluctuating temperatures on the development and survival of tropical and subtropical strains of Australian *Ae. aegypti*. Development was slow for all treatments but both strains were able to complete development to the adult stage, suggesting previous models underestimate the potential for the species to persist in eastern Australia. Results suggested that thermal buffering in large volume habitats, and water that persists for greater than 32 days, will facilitate completion of the life cycle during sub-tropical winters. Furthermore, we provide a non-linear estimate of the lower critical temperature for *Ae. aegypti* development that suggests the current threshold may be incorrect. Our study demonstrates that the current re-introduction of water storage containers such as rainwater tanks, into major Australian population centres will increase the risk of *Ae. aegypti* establishment by permitting year-round development in locations south of its current distribution.

Introduction

A key determinant of insect distribution and persistence is the ability of a species to tolerate micro-climates at a local scale [1]. Conditions within the core distribution of a species will be near-optimal, whereas less stable populations will persist around the margins of an insect's distribution. Population stability is mediated by access to food sources, the availability of suitable oviposition and resting sites and the abiotic factors such as temperature associated with these

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micro-habitats [2–4]. In recent years there has been renewed interest in predicting the spread of the mosquito *Aedes aegypti* (L.) into cool range margins, primarily due the increased variability in temperature and rainfall associated with climate change and the importance of the species as a disease vector [4]. In particular, rising temperatures, unpredictable rainfall and urban landscapes that are changing in response to climatic instability may impact mosquito distributions, daily activity patterns and peak annual population abundance in marginal habitats [2, 3, 5–8].

Aedes aegypti is a highly anthropophilic species [2, 9]. The continuous availability of oviposition sites and blood meals afforded by intra-domiciliary habitats can mitigate otherwise hostile environments and has allowed the species to achieve a global distribution [10, 11]. Pertinent to the spread of *Ae. aegypti* into parts of Australia, where the species was once common, is the increasing presence of large water storage containers, namely domestic rainwater tanks. These tanks can provide permanent aquatic habitats throughout the year, particularly in dry locations such as southern Queensland, where the species was once highly prevalent [3, 10, 12]. It is hypothesised that the presence of rainwater tanks may have contributed to historical distribution patterns of *Ae. aegypti* across temperate Australia [3]. The removal of this key larval habitat is thought to have contributed to the elimination of the species from southern Australia during the mid-1900s [3, 12]. The temperature-buffering effect of rainwater tanks may allow continual *Ae. aegypti* larval development, however this has not been tested in marginal habitats [2]. Water has a high specific heat capacity, low thermal inertia and, in large volumes, can resist large and rapid fluctuations in temperature [13]. We believe that the modern trend for the widespread installation of large water storage containers, in response to an unpredictable climate, may increase the risk of *Ae. aegypti* re-establishment and expansion.

The lower critical temperature for the development of larval stages of *Ae. aegypti* is widely accepted to be approximately 11.8°C [4, 14]. Methodologies to estimate thermal performance rely primarily on observations of development at constant temperatures and the use of linear regression to estimate lower critical thresholds [4]. Due to the difficulty of working at extremes in the thermal performance curve, little research has defined the lower critical threshold for *Ae. aegypti* using non-linear methods.

It has been recognized for some time that insects subjected to constant temperatures in the laboratory do not accurately reflect development and survival in the field [15]. Studies using fluctuations in temperature more accurately reflect the natural daily cycles experienced by insects. In mosquitoes, fluctuations in water temperature can alter immature mosquito development and survival around critical development temperatures. Studies suggest that thermal tolerance is improved when fluctuating temperature regimes are compared with those using constant temperatures [14, 16, 17]. For instance, Carrington et al. [14] found that a large diurnal thermal range of 18.6°C around a mean of 16°C significantly reduced development time (but not survival) of *Ae. aegypti* when compared to small (7.6°C) fluctuations or constant temperatures. There has been little research on the survival and development of *Ae. aegypti* in fluctuating temperatures and their relation to potential geographic distribution, particularly around the lower critical threshold.

To examine whether rainwater tanks encourage the survival and development of *Ae. aegypti* under temperate Australian conditions, the abiotic conditions typical of tanks (limited temperature fluctuations) and smaller containers (high fluctuations) were measured during winter in Brisbane, Australia. Those fluctuating temperatures were then replicated in environmental chambers and we assessed their impact on the larval development and survival of a tropical and subtropical strain of Australian *Ae. aegypti*. It was hypothesised that 1) the temperature buffering provided by rainwater tanks will increase survival and time to adult emergence when compared to smaller volume habitats and 2) that the subtropical *Ae. aegypti* population would

have higher survival and faster development under winter conditions than *Ae. aegypti* sourced from the tropics, due to adaptations to local conditions. Using our results and development rates sourced from the literature, we applied different non-linear equations to explore the lower critical temperature of development in *Ae. aegypti*.

Methods

Environmental observations

Temperature fluctuations in different container types were measured over winter in Brisbane (27.47° S, 153.03° E), from the start of June until September, 2014. HOBO Pendant data loggers were placed into ten rainwater tanks and ten buckets, and tank locations were classified into high (>66%), medium (33–65%) and low (0–32%) shade categories. Shade categories were estimated visually as the percentage of structural or vegetative shade covering each container. Air temperatures were measured outside in a high shade location. Tanks were checked fortnightly to ensure they remained sealed to the ingress of adult mosquitoes. Locations of tanks included: Greenslopes (27.51° S, 153.05° E), Moorooka (27.54° S, 153.03° E), Salisbury-Nathan (27.55° S, 153.03° E), Sunnybank (27.58° S, 153.06° E), Camira-Gailes (27.63° S, 152.91° E), Indooroopilly (27.50° S, 152.97° E) and St Lucia (27.50° S, 153.00° E; Fig 1). A black 9L bucket, representing the most common type of container in Brisbane backyards (Darbro, pers. comm.), was placed under similar shade conditions in a northerly position next to each tank, to ensure it caught the southern hemisphere sun.

Data loggers were attached to a floatation device in each tank, submerged to a depth of 30cm below the water surface to avoid warmer surface temperatures. Flotation devices were attached to a tape measure suspended from the roof of the tank. Water losses due to evaporation were noted fortnightly from tanks and buckets. Ten abiotic characteristics were measured from tanks including: temperature inside and out of each tank, total volume of water, humidity, dew point, pH, salinity, conductivity, total dissolved solids, and presence of larvae inside tanks and “first flush” devices (a separate pipe for collecting sediment before water enters a tank). The five-sweep netting technique was used to sample mosquito larvae from rainwater tanks [20]. As *Ae. aegypti* is not currently present in Brisbane, the native tree hole mosquito, *Aedes notoscriptus*, was used as an indicator species for tank productivity. After the first survey, two tanks were disconnected from input water sources to estimate evaporation rate and to prevent ingress of mosquito larvae and eggs. Archerfield (-27.57° S, 153.01° E) climate data was selected as it is the closest Bureau of Meteorology [18] station to the rainwater tanks surveyed. Informed, written consent was obtained from residents to permit the collection of eggs and rainwater tank data within private premises. Approval for this study was provided by the QIMR Berghofer Medical Research Institute human ethics committee.

