

RESEARCH ARTICLE

In Vitro Inhibition of NFAT5-Mediated Induction of CCL2 in Hyperosmotic Conditions by Cyclosporine and Dexamethasone on Human HeLa-Modified Conjunctiva-Derived Cells

Elise Warcoin^{1,2*}, Christophe Baudouin^{1,3}, Carole Gard⁴, Françoise Brignole-Baudouin^{1,5*}

1 Sorbonne Universités, UPMC Univ Paris 06, INSERM, CNRS, Institut de la Vision, Paris, France, **2** CHNO des Quinze-Vingts, Service Pharmacie, Paris, France, **3** CHNO des Quinze-Vingts, Service III, Paris, France, **4** Horus Pharma, Saint-Laurent du Var, France, **5** Faculté de Pharmacie de Paris, Univ Paris Descartes, Sorbonne Paris Cité, Paris, France

* elisewarcoin@gmail.com (EW); francoise.brignole-baudouin@parisdescartes.fr (FB)



OPEN ACCESS

Citation: Warcoin E, Baudouin C, Gard C, Brignole-Baudouin F (2016) In Vitro Inhibition of NFAT5-Mediated Induction of CCL2 in Hyperosmotic Conditions by Cyclosporine and Dexamethasone on Human HeLa-Modified Conjunctiva-Derived Cells. PLoS ONE 11(8): e0159983. doi:10.1371/journal.pone.0159983

Editor: Michele Madigan, Save Sight Institute, AUSTRALIA

Received: November 29, 2015

Accepted: July 12, 2016

Published: August 3, 2016

Copyright: © 2016 Warcoin et al. This is an open access article distributed under the terms of the [Creative Commons Attribution License](https://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Data Availability Statement: All relevant data are within the paper and its Supporting Information files.

Funding: Carole Gard is employed by Horus Pharma. Concerning her participation, as a co-author, we had regular scientific meetings with her and she participated in the design of the methods, as well as in the manuscript review. She also contributed to reagents, material and analysis tools. Horus Pharma provided support in the form of salary for author CG, but did not have any additional role in the study design, data collection and analysis, decision to

Abstract

Purpose

To investigate the pro-inflammatory intracellular mechanisms induced by an in vitro model of dry eye disease (DED) on a HeLa-modified conjunctiva-derived cells in hyperosmolarity (HO) stress conditions. This study focused on CCL2 induction and explored the implications of the nuclear factor of activated T-cells 5 (NFAT5) as well as mitogen-activated protein kinases (MAPK) and nuclear factor kappa B (NFκB). This work was completed by an analysis of the effects of cyclosporine A (CsA), dexamethasone (Dex) and doxycycline (Dox) on HO-induced CCL2 and NFAT5 induction.

Methods

A human HeLa-modified conjunctiva-derived cell line was cultured in NaCl-hyperosmolar medium for various exposure times. Cellular viability, CCL2 secretion, NFAT5 and CCL2 gene expression, and intracytoplasmic NFAT5 were assessed using the Cell Titer Blue[®] assay, enzyme-linked immunosorbent assay (ELISA), RT-qPCR and immunostaining, respectively. In selected experiments, inhibitors of MAPKs or NFκB, therapeutic agents or NFAT5 siRNAs were added before the hyperosmolar stimulations.

Results

HO induced CCL2 secretion and expression as well as NFAT5 gene expression and translocation. Adding NFAT5-siRNA before hyperosmolar stimulation led to a complete inhibition of CCL2 induction and to a decrease in cellular viability. p38 MAPK (p38), c-Jun NH₂-terminal kinase (JNK) and NFκB inhibitors, CsA and Dex induced a partial inhibition of HO-

publish, or preparation of the manuscript. The specific role of this author is articulated in the 'author contributions' section.

Competing Interests: This work was funded by an unrestricted grant from Horus Pharma. Carole Gard is employed by Horus Pharma. Christophe Baudouin: Financial support and consultant (Alcon; Allergan; Santen; Laboratoires Théa). There are no patents, products in development or marketed products to declare. This does not alter the authors' adherence to all the PLOS ONE policies on sharing data and materials.

induced CCL2, while Dox and extracellular signal-regulated kinase (ERK) inhibitor did not. Dex also induced a partial inhibition of HO-induced NFAT5 gene expression but not CsA or Dox.

Conclusions

These *in vitro* results suggest a potential role of CCL2 in DED and highlight the crucial role of NFAT5 in the pro-inflammatory effect of HO on HeLa-modified conjunctiva-derived cells, a rarely studied cellular type. This inflammatory pathway involving NFAT5 and CCL2 could offer a promising target for developing new therapies to treat DED, warranting further investigations to fully grasp the complete intracellular mechanisms.

Introduction

DED) is one of the most common ocular pathologies in the world, with a prevalence of 3–15% [1] in patients over the age of 50, although it is often underestimated because of its apparent harmlessness. However, patients with severe dry eye syndrome suffer from constant eye irritation symptoms as well as blurred and fluctuating vision [2,3] which can complicate daily tasks [4] and may in turn lead to anxiety and even depression [5]. DED is due to a dysfunction of the lachrymal functional unit resulting in decreased tear secretion and/or excessive evaporation of the aqueous tear phase. These effects then lead to an increase in tear film osmolarity, tear film instability and ultimately damages the ocular surface [6]. Tear HO and ocular surface inflammation are currently considered as the two key mechanisms underlying DED that maintain the vicious circle of the pathology on the ocular surface [1,7–9]. Clinical studies on dry eye patients reported an increase in pro-inflammatory cytokines and chemokines in tears and conjunctival cells such as interleukin (IL) -6, IL-8, TNF- α and IL-1 β ; a loss in conjunctival goblet cells [10,11]; and an increase in immune activation and infiltration in the conjunctiva [12–17]. To help understand the pathogenesis of DED, hyperosmolar conditions are often used because they reproduce the environment in contact with the ocular surface in the pathology. These experiments have shown that HO was responsible for ocular surface cell death [18,19], reactive oxygen species formation [20,21], activation of MAPKs such as p38, JNK and ERK [22–24] and increases in production of matrix metalloproteinases (MMP) [22], and pro-inflammatory cytokines such as IL-1 β , TNF- α , IL-8, IL-6 and CCL2 [25–30].

The molecular mechanism that regulates the transcription and secretion of these pro-inflammatory actors under hyperosmolar conditions is poorly understood. Among the actors involved, CCL2, a potent chemoattractant protein that attracts monocytes to the inflammation site [31], and its receptor CCR2 have been identified as potentially important actors in DED. Indeed, Goyal et al. discovered that a topical antagonist of CCR2 improved dry eye symptoms in *in vivo* experiments [32]. On other cell types such as renal tubular epithelial cells and peritoneal mesothelial cells, the induction of pro-inflammatory cytokines such as CCL2 by osmotic stress has been observed to depend on the NFAT5 transcription factor, also called the tonicity response element-binding protein (TonEBP) [33,34]. HO is already known to induce NFAT5 translocation to promote cellular adaptation and survival from hypertonic stress [35]. Its activation in response to HO most particularly regulates the transcription of target genes that lead to the accumulation of compatible osmolytes inside cells, promoting better protection. Concerning ocular studies, a study on human limbal cells showed that IL-1 β and TNF- α are induced in NaCl-induced hyperosmolar conditions via NFAT5 activation [36]. These

accumulated observations suggest that NFAT5 could play a preponderant role in the inflammation induced by hypertonic challenge. Except for the study on cell death [18], no work has been specifically dedicated to the epithelial conjunctival cell inflammatory responses. However, conjunctival cells collected from dry eye patients express MHC class II antigen, HLA-DR, ICAM-1 and CCR5, confirming their involvement in the pathological process [15,37–39] and conjunctival inflammation was also reported in several *in vivo* models of the pathology [40,41].

Topical CsA is actually the only drug specifically formulated to treat dry eye syndrome and approved by regulatory medical agencies (FDA, EMA). Restasis[®] (Allergan) and Ikervis[®] (Santen) provide an alternative to artificial tears, Dex and Dox. CsA has immunosuppressive and anti-inflammatory effects, but its exact therapeutic mechanisms on the ocular surface of dry eye patients remains unclear.

The number of patients suffering from DED is set to grow as the world population ages. A better understanding of the pathophysiology of DED and the mechanism of CsA could help find new targets for treating this pathology and relieve millions of patients from permanent and painful discomfort.

Therefore, our aim was to analyze and characterize *in vitro* conjunctival CCL2 induction in a hyperosmolar model of dry eye and to determine the relationship between NFAT5 and CCL2 on HeLa-modified conjunctiva-derived cells. Moreover, we investigated intracellular signaling pathways such as MAPKs (p38, JNK and ERK) and the NF κ B transcription factor. We also investigated the effects of the three major anti-inflammatory therapeutic agents: CsA, Dex and Dox on CCL2 and NFAT5 induction by HO (Fig 1). These molecules are used in DED patients as an acute or chronic treatment, and understanding their anti-inflammatory mechanism on the desiccated ocular surface would help develop new targeted therapies.

Materials and Methods

Cell Line

The Wong Kilbourne derivative of the Chang (WKD) HeLa-modified conjunctiva-derived epithelial cell line (clone 1-5c-4, American Type Culture Collection [ATCC] certified cell line [CCL], 20.2) was cultured under classic conditions (moist atmosphere, 5% CO₂, 37°C) in Dulbecco minimum essential medium supplemented with 10% fetal bovine serum, 1% glutamine (200 mM), 1% penicillin (10,000 units/mL) and streptomycin (10,000 μ g/mL) for 24 h to reach either 80% confluence before hyperosmolar stimulations and to reach 50% confluence before transfection for siRNA experiments. All reagents for cellular culture were purchased from Gibco (Gibco, Life technologies, Carlsbad, CA, USA). This cell line has previously been used for toxicological *in vitro* studies and was shown to respond similarly to the IOBA conjunctival cell line [42]. Despite the criticism on this cell line, immortalized by contact with cervical cancer HeLa cells, we decided to use it after having confirmed the conjunctival phenotype by a cytokeratin 13 flow cytometry analysis [43] and because it does not require supplementation of hydrocortisone. Moreover, a study comparing gene expression profiles of conjunctival cell lines, including WKD cells, with primary cultured conjunctival epithelial cells and human conjunctival tissue, explained that these cell lines may still be used to investigate some specific cellular mechanisms like those related to inflammation, as human tissues and primary conjunctival cells are often difficult to obtain [44].

