

Interspecific Variation in Compensatory Regrowth to Herbivory Associated with Soil Nutrients in Three *Ficus* (Moraceae) Saplings

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Abstract

Plant compensatory regrowth is an induced process that enhances plant tolerance to herbivory. Plant behavior against herbivores differs between species and depends on resource availability, thus making general predictions related to plant compensatory regrowth difficult. To understand how soil nutrients determine the degree of compensatory regrowth for different plant species, we selected saplings of three *Ficus* species and treated with herbivore insects and artificial injury in both glasshouse conditions and in the field at two soil nutrient levels. Compensatory regrowth was calculated by biomass, relative growth rate and photosynthetic characteristics. A similar pattern was found in both the glasshouse and in the field for species *F. hispida*, where overcompensatory regrowth was triggered only under fertile conditions, and full compensatory regrowth occurred under infertile conditions. For *F. auriculata*, overcompensatory regrowth was stimulated only under infertile conditions and full compensatory regrowth occurred under fertile conditions. *Ficus racemosa* displayed full compensatory regrowth in both soil nutrient levels, but without overcompensatory regrowth following any of the treatments. The three *Ficus* species differed in biomass allocation following herbivore damage and artificial injury. The root/shoot ratio of *F. hispida* decreased largely following herbivore damage and artificial injury, while the root/shoot ratio for *F. auriculata* increased against damage treatments. The increase of shoot and root size for *F. hispida* and *F. auriculata*, respectively, appeared to be caused by a significant increase in photosynthesis. The results indicated that shifts in biomass allocation and increased photosynthesis are two of the mechanisms underlying compensatory regrowth. Contrasting patterns among the three *Ficus* species suggest that further theoretical and empirical work is necessary to better understand the complexity of the plant responses to herbivore damage.

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Introduction

Growing evidence suggests that compensatory regrowth is a common tolerance strategy in plants in response to herbivore damage [1–4]. Compensatory regrowth is often achieved by mobilizing resource allocation or physiological function to reduce the impacts of damage on fitness relative to undamaged plants [1,2,5–7]. Following herbivore feeding, biomass of damaged plants could be larger (overcompensatory regrowth), equal (full compensatory regrowth) and less than (under compensatory regrowth) undamaged plants [2]. Differing from the constitutive resistance that plants invest in prior to herbivory damage, compensatory regrowth is an induced process following herbivore attack. It has been widely acknowledged that compensatory regrowth depends on resource availability in the plant's environment [1,8,9].

Compensatory regrowth is a heritable trait and varies markedly among plant species [2,10,11]. Early theories assumed that compensatory regrowth mostly occurred in herbs because of the faster growth rate compared to woody plants, but later studies demonstrated that many woody plants are also able to compensate or even overcompensate for biomass losses caused by herbivores

[10,12–15]. Plants along a successional gradient may also differ in their herbivore defense strategy and degree of tolerance [16–18]. Early to middle successional plants often experience higher levels of herbivory [19] and early successional plants tend to show rapid leaf turnover, comparatively little investment in defensive secondary compounds, and rapid regrowth when compensating for tissue loss [20]. In contrast, late successional plant species have intrinsically slower growth rates [21,22] and possibly lower levels of compensatory regrowth.

Many other factors may also determine the degree of compensatory regrowth such as type, frequency and severity of damage, and the availability of nutrients [8,20,21]. Previous studies have suggested that plant defense may be responsive to insect feeding but not to physical damage alone [22,23]. For example, a study on *Nicotiana sylvestris* (Solanaceae) indicated that higher concentrations of jasmonic acid (JA), which is known to mediate wound responses in plants, resulted mainly from herbivory by *Manduca sexta* (L.) larvae than by mechanical damage [24]. For a better understanding of induced defense by plants, it is therefore necessary to distinguish whether the induced response is stimulated by herbivores, mechanical injuries or both. A plant's

compensatory regrowth against herbivore damage also depends on surrounding resource availability. Some studies have shown that greater compensatory regrowth occurred in high-resource environments [2,25–30], while other studies displayed contradictory results in which plants showed greater compensatory regrowth in relatively stressful environments [29,31–33]. Until now, many attempts for a general explanation of plant compensatory regrowth under different environments have been proposed [8,9,20,34]; however, it appears difficult to make general predictions on compensatory regrowth following herbivore damage. For this reason, interspecific comparisons can help elucidate how plant species may evolve higher levels of compensatory regrowth [2].

We were particularly interested in examining how potential mechanisms of compensatory regrowth for woody plants are influenced by plant species and resource availability, since the majority of research in this field has focused on herbs and the conclusions may not pertain to woody species [7,35]. Woody species in general have a proportionately large capacity for storage of carbon and nutrient reserves compared with herbaceous species [36]. The allocation and accumulation of these reserves within the tree following defoliation is of particular interest because it may provide insights into why defoliation sometimes has little or no effect on growth [37,38]. In this study, we report the effect of resource availability on compensatory regrowth and the potential mechanisms of compensatory regrowth against herbivore damage in *Ficus* saplings.

Ficus (Moraceae) is one of the largest genera of woody plants in the tropics and shows diverse adaptations to different habitats [27]. Three *Ficus* species that are commonly distributed along rainforest edges or beside roads were studied: *F. hispida*, *F. racemosa* and *F. auriculata*. Previous findings suggest that these *Ficus* species receive significantly different levels of damage from herbivorous insects both in the field and in glasshouse experiments [39], and show interspecific variation in morphological and chemical defense [39,40]. In a common garden experiment, *F. hispida* suffered significantly more severe herbivore damage than the other two species [39]. And, the typical pioneer species *F. hispida* has considerably more pubescence on both the upper and lower leaves and lower C/N than the intermediate successional species *F. auriculata* [39]. Additionally, the amount of volatiles released by the three *Ficus* species were significantly different following herbivore damage [40].

To understand how soil nutrients determine the degree of compensatory regrowth of these *Ficus* species following herbivore damage and the potential mechanisms of compensatory regrowth, one field and one glasshouse experiment in which saplings of each species were treated with herbivores and artificial injury at two different soil nutrient levels were established. We predicted that: 1) compensatory regrowth of the three *Ficus* species would be greater under fertile conditions than infertile conditions; 2) different species may display different patterns in compensatory regrowth associated with resource availability; 3) herbivore damage causes different patterns in compensatory regrowth compared with artificial injury. Broadly, we wish to examine whether the three *Ficus* species show a general pattern in compensatory regrowth against herbivores under different levels of resource availability.

