Spatial and Temporal Variability of Macroinvertebrates in Spawning and Non-Spawning Habitats during a Salmon Run in Southeast Alaska

Emily Y. Campbell^{1*}, Richard W. Merritt¹, Kenneth W. Cummins², M. Eric Benbow^{1,3}

1 Department of Entomology, Michigan State University, East Lansing, Michigan, United States of America, **2** Department of Fisheries Biology, Humboldt State University, Arcata, California, United States of America, **3** Department of Biology, University of Dayton, Dayton, Ohio, United States of America

Abstract

Spawning salmon create patches of disturbance through redd digging which can reduce macroinvertebrate abundance and biomass in spawning habitat. We asked whether displaced invertebrates use non-spawning habitats as refugia in streams. Our study explored how the spatial and temporal distribution of macroinvertebrates changed during a pink salmon (*Oncorhynchus gorbuscha*) spawning run and compared macroinvertebrates in spawning (riffle) and non-spawning (refugia) habitats in an Alaskan stream. Potential refugia included: pools, stream margins and the hyporheic zone, and we also sampled invertebrate drift. We predicted that macroinvertebrates would decline in riffles and increase in drift and refugia habitats during salmon spawning. We observed a reduction in the density, biomass and taxonomic richness of macroinvertebrates in riffles during spawning. There was no change in pool and margin invertebrate communities, except insect biomass declined in pools during the spawning period. Macroinvertebrate density was greater in the hyporheic zone and macroinvertebrate density and richness increased in the drift during spawning. We observed significant invertebrate declines within spawning habitat; however in non-spawning habitat, there were less pronounced changes in invertebrate density and richness. The results observed may be due to spawning-related disturbances, insect phenology, or other variables. We propose that certain in-stream habitats could be important for the persistence of macroinvertebrates during salmon spawning in a Southeast Alaskan stream.

Citation: Campbell EY, Merritt RW, Cummins KW, Benbow ME (2012) Spatial and Temporal Variability of Macroinvertebrates in Spawning and Non-Spawning Habitats during a Salmon Run in Southeast Alaska. PLoS ONE 7(6): e39254. doi:10.1371/journal.pone.0039254

Editor: Björn Brembs, Freie Universitaet Berlin, Germany

Received: December 28, 2011; **Accepted:** May 22, 2012; **Published:** June 20, 2012

Copyright: © 2012 Campbell et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Funding: This research was supported by the USDA-CSREES National Research Initiative Competitive Grants Program (Ecosystems Program 2006-35101-16566) and the USDA Forest Service Pacific Northwest Research Station (Aquatic-Land Interactions Program). The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing Interests: The authors have declared that no competing interests exist.

* E-mail: campb613@msu.edu

‡ Current address: Department of Fisheries and Wildlife, Oregon State University, Corvallis, Oregon, United States of America

Introduction

The idea of the refuge is critically important and some argue that it should be considered an integrating concept in ecology and evolution [1]. This is because it encompasses a variety of phenomena including: enemy-free space, cover, crypsis, functional responses, factors of predator and prey behavior, competition for resources and shelter [1]. Past investigations of macroinvertebrate utilization of refugia in response to disturbance have largely focused on flow [2–4], while the use of refugia in response to salmon spawning is less well studied [5]. Macroinvertebrate distributional changes during salmon spawning could have important implications for the management and conservation of benthic macroinvertebrates and the organisms that consume them, both within streams and the adjacent riparian forests.

As anadromous and semelparous organisms, Pacific salmon (*Oncorhynchus* spp.) offer annual nutrient pulses to streams when they return to spawn and these subsidies can have ecological effects on lotic organisms over time [6,7]. Salmon resource subsidies have been documented to positively influence benthic macroinvertebrates [5,8], perhaps by the provision of nutrients and carbon that have been shown to increase during salmon runs

[9]. Stream environmental factors such as sediment size and large wood recruitment [9–11] along with spawning disturbance intensity [5,12] can alter the degree of marine-derived nutrient transfer available to benthic communities.