Hyperosmolar Condition Preparation and Control

Each hyperosmolar medium was prepared by adding sodium chloride (Sigma-Aldrich, Saint Louis, MO, USA) to supplemented medium. All osmolarity values were assessed using an osmometer (Roebing 13DR, Berlin, Germany) including the supplemented medium, which was found at 340 mOsM (mosm.L⁻¹) as expected from information provided by the

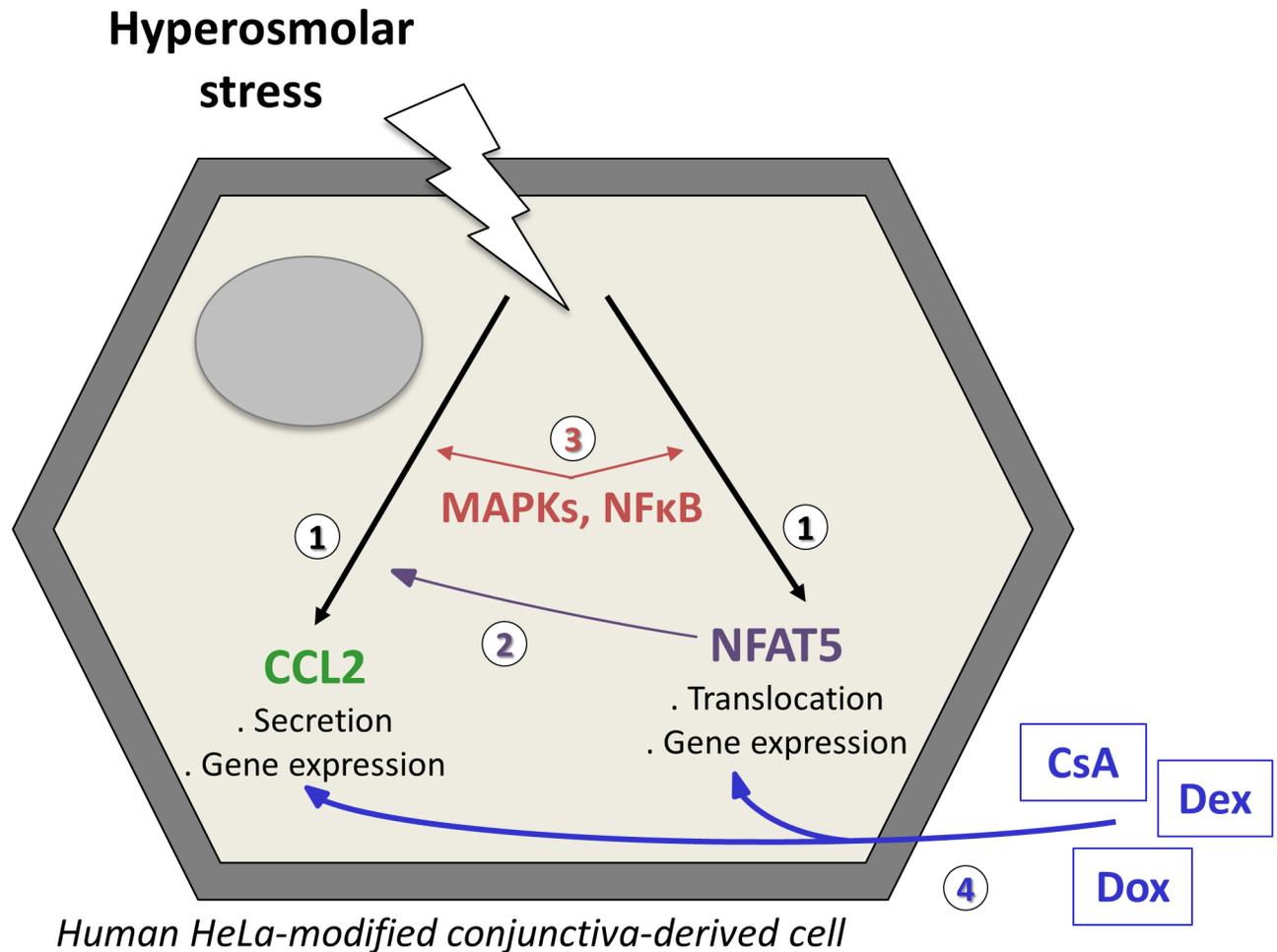


Fig 1. Schematic figure summarizing the hypotheses of the signaling mechanisms investigated in HeLa-modified conjunctiva-derived cells exposed to hyperosmolar conditions. 1) Is HO able to stimulate CCL2 or NFAT5? 2) Is NFAT5 involved in the CCL2 stimulation? 3) Are MAPKs or NFκB implicated in the CCL2 and NFAT5 stimulations? 4) What are the effects of CsA, Dex or Dox on these stimulations?

doi:10.1371/journal.pone.0159983.g001

manufacturer. We decided not to change this osmolarity level in order to keep the cell line in its regular medium even if this value is above the expected values of normal tear film, i.e., 302.2 ±8.3 mOsM, and in order to avoid any hypo-osmotic regulatory effects. In dry eye subjects, tear osmolarity is generally found at 336.4±22.3 mOsM [45] but some studies have reported higher tear hyperosmolarities that can reach 440 mosM [46,47] and spikes of 800–900 mOsM are thought to occur over the central cornea during tear film instability [8].

Cell Treatments

For experiments that analyzed a range of hyperosmolar conditions, cells were treated with supplemented medium (340 mOsM) or with hyperosmolar medium ranging from 400 mOsM to 600 mOsM for 4 h before collecting cell lysates for real-time quantitative PCR (RT-qPCR), or 24 h before collecting supernatants for ELISA, performing the cell viability test or fixing cells for immunostaining. For the time-course experiments, cells were treated with medium (340 mOsM) or with hyperosmolar medium (500 mOsM) for durations ranging from 30 min to 24 h before collecting cell lysates for RT-qPCR, supernatants for ELISA or performing the cell viability test.

This kind of protocol using a high osmolarity such as 500 mOsM during long duration as 24h on ocular surface cells is a common protocol often reported to study DED [19,36,48].

For the siRNA experiments, cells were treated with a mix of lipofectamine (Life Technologies, Carlsbad, CA, USA) and negative control siRNA (Life Technologies) or NFAT5 siRNA (Life Technologies) for 24 h, as recommended by the manufacturer. After this period of transfection, the cell medium was replaced by supplemented medium (340 mOsM) or hyperosmolar medium (500 mOsM) for 4 h before collecting cell lysates for RT-qPCR or 24 h before collecting supernatants for ELISA or performing the cell viability test.

For experiments testing the effects of therapeutic agents or cellular pathway inhibitors, cells were treated for 1 h with various dilutions of therapeutic agents. CsA was tested at 0.1, 1 and 10 $\mu\text{g}/\text{mL}$; the highest concentration of 10 $\mu\text{g}/\text{mL}$ corresponding to 1/100 and 1/50 of Ikervis[®] and Restasis[®], respectively. Dex was tested at 10^{-10} , 10^{-8} and 10^{-6}M and Dox at 10 $\mu\text{g}/\text{mL}$. A mix of CsA 10 $\mu\text{g}/\text{mL}$ and Dex 10^{-8}M was also tested. The inhibitors of p38 (SB203580), JNK (SP600125) and MEK/ERK (UO126) (inhibitor of MEK1 and hence of its downstream target ERK) were tested at 10 μM while the NF κ B inhibitor (PDTC) was tested at 50 μM . The concentrations tested were chosen regarding previous published studies [22,49,50] and after confirming their innocuity on HeLa-modified conjunctiva-derived cells with a cellular viability test, the CellTiter-Blue[®] assay (Promega, Madison, WI, USA). Therapeutic agents and inhibitors were all purchased from Sigma-Aldrich and were reconstituted with dimethylsulfoxide (DMSO) (Sigma-Aldrich) except for Dex that was reconstituted in water. They all were first diluted in DMSO and then in supplemented medium to achieve a final concentration of DMSO equal to 0.1%. The control condition for these experiments was DMSO 0.1% in supplemented medium. After a 1-h incubation, supplemented medium or hyperosmolar medium at 980 mOsM was added to induce a dilution of $\frac{3}{4}$ of the existing medium, leading to a final osmolarity of 340 mOsM and 500 mOsM, respectively. These conditions were applied for 4 h before collecting cell lysates for RT-qPCR, or 24 h before collecting supernatants for ELISA or performing the cell viability test. Pre-treating cells with therapeutic molecules or inhibitors before adding hyperosmolar condition is a classic protocol already reported on ocular surface cells [22,23,51].

For all these culture assays, we used one well for each condition concerning immunostaining (6-wells plate) or RTqPCR assays (24-wells plate) and at least two wells for ELISA and viability test (96-wells plate).

Cell Viability Test

Cellular viability was analyzed using resazurin dye to measure the metabolic capacity of cells, which is an indicator of cell viability, using the CellTiter-Blue[®] assay (Promega). After an incubation period under the above-described stress conditions, CellTiter-Blue[®] reagent was added to the cell culture following the protocol described by the manufacturer. Fluorescence intensity was then quantified using a microplate reader with an excitation wavelength of 560 nm and an emission wavelength of 590 nm (Infinite M1000, Tecan, Lyon, France). Fluorescence values were then normalized with respect to control cells considered as 100% viable.

Enzyme-Linked Immunosorbent Assay

Enzyme-linked immunosorbent assays (ELISAs) for human CCL-2 were performed using the commercial DuoSet ELISA Development kit (R&D Systems, Minneapolis, MN, USA). After the incubation period under different stress conditions, cellular supernatants were collected, centrifuged to remove potential cellular fragments, and were stored at -80°C . ELISAs were performed on supernatants according to the manufacturer's protocols. Absorbance was read at 450 nm using a microplate reader (Infinite M1000, Tecan) with a reference wavelength of 570 nm

Immunostaining

Cells were grown on round sterile cover glasses (diameter, 14 mm; Menzel GmbH, Braunschweig, Germany). After the incubation period under different stress conditions, the cells were washed and fixed in 4% paraformaldehyde-PBS (Sigma-Aldrich). They were then permeabilized in a 0.3% Triton (Triton X-100, Sigma-Aldrich) solution for intracellular staining, followed by a 1% bovine serum albumin (Calbiochem, Merck Millipore, Darmstadt, Germany) incubation for 30 min and an overnight period with the primary antibody anti-NFAT5 at a final concentration of $2 \mu\text{g}\cdot\text{mL}^{-1}$ (goat polyclonal antibody SC5499; Santa Cruz Biotechnology, Santa Cruz, CA, USA). Cells were then incubated for 1 h with the secondary antibody Alexa Fluor 488 rabbit anti-goat (Invitrogen, Life Technologies). Cover glasses were then mounted with Mountant PermaFluor[®] (Thermo Fisher Scientific, Courtaboeuf, France) before observation with an epifluorescence microscope (Leica DM6000B, Rueil-Malmaison, France).