Results

Compensatory regrowth

Relative growth rate (RGR) differed significantly among species ($F = 3.63$, $P = 0.009$ in 2009; $F = 64.52$, $P < 0.001$ in 2011), and RGRs of saplings under fertile condition were higher significantly than these in infertile condition ($F = 21.95$, $P < 0.001$ in 2009;

$F = 13.26$, $P < 0.001$ in 2011). But damage treatments did not affect RGRs significantly ($F = 1.06$, $P = 0.15$ in 2009; $F = 1.72$, $P = 0.19$ in 2011). RGRs were also significantly affected by the interaction between species and soil nutrient level ($F = 4.56$, $P < 0.001$ in 2009; $F = 7.07$, $P < 0.001$ in 2011), and the interaction among species, soil nutrient level and treatments ($F = 1.89$, $P = 0.022$ in 2009; $F = 6.07$, $P = 0.003$ in 2011). Twenty days after damage treatments were carried out, RGR of *F. hispida* and *F. auriculata* saplings under infertile conditions were lower than those under fertile conditions regardless of the herbivore treatment, but not for *F. racemosa* (Fig. 1). Under fertile conditions, RGR of *F. hispida* increased significantly after herbivore damage and artificial injury (Fig. 1A, D), while for *F. racemosa* and *F. auriculata*, RGR did not differ significantly among the three treatments (Fig. 1B, C, E, F). Of the saplings planted in the infertile soil, only *F. auriculata* showed a significant increase in RGR after herbivore damage and artificial injury (Fig. 1C, F).

Ficus hispida showed overcompensatory regrowth after herbivore damage and artificial injury under fertile soil conditions, but showed full compensatory regrowth under infertile conditions (Fig. 2). *F. racemosa* displayed full compensatory regrowth after herbivore damage and artificial injury under both soil nutrient levels. In contrast, *F. auriculata* displayed overcompensatory regrowth only in infertile soil after herbivore damage and artificial injury but full compensatory regrowth under fertile conditions.

By comparing the total biomass of undamaged saplings in glasshouse experiment, we also found that *F. hispida* and *F. auriculata* saplings were significantly negatively affected by soil nutrient level (*F. hispida*: 14.89 ± 0.67 g under fertile and 9.66 ± 0.49 g under infertile conditions ($n = 5$, $P < 0.0001$); *F. auriculata*: 14.06 ± 0.72 g under fertile and 7.01 ± 0.60 g under infertile conditions ($n = 5$, $P < 0.0001$)). However, the total biomass of undamaged saplings of *F. racemosa* was similar between the two soil nutrient levels (14.21 ± 1.86 g under fertile and 11.21 ± 1.10 g under infertile conditions ($n = 5$, $P = 0.20$)).

Biomass allocation

In the glasshouse experiment, root/shoot ratio was affected significantly by species ($F = 33.62$, $P < 0.001$), soil nutrient level ($F = 146.54$, $P < 0.001$), damage treatments ($F = 3.38$, $P = 0.04$), and interaction between species and nutrient level ($F = 3.95$, $P = 0.024$). In the field, species ($F = 20.71$, $P < 0.001$), soil nutrient level ($F = 17.71$, $P < 0.001$), treatments ($F = 8.22$, $P = 0.005$) and their interactions ($F = 4.95$, $P = 0.008$) had a significant effect on the root/shoot ratio. Root/shoot ratio of *F. hispida* saplings decreased remarkably following herbivore and artificial injury treatments in fertile soil (Fig. 3A, D). However, root/shoot ratio of *F. racemosa* did not change considerably after herbivore feeding or artificial injury under either soil nutrient level (Fig. 3B, E). Only *F. auriculata* saplings showed a prominent increase in root/shoot ratio after herbivore and artificial damage in infertile soils (Fig. 3C, F).

Natural herbivore feeding in the field

In the field experiment, the consumed area of the whole sample of leaves was not related to the *Ficus* species, soil nutrient or their interaction ($12.38 \pm 3.25\%$ for *F. hispida*, $5.88 \pm 1.16\%$ for *F. racemosa*, $6.50 \pm 1.28\%$ for *F. auriculata*; $n = 4$).

Biomass and Photosynthetic gas exchange

RGR increased linearly with increasing P_{sat} in *F. hispida* and *F. auriculata* (Fig. 4A). P_{sat} exhibited a positive relationship with G_s (Fig. 4B). A three-way ANOVA showed that P_{sat} was affected significantly by species ($F = 41.77$, $P < 0.001$), nutrient level ($F = 18.66$, $P < 0.001$), the interaction between species and nutrient

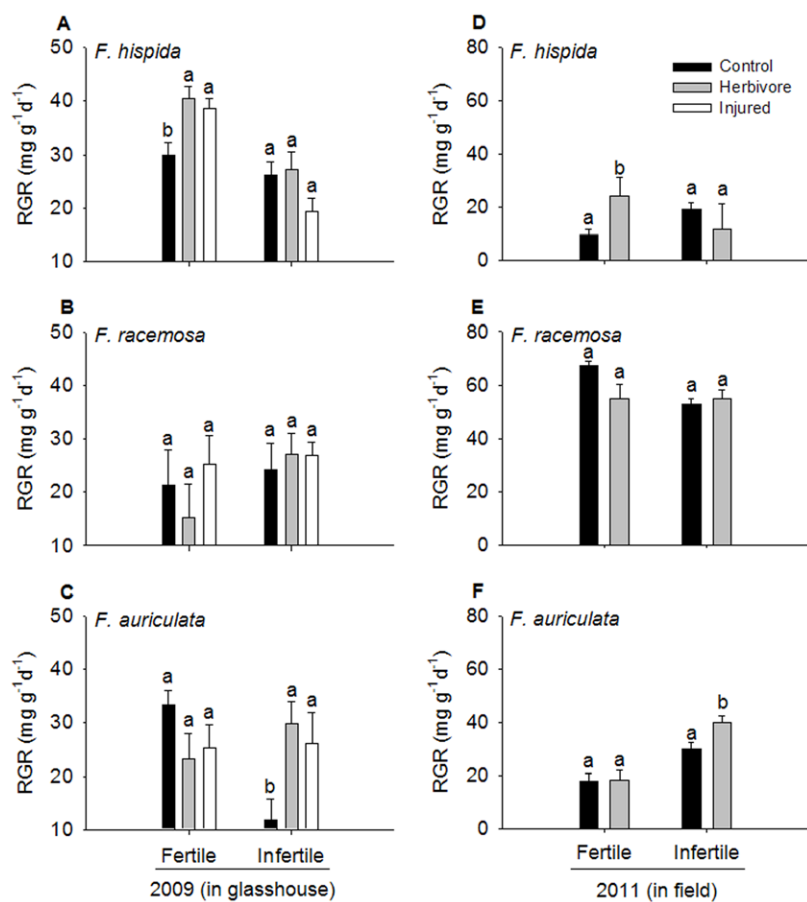


Figure 1. Effect of treatments on RGR of *Ficus* saplings under two soil nutrient levels. Significant differences between treatments are marked with different letters (mean \pm SE, $n=5$ in 2009, $n=12$ in 2011, $P<0.05$). RGR, relative growth rate ($\text{mg}\cdot\text{g}^{-1}\cdot\text{d}^{-1}$). doi:10.1371/journal.pone.0045092.g001

level ($F=6.14$, $P=0.005$), and the interaction among species, nutrient level, and treatments ($F=2.60$, $P=0.05$). Herbivory and artificial injury increased P_{sat} in *F. hispida* saplings in fertile soils, but had no effect in infertile conditions (Fig. 5A). Both herbivory and artificial injury remarkably decreased the P_{sat} of *F. racemosa* under fertile conditions, but this was not the case under infertile conditions (Fig. 5B). For *F. auriculata*, P_{sat} increased prominently after herbivory and artificial injury under infertile conditions but showed no change under fertile conditions (Fig. 5C).