In strong contrast to the nutrient enrichment effect, adult salmon disturb benthic communities in spawning habitat by redistributing substrata during redd (nest) construction [5,14]. Spawning disturbances can change the distribution, abundance and community composition of macroinvertebrates, causing substantial reductions in riffles where the redds are built [15–18]. Salmon redd construction can cause disturbances locally by displacing macroinvertebrates into drift at the excavation site due to suction and frictional forces; and also can disturb invertebrates downstream of the excavation site as displaced fine sediments can abrade and fill in interstitial spaces [5].

Habitat heterogeneity in streams is important as it sustains high biological diversity [13,19,20]. For example, riffles are often dominated by scrapers, such as heptageniid mayflies, that feed on benthic biofilm and collector filterers, such as simuliid dipterans, that collect fine particulate organic matter from stream drift. Pools however, typically sustain more shredders, such as limnephilid case

building caddisflies, and collector gatherers, such as some genera of Chironomidae [21]. Heterogeneity in streams can also offer organisms refugia, which we define here as distinct non-spawning habitats that are not normally disturbed [22,23].

Areas that are less likely to be influenced by spawners include: pools or other slack water habitats unusable for salmon eggs [7], stream margins which are too shallow for redd construction [2], and the hyporheic zone which is too deep to be disturbed [5,7]. Macroinvertebrates could use such habitats as refugia, avoiding the effects of bioturbation [14]. Organisms may inhabit refugia temporarily during disturbances and then disperse once it has passed [22,24,25], or they may stay within refugia after disturbances.

We investigated the spatial and temporal variability of macroinvertebrates in spawning and non-spawning habitats in a stream where salmon were spawning. We hypothesized that during spawning there would be: i) a reduction in the density, biomass and taxonomic richness of macroinvertebrates in riffles; ii) increased macroinvertebrate density, biomass and taxonomic richness in pools, margins and the hyporheic zone; and iii) increased daytime macroinvertebrate drift.

Results

The estimated density of adult salmon in our 300 m reach was 0.88 m^{-2} during this study (2008), compared to 0.51 m^{-2} in 2007 and 0.36 m^{-2} in 2006 [10,11]. Chlorophyll *a* was not statistically tested, but we observed a decline in pools and an increase in riffles and margins during the salmon run (Table 1).

Macroinvertebrates in Riffles

Macroinvertebrate assemblages in riffles significantly changed during salmon spawning. Regression analysis showed that invertebrate density was negatively correlated with that of salmon density ($R^2 = 0.84$, $p < 0.001$). A sharp decline was observed in macroinvertebrate density and insect biomass in riffles upon the arrival of salmon spawners (Figs. 1a, 1b). Repeated measures analysis of variance (rmANOVA) showed that macroinvertebrate density ($p < 0.001$, Fig. 2a), insect biomass ($p < 0.001$, Fig. 2b) and taxonomic richness ($p = 0.009$, Fig. 2c) declined in riffles during spawning.

We observed a change in functional feeding group structure in riffles during spawning. The density of collector-gatherers ($p < 0.001$, Fig. 3b), collector-filterers ($p < 0.001$, Fig. 3c) and scrapers ($p < 0.001$, Fig. 3d) declined in riffles during spawning. Genus-specific changes were also observed during spawning in riffle habitats (Fig. 4a–f). The dominant taxa were: *Chironomus* (19.9% of total taxa), *Swelsta* (7.6%), *Ameletus* (5.6%), *Baetis* (5.3%), *Cinygmula* (5.1%) and *Suwallia* (3.2%). *Swelsta* density ($p < 0.001$) was the only genus that increased in riffles during the salmon run (Figs. 4e). *Ameletus* ($p = 0.019$), *Baetis* ($p = 0.038$),

Cinygmula ($p < 0.001$) and *Suwallia* ($p < 0.001$) densities all declined, whereas *Chironomus* density did not change in riffles during spawning (Figs. 4a–f).

Macroinvertebrates in Drift

Drift density was positively correlated with salmon density ($R^2 = 0.61$, $p < 0.01$). The density ($p = 0.049$, Fig. 1c, Fig. 2a) and taxonomic richness ($p = 0.001$; Fig. 2c) of macroinvertebrates increased in stream drift during salmon spawning. The densities of collector-gatherers ($p = 0.05$, Fig. 3b), collector-filterers ($p = 0.02$, Fig. 3c) and predators ($p < 0.001$, Fig. 3e) increased in stream drift during spawning. *Swelsta* density ($p < 0.001$, Fig. 4e) increased, while all other dominant genera did not significantly change in stream drift during spawning.