Survival and development rate trial

Aedes aegypti colonies were established from eggs sourced from field sites in Cairns (tropical strain; -16.92°S, 145.78°E; coldest month July: mean high 25.8°C, mean low 17.1°C) and Gin Gin (subtropical strain; -24.99°S, 151.95°E; coldest month June: mean high 22.2°C, mean low 10.3°C), Queensland, in January, 2015. The subtropical colony originated from 30 eggs collected from a single ovitrap at three houses (separated by >100m), while the tropical colony was established from >500 eggs collected from a single ovitrap at five separate properties. North Queensland is within the optimal range of the species and has the highest genetic diversity of Australian populations [21]. A PCR test for the presence of *Wolbachia* [22] revealed absence in both colonies (n = 60). Colonies were maintained at QIMR Berghofer at >500 individuals per generation, and insectary conditions held at 26 ± 1°C, a 70% (± 10%) relative

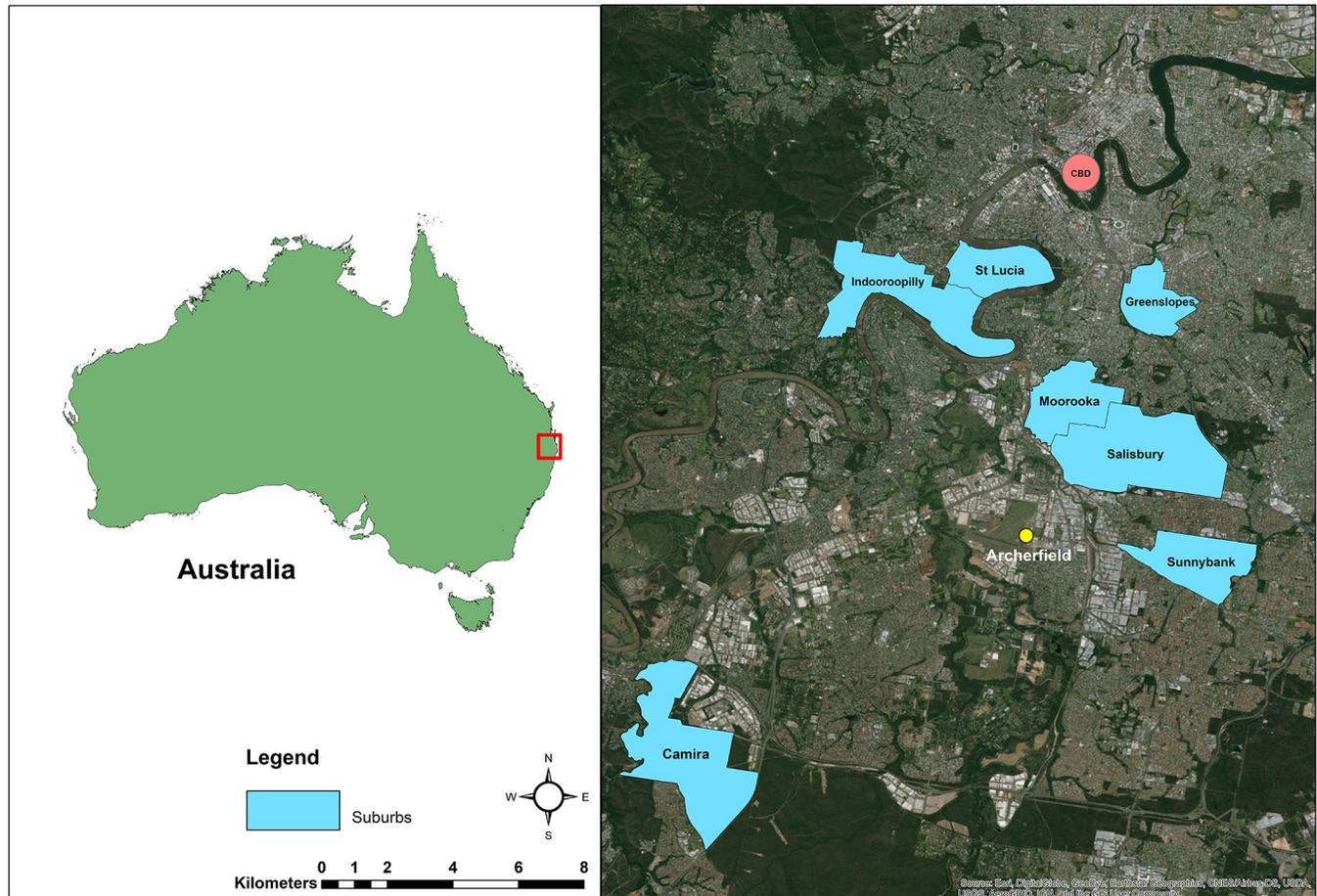


Fig 1. Location of study sites in Brisbane, Australia. Suburbs where temperatures in container habitats were measured during winter, 2014. Climate data was taken from Archerfield (yellow circle)[18]. Map Source: Base layer of Brisbane region sourced from Esri World Imagery [19]; Esri, DigitalGlobe, GeoEye, Earthstar Geographics, CNES/Airbus DS, USDA, USGS, AeroGRID, IGN, and the GIS User Community.

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humidity, with a 12:12 hour light cycle with twilight period. Adults were blood fed on an adult volunteer for 15 minutes, two days after emergence for three consecutive days (QIMR Berghofer Medical Research Institute human ethics form P2273). Eggs were collected from both colonies after generation two for use in environmental chamber experiments. Eggs were hatched synchronously using vacuum immersion in water at room temperature (24°C) for one hour.

A fluctuating temperature regime was derived from tank and bucket measurements during the coldest week in Brisbane during July 2014. These fluctuations were replicated in environmental chambers using two hourly intervals (S1 Table). A control treatment was set at 26°C ($\pm 1^\circ\text{C}$), 70% ($\pm 10\%$) and light regimes for all larval treatments were set at a 10:14h cycle, typical of Brisbane in July. Humidity for environmental chambers was set at 75% which is comparable to those observed in rainwater tanks during winter (S2 Table).

Fifty first-instar larvae from each mosquito strain were transferred into each of eight white, plastic containers (183 x 152 x 65mm) for a total of 400 larvae per strain, per temperature treatment, in a randomized block design and 500mL of tap water (aged 2 days) was placed into each container. Larvae were fed with TetraMin ground fish food (Tetra, Germany) standardized to the high diet treatment of Hugo et al. [23], with food concentrations estimated per larvae per volume each day and excess food removed daily before feeding. Containers were

topped up daily with pre-chilled water (equivalent temperature to each environmental chamber). Trays were rotated within environmental chambers and insectary shelves daily to prevent location bias. Containers were photographed each day to facilitate counting of all surviving instars and adult emergence was also recorded (defined as complete emergence from the pupal case).

Statistical analysis

To assess the effect of temperature on survival to adult in all treatments, Kaplan Meier (log-rank) survival analysis was used [24]. Student's t-tests and ANOVA were used to compare mean survival, development times and degree days for *Ae. aegypti* strains in tanks, buckets and controls. Student's t-tests were used to compare air temperature, humidity and dew point from measurements taken inside and outside the tanks. Heating degree day (HDD) models were constructed at 30 minute intervals, with a lower critical temperature of 11.78°C for the constant, tank and bucket temperature treatments. Statistical significance between each HDD model was compared with t-tests. For an estimate of cold stress, a cooling degree day (CDD) model was calculated for bucket treatments. To compare findings with published rates of development using non-linear equations, we selected the range of studies used by the Eisen et al review [4], which were accessible and provided relevant data around the lower critical threshold. To ensure we included variation at the population level we selected studies that sourced *Ae. aegypti* from different continents. We selected models that crossed the X axis to ensure the zero development point could be calculated and used the correlation of observed to fitted model residuals to compare models. All analyses were done using R version 3.2.2 [25] with the 'nlme', 'survival' libraries, 'survminer' for plotting survival curves and 'devRate' for comparing development rates. A map of Brisbane suburbs was created using ArcGIS 10.5 software by Esri (ESRI Inc., Redlands, CA, USA).

Results

Field observations

During winter 2014 (1st June until August 31st) Brisbane experienced average rainfall conditions, with above average maximum air temperatures, below average minimum temperatures and a minimum temperature of 0.5°C [18](S3 Table). Rainwater tanks had a mean temperatures of 16.8°C (range = 11.3°C, SD = 1.9) while buckets had a mean temperature of 16.3°C (range = 29.9°C, SD = 4.1; S2 Table). The relative difference between the mean weekly temperature in tanks consistently stayed above the mean weekly air temperature throughout the winter (mean relative difference = 1.3°C, SD = 0.14), while the relative difference in buckets was 0.55°C (SD = 0.14; Fig 2). The mean hourly tank temperature in high and low shade did not drop below the lower critical temperature during July (Fig 3). The minimum temperature of tanks only dropped below the lower critical temperature on 5.4% (5 of 92 days)(Fig 4). Temperatures below the lower critical temperature coincided with tanks in high shade or containing under 500L of water at the time of measurement (Fig 4, S2 Table).

In buckets the mean hourly temperature (high and low shade regimes) and mean daily minimum for all shade regimes dropped below the lower critical temperature throughout July (Fig 3) and for all months during winter (Fig 4), respectively. Daily temperatures in buckets dropped below the lower critical temperature 66.3% (61/92) and 93.3% (28/30) of the time in winter and July, respectively. During July, the lowest temperatures observed in buckets, tanks and air was 5.4°C, 11.2°C and 0.5°C respectively. Differences between mean internal (21.4°C, SD = 0.99) and external (21.7°C, SD = 0.94) air temperatures of tanks at different shade levels measured fortnightly were not significant ($F(1,12) = 0.26, P = 0.80$).