Species, soil nutrient, treatment and the interaction among three factors had significantly effect on G_s , C_i , Tr and WUE ($P<0.05$). The G_s , C_i and Tr of *F. hispida* increased considerably after herbivore damage and artificial injury under fertile conditions but decreased dramatically under infertile conditions (Fig. 5D–L). Herbivore damage and artificial injury significantly inhibited G_s , C_i and Tr in *F. racemosa* under fertile soil conditions but had no effect under infertile soil conditions. The G_s , C_i and Tr of *F. auriculata* increased intensely after herbivore damage under both soil nutrient levels and increased significantly after artificial injury under infertile conditions. WUE of *F. hispida* decreased intensely following herbivore damage and artificial injury under fertile conditions, and increased under infertile conditions (Fig. 5M). The WUE of *F. racemosa* increased following herbivore damage and artificial injury under fertile conditions, but did not change significantly under infertile conditions (Fig. 5N). Herbivore damage and artificial injury caused the WUE of *F. auriculata* to decrease notably under both soil nutrient levels (Fig. 5O).

Discussion

The three *Ficus* species showed great variation in soil nutrient associated compensatory regrowth. Overcompensatory regrowth in *F. hispida* was observed only under fertile soil conditions and *F. hispida* displayed full compensatory regrowth under fertile soil conditions. In *F. auriculata*, overcompensatory regrowth only occurred under infertile soil conditions, full compensatory regrowth were showed under fertile soil conditions. Meanwhile, *F. racemosa*, was less sensitive to the soil nutrient treatment and did not show significant overcompensatory regrowth under either soil nutrient condition. Both introduced insect herbivores and artificial damage had a significant effect on compensatory regrowth in any of the three *Ficus* species. This study provided experimental results for the conditionality of plant response to herbivore damage, and indicates that a general prediction on compensatory regrowth for all species is almost impossible.

Interspecific differences in compensatory regrowth

The saplings of three *Ficus* species displayed interspecific variation in the degree of compensatory regrowth following herbivore damage and artificial injury under both fertile and infertile conditions. The significant overcompensatory regrowth of *F. hispida* occurred only in the high nutrient soil (Fig. 2). This pattern has been demonstrated in many other studies [12,29]. However, based on a meta-analysis from Hawkes and Sullivan [41], compensatory regrowth in monocot plants is more common

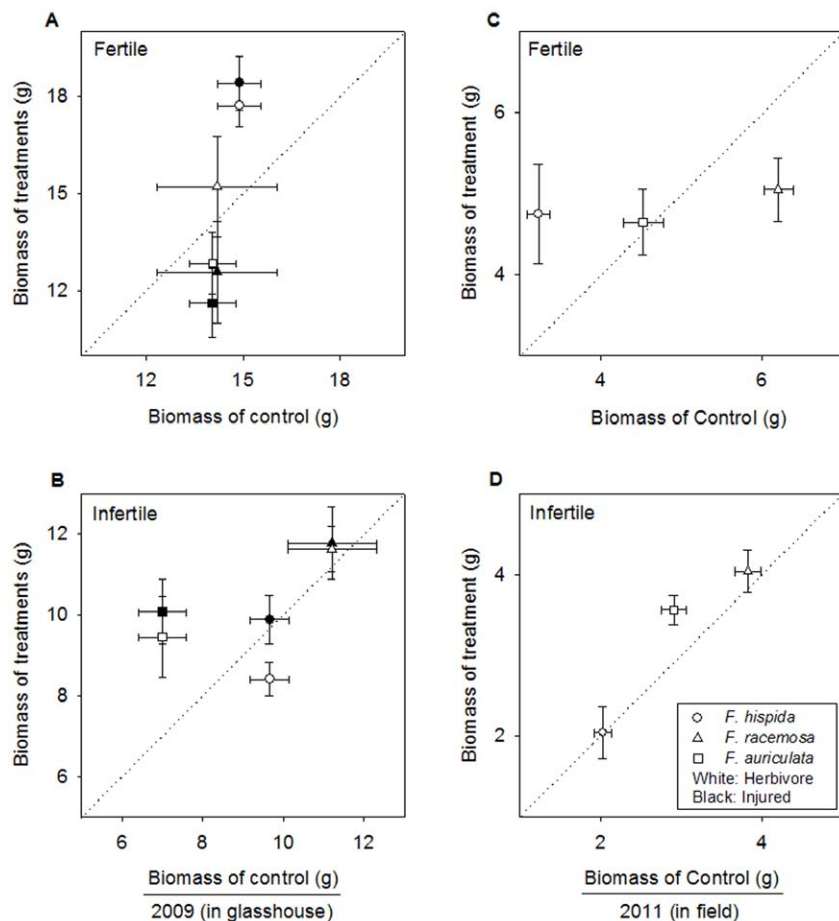


Figure 2. Compensatory growth of three *Ficus* species in response to herbivore treatment. Plant biomass above the line indicates overcompensatory regrowth, on the line indicates full compensatory regrowth, below the line indicates undercompensatory regrowth (Biomass in the damaged state = biomass in control state, slope = 1) (n = 5 in 2009, n = 12 in 2011). doi:10.1371/journal.pone.0045092.g002

when resources are high, but not for dicot herbs and woody plants. In another review [9], out of the 48 cases examined, only 31% showed greater tolerance in high-resource conditions. In contrast to *F. hispida*, overcompensatory regrowth in *F. auriculata* saplings was observed under low rather than high nutrient conditions, which is supported by other studies [9,34].

