

RESEARCH ARTICLE

# In vitro activity of seven antifungal agents against *Fusarium oxysporum* and expression of related regulatory genes

Dafeng Xu<sup>1</sup>, Kun Wang<sup>2</sup>, Tingchun Li<sup>1</sup>, Jingjing Wang<sup>3</sup>, Shiji Wang<sup>1</sup>, Fanna Kong<sup>1</sup>, Jie Dai<sup>3</sup>, Yuguo Liu<sup>4</sup>, Banfeng Ruan<sup>2\*</sup>, Benguo Zhou<sup>1\*</sup>

**1** Institute of Industrial Crops, Anhui Academy of Agricultural Sciences, Hefei, China, **2** School of Biological Food and Environment, Hefei University, Hefei, China, **3** Huainan Academy of Agricultural Sciences, Huainan, China, **4** Hachikō Legend Culture & Tourism Group, Huainan, China

\* [ruanbf@hfuu.edu.cn](mailto:ruanbf@hfuu.edu.cn) (BR); [bgzhou@163.com](mailto:bgzhou@163.com) (BZ)



## OPEN ACCESS

**Citation:** Xu D, Wang K, Li T, Wang J, Wang S, Kong F, et al. (2025) In vitro activity of seven antifungal agents against *Fusarium oxysporum* and expression of related regulatory genes. PLoS One 20(4): e0322206. <https://doi.org/10.1371/journal.pone.0322206>

**Editor:** Ravinder Kumar, ICAR - Indian Agricultural Research Institute, INDIA

**Received:** June 28, 2024

**Accepted:** March 18, 2025

**Published:** April 29, 2025

**Copyright:** © 2025 Xu et al. This is an open access article distributed under the terms of the [Creative Commons Attribution License](#), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

**Data availability statement:** The data-sets presented in this study are available from the Figshare database (10.6084/m9.figshare.27951405).

**Funding:** This study was supported by the Key Research and Development Program of Anhui, China (202204c06020060). The funders had

## Abstract

*Fusarium oxysporum* (*F. oxysporum*) is one of the main pathogenic fungus causing maize ear rot. In this study, the aims were to screen highly effective pesticides for *F. oxysporum*, reduce peasants' misunderstandings about pesticide application, improve disease control levels, and enhance economic efficiency. The toxicity of seven fungicides (carbendazim, pyraclostrobin, epoxiconazole, tricyclazole, azoxystrobin, difenoconazole, quintozeno) on *F. oxysporum* were determined by the mycelium growth rate and the spore germination method, and single and compound fungicides with effective inhibitory effects on mycelial growth were screened. The RT-qPCR method was used to detect the expression levels of chitin synthetase V (*ChsV*), folate uptake block T (*FUBT*), superoxide dismutase (*SOD*), and peroxidase dismutase (*POD*) genes in pathogenic bacteria treated with the selected agents and combination of fungicides. The results showed that all seven fungicides had inhibitory effects on mycelial growth hyphae and spore germination of *F. oxysporum*. Epoxiconazole had the strongest inhibitory effect on mycelium growth and spore germination of *F. oxysporum*, with effective concentrations ( $EC_{50}$ ) of 0.047 and 0.088  $\mu$ g/mL, respectively. The combination of pyraclostrobin and difenoconazole (P&D, combined at a mass ratio of 7:3) had the best inhibitory effect, with an  $EC_{50}$  of 0.094  $\mu$ g/mL and an SR of 2.650. Epoxiconazole and the combination P&D could inhibit mycelial growth and spore germination by down-regulating *ChsV*, *FUBT*, and *POD*, causing oxidative stress in *F. oxysporum*, and reducing the occurrence of maize ear rot.

## 1. Introduction

Maize ear rot, a prevalent and highly damaging fungal disease worldwide, significantly impacts maize yield and quality, and it brings great safety hazards to food and feed [1,2]. Changes such as the replacement of maize varieties, increased

no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

**Competing interests:** The authors have declared that no competing interests exist.

planting density, and alterations in cultivation practices have created favorable conditions for the occurrence and prevalence of maize ear rot [3,4]. This disease is caused by various fungal infections, with *Fusarium graminearum* (*F. graminearum*), *Fusarium oxysporum* (*F. oxysporum*), and *Fusarium verticillioides* (*F. verticillioides*) being the major pathogens [5,6]. Moreover, all three mentioned *Fusarium* fungi can produce fungal toxins that are associated with various diseases in both humans and animals [3,7]. *Fusarium* spp. can infect maize ears and grains, producing fungal toxins, a process regulated by related genes. The folate uptake block T gene (*FUBT*) can regulate the production of fusaric acid (FA) by *F. oxysporum* [8]. The chitin synthase gene (Chitin synthetase V, *ChsV*) can protect *Fusarium* from plant antimicrobial substances, and the absence of the *ChsV* gene leads to a loss of its pathogenicity [9]. *Fusarium* spp. itself possesses numerous protective enzyme genes, such as the superoxide dismutase (SOD) gene and the peroxidase dismutase (POD) gene [10]. SOD is a crucial enzyme in the antioxidant system, playing a central role in the elimination of reactive oxygen species [11], which helps defend *Fusarium* spp. from oxygen toxicity and oxidative damage. Under stress conditions, POD can efficiently eliminate H<sub>2</sub>O<sub>2</sub>, and its activity can reflect the metabolic status of the organism and its adaptability to the environment [12]. FA is a non-specific toxin produced by *F. oxysporum*, which can reduce host resistance by altering the permeability of host plant cell membranes, decreasing mitochondrial reactive oxygen species, inhibiting ATP synthesis, and suppressing plant root growth [13,14]. At present, research on *FUBT* and *ChsV* has made some progress in the fusarium wilt of watermelon, melon, and cotton caused by *F. oxysporum*, but there are few reports on maize ear rot [15,16].

As people's awareness of health increases, there is a growing concern about fungal toxin contamination in maize. Addressing and preventing maize ear rot disease and reducing fungal toxin contamination has become a current research focus. Currently, there are few varieties resistant to maize ear rot, and biological control methods are not yet mature [17,18]; chemical control is the most widely applied measure [19]. Fungicides can effectively control the occurrence and spread of maize ear rot in the short term, significantly reducing the content of toxins in the kernels [18]. However, there are a wide variety of fungicides on the market, each with different chemical structures and mechanisms of action. Improper use can easily lead to phytotoxicity issues. Therefore, conducting toxicity tests for different fungicides on maize ear rot is the primary task to ensure the selection of appropriate agents. In order to reduce the dosage and frequency of fungicide use, delay the development of resistance, and minimize phytotoxicity, it is essential to develop rational combinations of fungicides. This study focused on the toxicity and synergistic effects of seven commonly used fungicides for maize ear rot on *F. oxysporum*. The best single and compound fungicides were screened, and their effects on the expression levels of *ChsV*, *FUBT*, *SOD*, and *POD* genes in *F. oxysporum* are further analyzed, providing reference points for the effective control of maize ear rot.

## 2. Materials and methods

### 2.1 Materials

*Fusarium oxysporum* B (*F. oxysporum* B) was provided by the Plant Protection Laboratory of the Tobacco Research Institute, Anhui Academy of Agricultural Sciences. The potato dextrose agar medium (PDA), potato glucose broth medium (PDB), and water agar medium (WA) were prepared following the method outlined by Fang Zhongda [20].