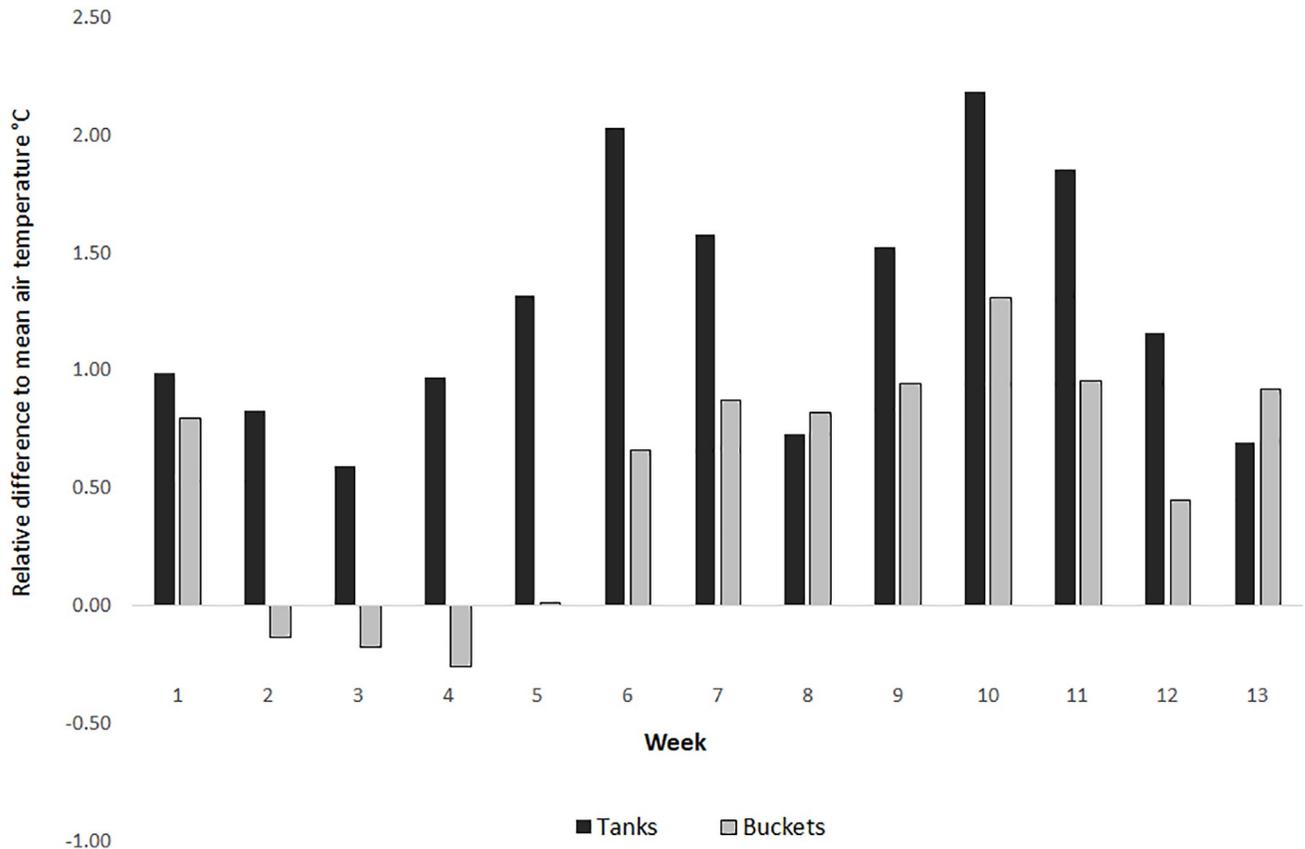


Fig 2. Relative differences between weekly container water and air temperatures. Relative difference in mean weekly water temperature for rainwater tanks (black) and buckets (grey) to air temperature in 100% shade during winter in Brisbane, 2014.

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Relative humidity was significantly higher inside rainwater tanks (mean = 78.1, SD = 11.0) than outside (mean = 48.8, SD = 10.7; $t(8) = -9.9$, $P < 0.001$). Likewise, fortnightly differences between the mean internal (17.2°C, SD = 4.2) and external (9.9°C, SD = 3.7) dew points were significant ($t(12) = -4.47$, $P < 0.001$). Applying evaporation rates observed in the low shade treatment (assuming a linear relationship over time), we estimate the water in a 9L bucket would take approximately 105 days to evaporate during winter. All abiotic conditions including humidity, dewpoint, salinity, total dissolved solids, changes in volume and evaporation for tanks and buckets are recorded in the Supporting Information (S2 Table).

The presence of mosquitoes was observed in tanks fortnightly (S4 Table). *Aedes notoscriptus* was the primary species observed, with a total of 1,820 (mean = 26/container, SD = 71.64) immature stages counted. Larvae were present in rainwater tanks in 12.5% to 100% of fortnightly surveys (S4 Table). The two tanks that were sealed against any further ingress of rainwater had *Ae. notoscriptus* larvae present only during the first 14 days. The total abundance of immature mosquitoes in first flush devices was 200 (mean = 4.8, SD = 18.7) and larval presence ranged from 12.5% to 62.5% of all surveys (S5 Table).

***Aedes aegypti* larval development and survival under fluctuating temperatures in simulated containers**

In environmental chambers, temperatures within containers differed from programmed air temperatures by 1°C (SD = 0.98 tanks, SD = 0.2 buckets). This was due to the thermal capacity