In this study, both *F. hispida* and *F. auriculata* showed a significant increase in biomass under fertile soil conditions compared to infertile soil conditions (Fig. 2), which is consistent with the finding that soil nutrient level is the limiting factor for these two species. On the other hand, *F. racemosa* did not show a significant change under different soil nutrient levels, which suggests that soil nutrient level is not a limiting factor for this species (Fig. 2). For *F. hispida*, soil nutrient level was the limiting factor, and herbivory on leaves decreased this limiting resource. Eventually, adding the resource (high soil nutrient level) ameliorated the impact of herbivory and increased tolerance. For *F. auriculata*, herbivore feeding might exacerbate carbon limitation, which is limiting under high nitrogen conditions. Therefore, tolerance of *F. auriculata* saplings in fertile soil was lower. In contrast, soil nutrient level was not a limiting factor for the saplings of *F. racemosa*. The consumed leaf area of herbivory insect might be insufficient to affect the utilization of soil nitrogen for *F. racemosa*, or it might only affect other local resources. Accordingly, *F. racemosa* showed equal tolerance at both high and low soil nutrient levels.

Photosynthesis and compensatory regrowth

An increase in photosynthetic capacity following herbivore feeding and artificial injury appeared to be one of the mechanisms for biomass increment in this study. Both herbivore damage and artificial injury enhanced P_{sat} , G_s and C_i in *F. hispida* (in fertile soil) and *F. auriculata* (in infertile soil), thus resulting in the observed increase in RGR (Fig. 1, 2). Herbivory-induced compensatory regrowth is a rather common phenomenon, although other research has indicated that insect herbivory may decrease the photosynthetic capacity in the remaining leaf tissue [42–45]. Compensatory regrowth may result from an increase in carboxylation efficiency or rate of transpiration [46,47]. In this study, C_i of *F. hispida* and *F. auriculata* increased intensely after herbivory feeding and artificial damage which confirmed the increase in utilization of carbon dioxide. Additionally, WUE of *F. hispida* and *F. auriculata* decreased largely following treatments, indicating a trade-off between photosynthesis and the utilization of water [48,49]. Herbivore damage and artificial injury increased the intercellular CO_2 and the rate of transpiration which resulted in the increase of photosynthesis (Fig 4, 5). The sink demand within the leaf may also be affected by herbivore damage, which is well documented in the extensive literature that exists on photosynthetic compensatory regrowth in response to arthropod herbivory [6,45]. Indirect alterations of photosynthesis have been identified across multiple plant systems and can be categorized by plant

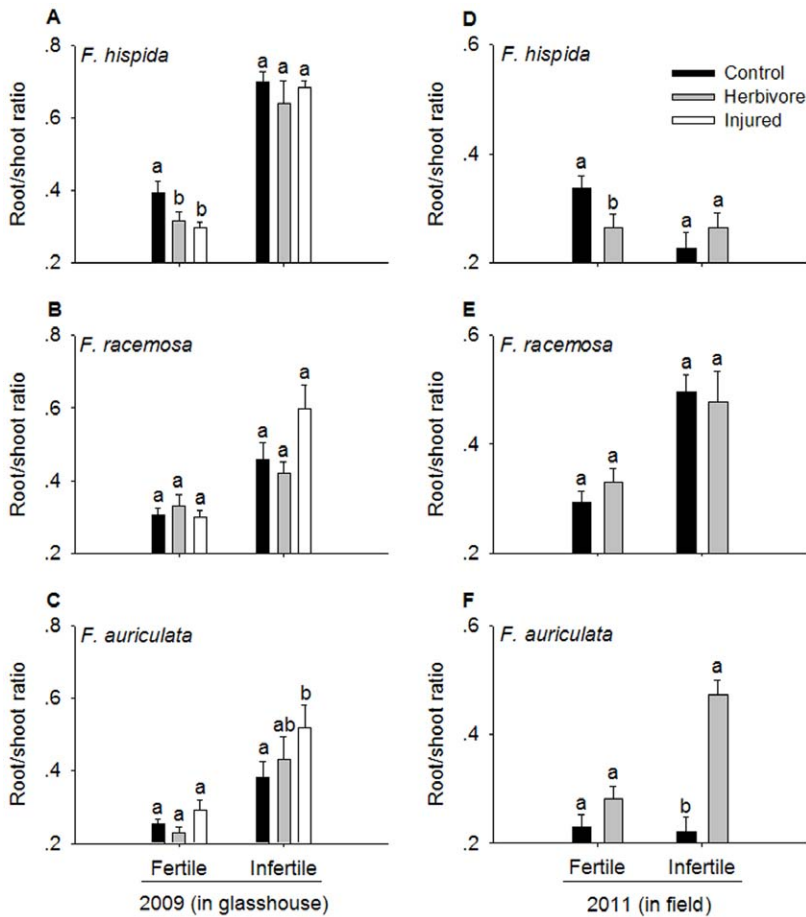


Figure 3. Effect of treatments on root/shoot ratio in *Ficus* saplings under two soil nutrient levels. Significant differences between treatments are marked with different letters (mean \pm SE, n=5 in 2009, n=12 in 2011, $P<0.05$). doi:10.1371/journal.pone.0045092.g003

responses [45]. Precisely how the indirect effect of photosynthesis propagates away from the point of damage remains unknown. While for *F. racemosa*, the results showed that there was no relationship between photosynthesis and RGR and indicated that the full compensatory regrowth of this species might result from modification of other characteristics such as leaf area.

Differences in biomass allocation

The differing biomass allocation of the saplings might be the other mechanism of compensatory regrowth in these three *Ficus* species. In the two species that showed significant overcompensatory regrowth, *F. hispida* and *F. auriculata*, the biomass allocation of the extra growth differed. In *F. hispida*, the enhancement in biomass occurred mostly above ground, while in *F. auriculata*, the enhancement occurred mostly below ground. Many other studies have also shown that the increase in biomass as a result of compensatory response was allocated to shoots [2,20,50]. In *Populus*, defoliation increased shoot biomass even at the expense of decreased root biomass [51].

The compensatory regrowth observed in *F. auriculata* is in contrast with the majority of published data on compensatory regrowth following herbivory [51–54]. The root/shoot ratio of *F. auriculata* increased in response to both herbivore damage and artificial injury, which indicates that this species allocated more resources to root biomass (Fig. 3). This overcompensatory regrowth occurred at the low nutrient level; therefore, an increase

in biomass of the root system may help damaged plants better acquire nutrients for regrowth [5,6,11]. From an ecological viewpoint, a temporary storage of biomass in the roots could be beneficial as a type of herbivore defense.