Fungicides: Carbendazim (95%, Shandong Huayang pesticide chemical industry group Co., Ltd.), Pyraclostrobin (98%, Anhui Kelihua Chemical Co., Ltd.), Epoxiconazole (98%, Ningxia Gree Fine Chemical Co., Ltd.), Tricyclazole (95%, Shandong Shangnong Agricultural Technology Co., Ltd.), Azoxystrobin (96%, Shandong Union Pesticide Industry Co., Ltd.), Difenoconazole (97%, Jiangsu Heben Biochemical Co., Ltd.), Quintozene (40%, Shanxi Nongfengbao Pesticide Co., Ltd.).

### 2.2 Toxicity determination of fungicides against *F. oxysporum* B

**2.2.1 Action of a single fungicide on the mycelium of *F. oxysporum* B.** The impact of different fungicides on the mycelial growth of *F. oxysporum* B was determined by the growth rate method [21]. Specifically, according to the method described by Kowalska Krochmal et al. [22], the minimum inhibitory concentration test was conducted to prepare PDA plates containing different doses (as shown in Table 1) of fungicides. A mycelial disc with a diameter of 6 mm was inoculated at the center of each plate with *F. oxysporum* B. Then the plates were cultivated in the dark at 25°C with three replicates for each treatment, and PDA plates without any fungicide served as the control. After 3 days, the colony diameter was measured, and the mycelial growth inhibition rate (MGIR) was calculated according to formula (1).

$$MGIR = \frac{\varphi_c - \varphi_t}{\varphi_t - 6} \times 100\% \quad (1)$$

Where: MGIR, mycelial growth inhibition rate;  $\varphi_c$ , colony diameter in control group;  $\varphi_t$ , colony diameter in the treatment group; 6, colony diameter of the initial mycelial disc.

The MGIR were converted into probability values of inhibition rate; the logarithm of the fungicide mass concentration was used as the horizontal axis, and the probability values were used as the vertical axis to fit the regression equation. Then the correlation coefficient ( $R^2$ ) and effective concentration ( $EC_{50}$ ) were calculated, and the toxicity levels of 7 fungicides against the mycelium of *F. oxysporum* B were compared.

**2.2.2 Action of a single fungicide on the spore of *F. oxysporum* B.** The impact of different fungicides at a single dose on the germination of spores of *F. oxysporum* B was determined by the spread plate method [23]. Specifically, *F. oxysporum* B was cultured on PDA plates for 3 days. The mycelium was rinsed with sterile water, filtered through double layers of sterile gauze, and the filtrate was centrifuged at 4,000 rpm for 10 min. Then the spore was resuspended in sterile deionized water to prepare a spore suspension with a concentration of  $10^6$  spores/mL. 100  $\mu$ L of spore suspension

**Table 1. Mass concentration of fungicides for inhibiting mycelial growth.**

Fungicide	Mass concentration ( $\mu$ g/mL)				
Carbendazim	0.30	0.40	0.50	0.60	0.70
Pyraclostrobin	0.10	1	10	100	500
Epoxiconazole	0.0125	0.025	0.05	0.10	0.20
Tricyclazole	6.25	12.50	25.00	50	100
Azoxystrobin	4.00	8.00	16.00	32.00	64.00
Difenoconazole	0.01	0.05	0.10	0.50	1.00
Quintozene	3.125	6.25	12.50	25	50

<https://doi.org/10.1371/journal.pone.0322206.t001>

was spread on WA plates containing different doses (as shown in [Table 2](#)) of fungicides and incubated at 25°C in the dark. With three replicates for each treatment, and WA plates without any fungicide served as the control. When the spore germination rate on the control WA plate reached 90% or more, the number of germinated spores was recorded for different mass concentrations of the fungicide treatment. The spore germination rate (SGR) and spore germination inhibition rate (SGIR) were calculated according to formulas (2) and (3), respectively.

$$SGR = \frac{n_t}{n_0} \times 100\% \quad (2)$$

$$SGIR = \frac{SGR_c - SGR_t}{SGR_c} \times 100\% \quad (3)$$

Where: SGR, spore germination rate;  $n_t$ , the number of germinated spores;  $n_0$ , the total number of spores; SGIR, spore germination inhibition rate;  $SGR_c$ , SGR of the control group;  $SGR_t$ , SGR of the treatment group.

The SGIR were converted into probability values of inhibition rate; the logarithm of the fungicide mass concentration was used as the horizontal axis, and the probability values were used as the vertical axis to fit the regression equation. Then the  $R^2$  and  $EC_{50}$  were calculated, and the toxicity levels of 7 fungicides against the spore of *F. oxysporum* B were compared.

**2.2.3 Action of compound fungicides on the mycelium of *F. oxysporum* B.** Based on the measurement results of 2.2.1 and 2.2.2, compound the fungicide. The effects of compound fungicides were determined by the growth rate method [\[21\]](#). Specifically, dilute each fungicide separately to 100  $\mu$ g/mL, and then prepare different proportions of mixed solutions

**Table 2. Mass concentration of fungicides for inhibiting spore germination.**

Fungicide	Mass concentration ( $\mu$ g/mL)				
Carbendazim	2	4	8	16	32
Pyraclostrobin	0.125	0.25	0.50	1	2
Epoxiconazole	0.05	0.10	0.20	0.40	0.80
Tricyclazole	12.50	25	50	100	200
Azoxystrobin	4	8	16	32	64
Difenoconazole	2	4	8	16	32
Quintozene	3.125	6.25	12.50	25	50

<https://doi.org/10.1371/journal.pone.0322206.t002>

**Table 3. Mass concentration of composite agents used to determine the inhibitory effect of mycelium growth.**

Mass ratio	Mass concentration ( $\mu$ g/mL)			
	Epoxiconazole: Carbendazim	Carbendazim: Quintozene	Pyraclostrobin: Difenoconazole	Pyraclostrobin: Carbendazim
1:9	0.05, 0.1, 0.2, 0.3, 0.4	1.8, 2.4, 3.0, 3.6, 4.2	0.03125, 0.0625, 0.125, 0.25, 0.5	0.1, 0.2, 0.4, 0.6, 0.8
2:8	0.05, 0.1, 0.2, 0.3, 0.4	0.6, 1.2, 1.8, 2.4, 3.0	0.03125, 0.0625, 0.125, 0.25, 0.5	0.1, 0.2, 0.4, 0.6, 0.8
3:7	0.05, 0.1, 0.2, 0.3, 0.4	0.6, 1.2, 1.8, 2.4, 3.0	0.125, 0.25, 0.5, 1, 2	0.2, 0.3, 0.4, 0.5, 0.6
4:6	0.025, 0.05, 0.1, 0.2, 0.3	0.4, 0.6, 0.8, 1.0, 1.2	0.125, 0.25, 0.5, 1, 2	0.4, 0.5, 0.6, 0.7, 0.8
5:5	0.025, 0.05, 0.1, 0.2, 0.3	0.4, 0.6, 0.8, 1.0, 1.2	0.03125, 0.0625, 0.125, 0.25, 0.5	0.4, 0.5, 0.6, 0.7, 0.8
6:4	0.025, 0.05, 0.1, 0.2, 0.4	0.4, 0.5, 0.6, 0.7, 0.8	0.0625, 0.125, 0.25, 0.5, 1	0.4, 0.6, 0.8, 1.0, 1.2
7:3	0.025, 0.05, 0.1, 0.2, 0.4	0.3, 0.4, 0.5, 0.6, 0.7	0.0625, 0.125, 0.25, 0.5, 1	0.4, 0.6, 0.8, 1.0, 1.2
8:2	0.0125, 0.025, 0.05, 0.1, 0.2	0.4, 0.5, 0.6, 0.7, 0.8	0.0625, 0.125, 0.25, 0.5, 1	0.4, 0.6, 0.8, 1.0, 1.2
9:1	0.0125, 0.025, 0.05, 0.1, 0.2	0.3, 0.4, 0.5, 0.6, 0.7	0.0625, 0.125, 0.25, 0.5, 1	0.25, 0.5, 1.0, 2.0, 3.0