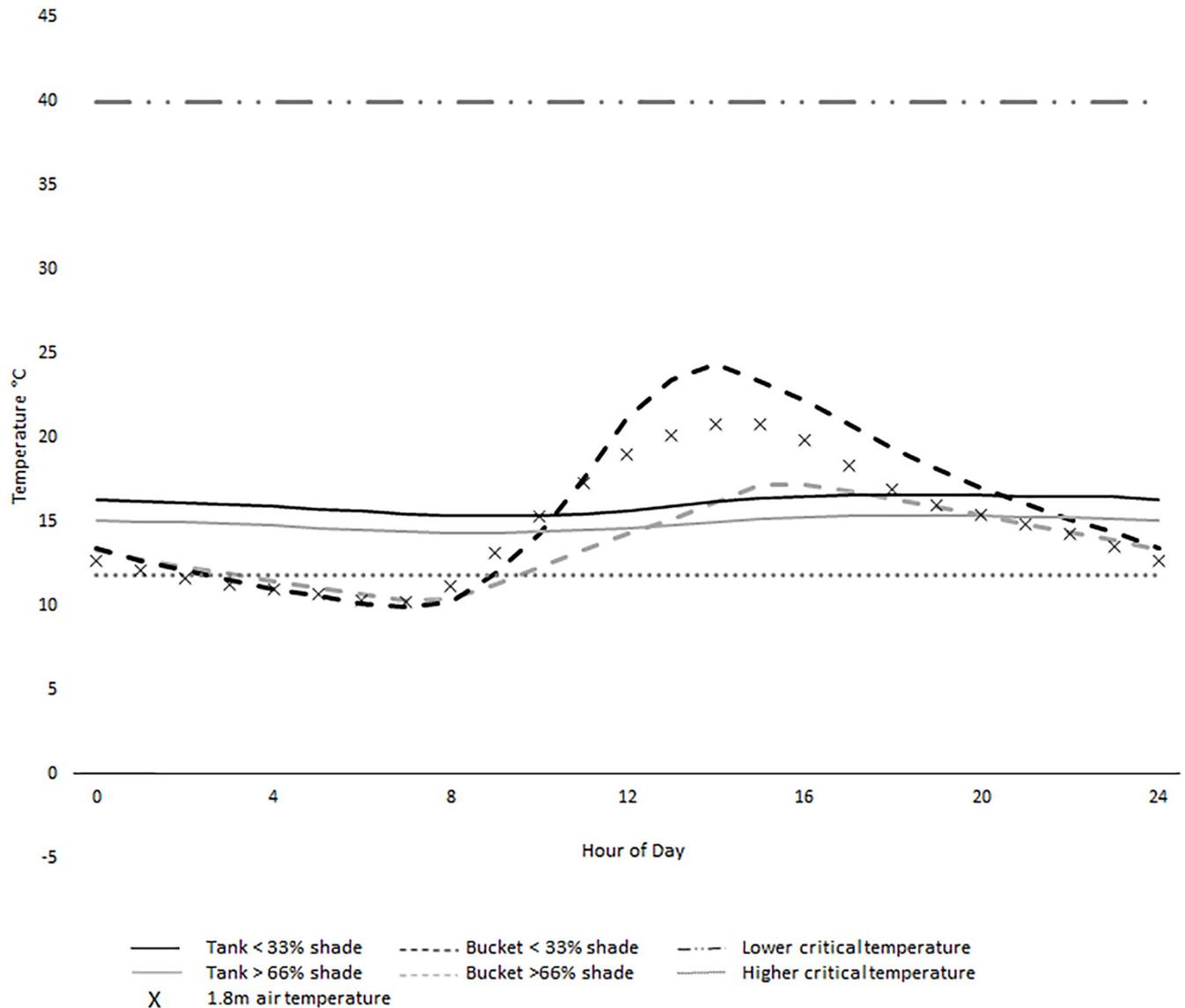


Fig 3. Mean hourly water temperatures from buckets and rainwater tanks during July in Brisbane, 2014. Fluctuations were recorded from high shade (black line) and low shade tanks (grey line), and high shade (black dashed line) and low shade buckets (grey dashed line). Air temperatures are recorded from 100% shade (crosses, 1.8m height) and upper (dash-dot line) and lower (dotted line) critical thresholds of *Aedes aegypti* are displayed.

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of the water stored within the chambers. Rainwater tank temperature simulations increased *Ae. aegypti* larval survival when compared to bucket simulations ($\chi^2 = 59.7$, $df = 1$, $P < 0.001$). This was true for tropical strains (Fig 5; S6 Table; $\chi^2 = 18.3$, $df = 1$, $P < 0.001$) and subtropical strains (Fig 5; S6 Table; $\chi^2 = 47.8$, $df = 1$, $P < 0.001$). *Aedes aegypti* larvae from the tropical strain had higher survival in both rainwater tanks ($\chi^2 = 5.2$, $df = 1$, $P = 0.022$) and buckets ($\chi^2 = 24.7$, $df = 1$, $P < 0.001$) when compared to the subtropical strain.

A comparison of tropical and sub-tropical temperature regimes from tanks showed no differences in mean time to adult emergence (32.5, SE = 0.19; 32.7, SE = 0.20). The same was true for comparisons of mean adult emergence from subtropical (32.2, SE = 0.23) and tropical (31.4, SE = 0.18) bucket temperatures ($F(1,29) = 1.48$, $P = 0.234$; S7 Table). Analysis indicated

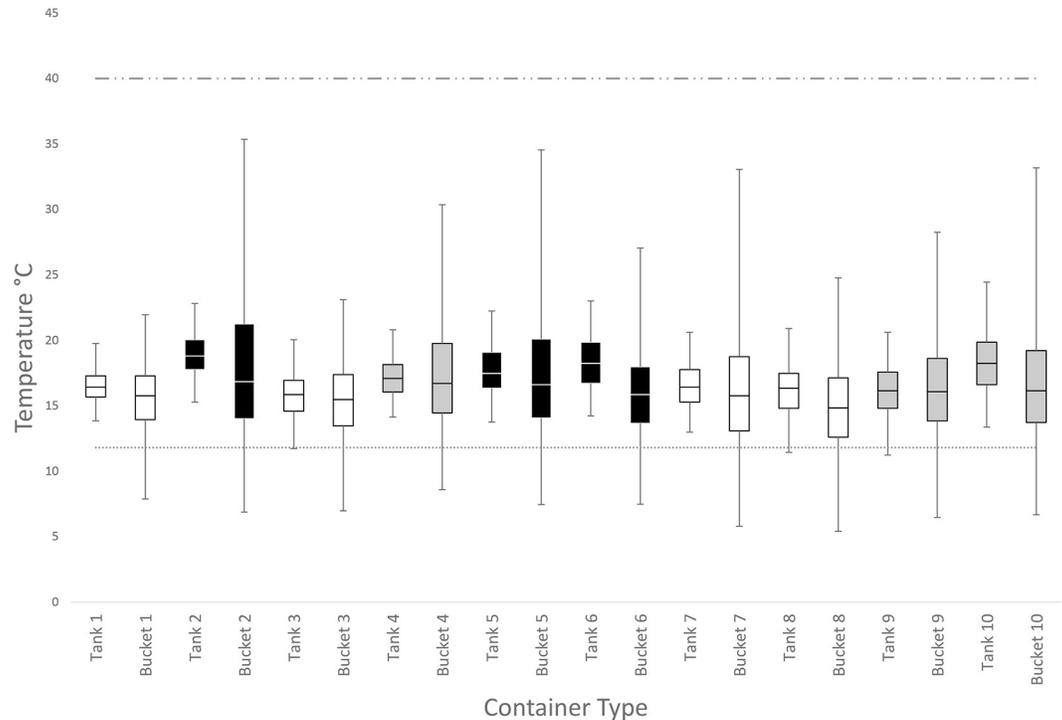


Fig 4. Interquartile ranges of water temperatures of rainwater tanks and buckets in Brisbane during winter 2014. Lower critical temperature threshold (dotted line, 11.8°C) for *Aedes aegypti* taken from Eisen et al. [4] and upper threshold (dash-dot line, 40.0°C) from Richardson et al. [26]. Black, grey and white boxes represent low, moderate and high shade conditions, respectively.

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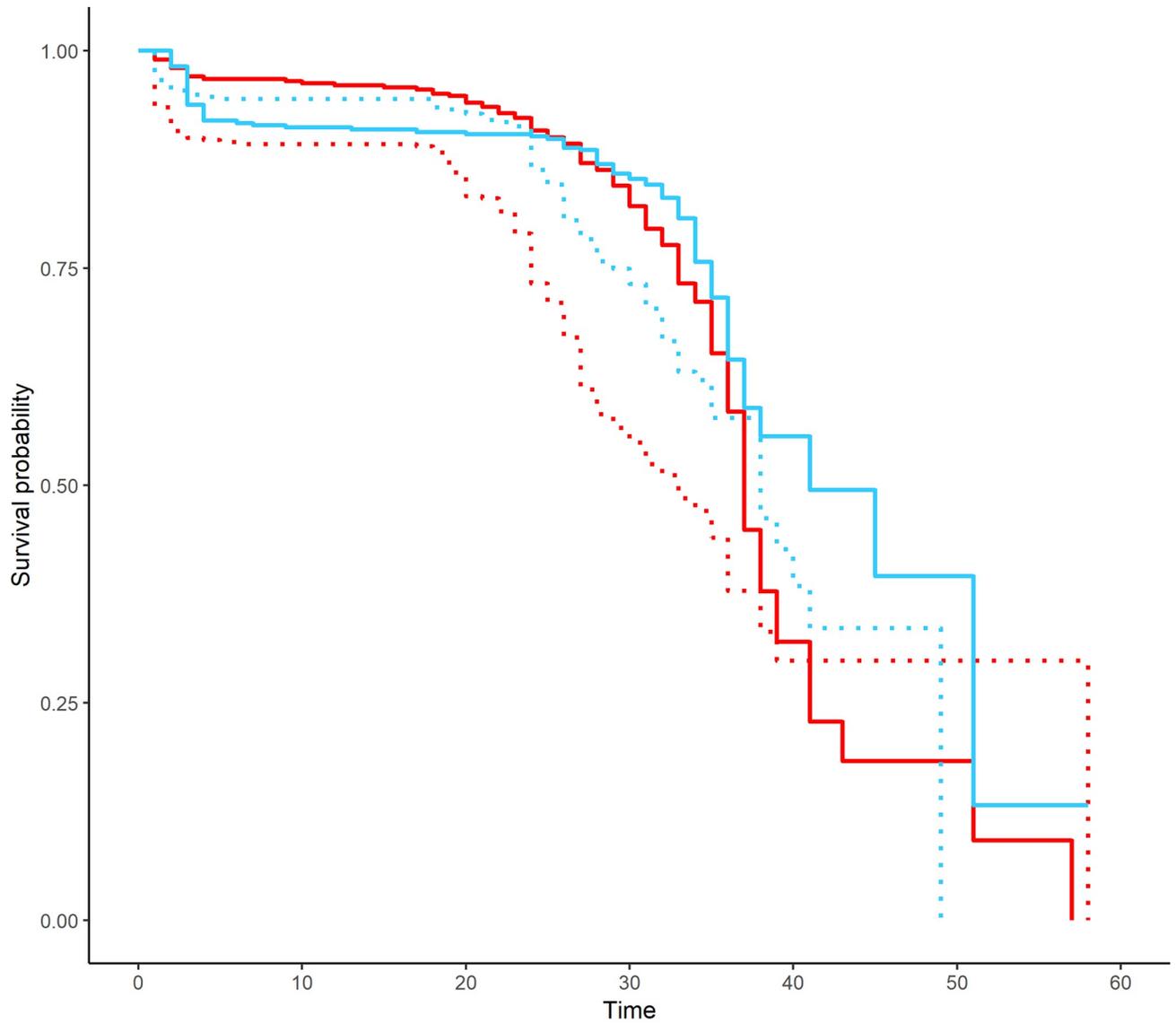
that strain and container type had no effect on mean larval development time ($F(1,29) = 0.646$, $P = 0.428$), and the interaction effect was not significant ($F(1,28) = 0.242$, $P = 0.627$).

