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# Diversity, *Leishmania* detection, and blood meal sources of sand flies from Iguatama, Minas Gerais, Brazil

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# Abstract

This study investigated the sand fly fauna of the municipality Iguatama, in the Midwest Region of Minas Gerais state, Brazil, including Leishmania infection rates and blood meal sources. Sand flies were collected during four periods over the course of a single year, encompassing both dry and rainy seasons, using CDC light traps placed in peridomiciles where dogs were seropositive for visceral leishmaniasis (VL). A total of 762 sand fly specimens, representing 12 species across seven genera, were collected. Lutzomyia longipalpis was the most abundant species, comprising 57.6% of the collected specimens, followed by Nyssomyia neivai (19.6%) and Nyssomyia whitmani (10.5%). Species richness and diversity varied among collection periods, with the highest diversity observed in January 2019. Molecular analysis detected Leishmania DNA in 12.5% of the sand fly specimens, with Le. infantum being the predominant species. Blood meal analysis revealed feeding on multiple vertebrate species, including humans, rats, dogs, and chickens. The presence of Leishmania DNA in sand flies, and the identification of human blood meals, highlight the potential role of these species in VL transmission. These findings underscore the importance of continued surveillance and control measures to prevent the spread of VL and reduce transmission risk in the region.

# Introduction

Sand flies (Diptera: Psychodidae), are of considerable ecological importance due to their crucial role in the transmission of various pathogens, including protistan parasites of the genus *Leishmania* Ross, 1903, the causative agent of leishmaniasis [1]. This disease, with both cutaneous (CL) and visceral (VL) forms, is highly endemic in Brazil and geographically widespread [2]. Leishmaniasis incidence rates in Brazil have had a significant impact on public health, with numerous reported cases and the involvement of multiple *Leishmania* species [3]. submit the manuscript for publication. GF, Conselho Nacional de Desenvolvimento Científico e Tecnológico, https://www.gov.br/cnpq/pt-br, The sponsor had no role in the study design, analysis and interpretation of data, or decision to submit the manuscript for publication. JDAF, Conselho Nacional de Desenvolvimento Científico e Tecnológico, https://www.gov.br/cnpq/pt-br, The sponsor had no role in the study design, analysis and interpretation of data, or decision to submit the manuscript for publication.

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In Southeast Brazil, the state of Minas Gerais (MG) is the most endemic for leishmaniasis [4,5]. Even municipalities previously known to be free of VL, such as those in the Midwest Region of MG, have witnessed a rapid spread of this disease [6]. The municipality of Iguatama was considered free of VL until 2013, when the first canine case was reported [7]. Subsequent canine serological surveys in the urban area of Iguatama revealed a seroprevalence of 8.3% in that year [8]. In response, approximately 80% of dogs seropositive for VL were euthanized as a measure to control the disease. Despite these efforts, a survey conducted in 2017 showed a seroprevalence of 7.4% [9], suggesting that euthanasia was not an effective measure in preventing new canine cases of VL. During this period, no human VL cases were reported; however, the first documented human case was reported in 2021, raising public health concerns [10]. Regarding CL, a total of 11 autochthonous cases were reported between 2007–2021 [4], highlighting that both clinical forms of the disease are occurring in Iguatama.

The wealth of epidemiological studies conducted in Iguatama, MG, has provided valuable insights into the prevalence of leishmaniasis, underscoring the significance of continuous research efforts aimed at monitoring and controlling disease transmission. The present study aimed to address the impact of leishmaniasis in this municipality by investigating the ecology of its sand fly fauna, identifying the presence of *Leishmania* parasites, elucidating factors that influence their transmission, and evaluating blood meal sources to gain insights into sand fly interactions with vertebrate hosts and their role in the transmission cycle.

## Material and methods

#### Study area, collection, and identification of sand flies

The municipality of Iguatama (20°10'26" S, 45°42'39" W), located in the Midwest Region of Minas Gerais State (MG), Brazil (Fig 1), is characterized by a tropical savannah climate, as classified by the Köppen-Geiger system [11]. The natural vegetation of the region is



Fig 1. Location of the study area. The municipality of Iguatama in the Midwest Region of Minas Gerais State, Brazil, and the collection sites throughout the municipality.

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predominantly Cerrado, and the rainy season typically extends from October to March, while the dry season lasts from April to September.

Sand flies were collected during four periods over the course of August 2018 and September 2019, with two collection campaigns in the dry season (May 2019 and September 2019) and two others in the rainy season (August 2018 and January 2019). Two CDC light traps were placed in 29 peridomiciles that housed dogs seropositive for VL, treated with Miltefosine [12,13], for two consecutive nights. All peridomiciles exhibited similar characteristics, including fruit trees and domestic animals (dogs, cats, and chickens). Collecting was conducted in accordance with the Sistema de Autorização e Informação em Biodiversidade–SISBIO (License N°15237–2), the Brazilian environmental agency.