<https://doi.org/10.1371/journal.pone.0322206.t003>

according to [Table 3](#), and prepare PDA plates containing 10% compound fungicide solution. The procedures outlined in Section 2.2.1 were repeated, *F. oxysporum* B mycelial disc (whose diameter was 6 mm) was inoculated, and the MGIR for the composite fungicide was calculated. Then we fitted a regression equation, and the R2 and EC<sub>50</sub> were calculated. Analyzing the synergistic enhancement effect of the combination agent based on the method proposed by Wadley [\[24,25\]](#), using the synergistic ratio (SR) for the analysis of combined enhancement effects (Formulas 4 and 5), SR<0.5 indicates antagonistic effects in the compound formulation of the two fungicides; 0.5≤SR≤1.5 indicates additive effects in the compound formulation of the two fungicides; SR>1.5 indicates synergistic effects in the compound formulation of the two fungicides.

$$SR = \frac{\text{Theory } EC_{50}}{\text{Actual } EC_{50}} \quad (4)$$

$$\text{Theory } EC_{50} = \frac{a + b}{\frac{a}{EC_{50A}} + \frac{b}{EC_{50B}}} \quad (5)$$

Where: SR, synergistic ratio; A (or B): one type of fungicide; a (or b): the mass (or volume) ratio of fungicide A (or B).

### 2.3 mRNA expression analysis

A single fungicide and a compound agent were selected that have inhibitory or synergistic effects on *F. oxysporum* B. Prepare PDA plates containing a fungicide or compound agent and cultivate *F. oxysporum* B; and the normal PDA plate was used as the control group. After 3 days, mycelium was collected. The mRNA levels of *ChsV* (*ChsV*-F: 5'-TCTTTCCCCA TCAAGTGTCT-3'; *ChsV*-R: 5'-GTGATGTTGGTGTTCGGTTGT-3'), *FUBT* (*FUBT*-F: 5'-GGAGCCTGAAGACAGATTGC-3'; *FUBT*-R: 5'-CCGATAATAGGGACGATCCA-3'), *SOD* (*SOD*-F: 5'-GGTCCTCACTTCAACCCTCA-3'; *SOD*-R: 5'-AGTCGG TGACAGAGCCCTTA-3'), *POD* (*POD*-F: 5'-CGAGGGATGGATCAAGGATA-3'; *POD*-R: 5'-GTAGCATCCTGCTGGTCGAT-3') [\[16,26\]](#) in the mycelium of *F. oxysporum* B were measured. The TRIzol (TIANGEN Biotech (Beijing) Co., Ltd.) method is utilized to extract mRNA from the mycelium. Then a 1μg mRNA of satisfactory quality is selected for reverse transcription to obtain cDNA (ReverTra Ace qPCR RT Master Mix with gDNA Remover; TOYOBO Co., Ltd.), which is subsequently subjected to qPCR. The relative mRNA abundance of the target genes was normalized to *Actin* (*Actin*-F: 5'-CCGTGACATCAA GGAGAACG-3'; *Actin*-R: 5'-GGAAAGTGGACAGGGAAAGCA-3') and was then calculated using the 2<sup>-ΔΔCt</sup> method.

### 2.4 Statistical analysis

Microsoft Excel 2010 software was used to process the data. Statistical analysis was conducted using SPSS 19.0 (SPSS, Inc., Chicago, USA). Significant differences were obtained by one-way ANOVA, and the difference was considered significant when *P* ≤ 0.05.

## 3 Results

### 3.1 Inhibitory effect of a single fungicide on the mycelial growth of *F. oxysporum* B

The effects of a single fungicide on mycelial growth are shown in [Table 4](#). It can be observed that the mycelial growth of *F. oxysporum* B is inhibited by all seven tested fungicides; the EC<sub>50</sub> is 0.047~35.089 μg/mL. And the three fungicides with the strongest inhibitory effect on mycelial growth are epoxiconazole, difenoconazole, and carbendazim. Their EC<sub>50</sub> are as follows: 0.047 μg/mL, 0.078 μg/mL, and 0.445 μg/mL. The inhibitory effects of azoxystrobin on mycelial growth are the weakest, with an EC<sub>50</sub> of 35.089 μg/mL. These results indicate that epoxiconazole, difenoconazole, and carbendazim are effective in inhibiting the mycelial growth of *F. oxysporum* B.

### 3.2 Inhibitory effect of a single fungicide on the spore germination of *F. oxysporum* B

The effects of a single fungicide on spore germination are shown in [Table 5](#). It can be observed that the mycelial growth of *F. oxysporum* B is inhibited by all seven tested fungicides; the EC<sub>50</sub> is 0.088~42.720 µg/mL. And the two fungicides with the strongest inhibitory effect on mycelial growth are epoxiconazole and pyraclostrobin; their EC<sub>50</sub> are as follows: 0.088 µg/mL, 0.249 µg/mL. The inhibitory effects of tricyclazole on mycelial growth are the weakest, with an EC<sub>50</sub> of 42.720 µg/mL. These results indicate that epoxiconazole and pyraclostrobin are effective in inhibiting the spore germination of *F. oxysporum* B.

### 3.3 Toxicity determination of compound fungicides on *F. oxysporum* B

**3.3.1 Inhibitory and synergistic effects of a mixture of epoxiconazole and carbendazim.** The range of EC<sub>50</sub> values for a mixture of epoxiconazole and carbendazim with different ratios is 0.046 to 0.282 µg/mL, and the EC<sub>50</sub> values of the mixture are all lower than the EC<sub>50</sub> values of carbendazim ([Table 6](#)). The inhibitory effects of the mixtures are stronger than those of carbendazim. When the mass ratio of epoxiconazole and carbendazim is 8:2, the EC<sub>50</sub> is 0.046 µg/mL, which is lower than the EC<sub>50</sub> of epoxiconazole (0.047 µg/mL, 95% confidence interval: 0.037~0.059 µg/mL). At this mass ratio (8:2), the inhibitory effect is best. The SR values for a mixture of epoxiconazole and carbendazim at different mass ratios range from 0.413 to 1.236. When the mass ratio is 4:6 and 7:3, the SRs are 0.464 and 0.413 (SR<0.5), indicating an antagonistic effect. When the mass ratio is 1:9, 2:8, 3:7, 5:5, 6:4, 8:2, and 9:1, the SR are 0.855, 0.656, 0.921, 0.868, 1.159, 1.236, and 0.537, suggesting an additive effect (0.5≤SR≤1.5).