Community Structure

The NMDS ordination and MRPP revealed a significant ($p < 0.001$) difference in macroinvertebrate community structure before and during the run (Fig. 5). A total of 83% of the variation in macroinvertebrate community structure was explained by a three axes solution: 1st axis = 36%, 2nd = 29% and 3rd = 18% and the mean stress was 15.3. We ran separate ordinations for the invertebrate assemblages in all habitat types before and during spawning, and did not find a significant difference. We therefore show the more robust ordination comparing overall before and during salmon invertebrate communities. The dominant indicator taxa in each habitat type before and during the salmon run are listed in Table 2.

Macroinvertebrates in Refugia

Macroinvertebrate density in margins, pools and the hyporheic zone were not correlated to salmon density. Within stream margins, macroinvertebrate density, biomass and taxonomic richness did not change (Figs. 2a–c). In pool habitats, the biomass of insects declined ($p = 0.002$, Fig. 2b), while invertebrate density (Fig. 2a) and richness (Fig. 2c) did not change during spawning. The hyporheic zone showed an increase in invertebrate density ($p = 0.019$, Fig. 2a) but no change in insect biomass (Fig. 2b) or taxonomic richness (Fig. 2c) during the salmon run.

Functional feeding group structure changed in non-spawning habitats during the salmon run. In stream margins, there was an increase in shredder ($p = 0.02$, Fig. 3a) and predator ($p < 0.001$, Fig. 3e) densities during the salmon run. In the hyporheic zone, collector-gatherer ($p = 0.007$, Fig. 3b) and predator ($p = 0.02$, Fig. 3e) densities increased. In stream margins, *Baetis* ($p < 0.001$), *Cinygmula* ($p = 0.012$) and *Suwallia* ($p < 0.001$) densities declined, while *Swelsta* ($p < 0.001$) density increased during the salmon run (Figs. 4b–e). In pools, the density of *Suwallia* ($p < 0.001$) declined and the densities of *Swelsta* ($p < 0.001$) and *Chironomus* ($p < 0.001$)

Table 1. Characteristics of riffles, margins and pools in Twelve Mile Creek, Alaska.

	Avg. Area (m ²)	Avg. Temp. (°C)	% Dissolved Oxygen	pH	Mean Sediment Size (mm)	% Canopy Cover	Mean Chlorophyll <i>a</i> (mg m ⁻²) Before Salmon	Mean Chlorophyll <i>a</i> (mg m ⁻²) During Salmon
Riffle	185.2 (0.25)	9.7 (0.09)	99.2 (0.54)	8.1 (0.02)	30.9 (3.03)	19 (5.79)	8.4 (1.22)	10.3 (1.15)
Margin	N/A	10.8 (0.54)	97.9 (0.85)	8.1 (0.06)	21.9 (5.1)	26 (7.04)	10.1 (1.8)	10.2 (1.6)
Pool	35.5 (0.23)	9.8 (0.16)	85.3 (9.7)	7.8 (0.2)	23.8 (3.49)	69 (12.39)	14.9 (1.78)	10.3 (1.08)

Means are presented and numbers in parenthesis represent standard errors.
doi:10.1371/journal.pone.0039254.t001

Figure 1. Invertebrates in riffles and drift throughout the salmon run. Invertebrate density (a) and insect biomass (b) in riffles, and invertebrate density (c) and insect biomass (d) in drift plotted against salmon density (right-hand axis). Note the differences in scale on the y-axis in all figures.

doi:10.1371/journal.pone.0039254.g001

increased (Figs. 4d–f). In the hyporheic zone, *Swelisia* ($p = 0.004$) density declined and the density of *Swelisia* ($p < 0.001$) increased during the run (Figs. 4d–e).