RNA Extraction, Reverse Transcription, and Quantitative Real-Time PCR

After the incubation period under different stress conditions, the cells were washed and lysed, and their total RNA was extracted using a NucleoSpin RNA II extraction kit (Macherey-Nagel, Düren, Germany). RNA content was measured using a NanoDrop detector (ND-1000 spectrophotometer) and cDNA was synthesized from equal amounts of RNA using Multiscribe reverse transcriptase (TaqMan Reverse Transcription Reagents, Applied Biosystems, Life Technologies). Concentrations of each sample were adjusted to 5 ng/ μL of cDNA. The reaction mixture containing 25 ng of cDNA per well was preheated at 95°C for 10 min, followed by 40 cycles (95°C/15 s and 60°C/1 min). Each assay was normalized by amplifying the housekeeping cDNA GAPDH (ID Hs9999905). Target cDNA was amplified using the 7300 Real-Time PCR system (Applied Biosystems, Life Technologies) with assays-on-demand primers for human CCL2 (Hs00234140) and NFAT5 (Hs00232437) (Applied Biosystems, Life Technologies). Changes in mRNA expression were calculated according to the $2^{-\Delta\Delta\text{CT}}$ method (CT, cycle threshold), with $\Delta\text{CT} = \text{CT}_{\text{target gene}} - \text{CT}_{\text{gapdh}}$ and $\Delta\Delta\text{CT} = \Delta\text{CT}_{\text{stimulated}} - \Delta\text{CT}_{\text{control}}$.

Statistical Analyses

All experiments were performed at least three times: immunostainings and RTqPCR assays were performed on one well per condition, in three separated experiments. Viability tests and ELISA were performed at least in two wells per condition, in three separated experiments. Their conditions were compared using one-way analysis of variance (ANOVA) followed by Dunnett's multiple comparison test, or two-way ANOVA followed by Sidak's or Tukey's multiple comparison test (GraphPad, GraphPad Software, La Jolla, CA, USA). Only p -values < 0.05 were considered statistically significant.

Results

Hyperosmolar conditions, from 450 or 500 mOsM, Induced Cell Death in an Osmo-Dependent Manner, CCL2 Secretion and Gene Expression, and NFAT5 Gene Expression and Translocation in HeLa-Modified Conjunctiva-Derived Cells (Fig 2)

Cell death. After a 24-h treatment, conjunctival cellular viability significantly decreased in the 500-mOsM hyperosmolar condition, with 16% of cell viability loss compared to control ($p \leq 0.001$) (Fig 2A). Higher hyperosmolar conditions, 550 and 600 mOsM, decreased viability even further, to values of 71 and 52%, respectively ($p \leq 0.0001$).

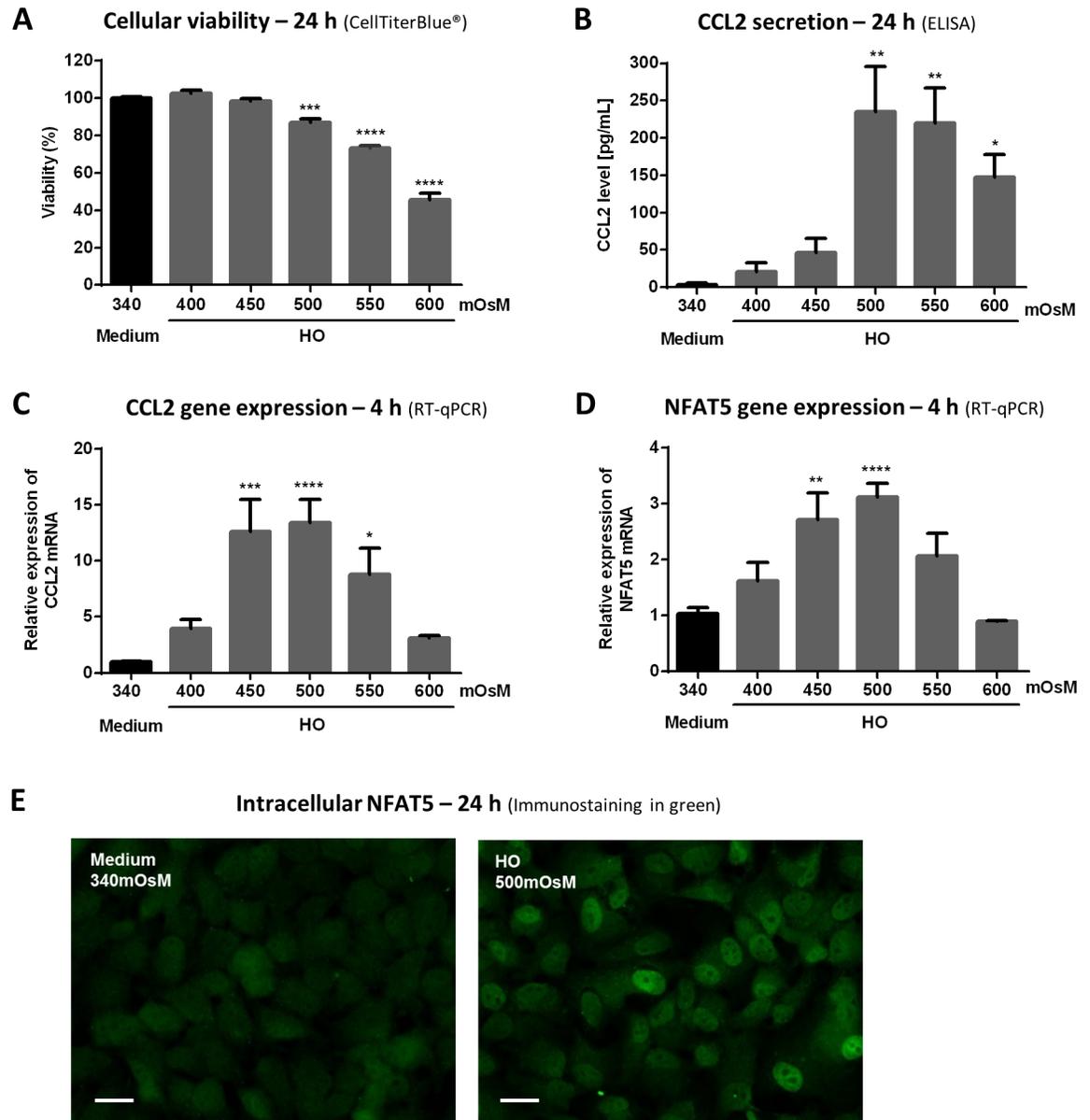


Fig 2. Hyperosmolar conditions, from 450 or 500 mOsM, induced cell death in an osmo-dependent manner, CCL2 secretion and gene expression, and NFAT5 gene expression and translocation in HeLa-modified conjunctiva-derived cells. Cells exposed to supplemented medium (340 mOsM) or different hyperosmolar conditions from 400 to 600 mOsM for 24 h were tested for cellular viability (A) with CellTiterBlue[®] assay and CCL2 secretion (B) by ELISA, and cells treated for 4 h were lysed for mRNA expression by RT-qPCR to determine the CCL2 and NFAT5 gene expression (C, D). Data show mean \pm SEM, $n = 3$ or more. Statistical analysis: one-way ANOVA followed by Dunnett's multiple comparison test. * $p \leq 0.05$; ** $p \leq 0.01$; *** $p \leq 0.001$; **** $p \leq 0.0001$, compared to 340 mOsM. Cells exposed to supplemented medium (340 mOsM) or hyperosmolar medium (500 mOsM) for 24 h were analyzed for NFAT5 translocation with NFAT5 immunofluorescent staining (E) in green. White scale bar: 20 μ m.

doi:10.1371/journal.pone.0159983.g002

CCL2 secretion and gene expression. Hyperosmolar conditions (500, 550 or 600 mOsM) applied for 24 h induced a significant secretion of CCL2 in supernatants by HeLa-modified conjunctiva-derived cells (235, 220, 148 pg/mL, respectively) ($p \leq 0.01$ or $p \leq 0.05$ compared to control cells at 3 pg/mL of CCL2 in supernatants), but no effect was detected for 400- and 450-mOsM conditions relative to the control osmolarity (Fig 2B).

CCL2 mRNA expression significantly increased in HO 450 mOsM, reaching a relative expression of 12.6 compared to the 340-mOsM control after 4 h ($p \leq 0.001$) (Fig 2C). This relative fold expression was also significantly higher than the control, in 500- and 550-mOsM conditions (13.4 and 8.8, respectively). Neither the highest (600 mOsM) nor the lowest (400 mOsM) HO tested modified CCL2 mRNA expression compared to the control medium.

NFAT5 gene expression and nuclear translocation. Four hours in hypertonic conditions at 450 and 500 mOsM induced an increase in NFAT5 gene expression, 2.7- and 3.1-fold respectively, compared to the 340-mOsM condition ($p \leq 0.01$ and $p \leq 0.0001$, respectively) (Fig 2D). The immunostaining of NFAT5 showed the presence of NFAT5 both in the cytoplasm and the nuclei for control cells in 340 mOsM with a fine diffuse fluorescent staining (Fig 2E). Cells in hyperosmotic conditions of 500 mOsM for 24 h displayed an intense fluorescent staining in the nuclei, indicating the nuclear translocation of this transcription factor.

Hyperosmolar Condition of 500 mOsM Induced Cell Death, Secretion and mRNA Expression of CCL2, and NFAT5 Gene Expression in HeLa-Modified Conjunctiva-Derived Cells in a Time-Dependent Manner (Fig 3)

Cell death. HeLa-modified conjunctiva-derived cells in a 500-mOsM medium started to significantly decrease their viability only after 9 h of stress (88% viability) (Fig 3A). The viability continued to decrease until 24 h of stress.