**3.3.2 Inhibitory and synergistic effects of a mixture of carbendazim and quintozone.** The EC<sub>50</sub> values for a mixture of carbendazim and quintozone with different ratios is 0.437 to 3.684 µg/mL ([Table 7](#)), and all of the EC<sub>50</sub> values for the mixture are lower than those for quintozone alone. Specifically, when the mass ratio of carbendazim and quintozone is 8:2, the EC<sub>50</sub> is 0.437 µg/mL, which is lower than the EC<sub>50</sub> of carbendazim (0.445 µg/mL, 95% confidence interval: 0.424~0.464 µg/mL). For all other combinations, the EC<sub>50</sub> values for the mixtures are higher than that of

**Table 4. Toxicity test results of 7 fungicides in a single dose on the mycelial growth of *F. oxysporum* B.**

Fungicide	Regression equation	EC <sub>50</sub> (µg/mL)	95% confidence interval (µg/mL)	R <sup>2</sup>
Carbendazim	y=6.639x+7.338	0.445	0.424~0.464	0.988
Pyraclostrobin	y=0.364x+4.761	4.533	2.037~9.385	0.933
Epoxiconazole	y=1.173x+6.558	0.047	0.037~0.059	0.983
Tricyclazole	y=1.585x+2.589	33.172	27.881~40.047	0.983
Azoxystrobin	y=0.448x+4.308	35.089	19.627~139.043	0.973
Difenoconazole	y=0.818x+5.905	0.078	0.054~0.109	0.990
Quintozone	y=0.958x+4.256	5.978	3.954~8.027	0.990

<https://doi.org/10.1371/journal.pone.0322206.t004>

**Table 5. Toxicity test results of 7 fungicides in a single dose on spore germination of *F. oxysporum* B.**

Fungicide	Regression equation	EC <sub>50</sub> (µg/mL)	95% confidence interval (µg/mL)	R <sup>2</sup>
Carbendazim	y=0.460x+4.431	17.285	9.813~62.681	0.970
Pyraclostrobin	y=1.184x+5.714	0.249	0.182~0.318	0.998
Epoxiconazole	y=0.327x+5.346	0.088	0.002~0.191	0.986
Tricyclazole	y=0.980x+3.402	42.720	31.869~55.890	0.953
Azoxystrobin	y=0.997x+3.573	26.963	20.610~30.005	0.935
Difenoconazole	y=1.417x+3.506	11.339	9.359~14.049	0.958
Quintozone	y=1.202x+3.487	18.142	14.102~22.641	0.960

<https://doi.org/10.1371/journal.pone.0322206.t005>

**Table 6. Toxicity effects of the epoxiconazole and carbendazim mixtures on *F. oxysporum* B.**

Fungicide combination	Mass ratio	Regression equation	R <sup>2</sup>	Actual EC <sub>50</sub> (µg/mL)	95% confidence interval (µg/mL)	Theory EC <sub>50</sub> (µg/mL)	SR
Epoxiconazole (A)	/	y=1.173x+1.558	0.983	0.047	0.037~0.059	/	/
Carbendazim (B)	/	y=6.639x+2.338	0.988	0.445	0.424~0.464	/	/
A+B	1:9	y=1.719x+5.946	0.993	0.282	0.203~0.286	0.241	0.855
A+B	2:8	y=1.291x+5.773	0.937	0.252	0.205~0.330	0.165	0.656
A+B	3:7	y=0.945x+5.817	0.939	0.136	0.098~0.180	0.126	0.921
A+B	4:6	y=0.944x+5.623	0.983	0.219	0.160~0.354	0.101	0.464
A+B	5:5	y=1.096x+6.105	0.977	0.098	0.077~0.126	0.085	0.868
A+B	6:4	y=0.963x+6.155	0.990	0.063	0.045~0.083	0.073	1.159
A+B	7:3	y=1.131x+0.915	0.981	0.155	0.122~0.207	0.064	0.413
A+B	8:2	y=1.170x+6.561	0.970	0.046	0.037~0.058	0.057	1.236
A+B	9:1	y=0.831x+5.846	0.990	0.096	0.069~0.154	0.052	0.537

Note: "/" indicates none or a value below 0.001

<https://doi.org/10.1371/journal.pone.0322206.t006>

**Table 7. Toxicity effects of the carbendazim and quintozene mixtures on *F. oxysporum* B.**

Fungicide combination	Mass ratio	Regression equation	R <sup>2</sup>	Actual EC <sub>50</sub> (µg/mL)	95% confidence interval (µg/mL)	Theory EC <sub>50</sub> (µg/mL)	SR
Carbendazim (A)	/	y=6.639x+2.338	0.988	0.445	0.424~0.464	/	/
Quintozene (B)	/	y=0.958x-0.744	0.990	5.978	3.954~8.027	/	/
A+B	1:9	y=3.081x+3.260	0.911	3.684	3.355~4.222	2.665	0.723
A+B	2:8	y=3.354x+4.324	0.968	1.591	1.290~1.930	1.714	1.078
A+B	3:7	y=3.165x+4.072	0.929	1.965	1.476~2.868	1.264	0.643
A+B	4:6	y=3.477x+4.919	0.979	1.055	0.965~1.187	1.001	0.949
A+B	5:5	y=4.915x+6.011	0.970	0.623	0.582~0.660	0.828	1.330
A+B	6:4	y=5.344x+5.647	0.986	0.757	0.711~0.825	0.707	0.934
A+B	7:3	y=4.599x+5.894	0.939	0.639	0.596~0.701	0.616	0.964
A+B	8:2	y=5.865x+7.111	0.968	0.437	0.402~0.464	0.546	1.251
A+B	9:1	y=4.409x+5.900	0.986	0.625	0.583~0.686	0.490	0.785

Note: "/" indicates none or a value below 0.001

<https://doi.org/10.1371/journal.pone.0322206.t007>

carbendazim. However, the SR values for the mixtures of carbendazim and quintozene at various mass ratios range from 0.643 to 1.330, indicating an additive effect (0.5≤SR≤1.5).

**3.3.3 Inhibitory and synergistic effects of a mixture of pyraclostrobin and difenoconazole.** The EC<sub>50</sub> values for a mixture of pyraclostrobin and difenoconazole at different mass ratios range from 0.044 to 0.176 µg/mL (Table 8). These values are significantly lower than the EC<sub>50</sub> value for pyraclostrobin (4.533 µg/mL, 95% confidence interval: 2.037~9.385 µg/mL), indicating that the inhibitory effects of these combinations are significantly better than those of the single pyraclostrobin. When the mass ratio of pyraclostrobin and difenoconazole is 2:8, 3:7, 4:6, and 6:4, the EC<sub>50</sub> values are 0.066 µg/mL, 0.044 µg/mL, 0.061 µg/mL, and 0.077 µg/mL. All these values are lower than the EC<sub>50</sub> of a single propiconazole; this indicates that these four combinations have a better inhibitory effect on *F. oxysporum* B than a single pyraclostrobin. For combinations of mass ratios at 3:7, 4:6, 6:4, 7:3, 8:2, and 9:1, the SRs are 2.494, 2.120, 2.481, 2.650, 2.386, and 3.903, respectively, and these combinations indicate a clear synergistic effect (SR >1.5). Additionally, at mass ratios of 1:9, 2:8, and 5:5, the SR are 0.972, 1.474, and 0.873, respectively, suggesting an additive effect (0.5≤SR≤1.5).