Herbivore damage vs. artificial injury

Our results showed that both herbivore damage and artificial injury stimulated similar responses in the three *Ficus* species under both fertile and infertile soil conditions. Previous studies have suggested that some plant species respond differently according to damage type. In some plants, defense genes may be responsive to insect feeding but not to physical damage alone [22,23], and such responses can often be mimicked with insect-derived cues found in regurgitant [55–57]. For example, *Nicotiana sylvestris* (Solanaceae) concentrated more jasmonic acid (JA) following herbivore feeding than mechanical damage [24], while some studies have shown that mechanical wounding and herbivore damage induce similar responses [58]. In our study, the three *Ficus* saplings responded to herbivore damage and artificial injury in the same way, indicating that *Ficus* saplings are sensitive to the loss of leaf area. The results also suggest that species may have responded similarly because the insect herbivore is not yet specialized or the plant species have not adapted locally to recognize the presence of this insect damage which has been recently demonstrated that even within a plant-herbivore system, a specific plant population can

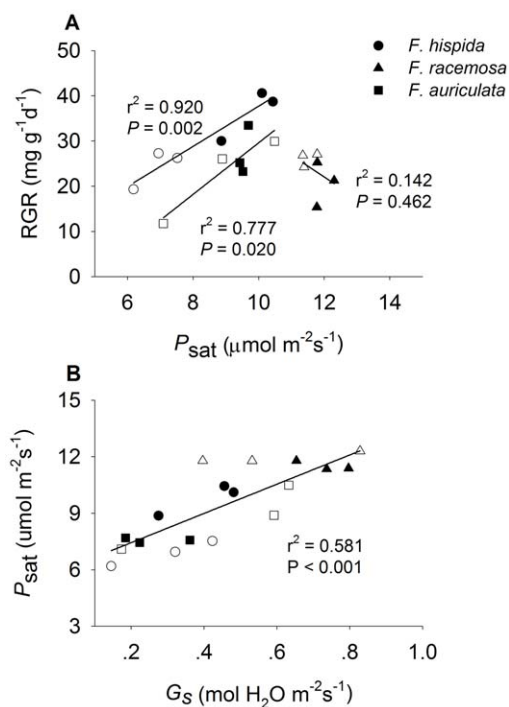


Figure 4. RGR as a function of P_{sat} and P_{sat} as a function of G_s in *Ficus* saplings under two soil nutrient levels. Significant differences between treatments are marked with different letters (mean \pm SE, $n=3$, $P<0.05$). Black shapes, fertile; white shapes, infertile. RGR relative growth rate ($\text{mg}\cdot\text{g}^{-1}\cdot\text{d}^{-1}$); P_{sat} light saturated photosynthetic rate ($\mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$); G_s , stomatal conductance ($\text{mol}\cdot\text{H}_2\text{O}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$). doi:10.1371/journal.pone.0045092.g004

differ in the ability to recognize its local versus foreign herbivore [59].

Conclusion

This study demonstrated that the compensatory regrowth, as a response to herbivory in saplings, and the effect of nutrient levels on such regrowth, varied across different species of the same genus, which is consistent with other studies [60–62]. The increase of photosynthesis and differences in biomass allocation appear to be the mechanisms underlying compensatory regrowth in *Ficus* saplings. The different strategies for three *Ficus* species sapling were consistent in both glasshouse and field experiment, which suggest the mechanism may operate in nature environments. However, our study did not manipulate the effect of water resources, which might limit the WUE. Future studies should consider other potential limiting factors in trying to elucidate the effect of resource availability on the response of plants following damage. *Ficus* species displayed different degrees of compensatory regrowth at the 20th d following damage treatments, and longer-term experiments should be considered to determine whether the degree of compensation varies with time after damage. Tropical rainforests are the most complex territorial ecosystems on the planet, and to explore the mechanisms for the maintenance of complexity and species coexistence in the tropical rainforest is one of the fundamental questions for ecology [12,63]. Future research is needed to explore how saplings utilize different defensive strategies to survive and maximize recruitment in nature and to determine the change in such strategies for different plant species throughout their ontogeny [64].

Materials and Methods

Study site, soil treatment and sapling preparation

All experiments were conducted at the Xishuangbanna Tropical Botanical Garden (XTBG), Chinese Academy of Sciences (21°41'N, 101°25'E; 570 m asl; annual mean temperature, 21.5°C; annual mean rainfall, 1560 mm). The study included two experiments: the first was done in an insect-proof glasshouse and the second was performed in the field. For the glasshouse experiment, all the saplings was placed in an insect-proof and rain-proof glass house. One white and two black nylon shade networks created 10% irradiance, similar to levels of irradiance in field conditions. For the experiment in the field, saplings were placed in a secondary natural forest with canopy coverage being approximately 80–90%.

Two levels of soil nutrient were set up for the experiments. For the low-level nutrient treatment, soil was collected from above the deep soil layer with a total N content of 0.99 g/kg, and for the high-level nutrient treatment, fertilizers (N-P-K = 15-15-15) were added to the low-level soil, yielding a total N content of 1.86 g/kg. The total N content was determined by Dumas combustion analysis [65] using an elemental analyser (Vario MAX CN, Germany) by the Biogeochemical Laboratory of the Kunming Division of the XTBG. All the experiments was conducted in pots (diameter, 20 cm) with one sapling per pot.

Seeds of the three *Ficus* species were collected from approximately 15–30 ripe fruits on 3–5 individual trees (5 trees for *F. hispida* and *F. racemosa*, 3 trees for *F. auriculata*) in the nearby forest. Seeds were germinated in washed sand. After 8 weeks, seedlings of approximately the same size (30~50 cm) from each species were transplanted into the pots and assigned randomly to one of two groups for the soil nutrient treatment.

Herbivore treatment and biomass measurement

About twelve weeks after transplant, five saplings in each soil nutrient condition (i.e., 10 saplings per species) were used to determine the initial biomass (measured separately by shoot and root biomass). The remaining saplings of each species in each soil nutrient condition were used for herbivore compensatory-regrowth experiments.

Third instar caterpillars of a common lepidopteran herbivore of *Ficus* (*Asota caricae* Fabricius) were prepared for the experiment in the glasshouse. The larvae were deprived of food for 24 h preceding the experiment to ensure that damage treatments could be completed within a single day.

The glasshouse experiment was conducted in August, 2009. Leaf damage was simulated for each *Ficus* species in both soil conditions using 5 duplicates as follows: A) Control: Saplings without damage by larvae or hole punch; B) Herbivore damage: Five prepared larvae were placed on the youngest mature leaf for 24 h and covered with insect-proof net; C) Artificial injury by hole punch (5 mm diameter): The first mature leaf was punched six times (we injured the youngest leaf once every 4 h and ensured that the size and shape of the removed area matched that in the herbivore damage treatment). Damage to the first mature leaf was about 30% in both herbivore treatments and artificial injured saplings, equal to about 7% of total leaf mass (more larvae were added if the removed leaf area was less than 30%).