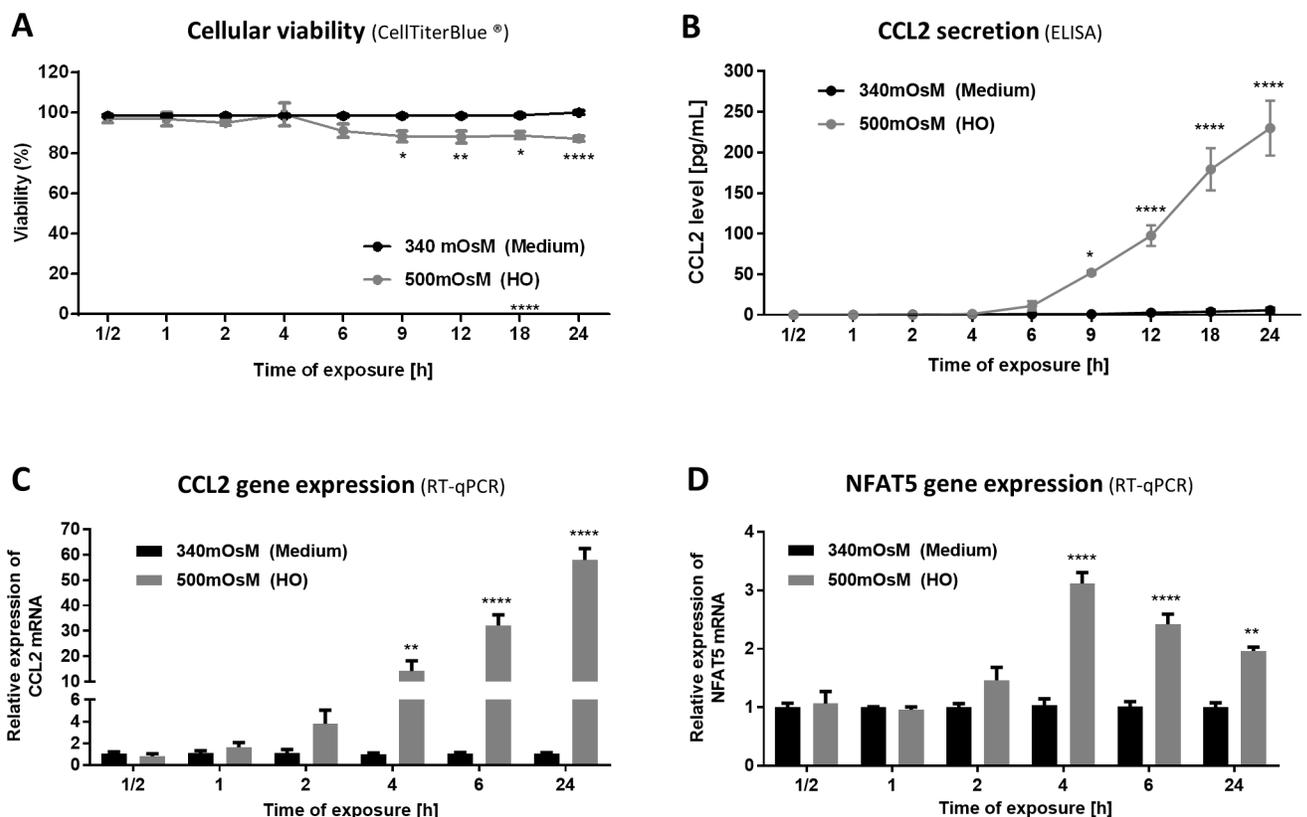


Fig 3. Hyperosmolar condition of 500 mOsM induced cell death, secretion and gene expression of CCL2 and NFAT5 gene expression in HeLa-modified conjunctiva-derived cells in a time-dependent manner. Cells exposed to supplemented medium (340 mOsM) or the hyperosmolar condition (500 mOsM) for different times from 30 min to 24 h were tested for cellular viability (A) with CellTiterBlue[®] assay, CCL2 secretion (B) with ELISA and were lysed for mRNA expression using RT-qPCR to determine CCL2 and NFAT5 gene expression (C, D). Data showing mean \pm SEM, $n = 3$ or more. Statistical analysis: two-way ANOVA followed by Sidak's multiple comparison test. * $p \leq 0.05$; ** $p \leq 0.01$; *** $p \leq 0.001$; **** $p \leq 0.0001$, compared to 340 mOsM at the same time.

doi:10.1371/journal.pone.0159983.g003

CCL2 secretion and gene expression. Similarly, CCL2 secretion significantly increased after 9 h in 500 mOsM (1 pg/mL vs 52 pg/mL) ($p \leq 0.05$) (Fig 3B). CCL2 secretion increased further to concentrations of 98 pg/mL, 180 pg/mL and 235 mg/mL after 12, 18 and 24 h of hyperosmolar treatment ($p \leq 0.0001$ compared to control cells).

CCL2 gene expression was also enhanced by the 500-mOsM medium with CCL2 mRNA expression increasing 14-fold compared to the 340-mOsM control after only 4 h in hyperosmolar conditions (Fig 3C). The mRNA expression increased 32-fold after 6 h and 58-fold after 24 h ($p \leq 0.0001$ for both conditions).

NFAT5 gene expression. NFAT5 gene expression increased 3.1-fold compared to control cells after 4 h in hyperosmolar conditions ($p \leq 0.0001$). NFAT5 mRNA expression significantly increased 2.4-fold and 2.0-fold following 6 and 24 h of treatment, respectively (Fig 3D).

NFAT5 siRNA Increased HO-Induced Cell Death and Inhibited HO-Induced CCL2 Secretion and Gene Expression in HeLa-Modified Conjunctiva-Derived Cells (Fig 4)

Cell death. The cell viability test did not reveal any toxic effect when using NFAT5 siRNA versus control siRNA in basal conditions of 340 mOsM (Fig 4A). However, in 500-mOsM hyperosmolar medium, the viability of cells treated with NFAT5 siRNA was significantly lower than cells treated with control siRNA (64% versus 79%).

CCL2 secretion and gene expression. Concerning the CCL2 secretion in 340 mOsM, control and NFAT5 siRNAs did not induce significantly different effects, with a low level of CCL2 in supernatant (37 and 16 pg/mL, respectively) (Fig 4B). In hyperosmolar conditions, adding NFAT5 siRNA induced a complete inhibition of HO-induced CCL2 secretion: HO induced a CCL2 concentration of 356 pg/mL in supernatant of cells treated with control siRNA and CCL2 concentration reached 32 pg/mL with NFAT5 siRNA ($p \leq 0.0001$). This low concentration did not significantly differ from that observed with NFAT5 siRNA in 340-mOsM conditions equal to 37 pg/mL (statistical analysis not shown).

The same inhibition profile was observed with CCL2 gene expression. The CCL2 mRNA expression with control siRNA increased 14.3-fold in 500-mOsM conditions compared to 340-mOsM conditions (Fig 4C). Treating cells with NFAT5 siRNA reduced this CCL2 mRNA expression to a value similar to those observed in basal conditions (0.4 vs 0.5).

NFAT5 gene expression. The specificity of the NFAT5 siRNA used was confirmed by RT-qPCR of NFAT5 (Fig 4D). Treating cells with NFAT5 siRNA significantly reduced NFAT5 mRNA expression compared to cells treated with control siRNA (0.3 vs 1.0 for 340 mOsM and 0.7 vs 2.0 for 500 mOsM).

The lack of toxicity of siRNAs was also verified using the CellTiter-Blue viability test. It showed that control siRNA and NFAT5 siRNA had no effect on the viability of HeLa-modified conjunctiva-derived cells compared to cells without siRNA in classic medium (data not shown).

Effects of a p38 Inhibitor (SB203580), a JNK Inhibitor (SP600125), a MEK/ERK Inhibitor (U0126) and a NF κ B Inhibitor (PDTC) on HO-Induced CCL2 Secretion, and CCL2 and NFAT5 Gene Expression in HeLa-Modified Conjunctiva-Derived Cells (Fig 5)

CCL2 secretion and gene expression. In hyperosmolar 500-mOsM conditions for 24 h, the control DMSO 0.1% induced CCL2 secretion in supernatants (257 pg/mL) (Fig 5A). This secretion significantly decreased for cells pretreated with the inhibitors SB203580, SP600125 and PDTC (162, 89, 130 pg/mL, respectively; $p \leq 0.0001$ for the three conditions). Treating cells with U0126 had no effect on this secretion.

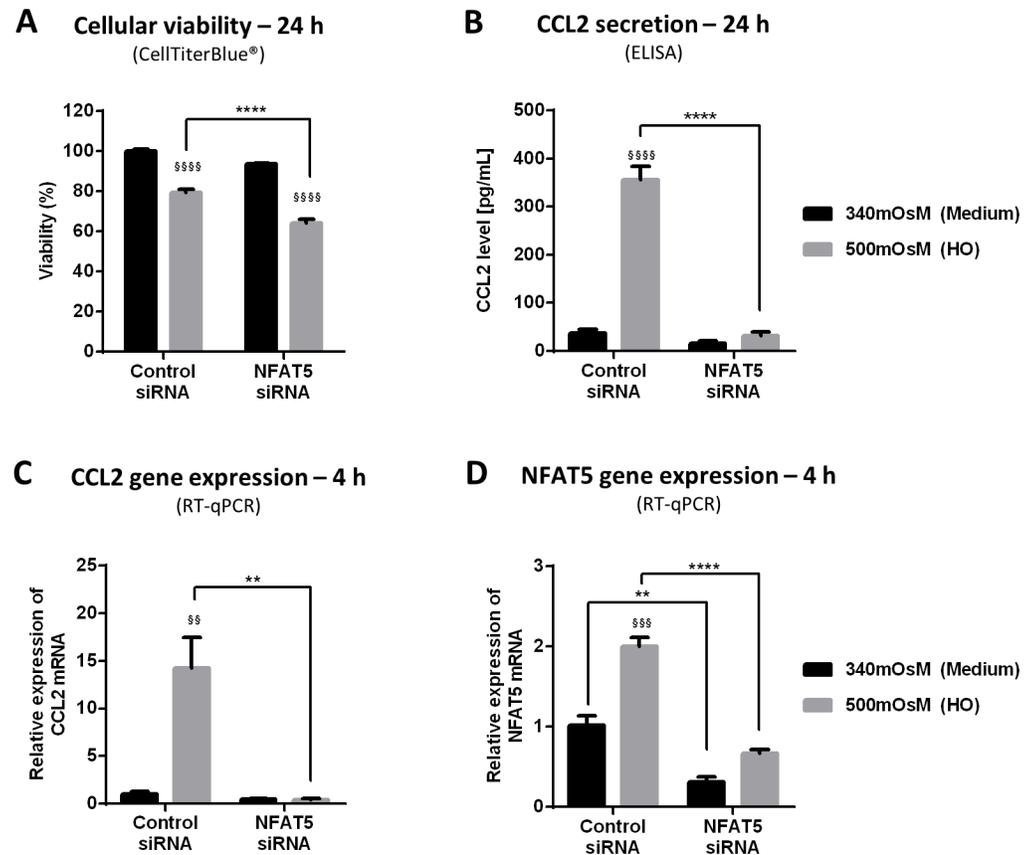


Fig 4. NFAT5 siRNA increased HO-induced cell death and inhibited HO-induced CCL2 secretion and CCL2 and NFAT5 gene expression in HeLa-modified conjunctiva-derived cells. Cells treated for 24 h with negative control siRNA or NFAT5 siRNA were then exposed to supplemented medium (340 mOsM) or hyperosmolar condition (500 mOsM). Cells exposed for 24 h were tested for cellular viability (A) with CellTiterBlue[®] assay, CCL2 secretion (B) by ELISA and cells exposed for 4 h were lysed for mRNA expression using RT-qPCR to determine the CCL2 and NFAT5 gene expression (C, D). Data showing mean \pm SEM, $n = 3$ or more. Statistical analysis: two-way ANOVA followed by Tukey's multiple comparison test. $\$ p \leq 0.05$; $\$\$ p \leq 0.01$; $\$\$\$ p \leq 0.001$; $\$\$\$\$ p \leq 0.0001$, compared to respective 340 mOsM. $* p \leq 0.05$; $** p \leq 0.01$; $*** p \leq 0.001$; $**** p \leq 0.0001$, compared to respective control siRNA.

doi:10.1371/journal.pone.0159983.g004

The same CCL2 mRNA inhibition profile was observed in hyperosmolar conditions following addition of the inhibitors SB203580, SP600125 and PDTC with a 7.4-, 11.3- and 3.5-fold mRNA increase, respectively, compared to the 18.4-fold increase for DMSO (Fig 5B). As observed during CCL2 secretion assays, U0126 had no effect on HO-induced CCL2 mRNA expression.