**Table 8. Toxicity effects of the pyraclostrobin and difenoconazole mixtures on *F. oxysporum* B.**

Fungicide combination	Mass ratio	Regression equation	R <sup>2</sup>	Actual EC <sub>50</sub> (µg/mL)	95% confidence interval (µg/mL)	Theory EC <sub>50</sub> (µg/mL)	SR
Pyraclostrobin (A)	/	y=0.364x-0.239	0.933	4.533	2.037~9.385	/	/
Difenoconazole (B)	/	y=0.818x+0.905	0.990	0.078	0.054~0.109	/	/
A+B	1:9	y=0.680x+5.714	0.999	0.089	0.054~0.130	0.087	0.972
A+B	2:8	y=0.646x+5.763	0.941	0.066	0.034~0.099	0.097	1.474
A+B	3:7	y=0.646x+5.874	0.972	0.044	0.006~0.098	0.111	2.494
A+B	4:6	y=0.670x+5.815	0.987	0.061	0.013~0.120	0.129	2.120
A+B	5:5	y=0.627x+5.474	0.966	0.176	0.101~0.265	0.153	0.873
A+B	6:4	y=0.677x+5.755	0.998	0.077	0.031~0.122	0.190	2.481
A+B	7:3	y=0.606x+5.621	0.969	0.094	0.038~0.151	0.250	2.650
A+B	8:2	y=0.661x+5.539	0.949	0.153	0.086~0.226	0.365	2.386
A+B	9:1	y=0.568x+5.434	0.970	0.173	0.091~0.502	0.675	3.903

Note: “/” indicates none or a value below 0.001

<https://doi.org/10.1371/journal.pone.0322206.t008>

All SR values are greater than 0.5 in different mass ratios of pyraclostrobin and difenoconazole; these combinations indicate additive and significant synergistic effects, thus holding promise for practical application and further research.

**3.3.4 Inhibitory and synergistic effects of a mixture of pyraclostrobin and carbendazim.** The EC<sub>50</sub> values for a mixture of pyraclostrobin and carbendazim at different mass ratios range from 0.260 to 0.824 µg/mL (Table 9). And when the mass ratio is 1:9, 2:8, and 3:7, the EC<sub>50</sub> values are 0.288 µg/mL, 0.260 µg/mL, and 0.379 µg/mL, all lower than the EC<sub>50</sub> of single carbendazim (0.445 µg/mL, P ≤ 0.05). The mixture of pyraclostrobin and carbendazim, in the range of 1:9–3:7, has a better inhibitory effect on *F. oxysporum* B than a single carbendazim. When the mass ratio of pyraclostrobin to carbendazim is between 7:3 and 9:1, as the proportion of pyraclostrobin increases, the EC<sub>50</sub> of the mixture gradually increases but remains significantly lower than the EC<sub>50</sub> of the single pyraclostrobin (4.533 µg/mL, 95% confidence interval: 2.037~9.385 µg/mL). The SR gradually decreases as the mass ratio of pyraclostrobin to carbendazim changes from 2:8–5:5. And at the ratios of 4:6 and 5:5, the SR is 1.305 and 1.219, respectively, indicating an additive effect. However, in other mass ratios, SR is greater than 1.5, demonstrating a significant synergistic effect. All SR values are greater than 0.5

**Table 9. Toxicity effects of pyraclostrobin and carbendazim mixtures on *F. oxysporum* B.**

Fungicide combination	Mass ratio	Regression equation	R <sup>2</sup>	Actual EC <sub>50</sub> (µg/mL)	95% confidence interval (µg/mL)	Theory EC <sub>50</sub> (µg/mL)	SR
Pyraclostrobin (A)	/	y=0.364x-0.239	0.933	4.533	2.037~9.385	/	/
Carbendazim (B)	/	y=6.639x+2.338	0.988	0.445	0.424~0.464	/	/
A+B	1:9	y=2.984x+6.614	0.961	0.288	0.204~0.386	0.241	0.855
A+B	2:8	y=2.312x+6.354	0.927	0.260	0.148~0.391	0.165	0.656
A+B	3:7	y=2.500x+6.054	0.897	0.379	0.282~0.515	0.126	0.921
A+B	4:6	y=1.950x+5.532	0.957	0.533	0.435~0.610	0.101	0.464
A+B	5:5	y=1.357x+5.240	0.942	0.665	0.548~1.171	0.085	0.868
A+B	6:4	y=1.502x+5.413	0.927	0.531	0.377~0.638	0.073	1.159
A+B	7:3	y=1.257x+5.391	0.987	0.489	0.283~0.613	0.064	0.413
A+B	8:2	y=0.551x+5.078	0.887	0.721	/	0.057	1.236
A+B	9:1	y=0.363x+5.031	0.926	0.824	0.215~2.099	0.052	0.537

Note: “/” indicates none or a value below 0.001

<https://doi.org/10.1371/journal.pone.0322206.t009>

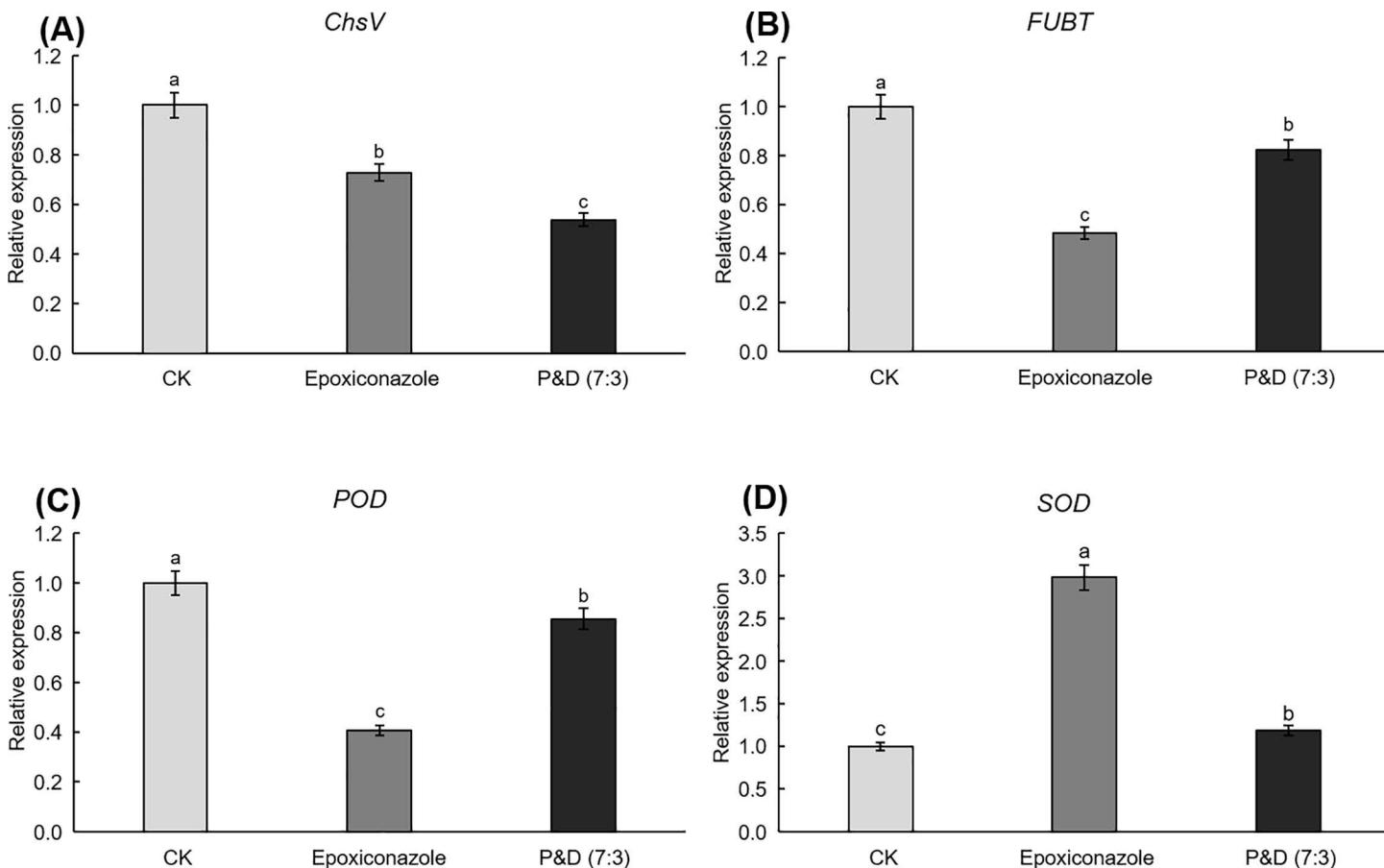
in different mass ratios of pyraclostrobin and carbendazim; these combinations indicate additive and significant synergistic effects. This combination formulation holds promise for practical application and further research.