Discussion

In this study, we quantified macroinvertebrate distributional changes in spawning (riffle) and non-spawning (pool, margin, hyporheic) habitats during a pink salmon (*Oncorhynchus gorbuscha*) spawning run. Consistent with the results of previous studies, macroinvertebrate density, biomass and taxonomic richness declined in riffles during the salmon run [5,11,13,26–28]. We speculate that the precipitous decline in riffle macroinvertebrate density and biomass was due to spawning-related activities. This explanation is bolstered by a controlled field experiment where we installed mesh enclosures in spawning habitat that prevented salmon disturbance, which were compared to areas where salmon had access. We found a greater abundance of macroinvertebrates in enclosure plots relative to control plots during spawning, suggesting that salmon were the causal mechanism for the observed invertebrate decline within spawning habitat [29].

We observed an increase in macroinvertebrate drift density, biomass and taxonomic richness during spawning as compared to pre-salmon related drift. This effect has been demonstrated in several other studies [5,15,30]. The sharp increase in drift density and biomass that occurred shortly after salmon arrival in this study

is likely due to salmon nest construction which displaced macroinvertebrates into drift at the excavation site due to suction and frictional forces [5]. There are other possible explanations however, for example, macroinvertebrates in Alaska may have prolonged drift periods in the summertime due to the long photoperiod causing higher drift densities. Additionally, the flood that occurred on 23 August could have accounted for the observed increase in invertebrate drift.

In comparison to riffle habitats, the macroinvertebrate communities in pool, margin and hyporheic habitats remained relatively stable throughout the salmon run. Invertebrate density and richness did not change in stream margins and pools, but insect biomass declined in pools during the salmon run. We propose that this was due to an emergence event. Insect emergence is high in the late summertime in Alaska and in high latitude streams in general, so it is possible that natural emergence could explain the reduction of insect biomass in pools during the run [30]. Overall and taxa-specific invertebrate density increased in the hyporheic zone during spawning, these results may be explained by invertebrates avoiding the physical disturbances from salmon spawners, but could also be due to insect life history cycles or food limitations in disturbed riffles causing invertebrates to vertically migrate into the hyporheic zone to feed.

Disturbance is a central organizing factor in stream communities [31] and is fundamental to the concept of patch dynamics, whereby the temporal and spatial variability of ecosystems are established by disturbance impacts [32]. We demonstrate that

Figure 2. Invertebrates in all habitats before and during spawning. Invertebrate density (a), insect biomass (b) and taxonomic richness (c) before (dark grey bars) and during (white bars) the salmon run in riffles, pools, margins, the hyporheic zone, and drift. Graphs show ± 1 s.e. and * means $p < 0.05$. doi:10.1371/journal.pone.0039254.g002

invertebrate assemblages undergo spatial shifts within the stream channel during spawning and suggest that adult salmon are the cause of the observed invertebrate reductions in patches (riffles) where spawning activities are greatest [15,17]. Managers can use these data as incentive to maintain channel complexity which may be an important factor regulating invertebrate persistence during disturbances [11]. The concept of a refuge is important in both basic and applied ecology, particularly as a stabilizing force during natural and anthropogenic disturbances [1]. Certain in-stream habitats may offer refuge to macroinvertebrates and be a fundamental determinant of macroinvertebrate resilience to bioturbation from spawning salmon in Southeast Alaska.

Materials and Methods

Study Sites

This study was conducted within a 300 m reach of Twelve Mile Creek (N55°482, W132°631) on Prince of Wales Island within the Tongass National Forest, Southeast Alaska, USA (Table 1). Catchments on Prince of Wales Island are composed of coniferous temperate rainforest that have been managed primarily for timber harvest. Much (68%) of the Twelve Mile Creek catchment has been harvested for timber, mostly in the 1960's. Dominant riparian tree species along the stream include red alder (*Alnus rubra* (Bong)), western hemlock (*Tsuga heterophylla* (Rafinesque)) and Sitka spruce (*Picea sitchensis* (Bongard)).

Macroinvertebrate Sampling and Processing

Macroinvertebrate samples were collected as five replicates each from riffles, pools, stream margins and hyporheic wells every 10 days from 27 June until 20 September 2008. The 10 sampling dates included five before the salmon arrived and five during their spawning. A flood prevented sampling on 23 August.