Under 340-mOsM basal conditions, the compounds tested had no effect on either the production of CCL2 by HeLa-modified conjunctiva-derived cells or its gene expression.

NFAT5 gene expression. Treating cells with PDTC increased NFAT5 gene expression 1.9-fold under basal 340-mOsM conditions (Fig 5C). A 3.0-fold increase in NFAT5 expression was recorded under hyperosmolar conditions containing DMSO. NFAT5 expression in 500-mOsM conditions increased more moderately when SB203580 and PDTC were added to medium (2.1- and 1.7-fold increase, respectively) contrary to SP600125 and U0126, which had no effect at all.

None of the compounds tested had a negative effect on cellular viability compared to their respective control conditions: DMSO 0.1% in 340 mOsM or in 500 mOsM (data not shown).

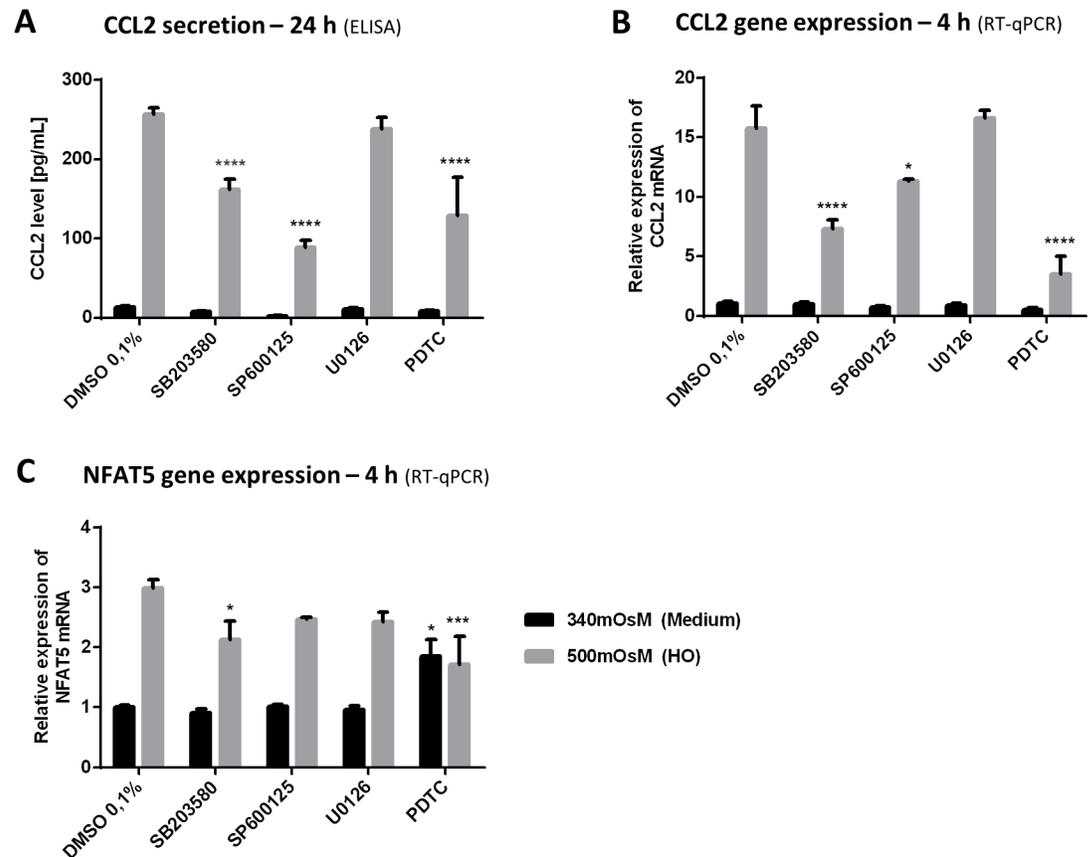


Fig 5. Effects of a p38 inhibitor (SB203580), a JNK inhibitor (SP600125), a MEK/ERK inhibitor (U0126) and a NFκB inhibitor (PDTC) on HO-induced CCL2 secretion and CCL2 and NFAT5 gene expression in HeLa-modified conjunctiva-derived cells. Cells were treated for 1 h with DMSO 0.1%, SB203580 10 μM, SP600125 10 μM, U0126 10 μM or PDTC 50 μM before adding supplemented medium 340 mOsM or hyperosmolar medium to reach 500 mOsM. Cells exposed to medium or HO for 24 h were tested for CCL2 secretion (A) by ELISA and cells exposed for 4 h were lysed for mRNA expression using RT-qPCR to determine the CCL2 and NFAT5 gene expression (B, C). Data showing mean ± SEM, $n = 3$ or more. Statistical analysis: two-way ANOVA followed by Tukey's multiple comparison test. * $p \leq 0.05$; ** $p \leq 0.01$; *** $p \leq 0.001$; **** $p \leq 0.0001$, compared to respective DMSO 0.1%.

doi:10.1371/journal.pone.0159983.g005

Effects of CsA, Dex and Dox on HO-Induced CCL2 Secretion as well as CCL2 and NFAT5 Gene Expressions in HeLa-Modified Conjunctiva-Derived Cells (Fig 6)

CCL2 secretion and gene expression. After 24 h in 500-mOsM conditions, CCL2 secretion with control DMSO 0.1% reached 250 pg/mL, whereas after 4 h, CCL2 mRNA expression increased 12.2-fold (Fig 6A and 6B). Both these effects were significantly inhibited in a concentration-dependent way by CsA and Dex. CsA 1 and 10 μg/mL decreased CCL2 levels to 193 and 144 pg/mL, respectively ($p \leq 0.0001$ compared to DMSO), and lowered the increase in CCL2 expression to 10.0-fold and 8.6-fold, respectively. The lowest dose tested, 0.1 μg/mL, had no observable impact on CCL2 levels. The lowest dose of Dex tested (10^{-10} M) also had no effect on CCL2 induction, but Dex 10^{-8} and 10^{-6} M had a significant inhibitory effect on CCL2 secretion (145 and 114 pg/mL, respectively, $p \leq 0.0001$ compared to DMSO) and on CCL2 mRNA expression (increasing it 6.7-fold and 5.7-fold, respectively, $p \leq 0.0001$ compared to DMSO) in 500-mOsM conditions. In contrast, Dox had no inhibitory effect on CCL2 secretion induced