### 3.4 Effect of fungicides on the expression of resistance genes in *F. oxysporum* B

Based on the inhibitory effects of single fungicides and compound fungicides on *F. oxysporum* B, the expression levels of *ChsV*, *FUBT*, *SOD*, and *POD* in the epoxiconazole group, pyraclostrobin and difenoconazole (7:3) group (P&D (7:3)), and control group (CK) were determined using RT-qPCR. The results are shown in Fig 1. Compared to the CK, both epoxiconazole and P&D (7:3) treatments significantly downregulated the relative expression levels of *ChsV*, *FUBT*, and *POD* in *F. oxysporum* B. Additionally, the relative expression level of *SOD* was significantly increased in both treatments.

## 4. Discussion

Maize ear rot occurs in the ears of corn, making it difficult to control. In maize-growing regions such as the United States, Canada, and South Africa, the primary causative agent of ear rot is often *Fusarium* [27–29]. In most regions of China, such as Jilin, Anhui, and Heilongjiang, the primary causative agent of maize ear rot is also *Fusarium* [30–33]. At present, there are many reports on the toxicity determination of pathogenic fungi such as *F. graminearum* and *F. verticillioides* in maize ear rot, while there are fewer reports on *F. oxysporum* [34,35]. This study found that epoxiconazole exhibited the



**Fig 1. Effect of fungicides on the expression level of genes related to *F. oxysporum* B.**

<https://doi.org/10.1371/journal.pone.0322206.g001>

most effective inhibition of the mycelial growth and spore germination of *F. oxysporum*, followed by difenoconazole and pyraclostrobin. While individual fungicides showed inhibitory effects on *F. oxysporum*, long-term use of a single fungicide can easily lead to resistance to the fungus. Therefore, it is recommended to use compound formulations to mitigate this issue. This experiment revealed that the combination of pyrazoxystrobin and difenoconazole exhibited significant inhibition and synergistic effects on the mycelial growth of *F. oxysporum*. When the mass ratio of pyrazoxystrobin to difenoconazole was 7:3 (P&D (7:3)), the EC<sub>50</sub> was 0.094 µg/mL, and the SR was 2.650.

Contains various pathogenic genes in *F. oxysporum* chitin synthase enzymes (Fochs). Among them, Fochs V and Fochs II play crucial roles in the pathogenicity of the strain [36]. Their absence results in a decrease in the pathogenic ability of the strain, and the loss of Fochs V leads to the loss of pathogenic capability in the strain [37]. Fusaric acid is a non-specific toxin that causes plant wilting. Research has demonstrated the presence of fusaric acid in cotton, leading to the occurrence of wilt disease. There is also evidence indicating a positive correlation between the virulence of *F. oxysporum* and the production of fusaric acid [13,14]. Additionally, *FUBT* has been shown to promote the production of fusaric acid. Knocking out the *FUBT* gene results in a significant reduction in fusaric acid production by *F. oxysporum* [8]. Antifungal agents can inhibit plant pathogens and cause damage by inducing the excessive production of reactive oxygen species (ROS) in the pathogens [38,39]. POD and SOD are critical enzymes in the reactive oxygen species (ROS) system. They have the ability to reduce or impede the damage caused by reactive oxygen-free radicals to organisms [40]. The activity of POD and SOD serves as important physiological indicators, reflecting the induced resistance of cells to the antifungal agent as well as their response to environmental stress [41,42]. Compared to the CK group, after treatment with epoxiconazole and P&D (7:3), the expression levels of *ChsV* and *FUBT* in *F. oxysporum* showed a significant decrease, suggesting a reduction in the pathogenicity and virulence of the fungus, which suggests a decreased likelihood of maize ear rot occurrence. And the expression level of the SOD exhibited an increasing trend, suggesting that the two treatments induced oxidative stress in *F. oxysporum*. Conversely, the expression level of the POD showed a significant decrease, indicating a reduction in the POD activity in *F. oxysporum*, leading to a weakened ability to eliminate H<sub>2</sub>O<sub>2</sub>. Therefore, the oxidative stress response in *F. oxysporum* was enhanced after treatment with epoxiconazole or P&D (7:3), leading to a decrease in its resistance to fungicides.

## 5. Conclusion

This study demonstrates the effectiveness of specific fungicides and their combinations in controlling *F. oxysporum*, a major pathogen causing maize ear rot. Epoxiconazole emerged as the most effective single fungicide, exhibiting strong inhibitory effects on both mycelial growth and spore germination. And compound formulations, particularly the combination of pyraclostrobin and difenoconazole (7:3 mass ratio), showed significant synergistic effects, providing superior control compared to individual fungicides. These fungicides downregulated key genes (*ChsV*, *FUBT*, and *POD*) and induced oxidative stress in *F. oxysporum*, reducing its environmental adaptability, infectivity, and pathogenicity. By optimizing fungicide combinations, this approach enhances disease control while minimizing pesticide use, offering a sustainable strategy to mitigate maize ear rot and its economic impact.