The field experiment was conducted in May 2011. Saplings of each of the three *Ficus* species were planted in both high and low nutrient soils (as described above), and exposed to two treatments: A) Control: herbivore-free, and, B) exposure to natural herbivory. All the saplings of *Ficus* species were departed into four groups. Each of the four groups included 6 saplings of each species in each

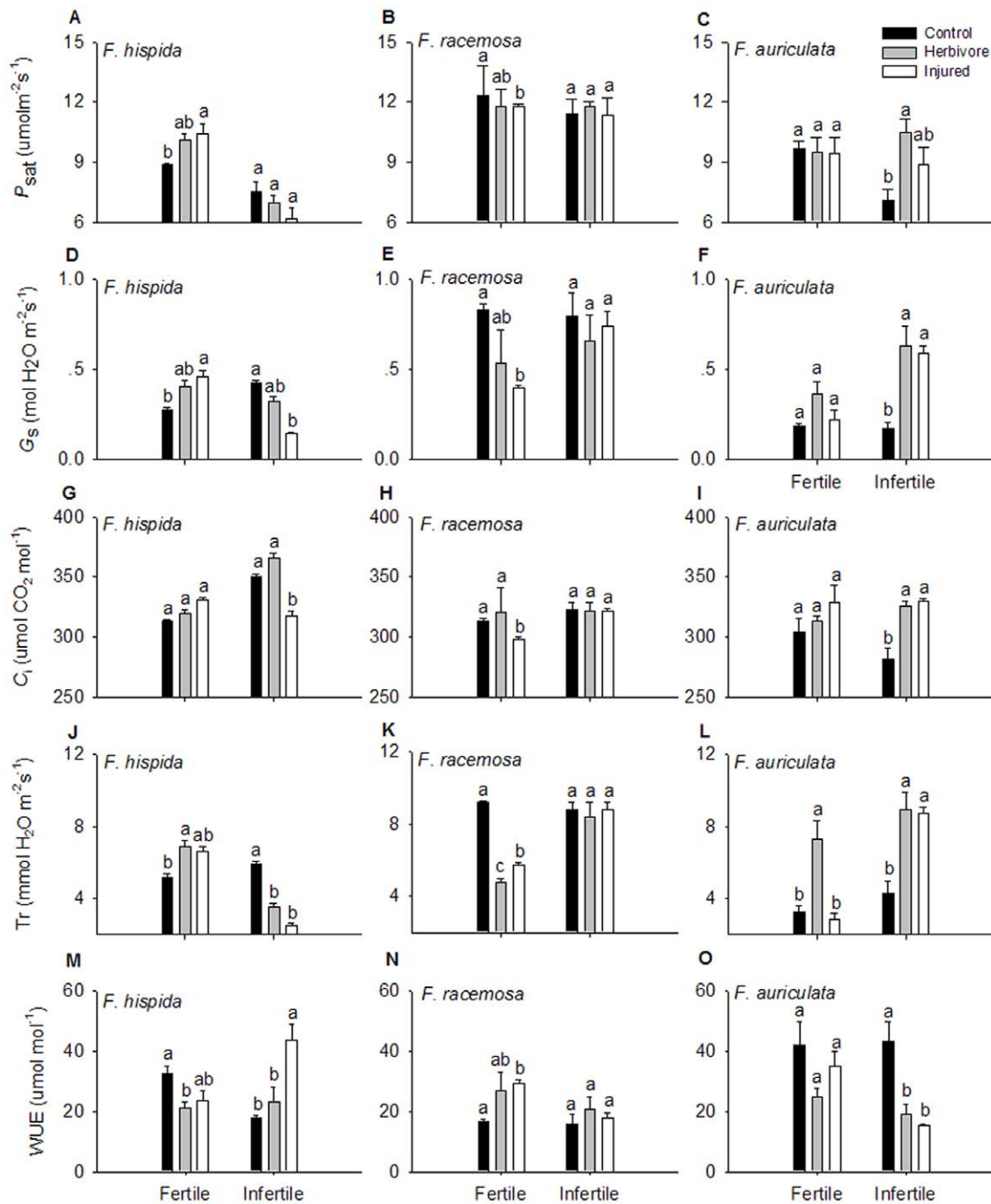


Figure 5. Effect of treatments on photosynthetic characteristics of *Ficus* saplings under two soil nutrient levels. Significant differences between treatments are marked with different letters (mean \pm SE, $n=3$, $P<0.05$). P_{sat} , light saturated photosynthetic rate ($\mu\text{mol m}^{-2}\text{s}^{-1}$); G_s , stomatal conductance ($\text{mol H}_2\text{O m}^{-2}\text{s}^{-1}$); WUE, water use efficiency ($\mu\text{mol mol}^{-1}$). doi:10.1371/journal.pone.0045092.g005

soil nutrient condition, within a group, placed >1 m apart, and at >30 m distances between groups.

All saplings were harvested 20 days after treatment both in the glasshouse and in the field. Leaf area consumed by natural herbivore (control) were measured by a LI-3000 portable area meter (Li-Cor, Lincoln, USA). Soil was cleaned from the roots. The saplings were oven-dried at 40°C until constant masses were reached and then were separated into shoot and root. Total

biomass was measured and the relative growth rate (RGR) was determined by the following equation [66]:

$$\text{RGR (mg}\cdot\text{g}^{-1}\cdot\text{d}^{-1}) = [\ln(\text{sapling mass at harvest}) - \ln(\text{initial sapling mass})] / [\text{duration of study (d)}].$$

The degree of compensatory regrowth was determined by comparing the total biomass of damaged and undamaged saplings. If the biomasses of damaged and undamaged saplings are similar, we can infer full compensatory regrowth of damaged saplings,

higher indicates overcompensatory regrowth and less indicates under compensatory regrowth.

Photosynthesis measurement

Photosynthetic characteristics were measured only in the glasshouse experiment using two Li-6400 portable photosynthesis systems (Li-Cor, Lincoln, Nebraska, USA) ($n = 3$). Three untreated leaves were measured on each sapling. The measurements were taken 20 days after the herbivore treatment, between 0800 and 1400 on 15 August. In the leaf chamber, each leaf was acclimated to $500 \mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$ Photosynthetic Photon Flux Density (PPFD) for 1–3 min and then to $1000 \mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$ for 1–3 min prior to gas exchange measurements. This method was found to induce plant photosynthesis without causing photoinhibition. Light saturated photosynthetic rates (P_{sat}), stomatal conductance (G_s), internal leaf CO_2 concentration (C_i), and transpiration rate (Tr) were recorded when the sample leaf was balanced for 200 s under saturated PPFD. Water use efficiency (WUE) was calculated as the ratio of P_{sat} to G_s . During photosynthesis measurements, cuvette air was maintained at $27\sim 29^\circ\text{C}$, $70\sim 80\%$ relative humidity, and $360 \mu\text{mol}\cdot\text{mol}^{-1}$ sample CO_2 partial pressure.