Non-linear estimate of *Aedes aegypti* larval lower critical temperature

We successfully fitted seven non-linear curves to *Ae. aegypti* larval development rates and temperatures derived from the published literature (Table 1). Correlations between observed and fitted values were similar across most scenarios. The model with the best correlation that allowed for an estimate of a larval zero development threshold was the Logan et al. model [27], which had a correlation of 0.899 (Table 1, Fig 6). This model does not have a parameter for the lower critical threshold, so the equation was solved for the zero development point on the X axis (9.21°C, Table 1, Fig 6). Others estimate this value between 6.55°C and 12.38°C and confidence intervals around these estimates vary considerably (Table 1). Parameter estimates were included for the Sharpe and DeMichele non-linear model [28] traditionally used in simulating *Ae. aegypti* development [29]. However, it is impossible to estimate zero development threshold with this model as it never crosses the zero development point on the X axis.

Degree day estimates of *Aedes aegypti* development

Based on the lower critical temperature of 11.8°C [4, 14], the tropical and subtropical strains of *Ae. aegypti* larvae required 125.7 (SE ± 0.57) and 123.4 (SE ± 0.45) HDDs to develop into adults at a constant 26°C temperature ($t(14) = 2.28$, $P = 0.046$). There was no significant difference in HDDs required at tank temperatures between the tropical (mean = 80.6, SE ± 2.04) and subtropical strains (mean = 81.1, SE ± 2.16) ($t(14) = -0.71$, $P = 0.401$). Nor was there any



Legend: •• Subtropical Bucket — Subtropical Tank •• Tropical Bucket — Tropical Tank

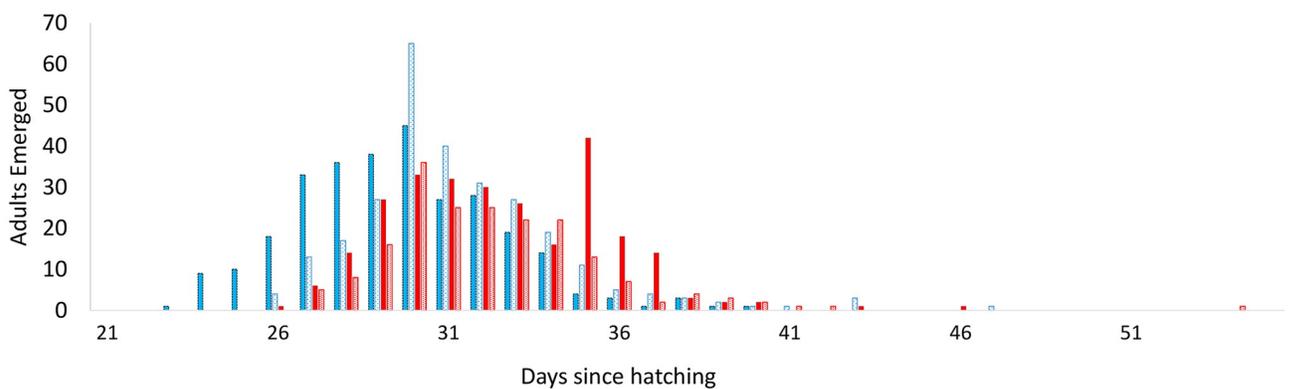


Fig 5. Survival curves and time to emergence of surviving adults comparing tropical (blue) and subtropical (red) *Aedes aegypti* strains in tank (solid fill) and bucket (spotted fill) temperature treatments.

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Table 1. Fit of non-linear models to published larval *Aedes aegypti* development rates. Models were selected based on highest correlation between observed and fitted residuals. Parameter estimates, estimations of upper and lower critical thresholds, confidence intervals and observed versus expected correlations (Cor) for non-linear models of *Aedes aegypti* development rates under constant and fluctuating temperatures sourced from scientific literature.

Model [Ref]	R: devRate Model	Cor	Lower	Upper	Parameter 1	Par 2	Par 3	Par 4	Par 5	Par 6
			Threshold (95% CI)	Threshold (95% CI)						
[33]	kontodimas_04	0.894	10.84 (1.90)	41.34 (0.95)	0.00005					
[34]	perf2_11	0.891	12.38 (1.95)	40.67 (0.81)	0.01349	0.193				
[35]	briere1_99	0.883	10.00 (3.08)	40.13 (0.26)	0.00010					
[35]	briere2_99	0.893	12.15 (2.55)	40.44 (0.87)	0.00007	1.439				
[36]	hilbertLogan_83	0.898	6.55 (31.3)	45.38 (197.8)	3.154	62.28	7.62			
[27]	logan10_76	0.899	9.21 (NA)	43.16 (11.2)	1.128	0.135	47.22	12.644		
[28]	sharpeDeMichele_77	0.900	N/A	N/A	32.2	14068.5	-264.96	-76397	282.52	87234

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difference in HDDs required for development between bucket temperatures; subtropical (mean = 107.2, SE ± 1.72), tropical (mean = 104.1, SE ± 1.64; $t(14) = -1.26, P = 0.23$). However, when HDDs were compared between containers, there were a significant differences between tropical ($t(14) = 8.99, P < 0.001$) and subtropical treatments ($t(14) = 9.44, P < 0.001$). In the bucket treatment, the tropical and subtropical *Ae. aegypti* larval strains were exposed to 47.2 (SE ± 0.27) and 48.5 (SE ± 0.35) CDD, respectively, while tank treatments never accumulated cold stress.

Discussion

Historically, *Ae. aegypti* was present in Brisbane during the early twentieth century when unsealed rainwater tanks and other forms of water storage were common [12, 37, 38].