For riffles, pools and margins, benthic macroinvertebrates were collected using a large PVC pipe sampler (diameter, 36 cm; area, 0.4 m²). The pipe was pushed down into the sediments (approx. 10–14 cm) to minimize water exchange. Samples were collected by disturbing the substratum within the pipe to a depth of about 10 cm for 30 s. A 250 μ m mesh net was then used to filter out organic matter and invertebrates and the material collected was preserved in 70% ethanol. A stream 'margin' was defined as the periphery of the channel with a width approximately equal to 10% the total stream width (mean channel width 13.05 m). 'Pools' were defined as low-velocity (0–0.02 m/s) areas caused by large wood debris or boulders, that were connected to the main channel at base flow, were not too deep (<0.65 m) and were adjacent to spawning habitats.

Hyporheic macroinvertebrates were collected using 0.6 m long PVC hyporheic wells installed 30 cm into the sediment at random within each riffle (1 well per riffle, 5 wells total). All wells were capped to avoid surface and water column invertebrates from intruding. The bottom 0.15 m of the wells had 30 holes (8 mm diameter) drilled into the sides to allow invertebrates to be withdrawn from a larger volume of water surrounding the bottom of the wells [33]. A bilge pump was used to pump 2 L of hyporheic water per well, which was then filtered through a 250 μ m sieve and preserved in 70% ethanol.

Three drift nets (mesh, 250 μ m) evenly spaced across the channel and 3 cm above the bed [33] were used to collect macroinvertebrate drift for 30 min. Mean current velocity (at beginning and end of 30 min) and water depth were measured and the volume of water filtered was estimated. Drifting material was preserved in 70% ethanol. As behavioral drift is greatest at dawn and dusk [34], drift was taken at noon to collect organisms principally dislodged due to spawning activities. Insects were identified to species or genus, non-insects to family or order and functional group designation based on Merritt et al. [21]. The lengths (nearest 0.5 mm) of insects were measured to allow estimation of biomass based on INVERTCALC, using length-weight regressions [35].

Salmon

Summer runs in Twelve Mile Creek were dominated by pink (*Oncorhynchus gorbuscha* (Walbaum)) salmon, with chum salmon (*O. keta* (Walbaum)) being the next dominate salmon species present during the study. Resident juvenile coho salmon (*O. kisutch* (Walbaum)), Dolly Varden char (*Salvelinus malma* (Walbaum)) and sculpin (*Cottus* spp.) were also present. All live pink and chum salmon

Figure 3. Functional feeding group differences before and during spawning. Shredder (a), Collector-gatherer (b), Collector-filterer (c), Scraper (d), and Predator (e) densities before (dark grey bars) and during (white bars) the salmon run in riffles, pools, margins, the hyporheic zone, and drift. Graphs show ± 1 s.e. and * means $p < 0.05$.

doi:10.1371/journal.pone.0039254.g003

were quantified in 4 meter wide belt transects perpendicular to stream flow every 10 m for the entire 300 m reach. These counts were then scaled up to estimate the total number of salmon present in the 300 m stream reach on each date. Salmon were counted approximately twice weekly from the start of the spawning run on 5 August until only carcasses remained in the stream. Live salmon density increased from the start of the run throughout most of the summer and began to decline about 2 weeks before our last sampling date on 24 September 2008.

Habitat Characteristics

In each habitat, we measured pH, % dissolved oxygen and specific conductivity using a Hydrolab MS 5 Mini Sonde (HACH Environmental, Loveland, Colorado, USA). Habitat area was measured with an electronic distance measurer (EDM), % canopy

cover using a spherical densiometer, sediment size using a Wentworth scale gravelometer and water velocity and depth using a digital flow meter [33], each three times throughout the study. Additionally, we estimated benthic algal biomass (as chlorophyll *a*) by randomly selecting five cobbles which were immediately transported in a cooler to the laboratory and processed within 6 hours. The cobbles were scrubbed across their entire surface onto pre-ashed glass filters (0.7 μ m) type A/E (Pall Corporation, Ann Arbor, Michigan, USA) and then analyzed with a Trilogy Turner Design Fluorometer (Turner Designs, Inc., Sunnyvale, California, USA). Surface area of the cobbles was estimated by measuring length, width and depth and assuming an ellipse. The chlorophyll *a* per unit area was then pooled across the five cobbles to obtain a mean.