Statistical analysis

Three-way ANOVA was performed to test the effects of species, soil nutrient level, treatments, and all possible interactions between these factors on growth, biomass allocation and photosynthetic characters (The data of RGR, root/shoot ratio, P_{max} , C_i , Tr and WUE were normally distributed and homogenous. Data of G_s were log₁₀-transformed which made the residuals reasonably normal and homogenous. And the data of one sapling of *F. racemosa* which was totally consumed in field were deleted to avoid bias of results.). Because the effect of interactions between these factors on these variables were significant, one-way analysis and multi-comparison (Tukey-HSD) of variance was performed to test

treatment effects on RGR, root/shoot ratio and photosynthetic characters for each species under each soil nutrient level. To determine whether soil nutrient level was the limiting resource for all three *Ficus* saplings, *t*-tests were performed on the total mass of the undamaged saplings of each species. One-way ANCOVA was used to test differences among the three *Ficus* species in the correlation between P_{sat} and RGR, P_{sat} and G_s , with species as a fixed factor, and variables indicated by y - and x -axes as dependent variable and covariate, respectively. If the difference was significant, we then tested for the significance of the correlation (Pearson correlation, two-tailed) for the three *Ficus* species separately; otherwise, we pooled data from all species to test for the significance of correlation. To evaluate the effects of species, soil nutrient level and their interactions on consumed leaf area in the field experiment, two-way ANOVA and multi-comparison (Tukey-HSD) were performed, where the average consumed leaf area of each species with each soil nutrient level for each group were analyzed ($n = 4$). The data were normally distributed and homogenous. All statistical analyses were performed using SPSS (SPSS 13.0, Chicago, USA).

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Author Contributions

Conceived and designed the experiments: JZ JC. Performed the experiments: JZ. Analyzed the data: JZ JC. Contributed reagents/materials/analysis tools: JZ. Wrote the paper: JZ JC.

References

- McNaughton SJ (1983) Compensatory plant growth as a response to herbivory. *Oikos* 40: 329–336.
- Strauss SY, Agrawal AA (1999) The ecology and evolution of plant tolerance to herbivory. *Trends Ecol Evol* 14: 179–185.
- Stowe KA, Marquis RJ, Hochwender CG, Simms EL (2000) The evolutionary ecology of tolerance to consumer damage. *Ann Rev Ecol Syst* 31: 565–595.
- Foroni J (2011) Ecological and evolutionary implications of plant tolerance to herbivory. *Functional Ecology* 25: 399–407.
- Chapin III FS, McNaughton SJ (1989) Lack of compensatory growth under phosphorus deficiency in grazing-adapted grasses from the Serengeti Plains. *Oecologia* 79: 551–557.
- Trumble JT, Kolodny-Hirsch DM, Ting IP (1993) Plant compensation for arthropod herbivory. *Ann Rev Entomol* 38: 93–119.
- Tiffin P (2000) Mechanisms of tolerance to herbivore damage: what do we know? *Ecol* 14: 523–536.
- Wise MJ, Abrahamson WG (2005) Beyond the compensatory continuum: environmental resource levels and plant tolerance of herbivory. *Oikos* 109: 417–428.
- Wise MJ, Abrahamson WG (2007) Effects of resource availability on tolerance of herbivory: a review and assessment of three opposing models. *Am Nat* 169: 443–454.
- Lehtilä K, Haukioja E, Kaitaniemi P, Laine KA (2000) Allocation of resources within mountain birch canopy after simulated winter browsing. *Oikos* 90: 160–170.
- Glynn C, Herms DA, Egawa M, Hansen R, Mattson WJ (2003) Effects of nutrient availability on biomass allocation as well as constitutive and rapid induced herbivore resistance in poplar. *Oikos* 101: 385–397.
- Coley PD, Bryant JP, Chapin FS (1985) Resource availability and plant antiherbivore defense. *Science* 230: 895–899.
- Gleeson SK, Tilman D (1994) Plant allocation, growth rate, and successional status. *Funct Ecol* 8: 543–550.
- Van der Meijden E, Wijn M, Verkaar HJ (1988) Defense and regrowth, alternative plant strategies in the struggle against herbivores. *Oikos* 51: 355–363.
- Leps J, Novotny V, Basset Y (2001) Habitat and successional status of plants in relation to the communities of their leafchewing herbivores in Papua New Guinea. *J Ecol* 89: 186–199.
- Edwards-Jones G, Brown VK (1993) Successional trends in insect herbivore population densities: a field test of a hypothesis. *Oikos* 66: 463–471.
- Coley PD (1988) Effects of plant growth on leaf lifetime and on the amount and type of antiherbivore defense. *Oecologia* 74: 531–536.
- Grime JP (1977) Evidence for the existence of three primary strategies in plants and its relevance to ecological and evolutionary theory. *Am Nat* 111: 1169–1194.
- Coley PD (1983) Herbivory and defensive characteristics of tree species in a lowland tropical forest. *Ecol Monogr* 53: 209–233.
- Maschinski J, Whitham TG (1989) The continuum of plant responses to herbivory: the influence of plant association, nutrient availability and timing. *Am Nat* 134: 1–19.
- Pinkard EA, Battaglia M, Mohammed CL (2007) Defoliation and nitrogen effects on photosynthesis and growth of *Eucalyptus globulus*. *Tree Physiol* 27: 1053–1063.
- Hermesmeier D, Schittko U, Baldwin IT (2001) Molecular interactions between the specialist herbivore *Manduca sexta* (Lepidoptera, Sphingidae) and its natural host *Nicotiana attenuata*. I. Large-scale changes in the accumulation of growth- and defense-related plant mRNAs. *Plant Physiol* 125: 683–700.
- Reymond P, Bodenhausen N, Van Poecke RMP, Krishnamurthy V, Dicke M, et al. (2004) A conserved transcript pattern in response to a specialist and a generalist herbivore. *Plant Cell* 16: 3132–3147.
- McCloud ES, Baldwin IT (1997) Herbivory and caterpillar regurgitants amplify the wound-induced increases in jasmonic acid but not nicotine in *Nicotiana sylvestris*. *Planta* 203: 430–435.
- Belsky AJ, Carson WP, Jensen CL (1993) Overcompensation by plants: herbivore optimization or red herring? *Evol Ecol* 7: 109–121.
- Rosenthal JP, Kotanen PM (1994) Terrestrial plant tolerance to herbivory. *Trends Ecol Evol* 9: 145–148.
- Mutikainen P, Walls M (1995) Growth, reproduction and defence in nettles: Responses to herbivory modified by competition and fertilization. *Oecologia* 104: 487–495.
- Fay PA, Hartnett DC, Knapp AK (1996) Plant tolerance of gall-insect attack and gall-insect performance. *Ecology* 77: 521–534.
- Crawley MJ (1997) *Plant ecology*. Blackwell, Oxford, p. 736.