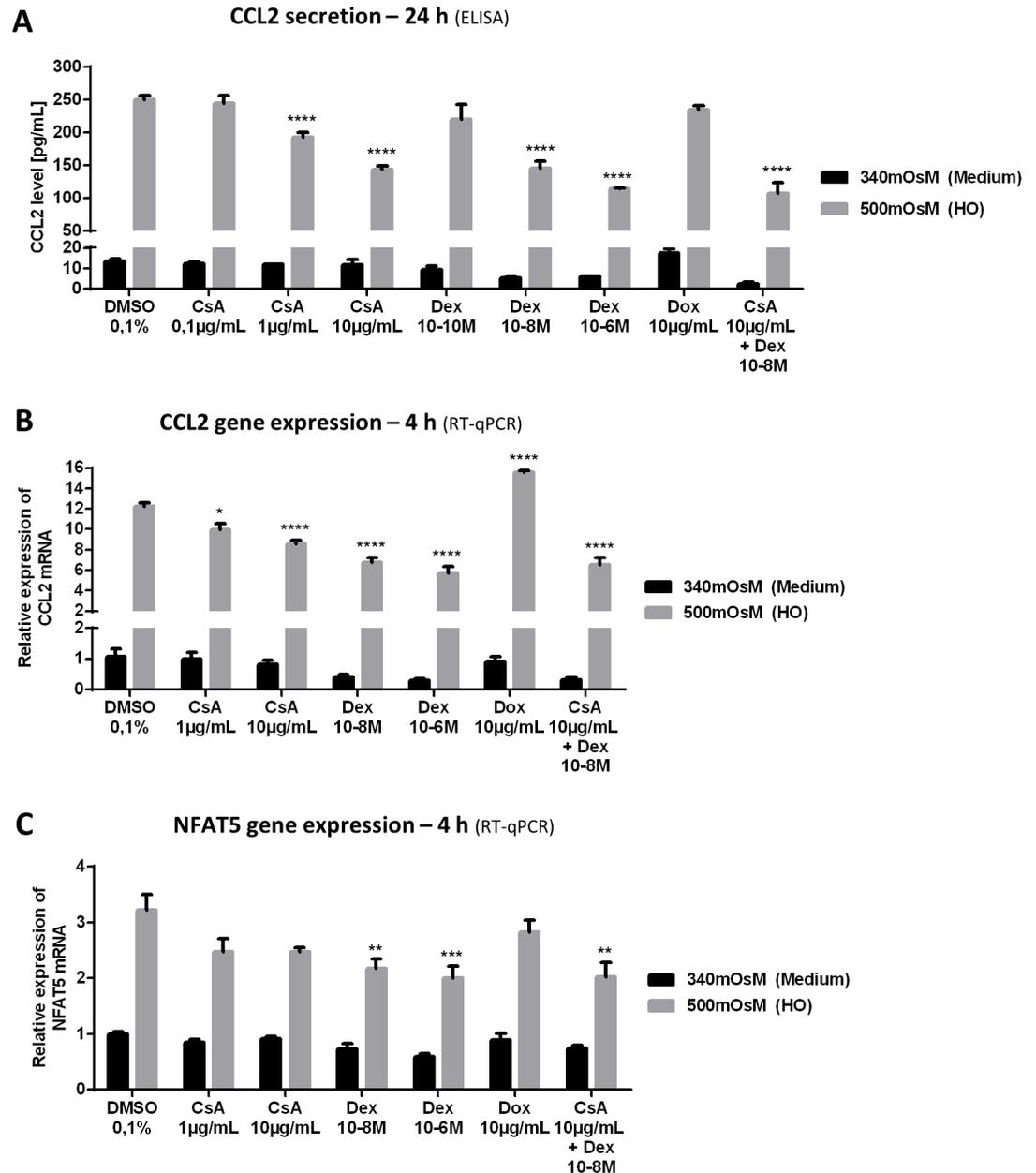


Fig 6. Effects of CsA, Dex and Dox on HO-induced CCL2 secretion and CCL2 and NFAT5 gene expression in HeLa-modified conjunctiva-derived cells. Cells were treated for 1 h with DMSO 0.1%, CsA (0.1, 1 and 10 µg/mL), Dex (10⁻¹⁰, 10⁻⁸ and 10⁻⁶M), Dox 10 µg/mL or a mix of CsA 10 µg/mL and Dex 10⁻⁸M before adding supplemented medium 340 mOsM or HO to reach 500 mOsM. Cells exposed to medium or HO for 24 h were tested for CCL2 secretion (A) by ELISA and cells exposed for 4 h were lysed for mRNA expression using RT-qPCR to determine the CCL2 and NFAT5 gene expression (B, C). Data showing mean ± SEM, *n* = 3 or more. Statistical analysis: two-way ANOVA followed by Tukey's multiple comparison test. * *p* ≤ 0.05; ** *p* ≤ 0.01; *** *p* ≤ 0.001; **** *p* ≤ 0.0001, compared to respective DMSO 0.1%.

doi:10.1371/journal.pone.0159983.g006

by HO and even led to a significant increase in HO-induced CCL2 gene expression compared to DMSO (15.6-fold, *p* ≤ 0.0001).

NFAT5 gene expression. HO conditions increased NFAT5 mRNA expression 3.2-fold. This HO-induced increase was significantly inhibited by Dex 10⁻⁸ and 10⁻⁶M (2.2-fold and

2.0-fold, respectively) (Fig 6C). CsA and Dox induced no significant effect on NFAT5 gene expression.

Mixing Dex and CsA induced no effect that differed significantly from each of the conditions alone (statistics not shown; two-way ANOVA followed by Tukey's multiple comparison test). Under 340-mOsM conditions, none of the compounds tested had any effect on the production and gene expression of CCL2, or on NFAT5 gene expression by HeLa-modified conjunctiva-derived cells. None of the compounds tested affected cellular viability compared to DMSO 0.1% conditions under 340 mOsM or 500 mOsM, respectively (data not shown).

Discussion

Despite its worldwide high prevalence and its significant impact on everyday life for patients, DED actually has few symptomatic and curative treatments. It is therefore important to improve the understanding of the pathogenesis mechanisms with the ultimate goal of developing new targeted therapeutic strategies. Conjunctiva could be an important tissue to study as it covers the major part of the ocular surface and is the site where inflammatory reactions take place. Moreover, conjunctival cells themselves are able to secrete inflammatory cytokines and to participate in inflammatory processes [52]. Therefore, we used a HeLa-modified cell line derived from conjunctiva in a hyperosmolar NaCl-induced in vitro model of dry eye. We wished to characterize the mechanism of the induction of the CCL2 pro-inflammatory chemokine through cellular pathways, such as MAPK and NF κ B, with a focus on the NFAT5 transcription factor. We also investigated the effects of different drugs used in DED, among which CsA, on CCL2 and NFAT5 induction.

We demonstrated that HeLa-modified conjunctiva-derived cells respond to hyperosmotic stress by inducing gene expression and secretion of CCL2, just as corneal cells or other cell types do [53,54]. This induction is dependent on the osmolarity level, and with the highest range of osmolarity, this induction decreases, probably due to the substantial cell death that occurs under these conditions. The lowest osmolarity tested that induced both secretion and gene expression of CCL2 was 500 mOsM, a classical osmolarity level used in previous studies on ocular surface cells [55]. Our results tend to show that HO induces CCL2 by stimulating its gene expression and that there is no preformed stock of this chemokine in these cells, as under hyperosmolar condition, gene expression of CCL2 was induced before its secretion. By secreting this pro-inflammatory chemokine, conjunctival cells could therefore be responsible for the attraction of immune cells, namely monocytes/macrophages that will further nourish the inflammatory process on the ocular surface. Blocking this inflammatory pathway could then potentially be of therapeutic interest, as Goyal et al. have demonstrated in an in vivo model of dry eye [32]. Indeed, animals with dry eye receiving CCR2 antagonist presented a decrease in corneal alterations and in pro-inflammatory cytokines TNF- α and IL-1 β on the ocular surface, confirming the importance of CCL2 in the pathology.

CsA displayed a concentration-dependent inhibitory effect on the HO-induced CCL2 production in our model. This was also observed with Dex, an effect previously reported in a study on corneal cells [29]. Conversely, Dox induced an elevation on CCL2 gene expression. Reported inhibitory effects of anti-inflammatory drugs on CCL2 induction are various. Regarding only studies on ocular cells, Dex was reported to inhibit CCL2 induced by IL-1 β on retinal cells [56] but not on corneal cells [49]. In retinal epithelial cells, CsA has inhibited the CCL2 induction caused by TNF α but not by IL-1 β [56]. In lung epithelial cells, Dox inhibited CCL2 production induced by a mix of cytokines [57]. In addition, we observed that in spite of having different mechanisms of action, CsA and Dex inhibit CCL2 induction probably through the same molecular pathway, since combining these two molecules did not result in a stronger inhibitory effect.

It is noteworthy that the drug concentrations tested are lower than those actually present in eye-drops. Our results further highlight that the CCL2 induction pathway could represent an advantageous therapeutic target, as the anti-inflammatory effect of CsA and Dex observed in DED patients could partly be explained by its inhibition. It is therefore important to understand which molecular actors of this pathway lead to CCL2 secretion.

The three MAPKs, p38, JNK and ERK, as well as the transcription factor NF κ B are activated by HO in several types of mammalian cells [33,58–61]. Previous studies on corneal cells showed that NaCl-induced HO activates p38 [51,62], JNK [19,22,51,63,64] and ERK [19] as well as NF κ B [65], and a recent study showed that HO activates JNK on primary conjunctival cells as well [66]. In our experimental conditions, we demonstrated that CCL2 stimulation by HO is related to p38, JNK and NF κ B but not to ERK. These results confirmed those of other studies conducted on corneal cells, which found that some effects of HO were directly linked to these pathways: induction of MMP [22], apoptosis [19] and induction of cytokine secretion such as IL-1 β [50]. These pathways are therefore potential factors to consider for developing new drugs for DED and they are known to be repressed by CsA and Dex, including in ocular cell studies [29,67,68].

HO is known to activate NFAT5, a transcription factor that plays a key role in osmoprotection by driving osmoprotective gene expression, thus promoting cell survival and protecting cells from the deleterious effects of shrinkage. In addition to this fundamental role, it has a wider range of functions, such as inflammatory cytokine production [69,70] or growth factor stimulation [71]. In our experiments, we observed that HO induces NFAT5 translocation and mRNA expression, and we demonstrated that the CCL2 stimulation induced by HO is entirely mediated by NFAT5. Lee et al. showed that, in limbal cells, the induction of IL-1 β and TNF- α by HO was also related to NFAT5 [36] and a relation between HO, CCL2 and NFAT5 has been reported in rat kidney cells and mesothelial cells [33,34]. In our experiments, we also confirmed the protective effect of NFAT5 on cell survival under hyperosmolar stress. These findings on HeLa-modified conjunctiva-derived cells highlight the potentially crucial role of NFAT5 in dry eye syndrome and the necessity to further investigate its mechanisms and effects in ocular cells. We also determined that the HO-induced NFAT5 expression was linked to p38 activity and that NF κ B has an important dual role in NFAT5 expression, with a stimulatory effect in basal conditions and an inhibitory effect in HO. Another study reported that NFAT5 modulates NF κ B activity in hyperosmolar conditions [72], confirming the important interaction between these two actors activated by hyperosmolar stress. We showed that Dex was able to decrease HO-induced NFAT5 mRNA expression. This effect could explain its inhibitory action on HO-induced CCL2 stimulation. CsA also induces a decrease on it but in a non-statistically significant manner. Other studies on corneal or collecting duct cells have shown a lack of effect of CsA on NFAT5 expression [73,74]. Overall, our results tend to show similar effects induced by CsA and Dex on HO-induced response in HeLa-modified conjunctiva-derived cells, especially on CCL2 induction.

All these results obtained with HeLa-modified-WKD cells should be compared and confirmed with primary human conjunctival cells. Nevertheless, our results highlight the potentially important role of conjunctival cells in the response of the ocular surface to a modification of the tear film. They are able to secrete pro-inflammatory mediators that could participate in the vicious circle of DED by interfering with the inflammatory process stimulated by tear film HO. The inflammatory cellular pathway of NFAT5 translocation, a typical feature of hyperosmolar stress, leading to CCL2 secretion through other factors such as MAPKs and NF κ B, constitutes a biological cascade with potentially promising new targets of interest for DED therapy.

Supporting Information

S1 Table. Raw data from Fig 2.
(XLSX)

S2 Table. Raw data from Fig 3.
(XLSX)

S3 Table. Raw data from Fig 4.
(XLSX)

S4 Table. Raw data from Fig 5.
(XLSX)

S5 Table. Raw data from Fig 6.
(XLSX)

Acknowledgments

We thank Benedict O'Donnell and Linda Northrup for editing advice and Georges Kalouche for providing technical assistance for in vitro experiments. We also thank the Institut National de la Santé Et de la Recherche Médicale.

Author Contributions

Conceptualization: EW CB CG FB.

Formal analysis: EW FB.

Funding acquisition: CB FB.

Investigation: EW.

Methodology: EW CB FB.

Project administration: CB FB.

Resources: CG.

Supervision: FB CB.

Validation: EW FB.

Visualization: EW FB.

Writing - original draft: EW FB.

Writing - review & editing: EW CB CG FB.