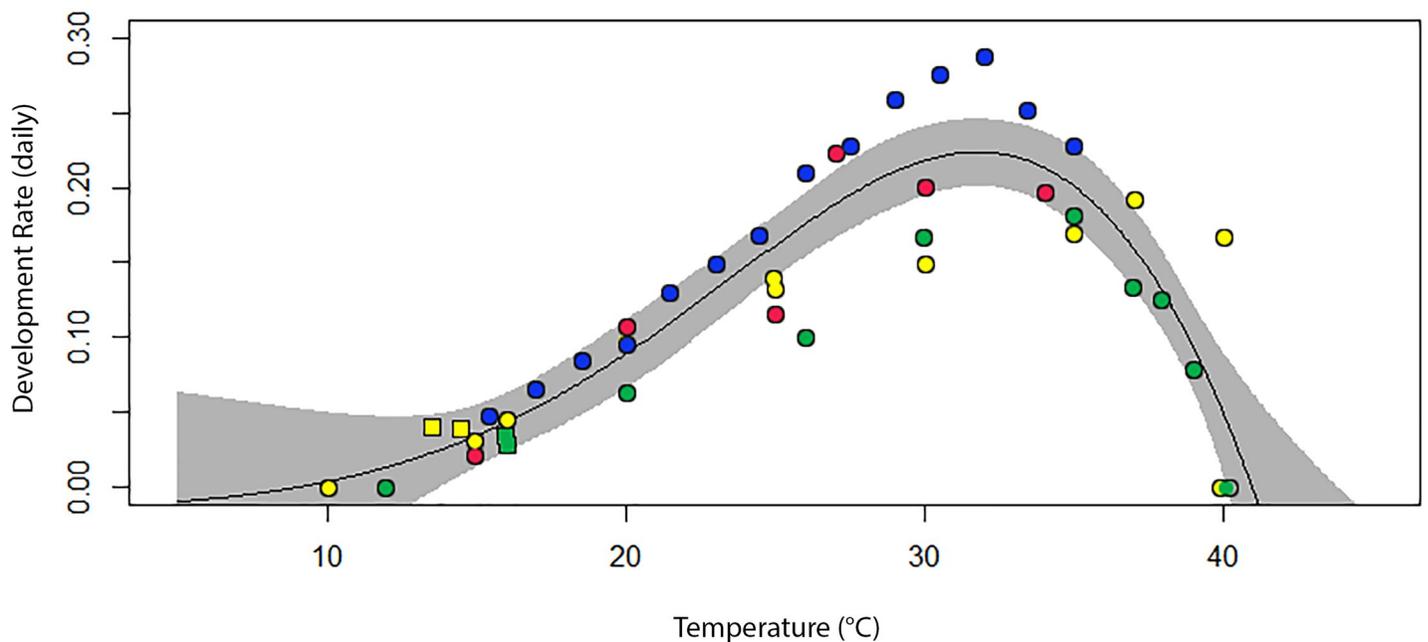


Fig 6. Non-linear development model for larval *Aedes aegypti*. Comparison of the mean and median time to pupation from published larval development rates, including the current study [14, 26, 30, 31, 32]. To these estimates we fit the temperature dependent model developed by Logan et al. [27]. Colours represent different continents; Africa (blue), Australia (yellow), South East Asia (green) and North America (red); spots represent constant and squares fluctuating temperatures.

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Modelling has suggested that conditions in Brisbane are currently inhospitable for the species during winter and there is contention to whether the species is able to persist in areas south of its current distribution under normal rainfall conditions [2, 39]. Our results suggest that the presence of water storage containers, such as rainwater tanks, may change these survival prospects by providing a semi-permanent larval habitat that is thermally buffered in a low rainfall, subtropical climate.

During winter, tank and bucket water temperatures were comparable, with differences in relative weekly mean temperature differing by less than 2.2°C throughout winter. The largest difference in relative temperature occurred between internal and external air temperatures, with tanks consistently retaining a higher mean air temperature than external air temperatures. Humidity levels of ~70% and high dewpoints over the surface of the water in rainwater tanks suggests that the air cavity may protect mosquito lifecycle stages when conditions outside are unfavourable. It is likely that these conditions may protect eggs and adults from desiccating during periods of low humidity that occur during Australian winters [40, 41].

Our results suggest *Ae. aegypti* larvae can develop and survive throughout winter in Brisbane, in both rainwater tanks and buckets. When larvae were reared under fluctuating temperature regimes derived from direct observations during the coldest winter month, approximately 50% and 70% survived to adults in buckets and rainwater tanks, respectively. The low thermal inertia exhibited in rainwater tanks resulted in mean weekly minimum and maximum water temperature rarely fluctuating more than $\pm 5^\circ\text{C}$ and seldom breaching the lower critical threshold for development. Rainwater tanks provided a buffered environment, with lower thermal stress, and we suggest that tanks provide a more stable habitat for *Ae. aegypti* larval development when compared to smaller volume containers such as buckets.

Evidence supported the null hypothesis that *Ae. aegypti* larvae could survive in simulated winter bucket temperatures from Brisbane. Specifically, 48–67% of larvae were observed to survive in bucket treatments representing the coldest week observed, where the mean temperature (13.5°C) was close to the lower critical temperature [4]. The dark colouring of buckets likely lowered solar reflectance, therefore enhancing absorption of solar radiation and increasing the upper fluctuation into the optimal temperature range during the day. However, at night buckets of all shade regimes stabilized with air temperature around 12am, and at times resulting in minimum temperatures dropping below the lower critical threshold. Conditions in our study had consistent volumes and ideal nutrition and water quality, so results should be interpreted as optimal conditions for survival within the temperatures tested. Although we exposed larvae to temperatures down to 4.5°C for short periods of time, the large fluctuation likely resulted in higher survival than was expected in these smaller containers.

Fluctuating temperatures can have varying effects on life span, particularly due to the length of time and amplitude of exposure outside of optimal conditions [15]. This suggests that the higher larval survival observed in the tank treatments (67% - 76%) was the result of lower cold stress (CDD) when compared with bucket treatments. Low temperature stresses may account for differences between our survival results and those contrasting results of Carrington et al. [14], who observed a higher survival in their large fluctuation treatment. Temperature fluctuations in the current study had lower maximum and minimum mean temperatures (22.8°C and 4.5°C respectively for the large fluctuation) which may have resulted in larvae experiencing longer periods below the lower critical threshold when compared with Carrington et al. [14] (range 25.3°C to 6.7°C). Furthermore, our study did not compare treatments with similar means due to differences in volumes observed in field containers and on which we based our temperature models. Our differences in larval survival support the hypothesis that rainwater tanks (small fluctuation) can provide a thermally buffered habitat during periods of stress, and

are likely to increase survival of immatures to adult emergence when compared to containers of smaller volume.

Although there were differences in larval survival between strains, results for temperature regimes in our study were above those observed in previous studies where constant temperatures were applied [14, 26, 30]. Previous studies sourced their strain from a similar location in North Queensland [26, 30] so it is unlikely that variations observed between the current and previous studies were attributable to population differences. Higher survival at lower temperature in the tropical strain suggests that results do not support the hypothesis that southern populations are better adapted to colder temperatures. We acknowledge that establishing a colony with 30 eggs is unlikely to be representative of a sub-tropical population but due to the small population size in the town, this was all that was available. As far as we are aware there are no records of temperature adaptation in *Ae. aegypti* so this is perhaps unsurprising. As adaptation is facilitated by genetic diversity and driven by the accumulation of beneficial mutations, so it is possible that small remnant or recently established southern Queensland populations may have very low genetic diversity [42] and therefore little potential for adaptation. We therefore suggest that fluctuating temperatures, whether high or low, can increase the survival of all *Ae. aegypti* larvae around lower critical thresholds for development.

The lower critical temperature for *Ae. aegypti* larvae is clearly less than what is currently accepted as the value for this limit (11.78 ~11.8°C) which is typically calculated using linear regression to estimate the point where the function crosses the temperature-axis [4, 14, 30]. Estimating the lower critical temperature via constant temperatures is highly artificial [16]. It may be that the fluctuating temperature studies we designed also give more accurate empirical thresholds. In reality, development rates at the lower threshold tend to decay in an exponential fashion and thermal minimums estimated using linear functions will have a high margin of error compared to estimations using non-linear methods [16]. Thus our findings, which indicated a threshold around 10°C, suggest that the lower critical threshold for *Ae. aegypti* larvae is likely lower than previous estimates and studies (particularly those using degree days) may have underestimated the ability of the species to endure colder temperatures.

Our estimates of larval development time in colder temperatures were consistent with other studies of Australian *Ae. aegypti* populations [26, 30]. These studies estimated mean emergence times of 30–39 days for constant temperatures regimes of 15°C and 16°C [26, 30]. Interestingly, no significant differences in mean development time were observed between the fluctuating temperature regimes in the current study. However, we did observe differences in the number of HDDs required for development when applying the traditional lower critical temperature, with mosquitoes under the tank regime requiring significantly fewer degree days than the bucket regime. It is likely that HDD estimates will be inaccurate when assuming the 11.8°C threshold. For example, the number of HDDs calculated for a small fluctuation around an inaccurate lower critical temperature, will tend to underestimate the total HDDs required for development compared to rates entering the linear part of the developmental curve [16]. Thus, interpreting development rates that apply linear relationships around thermal minimums must be done with caution.

Aedes notoscriptus larvae, the native container inhabiting species, were consistently present in sealed tanks throughout the winter. This suggests that roof guttering or piping may play an important role in 'seeding' tanks with larval mosquitoes during rainfall events. The role that gutters play as a source of container inhabiting mosquitoes has been identified previously in north Queensland [43]. In our study, gutters with high levels of organic matter or sitting water were likely responsible for eggs or larvae being washed into tanks during frequent rainfall events (S1 Fig). When tanks are sealed, there is little chance of adult mosquitoes escaping but any small gap in the multiple seals, mesh openings and plastic covers typical of rainwater

tanks, would convert them into highly productive habitats. Furthermore, the presence of *Ae. notoscriptus* larvae was observed in first flush devices throughout winter, suggesting that attendant infrastructure such as these containers, could seed larvae into tanks and contribute to local mosquito populations.