30. Pilson D (2000) The evolution of plant response to herbivory: si- multaneously considering resistance and tolerance in *Brassica rapa*. *Evol Ecol* 14: 457–489.
31. Meyer GA, Root RB (1993) Effects of herbivorous insects and soil fertility on reproduction of goldenrod. *Ecology* 74: 1117–1128.
32. Ferraro DO, Oesterheld M (2002) Effect of defoliation on grass growth. A quantitative review. *Oikos* 98: 125–133.
33. Rand TA (2004) Competition, facilitation, and compensation for insect herbivory in an annual salt marsh forb. *Ecology* 85: 2046–2052.
34. Hilbert DW, Swift DM, Detling JK, Dyer MI (1981) Relative growth rates and the grazing optimization hypothesis. *Oecologia* 51: 14–18.
35. Haukioja E, Koricheva J (2000) Tolerance to herbivory in woody vs. herbaceous plants. *Evol Ecol* 14: 551–562.
36. Kozlowski TT (1992) Carbohydrate sources and sinks in woody plants. *Bot Rev* 58: 107–222.
37. Bassman JH, Dickmann DI (1982) Effects of defoliation in the developing leaf zone on young *Populus neuramericana* plants. I. Photosynthetic physiology, growth, and dry weight partitioning. *Forest Sci* 28:599–612.
38. Anttonen S, Piispanen R, Ovaska J, Mutikainen P, Saranpää P, et al. (2002) Effects of defoliation on growth, biomass allocation, and wood properties of *Betula pendula* clones grown at different nutrient levels. *Can J Forest Res* 32: 498–508.
39. Xiang H, Chen J (2004) Interspecific variation of plant traits associated with resistance to herbivory among four species of *Ficus* (Moraceae). *Ann Bot* 94: 377–384.
40. Xia SW, Chen J (2007) Effect upon volatiles release in five *Ficus* (Moraceae) species by herbivores and artificial damage. *Acta Botanica Yunanica* 29: 694–700.
41. Hawkes CV, Sullivan JJ (2001) The impact of herbivory on plants in different resource conditions: a meta-analysis. *Ecology* 82: 2045–2058.
42. Zangerl AR, Hamilton JG, Miller TJ, Crofts AR, Oxborough K, et al (2002) Impact of folivory on photosynthesis is greater than the sum of its holes. *PNAS* 99: 1088–1091.
43. Aldea M, Hamilton JG, Resti JP, Zangerl AR, Berenbaum MR, et al (2006) Comparison of photosynthetic damage from arthropod herbivory and pathogen infection in understory hardwood samplings. *Oecologia* 149: 221–232.
44. Dungan RJ, Turnbull MH, Kelly D (2007) The carbon costs for host trees of a phloem-feeding herbivore. *J Ecol* 95: 603–613.
45. Nability PD, Zavala JA, DeLucia EH (2009) Indirect suppression of photosynthesis on individual leaves by arthropod herbivory. *Ann Bot* 103: 655–663.
46. Thomson VP, Cunningham SA, Ball MC, Nicotra AB (2003) Compensation for herbivory by *Cucumis sativus* through increased photosynthetic capacity and efficiency. *Oecologia* 134: 167–175.
47. Ozaki K, Saito H, Yamamuro K (2004) Compensatory photosynthesis as a response to partial debudding in *ezo* spruce, *Picea jezoensis*, seedlings. *Ecol Res* 19: 225–231.
48. Cowan IR (1982) Regulation of water use in relations to carbon gain in higher plants. Lange OL, Nobel PS, Osmond CB, Ziegler H. *Encyclopedia of plant physiology*. Springer, New York, New York, USA. p. 589–613.
49. Zeiger EJ, Farquhar GD, Cowan IR (1987) Stomatal function. Stanford University Press, Stanford, California, USA.
50. Mabry CM, Wayne PW (1997) Defoliation of the annual herb *Abutilon theophrasti*: mechanisms underlying reproductive compensation. *Oecologia* 111: 225–232.
51. Snyder KA, Williams D (2007) Root allocation and water uptake patterns in riparian tree saplings: responses to irrigation and defoliation. *Forest Ecol Manag* 246: 222–231.
52. Hermans C, Hammond JP, White PJ, Verbruggen N (2006) How do plants respond to nutrient shortage by biomass allocation? *Trends Plant Sci* 11: 1360–1385.
53. Barton KE (2008) Phenotypic plasticity in seedling defense strategies: compensatory growth and chemical induction. *Oikos* 117: 917–925.
54. Eyles A, Pinkard EA, Mohammed C (2009) Shifts in biomass and resource allocation patterns following defoliation in *Eucalyptus globulus* growing with varying water and nutrient supplies. *Tree Physiol* 29: 753–764.
55. Schittko U, Hermsmeier D, Baldwin IT (2001) Molecular interactions between the specialist herbivore *Manduca sexta* (Lepidoptera, Sphingidae) and its natural host *Nicotiana attenuata*. II. Accumulation of plant mRNAs in response to insect-derived cues *Plant Physiol* 125: 701–710.
56. Halitschke R, Gase K, Hui DQ, Schmidt DD, Baldwin IT (2003) Molecular interactions between the specialist herbivore *Manduca sexta* (Lepidoptera, Sphingidae) and its natural host *Nicotiana attenuata*. VI. Microarray analysis reveals that most herbivore-specific transcriptional changes are mediated by fatty acid-amino acid conjugates. *Plant Physiol* 131: 1894–1902.
57. Roda A, Halitschke R, Steppuhn A, Baldwin IT (2004) Individual variability in herbivore-specific elicitors from the plant's perspective. *Mol Ecol* 13: 2421–2433.
58. Green TR, Ryan CA (1971) Wound-induced proteinase inhibitor in plant leaves: a possible defense mechanism against insects. *Science* 175: 776–777.
59. Garrido E, Andraca-Gomez G, Fornoni J (2012) Local adaptation: simultaneously considering herbivores and their host plants. *New Phytologist* 193: 445–453.
60. Hoffmann WA, Franco AC (2003) Comparative growth analysis of tropical forest and savanna woody plants using phylogenetically independent contrasts. *J Ecol* 91: 475–484.
61. Sixto H, Grau JM, Alba N (2005) Response to sodium chloride in different species and clones of genus *Populus* L. *Forestry* 78: 93–104.
62. Valladares F, Sánchez-Gómez D (2006) Ecophysiological traits associated with drought in Mediterranean tree seedlings: individual responses versus interspecific trends in eleven species. *Plant Biology* 8: 688–697.
63. Eichhorn MP, Nilus R, Compton SG, Hartley SE, Burslem DFRP (2010) Herbivory of tropical rain forest tree seedlings correlates with future mortality. *Ecology* 91: 1092–1101.
64. Boege K, Marquis RJ (2005) Facing herbivory as you grow up: the ontogeny of resistance in plants. *Trends in Ecology and Evolution* 20: 441–448.
65. Dumas JBA (1831) *Procédes de lanalyse organique*. *Annales de Chimie et de Physique* 247: 198–213.
66. Evans GC (1972) *The Quantitative Analysis of Plant Growth*. Oxford: Blackwell Scientific Publications, 734.