References

1. DEWS. The definition and classification of dry eye disease: report of the Definition and Classification Subcommittee of the International Dry Eye WorkShop (2007). *Ocul Surf.* 2007; 5:75–92. PMID: [17508116](#)
2. Goto E, Yagi Y, Matsumoto Y, Tsubota K. Impaired functional visual acuity of dry eye patients. *Am J Ophthalmol.* 2002; 133:181–6. PMID: [11812420](#)
3. Bron AJ, Tomlinson A, Foulks GN, Pepose JS, Baudouin C, Geerling G, et al. Rethinking dry eye disease: a perspective on clinical implications. *Ocul Surf.* 2014; 12:S1–31. doi: [10.1016/j.jtos.2014.02.002](#) PMID: [24725379](#)

4. Miljanović B, Dana R, Sullivan D a., Schaumberg D a. Impact of Dry Eye Syndrome on Vision-Related Quality of Life. *Am J Ophthalmol.* 2007; 143:409–15. PMID: [17317388](#)
5. Labbé A, Wang YX, Jie Y, Baudouin C, Jonas JB, Xu L. Dry eye disease, dry eye symptoms and depression: the Beijing Eye Study. *Br J Ophthalmol.* 2013; 97:1399–403. doi: [10.1136/bjophthalmol-2013-303838](#) PMID: [24013959](#)
6. Baudouin C. A new approach for better comprehension of diseases of the ocular surface. *J Fr Ophtalmol.* 2007; 30:239–46. PMID: [17417148](#)
7. Baudouin C, Aragona P, Messmer EM, Tomlinson A, Calonge M, Boboridis KG, et al. Role of Hyperosmolarity in the Pathogenesis and Management of Dry Eye Disease: Proceedings of the OCEAN Group Meeting. *Ocul Surf.* 2013; 11:246–58. doi: [10.1016/j.jtos.2013.07.003](#) PMID: [24112228](#)
8. Liu H, Begley C, Chen M, Bradley A, Bonanno J, McNamara NA, et al. A link between tear instability and hyperosmolarity in dry eye. *Invest Ophthalmol Vis Sci.* 2009; 50:3671–9. doi: [10.1167/iovs.08-2689](#) PMID: [19324847](#)
9. Stern ME, Pflugfelder SC. Inflammation in dry eye. *Ocul Surf.* 2004; 2:124–30. PMID: [17216083](#)
10. Pisella P, Brignole F, Debbasch C, Lozato P, Creuzot-garcher C, Bara J, et al. Flow Cytometric Analysis of Conjunctival Epithelium in Ocular Rosacea and Keratoconjunctivitis Sicca. *Ophthalmology.* 2000; 101:1841–9.
11. Ralph RA. Conjunctival goblet cell density in normal subjects and in dry eye syndromes. *Invest Ophthalmol.* 1975; 14:299–302. PMID: [1123285](#)
12. Barabino S, Montaldo E, Solignani F, Valente C, Mingari MC, Rolando M. Immune response in the conjunctival epithelium of patients with dry eye. *Exp Eye Res.* 2010; 91:524–9. doi: [10.1016/j.exer.2010.07.008](#) PMID: [20654615](#)
13. Massingale ML, Li X, Vallabhajosyula M, Chen D, Wei Y, Asbell P a. Analysis of inflammatory cytokines in the tears of dry eye patients. *Cornea.* 2009; 28:1023–7. doi: [10.1097/ICO.0b013e3181a16578](#) PMID: [19724208](#)
14. Yoon K-C, Jeong I-Y, Park Y-G, Yang S-Y. Interleukin-6 and tumor necrosis factor-alpha levels in tears of patients with dry eye syndrome. *Cornea.* 2007; 26:431–7. PMID: [17457192](#)
15. Brignole F, Pisella P, Goldschild M, Saint Jean MD, Goguel A, Baudouin C. Flow Cytometric Analysis of Inflammatory Markers in Conjunctival Epithelial Cells of Patients with Dry Eyes. *Invest Ophthalmol Vis Sci.* 2000; 41:1356–63. PMID: [10798650](#)
16. Kunert KS, Tisdale AS, Gipson IK. Goblet cell numbers and epithelial proliferation in the conjunctiva of patients with dry eye syndrome treated with cyclosporine. *Arch Ophthalmol.* 2002; 120:330–7. PMID: [11879137](#)
17. Lam H, Bleiden L, De Paiva CS, William F, E SM, Pflugfelder SC. Tear cytokine profiles in dysfunctional tear syndrome. *Am J Ophthalmol.* 2009; 147:1–19.
18. Clouzeau C, Godefroy D, Riancho L, Rostène W, Baudouin C, Brignole-Baudouin F. Hyperosmolarity potentiates toxic effects of benzalkonium chloride on conjunctival epithelial cells in vitro. *Mol Vis.* 2012; 18:851–63. PMID: [22529703](#)
19. Luo L, Li D-Q, Pflugfelder SC. Hyperosmolarity-induced apoptosis in human corneal epithelial cells is mediated by cytochrome c and MAPK pathways. *Cornea.* 2007; 26:452–60. PMID: [17457195](#)
20. Chen Y, Li M, Li B, Wang W, Lin A, Sheng M. Effect of reactive oxygen species generation in rabbit corneal epithelial cells on inflammatory and apoptotic signaling pathways in the presence of high osmotic pressure. *PLoS One.* 2013; 8:e72900. doi: [10.1371/journal.pone.0072900](#) PMID: [23977369](#)
21. Deng R, Hua X, Li J, Chi W, Zhang Z, Lu F, et al. Oxidative stress markers induced by hyperosmolarity in primary human corneal epithelial cells. *PLoS One.* 2015; 10:e0126561. doi: [10.1371/journal.pone.0126561](#) PMID: [26024535](#)
22. Li D-Q, Chen Z, Song XJ, Luo L, Pflugfelder SC. Stimulation of matrix metalloproteinases by hyperosmolarity via a JNK pathway in human corneal epithelial cells. *Invest Ophthalmol Vis Sci.* 2004; 45:4302–11. PMID: [15557436](#)
23. Li D, Luo L, Chen Z, Kim H-S, Song XJ, Pflugfelder SC. JNK and ERK MAP kinases mediate induction of IL-1 β , TNF- α and IL-8 following hyperosmolar stress in human limbal epithelial cells. *Exp Eye Res.* 2006; 82:588–96. PMID: [16202406](#)
24. Pan Z, Wang Z, Yang H, Zhang F, Reinach PS. TRPV1 activation is required for hypertonicity-stimulated inflammatory cytokine release in human corneal epithelial cells. *Invest Ophthalmol Vis Sci.* 2011; 52:485–93. doi: [10.1167/iovs.10-5801](#) PMID: [20739465](#)
25. Luo L, Li D-Q, Corrales RM, Pflugfelder SC. Hyperosmolar saline is a proinflammatory stress on the mouse ocular surface. *Eye Contact Lens.* 2005; 31:186–93. PMID: [16163009](#)

26. Zhu L, Shen J, Zhang C, Park CY, Kohanim S, Yew M, et al. Inflammatory cytokine expression on the ocular surface in the Botulium toxin B induced murine dry eye model. *Mol Vis*. 2009; 15:250–8. PMID: [19190733](#)
27. Hua X, Su Z, Deng R, Lin J, Li D-Q, Pflugfelder SC. Effects of L-Carnitine, Erythritol and Betaine on Pro-inflammatory Markers in Primary Human Corneal Epithelial Cells Exposed to Hyperosmotic Stress. *Curr Eye Res*. 2015; 40:657–67. doi: [10.3109/02713683.2014.957776](#) PMID: [25271595](#)
28. Cavet ME, Harrington KL, Vollmer TR, Ward KW, Zhang J-Z. Anti-inflammatory and anti-oxidative effects of the green tea polyphenol epigallocatechin gallate in human corneal epithelial cells. *Mol Vis*. 2011; 17:533–42. PMID: [21364905](#)
29. Cavet ME, Harrington KL, Ward KW, Zhang J-Z. Mapracorat, a novel selective glucocorticoid receptor agonist, inhibits hyperosmolar-induced cytokine release and MAPK pathways in human corneal epithelial cells. *Mol Vis*. 2010; 16:1791–800. PMID: [20824100](#)
30. Chen L, Li J, Guo T, Ghosh S, Koh SK, Tian D, et al. Global Metabonomic and Proteomic Analysis of Human Conjunctival Epithelial Cells (IOBA-NHC) in Response to Hyperosmotic Stress. *J Proteome Res*. 2015; 14:3982–95. doi: [10.1021/acs.jproteome.5b00443](#) PMID: [26260330](#)
31. Deshmane SL, Kremlev S, Amini S, Sawaya BE. Monocyte chemoattractant protein-1 (MCP-1): an overview. *J Interf cytokine Res*. 2009; 29:313–26.
32. Goyal S, Chauhan SK, Zhang Q, Dana R. Amelioration of Murine Dry Eye Disease by Topical Antagonist to Chemokine Receptor 2. *Arch Ophthalmol*. 2009; 127:882–7. doi: [10.1001/archophthalmol.2009.125](#) PMID: [19597109](#)
33. Kojima R, Taniguchi H, Tsuzuki A, Nakamura K, Sakakura Y, Ito M. Hypertonicity-induced expression of monocyte chemoattractant protein-1 through a novel cis-acting element and MAPK signaling pathways. *J Immunol*. 2010; 184:5253–62. doi: [10.4049/jimmunol.0901298](#) PMID: [20368270](#)
34. Küper C, Beck F-X, Neuhofer W. NFAT5 contributes to osmolality-induced MCP-1 expression in mesothelial cells. *Mediators Inflamm*. 2012; 2012:513015. doi: [10.1155/2012/513015](#) PMID: [22619484](#)
35. Miyakawa H, Woo SK, Dahl SC, Handler JS, Kwon HM. Tonicity-responsive enhancer binding protein, a rel-like protein that stimulates transcription in response to hypertonicity. *Proc Natl Acad Sci U S A*. 1999; 96:2538–42. PMID: [10051678](#)
36. Lee JH, Kim M, Im YS, Choi W, Byeon SH, Lee HK. NFAT5 induction and its role in hyperosmolar stressed human limbal epithelial cells. *Invest Ophthalmol Vis Sci*. 2008; 49:1827–35. doi: [10.1167/iovs.07-1142](#) PMID: [18436816](#)
37. Tsubota K, Fujihara T, Saito K, Takeuchi T. Conjunctival epithelium expression of HLA-DR in dry eye patients. *Ophthalmologica*. 1999; 213:16–9. PMID: [9838252](#)
38. Baudouin C, Haouat N, Brignole F, Bayle J, Gastaud P. Immunopathological findings in conjunctival cells using immunofluorescence staining of impression cytology specimens. *Br J Ophthalmol*. 1992; 76:545–9. PMID: [1420060](#)
39. Baudouin C, Liang H, Bremond-Gignac D, Hamard P, Hreiche R, Creuzot-Garcher C, et al. CCR 4 and CCR 5 expression in conjunctival specimens as differential markers of T(H)1/ T(H)2 in ocular surface disorders. *J Allergy Clin Immunol*. 2005; 116:614–9. PMID: [16159632](#)
40. Contreras-Ruiz L, Regenfuss B, Mir FA, Kearns J, Masli S. Conjunctival inflammation in thrombospondin-1 deficient mouse model of Sjögren's syndrome. *PLoS One*. 2013; 8:e75937. doi: [10.1371/journal.pone.0075937](#) PMID: [24086667](#)
41. Xiao B, Wang Y, Reinach PS, Ren Y, Li J, Hua S, et al. Dynamic Ocular Surface and Lacrimal Gland Changes Induced in Experimental Murine Dry Eye. *PLoS One*. 2015; 10:e0115333. doi: [10.1371/journal.pone.0115333](#) PMID: [25590134](#)
42. Brasnu E, Brignole-Baudouin F, Riancho L, Warnet J-M, Baudouin C. Comparative study on the cytotoxic effects of benzalkonium chloride on the Wong-Kilbourne derivative of Chang conjunctival and IOBA-NHC cell lines. *Mol Vis*. 2008; 14:394–402. PMID: [18334956](#)
43. Ramirez-miranda A, Nakatsu MN, Zarei-ghanavati S, Nguyen C V, Deng SX. Keratin 13 is a more specific marker of conjunctival epithelium than keratin 19. *Mol Vis*. 2011; 17:1652–61. PMID: [21738394](#)
44. Tong L, Diebold Y, Calonge M, Gao J, Stern ME, Beuerman RW. Comparison of gene expression profiles of conjunctival cell lines with primary cultured conjunctival epithelial cells and human conjunctival tissue. *Gene Expr*. 2009; 14:265–78. PMID: [19630270](#)
45. Sullivan BD, Whitmer D, Nichols KK, Tomlinson A, Foulks GN, Geerling G, et al. An objective approach to dry eye disease severity. *Invest Ophthalmol Vis Sci*. 2010; 51:6125–30. doi: [10.1167/iovs.10-5390](#) PMID: [20631232](#)
46. Gilbard JP, Farris RL, Santamaria J. Osmolarity of tear microvolumes in keratoconjunctivitis sicca. *Arch Ophthalmol*. 1978; 96:677–81. PMID: [646697](#)