Figure 4. Genus-level differences throughout the salmon run. Densities of the six dominant taxa: *Ameletus* (a), *Baetis* (b), *Cinygmula* (c), *Suwallia* (d) *Sweltsa* (e) and *Chironomus* (f) throughout the salmon run in riffles, pools, margins, the hyporheic zone and drift.
doi:10.1371/journal.pone.0039254.g004

No specific permits were required for the described field of studies. The study location is not privately-owned or protected in any way and the field studies did not involve endangered or protected species.

Statistical Analysis

Linear regression was performed to analyze whether salmon density influenced macroinvertebrate density in the different habitat types and in the drift. The density and biomass of

invertebrates in riffles and in the drift were plotted over the course of the salmon run to determine how invertebrates responded to salmon in different habitats and the drift. The densities of the six dominant genera: *Chironomus* (Diptera: Chironomidae), *Sweltsa* (Plecoptera: Chloroperlidae), *Ameletus* (Ephemeroptera: Ameletidae), *Baetis* (Ephemeroptera: Baetidae), *Cinygmula* (Ephemeroptera: Heptageniidae) and *Suwallia* (Plecoptera: Chloroperlidae), were also plotted over the course of the sampling period to determine how specific taxa responded to spawning salmon.

Figure 5. Macroinvertebrate communities before and during spawning. Non-Metric Multi-Dimensional Scaling ordination showing the separation of invertebrate community structure before and during the salmon run. A total of 65% of the variation in macroinvertebrate community structure was explained by: 1st axis = 36% and 2nd = 29%.
doi:10.1371/journal.pone.0039254.g005

Repeated measures analysis of variance (rmANOVA) was performed with a Bonferroni correction to determine whether salmon presence (before and during) altered macroinvertebrate abundance and taxonomic composition. The two main effects were presence of salmon and habitat, where salmon was treated as the repeated factor and the replicates of each habitat were treated as random effects. A compound symmetric covariance structure was specified using SAS (Version 11; SAS Institute, Cary, North Carolina, USA). Macroinvertebrate response variables were total density, total biomass, taxonomic richness, the densities of shredders, collector-gatherers, collector-filterers, scrapers and predators, and the densities of the six dominant genera. Violations of the assumptions of ANOVA were corrected by transforming the data (logarithmic or exponential, as appropriate). Results were considered significant when $\alpha < 0.05$.

A Non-Metric Multi-Dimensional Scaling (NMDS) ordination was carried out to evaluate differences in macroinvertebrate community structure among the different habitats, and in the drift,

before and during the salmon run [36] using PC ORD (version 5; MJM software, Gleneden Beach, Oregon, USA). We ran a total of 250 iterations for the real data with a random seed start. A multiple response permutation procedure (MRPP), using Sørensen distances, was performed to test for significant differences in community structure among habitat types before and during the salmon run. When significant differences were found in macroinvertebrate community structure, Indicator Species Analysis (ISA) was used to determine which taxa were significant indicators of the communities in the different habitat types. Taxa were considered significant in the ISA when indicator values (% of perfect indication) were $> 55\%$ with $p < 0.001$. All aquatic insect taxa that represented $> 3\%$ of all samples were used in the ordination procedures.

Acknowledgments

We are grateful to Angeline Kosnik, Brittany Tanis, Ann-Marie Larquier, Winnie Winikoff, Peter Levi, Janine Rüegg, Scott Tiegs and John Hudson for assistance with laboratory and field work; the Thorne Bay and Craig Ranger Districts (USDA Forest Service); Dominic Chaloner, Gary Lamberti, Jennifer Tank, Aaron and K.K. Prussian, Erik Norberg, Rick Edwards and Steve McCurdy for insight about sampling design and streams on Prince of Wales Island; Bridgette Kolouch for calculating insect biomass; Wei Wang for statistical advice and consultation; thanks to Janine Rüegg and Dominic Chaloner for comments in previous drafts of this manuscript; furthermore we thank the anonymous reviewers and editor for their comments.