## Author contributions

**Conceptualization:** Dafeng Xu, Kun Wang.

**Data curation:** Fanna Kong.

**Formal analysis:** Jingjing Wang, Jie Dai.

**Funding acquisition:** Dafeng Xu, Benguo Zhou.

**Investigation:** Shiji Wang.

**Methodology:** Kun Wang, Jie Dai.

**Project administration:** Yuguo Liu.

**Resources:** Jingjing Wang, Shiji Wang, Banfeng Ruan.

**Software:** Tingchun Li.

**Supervision:** Fanna Kong, Yuguo Liu.

**Visualization:** Tingchun Li, Jingjing Wang.

**Writing – original draft:** Dafeng Xu.

**Writing – review & editing:** Banfeng Ruan, Benguo Zhou.

## References

1. Goswami RS, Kistler HC. Heading for disaster: *Fusarium graminearum* on cereal crops. *Mol Plant Pathol*. 2004;5(6):515–25. <https://doi.org/10.1111/j.1364-3703.2004.00252.x> PMID: 20565626
2. Yin Z, Wei X, Cao Y, Dong Z, Long Y, Wan X. Regulatory balance between ear rot resistance and grain yield and their breeding applications in maize and other crops. *J Adv Res*. 2024;S2090-1232(24)00479-X. <https://doi.org/10.1016/j.jare.2024.10.024> PMID: 39447642
3. Santiago R, Cao A, Malvar RA, Butrón A. Genomics of Maize Resistance to *Fusarium* Ear Rot and Fumonisin Contamination. *Toxins (Basel)*. 2020;12(7):431. <https://doi.org/10.3390/toxins12070431> PMID: 32629954
4. Duan C, Wang X, Song F, Sun S, Zhou D, Zhu Z. Advances in research on maize resistance to ear rot. *Scientia Agricultura Sinica*. 2015;48(11):2152–64. <https://doi.org/10.3864/j.issn.0578-1752.2015.11.007>
5. Mesterhazy A. Food Safety Aspects of Breeding Maize to Multi-Resistance against the Major (*Fusarium graminearum*, *F. verticillioides*, *Aspergillus flavus*) and Minor Toxicogenic Fungi (*Fusarium* spp.) as Well as to Toxin Accumulation, Trends, and Solutions-A Review. *J Fungi (Basel)*. 2024;10(1):40. <https://doi.org/10.3390/jof10010040> PMID: 38248949
6. Li L, Qu Q, Cao Z, Guo Z, Jia H, Liu N, et al. The Relationship Analysis on Corn Stalk Rot and Ear Rot According to *Fusarium* Species and Fumonisin Contamination in Kernels. *Toxins (Basel)*. 2019;11(6):320. <https://doi.org/10.3390/toxins11060320> PMID: 31195636
7. Bani M, Rispail N, Evidente A, Rubiales D, Cimmino A. Identification of the main toxins isolated from *Fusarium oxysporum* f. sp. *pisi* race 2 and their relation with isolates' pathogenicity. *J Agric Food Chem*. 2014;62(12):2574–80. <https://doi.org/10.1021/jf405530g> PMID: 24568659
8. Crutcher FK, Liu J, Puckhaber LS, Stipanovic RD, Bell AA, Nichols RL. FUBT, a putative MFS transporter, promotes secretion of fusaric acid in the cotton pathogen *Fusarium oxysporum* f. sp. *vasinfectum*. *Microbiology (Reading)*. 2015;161(Pt 4):875–83. <https://doi.org/10.1099/mic.0.000043> PMID: 25627440
9. Zhang D, Ren L, Wang Q, Wenjing Li, Song Z, Jin X, et al. Systematic assessment of the antifungal mechanism of soil fumigant methyl isothiocyanate against *Fusarium oxysporum*. *Environ Pollut*. 2024;341:122791. <https://doi.org/10.1016/j.envpol.2023.122791> PMID: 37940016
10. Gao T, Zhang Y, Shi J, Mohamed SR, Xu J, Liu X. The Antioxidant Guaiacol Exerts Fungicidal Activity Against Fungal Growth and Deoxynivalenol Production in *Fusarium graminearum*. *Front Microbiol*. 2021;12:762844. <https://doi.org/10.3389/fmicb.2021.762844> PMID: 34867894
11. Jiang W, Yang L, He Y, Zhang H, Li W, Chen H, et al. Genome-wide identification and transcriptional expression analysis of superoxide dismutase (SOD) family in wheat (*Triticum aestivum*). *PeerJ*. 2019;7:e8062. <https://doi.org/10.7717/peerj.8062> PMID: 31763072
12. Cai L, Zhang W, Jia H, Feng H, Wei X, Chen H, et al. Plant-derived compounds: A potential source of drugs against Tobacco mosaic virus. *Pestic Biochem Physiol*. 2020;169:104589. <https://doi.org/10.1016/j.pestbp.2020.104589> PMID: 32828361
13. Liu S, Li J, Zhang Y, Liu N, Viljoen A, Mostert D, et al. Fusaric acid instigates the invasion of banana by *Fusarium oxysporum* f. sp. *cubense* TR4. *New Phytol*. 2020;225(2):913–29. <https://doi.org/10.1111/nph.16193> PMID: 31513293
14. López-Díaz C, Rahjoo V, Sulyok M, Ghionna V, Martín-Vicente A, Capilla J, et al. Fusaric acid contributes to virulence of *Fusarium oxysporum* on plant and mammalian hosts. *Mol Plant Pathol*. 2018;19(2):440–53. <https://doi.org/10.1111/mpp.12536> PMID: 28093838
15. Jackson E, Li J, Weerasinghe T, Li X. The Ubiquitous Wilt-Inducing Pathogen *Fusarium oxysporum*-A Review of Genes Studied with Mutant Analysis. *Pathogens*. 2024;13(10):823. <https://doi.org/10.3390/pathogens13100823> PMID: 39452695
16. Liu C. Allelopathy of root exudates of different resistant muskmelon cultivars on *Fusarium* wilt. Master's theses, Shenyang Agricultural University, Shenyang. 2018. Available from: [https://kns.cnki.net/kcms2/article/abstract?v=WStw-PbchowPmmYTjp7XPzj4bwwh-Mq8Gm1wjx55VE2kU-Wr50jsQDf5aC7pQ4VIX\\_0uFv2F\\_vBStQpL5-Ud4jhBPY8gJbesc1S2era05VNNeKy8FRFF1SNdqfoGzytP6ft1IE3ix3vLJy62N\\_i8Vktc4STSy7XBF-PzXHc4mHOogL\\_nQB9ayVa1HulWNPqYAP9tQ3vNXs=&uniplatform=NZKPT&language=CHS](https://kns.cnki.net/kcms2/article/abstract?v=WStw-PbchowPmmYTjp7XPzj4bwwh-Mq8Gm1wjx55VE2kU-Wr50jsQDf5aC7pQ4VIX_0uFv2F_vBStQpL5-Ud4jhBPY8gJbesc1S2era05VNNeKy8FRFF1SNdqfoGzytP6ft1IE3ix3vLJy62N_i8Vktc4STSy7XBF-PzXHc4mHOogL_nQB9ayVa1HulWNPqYAP9tQ3vNXs=&uniplatform=NZKPT&language=CHS)
17. Mesterhazy A, Szieberth D, Toldine ET, Nagy Z, Szabó B, Herczeg B, et al. Updating the Methodology of Identifying Maize Hybrids Resistant to Ear Rot Pathogens and Their Toxins-Artificial Inoculation Tests for Kernel Resistance to *Fusarium graminearum*, *F. verticillioides*, and *Aspergillus flavus*. *J Fungi (Basel)*. 2022;8(3):293. <https://doi.org/10.3390/jof8030293> PMID: 35330295
18. Blandino M, Scarpino V, Testa G, Vanara F, Reyneri A. The Effect of Foliar Fungicide and Insecticide Application on the Contamination of Fumonisins, Moniliformin and Deoxynivalenol in Maize Used for Food Purposes. *Toxins (Basel)*. 2022;14(7):422. <https://doi.org/10.3390/toxins14070422> PMID: 35878160
19. He D, Shi J, Qiu J, Hou Y, Du Y, Gao T, et al. Antifungal activities of a novel triazole fungicide, mefenitrifluconazole, against the major maize pathogen *Fusarium verticillioides*. *Pestic Biochem Physiol*. 2023;192:105398. <https://doi.org/10.1016/j.pestbp.2023.105398> PMID: 37105621