47. Farris RL. Tear osmolarity—a new gold standard? *Adv Exp Med Biol.* 1994; 350:495–503. PMID: [8030524](#)
48. Zheng Q, Ren Y, Reinach PS, Xiao B, Lu H, Zhu Y, et al. Reactive oxygen species activated NLRP3 inflammasomes initiate inflammation in hyperosmolarity stressed human corneal epithelial cells and environment-induced dry eye patients. *Exp Eye Res.* 2015; 134:133–40. doi: [10.1016/j.exer.2015.02.013](#) PMID: [25701684](#)
49. Djalilian AR, Nagineni CN, Mahesh SP, Smith J a, Nussenblatt RB, Hooks JJ. Inhibition of inflammatory cytokine production in human corneal cells by dexamethasone, but not cyclosporin. *Cornea.* 2006; 25:709–14. PMID: [17077666](#)
50. Chen M, Hu D-N, Pan Z, Lu C-W, Xue C-Y, Aass I. Curcumin protects against hyperosmoticity-induced IL-1beta elevation in human corneal epithelial cell via MAPK pathways. *Exp Eye Res.* 2010; 90:437–43.
51. Corrales RM, Luo L, Chang EY, Pflugfelder SC. Effects of osmoprotectants on hyperosmolar stress in cultured human corneal epithelial cells. *Cornea.* 2008; 27:574–9. doi: [10.1097/ICO.0b013e318165b19e](#) PMID: [18520508](#)
52. Pflugfelder SC, Jones D, Ji Z, Afonso A, Monroy D. Altered cytokine balance in the tear fluid and conjunctiva of patients with Sjögren's syndrome keratoconjunctivitis sicca. *Curr Eye Res.* 1999; 19:201–11. PMID: [10487957](#)
53. Hua X, Deng R, Li J, Chi W, Su Z, Lin J, et al. Protective Effects of L-Carnitine Against Oxidative Injury by Hyperosmolarity in Human Corneal Epithelial Cells. *Investig Ophthalmology Vis Sci.* 2015; 56:5503–11.
54. Matsuo H, Tamura M, Kabashima N, Serino R, Tokunaga M, Shibata T, et al. Prednisolone inhibits hyperosmolarity-induced expression of MCP-1 via NF-kappaB in peritoneal mesothelial cells. *Kidney Int.* 2006; 69:736–46. PMID: [16518329](#)
55. Narayanan S, Manning J, Proske R, McDermott AM. Effect of Hyperosmolality on β -Defensin Gene Expression by Human Corneal Epithelial Cells. *Cornea.* 2006; 25:1063–8. PMID: [17133055](#)
56. Kurtz RM, Elner VM, Bian ZM, Strieter RM, Kunkel SL, Elner SG. Dexamethasone and cyclosporin A modulation of human retinal pigment epithelial cell monocyte chemoattractant protein-1 and interleukin-8. *Invest Ophthalmol Vis Sci.* 1997; 38:436–45. PMID: [9040477](#)
57. Raza M, Ballering JG, Hayden JM, Robbins RA, Hoyt JC. Doxycycline decreases monocyte chemoattractant protein-1 in human lung epithelial cells. *Exp Lung Res.* 2006; 32:15–26. PMID: [16809218](#)
58. Duzgun S a, Rasque H, Kito H, Azuma N, Li W, Basson MD, et al. Mitogen-activated protein phosphorylation in endothelial cells exposed to hyperosmolar conditions. *J Cell Biochem.* 2000; 76:567–71. PMID: [10653976](#)
59. Matsuda S, Kawasaki H, Moriguchi T, Gotoh Y, Nishida E. Activation of protein kinase cascades by osmotic shock. *J Biol Chem.* 1995; 270:12781–6. PMID: [7759532](#)
60. Németh ZH, Deitch E a, Szabó C, Haskó G. Hyperosmotic stress induces nuclear factor-kappaB activation and interleukin-8 production in human intestinal epithelial cells. *Am J Pathol.* 2002; 161:987–96. PMID: [12213727](#)
61. Hao CM, Yull F, Blackwell T, Kömhoff M, Davis LS, Breyer MD. Dehydration activates an NF- κ B-driven, COX2-dependent survival mechanism in renal medullary interstitial cells. *J Clin Invest.* 2000; 106:973–82. PMID: [11032857](#)
62. Wang L, Dai W, Lu L. Hyperosmotic stress-induced corneal epithelial cell death through activation of Polo-like kinase 3 and c-Jun. *Invest Ophthalmol Vis Sci.* 2011; 52:3200–6. doi: [10.1167/ovs.10-6485](#) PMID: [21296815](#)
63. Chen Z, Tong L, Li Z, Yoon K-C, Qi H, Farley W, et al. Hyperosmolarity-induced cornification of human corneal epithelial cells is regulated by JNK MAPK. *Invest Ophthalmol Vis Sci.* 2008; 49:539–49. doi: [10.1167/ovs.07-0569](#) PMID: [18234997](#)
64. Capó-Aponte JE, Wang Z, Bildin VN, Pokorny KS, Reinach PS. Fate of hypertonicity-stressed corneal epithelial cells depends on differential MAPK activation and p38MAPK/Na-K-2Cl cotransporter1 interaction. *Exp Eye Res.* 2007; 84:361–72. PMID: [17140565](#)
65. Chang E-J, Im YS, Kay EP, Kim JY, Lee JE, Lee HK. The Role of Nerve Growth Factor in Hyperosmolar Stress Induced Apoptosis. *J Cell Physiol.* 2008; 216:69–77. doi: [10.1002/jcp.21377](#) PMID: [18300262](#)
66. Kang S, Ha S, Kim E, Shin J, Kim JY, Tchah H. Effect of Nerve Growth Factor on the In Vitro Induction of Apoptosis of Human Conjunctival Epithelial Cells by Hyperosmolar Stress. *Invest Ophthalmol Vis Sci.* 2014; 55:535–41. doi: [10.1167/ovs.13-12459](#) PMID: [24327613](#)
67. Matsuda S, Koyasu S. Mechanisms of action of cyclosporine. *Immunopharmacology.* 2000; 47:119–25. PMID: [10878286](#)

68. Mattila PS, Ullman KS, Fiering S, Emmel E a, McCutcheon M, Crabtree GR, et al. The actions of cyclosporin A and FK506 suggest a novel step in the activation of T lymphocytes. *EMBO J.* 1990; 9:4425–33. PMID: [1702384](#)
69. López-Rodríguez C, Antos CL, Shelton JM, Richardson J a, Lin F, Novobrantseva TI, et al. Loss of NFAT5 results in renal atrophy and lack of tonicity-responsive gene expression. *Proc Natl Acad Sci U S A.* 2004; 101:2392–7. PMID: [14983020](#)
70. Arambu J, Drews-Elger K, Estrada-Gelonch A, Minguillon J, Morancho B, Santiago V, et al. Regulation of the hypertonic stress respons and the other cellular functions by the Rel-like transcription factor NFAT5. *Biochem Pharmacol.* 2006; 72:1597–604. PMID: [16904650](#)
71. Hollborn M, Vogler S, Reichenbach A, Wiedemann P, Bringmann A, Kohen L. Regulation of the hyperosmotic induction of aquaporin 5 and VEGF in retinal pigment epithelial cells : Involvement of NFAT5. *Mol Vis.* 2015; 21:360–77. PMID: [25878490](#)
72. Roth I, Leroy V, Martin P, Féraille E, Hasler U. Osmoprotective transcription factor NFAT5/TonEBP modulates nuclear factor-kappaB activity. *Mol Biol Cell.* 2010; 21:3459–74. doi: [10.1091/mbc.E10-02-0133](#) PMID: [20685965](#)
73. Lee JH, Kim JW, Im YS, Seong GJ, Lee HK. Cyclosporine A induces nerve growth factor expression via activation of MAPK p38 and NFAT5. *Cornea.* 2011; 30:S19–24. doi: [10.1097/ICO.0b013e3182281028](#) PMID: [21912224](#)
74. Rinschen MM, Klokkers J, Pavenstädt H, Neugebauer U, Schlatter E, Edemir B. Different effects of CsA and FK506 on aquaporin-2 abundance in rat primary cultured collecting duct cells. *Pflugers Arch.* 2011; 462:611–22. doi: [10.1007/s00424-011-0994-6](#) PMID: [21773745](#)