Author Contributions

Conceived and designed the experiments: EYC RWM KWC MEB. Performed the experiments: EYC RWM KWC MEB. Analyzed the data: EYC RWM KWC MEB. Contributed reagents/materials/analysis tools: EYC RWM KWC MEB. Wrote the paper: EYC RWM KWC MEB.

Table 2. Dominant indicator taxa based on indicator species analysis (ISA) pooling before and during the salmon run in each habitat type and drift.

	3 Most Significant Taxa	Indicator Values
Riffle	<i>Planaria</i> , <i>Epeorus</i> , <i>Baetis</i>	40%, 43%, 37%
Margin	<i>Ameletus</i> , <i>Cinygmula</i> , <i>Suwallia</i>	40%, 35%, 28%
Pool	<i>Oligochaete</i> , <i>Prodezzia</i> , <i>Oreodytes</i>	61%, 53%, 50%
Hyporheic	<i>Ostracoda</i> , <i>Copepoda</i> , <i>Paraleptophlebia</i>	47%, 36%, 22%
Drift	<i>Chironomidae</i> , <i>Acari</i> , <i>Araneae</i>	84%, 54%, 42%

doi:10.1371/journal.pone.0039254.t002

References

- Berryman AA, Hawkins BA (2006) The refuge as an integrating concept in ecology and evolution. *Oikos* 115: 192–196.
- Negishi JN, Inoue M, Nunokawa M (2002) Effects of channelization on stream habitat in relation to spate and flow refugia for macroinvertebrates in northern Japan. *Freshwater Biology* 47: 1515–1529.
- Lancaster J, Hildrew AG (1993) Flow refugia and the micro distribution of lotic macroinvertebrates. *Journal of the North American Benthological Society* 12(4): 385–393.
- Palmer MA, Bely AE, Berg KE (1991) Response of invertebrates to lotic disturbance: a test of the hyporheic refuge hypothesis. *Oecologia* 89(2): 182–194.
- Monaghan KA, Milner AM (2009) Effect of Anadromous salmon redd construction on macroinvertebrate communities in a recently formed stream in coastal Alaska. *Journal of the North American Benthological Society* 28: 153–166.
- Jones CG, Lawton JH, Shachak M (1994) Organisms as ecosystem engineers. *Oikos* 69: 373–386.
- Quinn TP (2005) *The Behavior and Ecology of Pacific Salmon and Trout*. American Fisheries Society, University of Washington Press.
- Gende SM, Edwards RT, Willson MF, Wipfli MS (2002) Pacific salmon in aquatic and terrestrial ecosystems. *BioScience* 52: 917–928.
- Levi PS, Tank JL, Tieggs SD, Ruetgg J, Chaloner DT, et al. (2011) Does timber harvest influence the dynamics of marine-derived nutrients in Southeast Alaska streams? *Canadian Journal of Fisheries and Aquatic Sciences* 68(8): 1316–1329.
- Tieggs SD, Chaloner DT, Levi PS, Ruetgg J, Tank JL, et al. (2008) Timber harvest transforms ecological roles of salmon in Southeast Alaska rainforest streams. *Ecological Applications* 18(1): 4–11.
- Campbell EY, Benbow ME, Tieggs SD, Hudson JP, Merritt RW, et al. (2011) Timber harvest intensifies spawning salmon disturbance of macroinvertebrates in Southeast Alaska streams. *Journal of the North American Benthological Society* 30(1): 49–59.
- Ruetgg J, Tieggs SD, Chaloner DT, Levi PS, Tank JL, et al. (2011) Salmon subsidies alleviate nutrient limitation of benthic biofilms in southeast Alaska streams. *Canadian Journal of Aquatic Sciences* 68: 277–287.
- Moore JW, Schindler DE (2008) Biotic disturbance and benthic community dynamics in salmon bearing streams. *Journal of Animal Ecology* 77: 275–284.
- Minakawa N, Gara RI (2003) Effects of chum salmon redd excavation on benthic communities in a stream in the Pacific Northwest. *Transactions of the American Fisheries Society* 132: 598–604.
- Peterson DP, Foote CJ (2000) Disturbance of small-stream habitat by spawning sockeye salmon in Alaska. *Transactions of the American Fisheries Society* 129: 924–934.