Our findings have important implications for estimating the potential distribution of *Ae. aegypti* and demonstrating the risk of re-establishment in southern Queensland where larval habitat is readily available in the form of rainwater tanks [8]. Considering that eggs and adults likely tolerate lower daily temperature extremes of $\sim -10^{\circ}\text{C}$ and $\sim 0^{\circ}\text{C}$ respectively [44], and a historical presence as far south as the Victorian border, it is no surprise that *Ae. aegypti* larvae are capable of completing development during winter temperatures in Brisbane. Kearney et al. [2] and Richardson et al. [26] postulated that rainfall was not sufficient in Brisbane for small containers, such as buckets, to act as larval habitat for *Ae. aegypti* throughout the year and it must be noted that limited larval nutrition may influence development in some containers [45]. However, our field observations indicated that any container holding $>3\text{L}$ or that retains water for >32 days in winter, is potentially productive during even the coldest months, with buckets representing one of the most common and productive containers for urban mosquitoes in Brisbane [8]. Considering that temperature alone is not limiting in Brisbane, it is likely that larval stages will be able to persist through colder weather, particularly if not limited by water volume or nutrition.

We conclude that rainwater tanks and buckets provide suitable larval habitat for the (re) establishment and persistence of invasive mosquito species in areas where low rainfall and temperatures might make establishment difficult. These containers ensure that sub-optimal landscapes can be unwittingly manipulated by human behaviours to support the establishment of invasive disease vectors across new urban areas. Drought conditions have resulted in the mass re-introduction of rainwater tanks into Brisbane and other major Australian cities. The last time Brisbane had large numbers of unregulated rainwater tanks *Ae. aegypti* and epidemics of dengue were common [10, 12]. This points to the importance of public education and the need for ongoing surveillance and management of rainwater tanks to ensure they remain compliant with regulatory standards. If rainwater tanks and other key containers are not managed appropriately, large areas of southern Australia may see the return of *Ae. aegypti* with tremendous implications for public health and the management of imported cases of dengue, chikungunya, Zika and yellow fever.

Supporting information

S1 Table. Temperature regimes. Environmental chamber temperatures used to determine survival of *Aedes aegypti* in different container categories from Brisbane, Australia. (DOCX)

S2 Table. Rainwater tank conditions. Abiotic conditions within rainwater tanks and buckets in Brisbane during winter (July) 2014. Levels represent 0–32% shade cover (1), 33–65% (2) and 66–100% (3). (DOCX)

S3 Table. Weather records. Temperatures recorded in air at Archerfield Airport, (-27.57°S , 153.01°E), tanks and buckets from Brisbane during winter (1st June until 31st August), 2014. (DOCX)

S4 Table. Presence/absence of *Aedes notoscriptus* immatures in sealed rainwater tanks during winter in Brisbane, 2014. Shading represents presence during larval surveys conducted fortnightly. All tanks were sealed, and tanks 1 and 8 had inflows of water removed after the

first survey.
(DOCX)

S5 Table. Mosquito presence in rainwater tanks. Presence of *Aedes notoscriptus* immatures (grey shading) in first flush devices during winter in Brisbane, 2014. Numbers correspond to the tank which contained devices. For example, tank 10 had two separate first flush devices (10a,10b) on downpipes entering tank. Volume measures the mean volume found in each device throughout the field survey. Presence represents the percentage of surveys where at least one *Ae. notoscriptus* immature was sampled from the device.
(DOCX)

S6 Table. *Aedes aegypti* survival. Survival of tropical and subtropical *Aedes aegypti* strains in rainwater tank (small fluctuation), buckets (large fluctuation) and 26°C control (constant) treatments.
(DOCX)

S7 Table. *Aedes aegypti* development. Mean, standard error, minimum and maximum development time for tropical and subtropical *Aedes aegypti* strains in rainwater tank (small fluctuation), buckets (large fluctuation) and 26°C control (constant) treatments.
(DOCX)

S1 Fig. Productive infrastructure. Roof gutters observed that likely increased productivity of rainwater tanks during winter in Brisbane, 2014.
(DOCX)

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References

1. Kearney M, Porter W. Mechanistic niche modelling: combining physiological and spatial data to predict species' ranges. *Ecol Lett*. 2009; 12(4):334–50. <https://doi.org/10.1111/j.1461-0248.2008.01277.x> PMID: 19292794
2. Kearney M, Porter WP, Williams C, Ritchie S, Hoffmann A. Integrating biophysical models and evolutionary theory to predict climatic impacts on species' ranges: the dengue mosquito *Aedes aegypti* in Australia. *Func Ecol*. 2009; 23(3):528–38. <https://doi.org/10.1111/j.1365-2435.2008.01538.x>
3. Russell RC, Currie BJ, Lindsay MD, Mackenzie JS, Ritchie SA, Whelan PI. Dengue and climate change in Australia: predictions for the future should incorporate knowledge from the past. *Med J Aust*. 2009; 190(5):265–8. PMID: 19296793
4. Eisen L, Monaghan AJ, Lozano-Fuentes S, Steinhoff DF, Hayden MH, Bieringer PE. The impact of temperature on the bionomics of *Aedes (Stegomyia) aegypti*, with special reference to the cool geographic range margins. *J Med Entomol*. 2014; 51(3):496–516. <https://doi.org/10.1603/ME13214> PMID: 24897844
5. Jetten TH, Focks DA. Potential changes in the distribution of dengue transmission under climate warming. *Am J Trop Med Hyg*. 1997; 57(3):285–97. <https://doi.org/10.4269/ajtmh.1997.57.285> PMID: 9311638
6. Bader CA, Williams CR. Mating, ovariole number and sperm production of the dengue vector mosquito *Aedes aegypti* (L.) in Australia: broad thermal optima provide the capacity for survival in a changing climate. *Physiol Entomol*. 2012; 37(2):136–44. <https://doi.org/10.1111/j.1365-3032.2011.00818.x>
7. Beebe NW, Cooper RD, Mottram P, Sweeney AW. Australia's dengue risk driven by human adaptation to climate change. *PLoS NTD*. 2009; 3(5):e429. <https://doi.org/10.1371/journal.pntd.0000429> PMID: 19415109
8. Trewin BJ, Kay BH, Darbro JM, Hurst TP. Increased container-breeding mosquito risk owing to drought-induced changes in water harvesting and storage in Brisbane, Australia. *Int Health*. 2013; 5(4):251–8. <https://doi.org/10.1093/inthealth/ihnt023> PMID: 24225151
9. Jansen CC, Beebe NW. The dengue vector *Aedes aegypti*: what comes next. *Microbes Infect*. 2010; 12(4):272–9. <https://doi.org/10.1016/j.micinf.2009.12.011> PMID: 20096802
10. Lumley GF, Taylor FH. *Dengue: School of Public Health and Tropical Medicine (University of Sydney)*; 1943.
11. Russell RC. Mosquito-borne arboviruses in Australia: the current scene and implications of climate change for human health. *Int J Parasitol*. 1998; 28(6):955–69. [https://doi.org/10.1016/S0020-7519\(98\)00053-8](https://doi.org/10.1016/S0020-7519(98)00053-8) PMID: 9673874
12. Trewin BJ, Darbro JM, Jansen CC, Schellhorn NA, Zalucki MP, Hurst TP, et al. The elimination of the dengue vector, *Aedes aegypti*, from Brisbane, Australia: The role of surveillance, larval habitat removal and policy. *PLoS NTD*. 2017; 11(8):e0005848. <https://doi.org/10.1371/journal.pntd.0005848> PMID: 28846682
13. Maréchal Y. *The hydrogen bond and the water molecule: The physics and chemistry of water, aqueous and bio-media*: Elsevier; 2006.
14. Carrington LB, Armijos MV, Lambrechts L, Barker CM, Scott TW. Effects of fluctuating daily temperatures at critical thermal extremes on *Aedes aegypti* life-history traits. *PLoS One*. 2013; 8(3):e58824. <https://doi.org/10.1371/journal.pone.0058824> PMID: 23520534
15. Colinet H, Sinclair BJ, Vernon P, Renault D. Insects in fluctuating thermal environments. *Ann Rev Entomol*. 2015; 60:123–40. <https://doi.org/10.1146/annurev-ento-010814-021017> PMID: 25341105
16. Allsopp PG, Daghli GJ, Taylor MFJ, Gregg PC, Zalucki MP. Measuring development of *Heliothis* species. *Heliothis: research methods and prospects*. 1991:90–101. https://doi.org/10.1007/978-1-4612-3016-8_8
17. Ruel JJ, Ayres MP. Jensen's inequality predicts effects of environmental variation. *Trends Ecol Evol*. 1999; 14(9):361–6. [https://doi.org/10.1016/S0169-5347\(99\)01664-X](https://doi.org/10.1016/S0169-5347(99)01664-X) PMID: 10441312
18. Bureau of Meteorology. *Climate Data Online: Bureau of Meteorology*; 2015 [Available from: <http://www.bom.gov.au/climate/data/>].