20. Fang ZD. Research methods for plant diseases. Beijing: China Agriculture Press, 1996.
21. Mu LY. The research technique of plant chemical protection. Beijing: China Agriculture Press, 1994.
22. Kowalska-Krochmal B, Dudek-Wicher R. The Minimum Inhibitory Concentration of Antibiotics: Methods, Interpretation, Clinical Relevance. Pathogens. 2021;10(2):165. <https://doi.org/10.3390/pathogens10020165> PMID: 33557078
23. Song Y, Chen X, Sun J, Bai Y, Jin L, Lin Y, et al. *In Vitro* Determination of Sensitivity of *Fusarium fujikuroi* to Fungicide Azoxystrobin and Investigation of Resistance Mechanism. J Agric Food Chem. 2022;70(31):9760–8. <https://doi.org/10.1021/acs.jafc.2c02663> PMID: 35901518
24. Jiang J, Chen JP, Wei JQ, Guo XH, Chen ZP, Tian YE. Synergistic effect of fludioxonil and tebuconazole against *Fusarium graminearum*. Chinese Agricultural Science Bulletin. 2022;38(6):116–20. <https://doi.org/10.11924/j.issn.1000-6850.casb2021-0340>
25. Yao X, Qin W. Joint-toxicity and storage control efficacy of difenoconazole-iprodione mixtures against *Coniella granati*. Journal of Fruit Science. 2017;34(8):1033–42. <https://doi.org/10.13925/j.cnki.gsxb.20160403>
26. Qi X, Yang L, Huang J. Cloning and expression analysis of two catalase gene of FOC4 and the study on the oxidativ burst in banana by FOC4. Chinese Agricultural Science Bulletin. 2012;28(15):163–9. <https://doi.org/10.3969/j.issn.1000-6850.2012.15.032>
27. Lana FD, Madden LV, Carvalho CP, Paul PA. Impact of Gibberella Ear Rot on Grain Quality and Yield Components in Maize as Influenced by Hybrid Reaction. Plant Dis. 2022;106(12):3061–75. <https://doi.org/10.1094/PPDIS-01-22-0148-RE> PMID: 35536201
28. Smith JL, Limay-Rios V, Hooker DC, Schaafsma AW. *Fusarium graminearum* Mycotoxins in Maize Associated With *Striacosta albicosta* (Lepidoptera: Noctuidae) Injury. J Econ Entomol. 2018;111(3):1227–42. <https://doi.org/10.1093/jee/toy005> PMID: 29547905
29. Price J-L, Visagie CM, Meyer H, Yilmaz N. Fungal Species and Mycotoxins Associated with Maize Ear Rots Collected from the Eastern Cape in South Africa. Toxins (Basel). 2024;16(2):95. <https://doi.org/10.3390/toxins16020095> PMID: 38393173
30. Duan C, Qin Z, Yang Z, Li W, Sun S, Zhu Z, et al. Identification of Pathogenic *Fusarium* spp. Causing Maize Ear Rot and Potential Mycotoxin Production in China. Toxins (Basel). 2016;8(6):186. <https://doi.org/10.3390/toxins8060186> PMID: 27338476
31. Chai H, Jia J, Bai X, Meng L, Zhang W, Jin R. Identification of pathogenic *Fusarium* spp. causing maize ear rot and susceptibility of some strains to fungicides in Jilin province. Scientia Agricultura Sinica. 2023;56(1):64–78. <https://doi.org/10.3864/j.issn.0578-1752.2023.01.005>
32. Wei Q, Liao L, Chen L, Qi Y. Isolation and identification of main *Fusarium* species causing maize ear rot in Anhui province. Plant Protection. 2019;45(5):221–5. <https://doi.org/10.16688/j.zwzh.2018362>
33. Wang B, Bi S, Xiao M, Zhang D, Yan Q, Zhang Y, et al. Isolation and identification of pathogenic *Fusarium* spp. causing maize ear rot and analysis of their toxin-producing genotype in Heilongjiang province. Acta Prataculturae Sinica. 2020;29(1):163–74. <https://doi.org/10.11733/j.issn.1007-0435.2014.01.028>
34. Yin M, Gao X, Qian L, Jiang J, Zhang C, Liu S. Inhibitory activity of triticonazole and its mixture against *Fusarium graminearum* in Henan Province and its control efficacy on *Fusarium* head blight of wheat. Chinese Journal of Pesticide Science. 2024;26(5):974–82. <https://doi.org/10.16801/j.issn.1008-7303.2024.0092>
35. Su Y, Liu P, Gao K, Song J, Yang M, Lu H. Research progress on ear rot in maize. Heilongjiang Agricultural Sciences. n.d.;9(9):103–13. <https://doi.org/10.11942/j.issn1002-2767.2024.09.0103>
36. Soulié M-C, Perino C, Piffeteau A, Choquer M, Malfatti P, Cimerman A, et al. *Botrytis cinerea* virulence is drastically reduced after disruption of chitin synthase class III gene (Bccchs3a). Cell Microbiol. 2006;8(8):1310–21. <https://doi.org/10.1111/j.1462-5822.2006.00711.x> PMID: 16882034
37. Martín-Udíroz M, Madrid MP, Roncero MIG. Role of chitin synthase genes in *Fusarium oxysporum*. Microbiology (Reading). 2004;150(Pt 10):3175–87. <https://doi.org/10.1099/mic.0.27236-0> PMID: 15470098
38. Delattin N, Cammue BPA, Thevissen K. Reactive oxygen species-inducing antifungal agents and their activity against fungal biofilms. Future Med Chem. 2014;6(1):77–90. <https://doi.org/10.4155/fmc.13.189> PMID: 24358949
39. Oiki S, Nasuno R, Urayama S-I, Takagi H, Hagiwara D. Intracellular production of reactive oxygen species and a DAF-FM-related compound in *Aspergillus fumigatus* in response to antifungal agent exposure. Sci Rep. 2022;12(1):13516. <https://doi.org/10.1038/s41598-022-17462-y> PMID: 35933435
40. Zhang J, Zhang L, Liang D, Yang Y, Geng B, Jing P, et al. ROS accumulation-induced tapetal PCD timing changes leads to microspore abortion in cotton CMS lines. BMC Plant Biol. 2023;23(1):311. <https://doi.org/10.1186/s12870-023-04317-5> PMID: 37308826
41. Zhang X, Cheng Z, Yao W, Gao Y, Fan G, Guo Q, et al. Overexpression of *PagERF072* from Poplar Improves Salt Tolerance. Int J Mol Sci. 2022;23(18):10707. <https://doi.org/10.3390/ijms231810707> PMID: 36142609
42. Xiong J, Zhang W, Zheng D, Xiong H, Feng X, Zhang X, et al. ZmLBD5 Increases Drought Sensitivity by Suppressing ROS Accumulation in Arabidopsis. Plants (Basel). 2022;11(10):1382. <https://doi.org/10.3390/plants11101382> PMID: 35631807