- Chaloner DT, Lamberti GA, Merritt RW, Mitchell NL, Ostrom PH, et al. (2004) Variation in responses of spawning Pacific salmon among three southeastern Alaska streams. *Freshwater Biology* 49: 587–599.
- Moore JW, Schindler DE, Scheuerell MD (2004) Disturbance of freshwater habitats by anadromous salmon in Alaska. *Oecologia* 139: 298–308.
- Lessard JL, Merritt RW, Berg MB (2009) Investigating the effect of marine-derived nutrients from spawning salmon on macroinvertebrate secondary production in southeast Alaskan streams. *Journal of the North American Benthological Society* 28(3): 683–693.
- Kerans BL, Karr JR (1992) Aquatic invertebrate assemblages: spatial and temporal differences among sampling protocols. *Journal of the North American Benthological Society* 11(4): 377–390.
- Gjerlov C, Hildrew AG, Jones JI (2003) Mobility of stream invertebrates in relation to disturbance and refugia: a test of habitat template theory. *Journal of the North American Benthological Society* 22(2): 207–223.
- Merritt RW, Cummins KW, Berg MB (2008) *An Introduction to the Aquatic Insects of North America*. Dubuque, IA, USA Kendall/Hunt Press.
- Sedell JR, Reeves GH, Hauer FR, Stanford JA, Hawkins CP (1990) Role of refugia in recovery from disturbances: modern fragmented and disconnected river systems. *Environmental Management* 14(5): 711–724.
- Winterbottom JH, Orton SE, Hildrew AG (1997) Field experiments on the mobility of benthic invertebrates in a southern English stream. *Freshwater Biology* 38: 37–47.
- Oliver MJ, Marmonier P, Befly JL (1997) Response of invertebrates to lotic disturbance: is the hyporheic zone a patchy refugium? *Freshwater Biology* 37: 257–276.
- Rosario RBD, Resh VH (2000) Invertebrates in intermittent and perennial streams: is the hyporheic zone a refuge from drying? *Journal of the North American Benthological Society* 50: 839–853.
- Lessard JL, Merritt RW (2006) Influence of marine-derived nutrients from spawning salmon on aquatic insect communities in southeast Alaskan streams. *Oikos* 113: 334–343.
- Hildebrand SG (1971) The effect of Coho spawning on the benthic invertebrates of the Platte River, Benzie County, Michigan. *Transactions of the American Fisheries Society* 100: 61–68.
- Field-Dodgson MS (1987) The effect of salmon red excavation on stream substrate and a benthic community of two salmon spawning streams in Canterbury, New Zealand. *Hydrobiologia* 154: 3–11.
- Tieggs SD, Campbell EY, Levi PS, Ruetgg J, Benbow ME, et al. (2009) Separating physical disturbance and nutrient enrichment caused by Pacific salmon in stream ecosystems. *Freshwater Biology* 54: 1864–1875.
- Moore JW, Schindler DE (2010) Spawning salmon and the phenology of emergence in stream insects. *Proceedings of the Royal Society* 277: 1695–1703.
- Resh VH, Brown AV, Covich AP, Gurtz ME, Li HW, et al. (1988) The role of disturbance in stream ecology. *Journal of the North American Benthological Society* 7: 433–455.
- White PS, Pickett STA (1985) *The Ecology of Natural Disturbance and Patch Dynamics*. Academic Press, Orlando, USA.
- Hauer FR, Lamberti GA (2007) *Methods in Stream Ecology*, second edition. Academic Press, Elsevier Inc.
- Hynes HBN (1975) Downstream drift of invertebrates in a river in southern Ghana. *Freshwater Biology* 5: 515–532.
- Benke AC, Hurn AD, Smock LA, Wallace JB (1999) Length-mass relationships for freshwater macroinvertebrates in North America with particular reference to the southeastern United States. *Journal of the North American Benthological Society*, 18: 308–343.
- McCune B, Grace JB, Urban DL (2002) *Analysis of Ecological Communities*. MjM Software Design.