19. ESRI World Imagery, cartographer World Topographic Map2018. "Topographic" [basemap]. "World Topographic Map". November 7, 2018. <http://www.arcgis.com/home/item.html?id=30e5fe3149c34df1ba922e6f5bbf808f>. Accessed November 19, 2018.
20. Knox TB, Yen NT, Nam VS, Gattón ML, Kay BH, Ryan PA. Critical Evaluation of Quantitative Sampling Methods for *Aedes aegypti* (Diptera: Culicidae) Immatures in Water Storage Containers in Vietnam. *J Med Entomol.* 2007; 44(2):192–204. <https://doi.org/10.1093/jmedent/44.2.192> PMID: 17427686
21. Rasic G, Endersby NM, Williams C, Hoffmann AA. Using *Wolbachia*-based release for suppression of *Aedes* mosquitoes: insights from genetic data and population simulations. *Ecol Appl.* 2014; 24(5):1226–34. <https://doi.org/10.1890/13-1305.1> PMID: 25154109
22. Ulrich JN, Beier JC, Devine GJ, Hugo LE. Heat Sensitivity of wMel *Wolbachia* during *Aedes aegypti* Development. *PLoS NTD.* 2016; 10(7):e0004873. <https://doi.org/10.1371/journal.pntd.0004873> PMID: 27459519
23. Hugo LE, Kay BH, O'Neill SL, Ryan PA. Investigation of environmental influences on a transcriptional assay for the prediction of age of *Aedes aegypti* (Diptera: Culicidae) mosquitoes. *J Med Entomol.* 2010; 47(6):1044–52. <https://doi.org/10.1603/ME10030> PMID: 21175052
24. Pepe MS, Fleming TR. Weighted Kaplan-Meier statistics: a class of distance tests for censored survival data. *ISO 4.* 1989:497–507.
25. R Core Team. R: A language and environment for statistical computing.2018; (3.5.1). Available from: <http://www.R-project.org/>.
26. Richardson K, Hoffmann AA, Johnson P, Ritchie S, Kearney MR. Thermal Sensitivity of *Aedes aegypti* From Australia: Empirical Data and Prediction of Effects on Distribution. *J Med Ent.* 2011; 48(4):914–23. <https://doi.org/10.1603/me10204>
27. Logan JA, Wollkind DJ, Hoyt SC, Tanigoshi LK. An Analytic Model for Description of Temperature Dependent Rate Phenomena in Arthropods 1. *Enviro Entomol.* 1976; 5(6):1133–40. <https://doi.org/10.1093/ee/5.6.1133>
28. Sharpe PJ, DeMichele DW. Reaction kinetics of poikilotherm development. *J Theor Biol.* 1977; 64(4):649–70. [https://doi.org/10.1016/0022-5193\(77\)90265-X](https://doi.org/10.1016/0022-5193(77)90265-X) PMID: 846210
29. Focks DA, Daniels E, Haile DG, Keesling JE. A simulation model of the epidemiology of urban dengue fever: literature analysis, model development, preliminary validation, and samples of simulation results. *Am J Trop Med Hyg.* 1995; 53(5):489–506. <https://doi.org/10.4269/ajtmh.1995.53.489> PMID: 7485707
30. Tun-Lin W, Burkot TR, Kay BH. Effects of temperature and larval diet on development rates and survival of the dengue vector *Aedes aegypti* in north Queensland, Australia. *Med Vet Ent.* 2000; 14(1):31–7. <https://doi.org/10.1046/j.1365-2915.2000.00207.x>
31. Gilpin ME, McClelland GA. Systems analysis of the yellow fever mosquito *Aedes aegypti*. *Fortschr Zool.* 1979; 25(2–3):355. PMID: 535875
32. Rueda LM, Patel KJ, Axtell RC, Stinner RE. Temperature-Dependent Development and Survival Rates of *Culex quinquefasciatus* and *Aedes aegypti* (Diptera: Culicidae). *J Med Ent.* 1990; 27(5):892–8. <https://doi.org/10.1093/jmedent/27.5.892>
33. Kontodimas DC, Eliopoulos PA, Stathas GJ, Economou LP. Comparative Temperature-Dependent Development of *Nephus includens* (Kirsch) and *Nephus bisignatus* (Boheman) (Coleoptera: Coccinellidae) Preying on *Planococcus citri* (Risso) (Homoptera: Pseudococcidae): Evaluation of a Linear and Various Nonlinear Models Using Specific Criteria. *Enviro Entomol.* 2004; 33(1):1–11. <https://doi.org/10.1603/0046-225X-33.1.1>
34. Shi P, Ge F, Sun Y, Chen C. A simple model for describing the effect of temperature on insect developmental rate. *J Asia Pac Entomol.* 2011; 14(1):15–20. <https://doi.org/10.1016/j.aspen.2010.11.008>
35. Briere J-F, Pracros P, Le Roux A-Y, Pierre J-S. A Novel Rate Model of Temperature-Dependent Development for Arthropods. *Enviro Entomol.* 1999; 28(1):22–9. <https://doi.org/10.1093/ee/28.1.22>
36. Hilbert DW, Logan JA. Empirical Model of Nymphal Development for the Migratory Grasshopper, *Melanoplus sanguinipes* (Orthoptera: Acrididae) 1. *Enviro Entomol.* 1983; 12(1):1–5. <https://doi.org/10.1093/ee/12.1.1>
37. Cooling LE. Report on Mosquito Survey of the Brisbane Metropolitan Area, 1923. Australia Dept Health. 1923.
38. Hamlyn-Harris R. The elimination of *Aedes argenteus* Poiret as a factor in dengue control in Queensland: Liverpool School of Tropical Medicine; 1931.
39. Williams CR, Bader CA, Kearney MR, Ritchie SA, Russell RC. The extinction of dengue through natural vulnerability of its vectors. *PLoS NTD.* 2010; 4(12):e922. <https://doi.org/10.1371/journal.pntd.0000922> PMID: 21200424

40. Russell B, Kay B, Shipton W. Survival of *Aedes aegypti* (Diptera: Culicidae) eggs in surface and subterranean breeding sites during the northern Queensland dry season. *J Med Ent.* 2001; 38(3):441–5. <https://doi.org/10.1603/0022-2585-38.3.441>
41. Faull KJ, Williams CR. Intraspecific variation in desiccation survival time of *Aedes aegypti* (L.) mosquito eggs of Australian origin. *J Vect Ecol.* 2015; 40(2):292–300. <https://doi.org/10.1111/jvec.12167> PMID: 26611964
42. Endersby-Harshman NM. *Aedes aegypti* from Gin Gin, Queensland—potential for a successful suppression/extinction Wolbachia program based on population genetic data. 2014.
43. Montgomery BL, Ritchie SA. Roof gutters: a key container for *Aedes aegypti* and *Ochlerotatus notoscriptus* (Diptera: Culicidae) in Australia. *Am J Trop Med Hyg.* 2002; 67(3):244–6. <https://doi.org/10.4269/ajtmh.2002.67.244> PMID: 12408662
44. Focks D. A., Haile D. G., Daniels E., & Mount G. A. 1993. Dynamic life table model for *Aedes aegypti* (Diptera: Culicidae): analysis of the literature and model development. *Journal of medical entomology*, 30(6), 1003–1017. PMID: 8271242
45. Hancock P. A., Ritchie S. A., Koenraad C. J., Scott T. W., Hoffmann A. A., & Godfray H. C. J. (2018). Predicting the spatial dynamics of Wolbachia infections in *Aedes aegypti* arbovirus vector populations in heterogeneous landscapes. bioRxiv, 458794.