The collected sand flies were stored at -20°C in 1.5mL tubes containing 70% ethanol solution until dissection. Female specimens were dissected on glass slides containing phosphatebuffered saline, with the head and last three abdominal segments being removed for morphological identification of species. The remaining thorax and abdomen were individually stored dry at -20°C until DNA extraction. Male specimens were mounted on glass slides using Berlese liquid.

All sand flies were identified following the classification proposed by Galati [14]. Females of *Evandromyia cortelezzii* (Brethes, 1923) were identified to the species level since no other species of the Cortelezzii complex [15] were collected. The abbreviations for sand fly genera followed Marcondes [16].

#### Ecological assessments of the sand fly fauna

Ecological assessments of the sand fly fauna were conducted to analyze population patterns and diversity indices using Microsoft Excel (Office 2016). The cumulative number of species was calculated to estimate species richness over the study period and assess the adequacy of sampling efforts with CDC light traps. Non-linear logarithmic regression analysis was performed to examine the potential influence of extrapolated hypothetical collections on species richness. Species richness was also estimated using the classical formula of the *Chao1* Index. Ecological analyses of the collection sites utilized Shannon (H) and Simpson (D) diversity indices, as well as the Pielou Evenness Index (J) [17,18]. Constancy Index was used to evaluate the frequency of each species throughout the study period [19]. All ecological indices were calculated using EstimateS software version 9.1.0.

#### Molecular detection of Leishmania

For molecular detection of *Leishmania* DNA, whole DNA of non-engorged individual female sand flies was extracted using the Gentra Puregene Cell and Tissue Kit (Qiagen, Valencia, CA), following the manufacturer's protocol. The extracted DNA was then stored at -20°C until further molecular assays. Negative control groups consisting of *Lutzomyia longipalpis* males were included during DNA extraction to avoid potential cross-contamination. Instruments and working areas were decontaminated using DNAZap (Ambion Life Technologies, Inc.) to ensure accuracy of results.

A PCR assay was conducted to detect the presence of *Leishmania* DNA, using primers 150 (5' GGGKAGGGGCGTTCTSCGAA 3') and 152 (5' SSSWCTATWTTACACCAACCCC3'), targeting a conserved region of the kinetoplast DNA minicircle, following conditions previously described [20,21]. A positive control was included in all PCR assays, consisting of the reference strain of *Leishmania infantum* (MHOM/BR/1974/PP75), along with a negative control of non-template samples. PCR-positive products were purified using the ExoSAP-IT<sup>TM</sup> PCR Product Cleanup Reagent (Thermo Fisher, California, USA), and subsequently subjected to Sanger

sequencing [22]. All obtained sequences were analyzed using Finch TV software (Geospiza, Inc., Seattle, USA), and amplicons were compared with sequences from the GenBank database using the BLAST tool.

#### **Blood meal analysis**

Blood meal analysis was conducted to identify the source of engorged female sand flies' blood meals. Engorged female sand flies were dissected as previously described for non-engorged females. The remaining parts were stored dry at -20°C until DNA extraction. Whole DNA was extracted using the QIAamp<sup>®</sup> Blood Kit (Qiagen, USA), following the manufacturer's protocol. Precautions were taken to prevent cross-contamination during DNA extraction [23]. A PCR assay was conducted using the set of primers *cytb1* (5'- CCATCCAACATCTCAGCATGAT GAAA-3') and *cytb2* (5'- GCCCCTCAGAATGATATTTGTCCTCA-3') targeting a 359bp fragment of the Cytochrome B gene (*cytb*) to identify the source of blood meals, following conditions previously described [24]. Positive controls consisted of DNA extracted from a blood sample of *Gallus gallus*, while non-template samples were included as negative controls. PCR-positive products were purified using the ExoSAP-IT<sup>™</sup> PCR Product Cleanup Reagent (Thermo Fisher, California, USA) and subsequently subjected to Sanger sequencing. The obtained sequences were analyzed and compared to those deposited in the GenBank database.

#### Results

A total of 762 sand fly specimens were collected, comprising 12 species from seven genera. The predominant species was *Lu. longipalpis*, which accounted for 57.6% of all collected specimens, followed by *Ny. neivai* and *Ny. whitmani*, which represented 19.6% and 10.5%, respectively. These three species together represented 88% of all captured sand flies (Table 1). The highest species diversity was observed in January 2019, with 10 species (H = 1.35; D = 0.35), followed by May 2019 with eight species (H = 1.31; D = 0.31), and August 2018 with six species (H = 0.92; D = 0.53). The species accumulation curves indicated that richness reached its maximum after the third collection, with stabilization of the logarithmic curve. However, additional species were expected in the study area, suggesting a need for increased sampling effort (R<sup>2</sup> = 0.9492) (Fig 2). The estimated real richness value, as indicated by the *Chao1* Index, further supported the presence of additional species beyond those observed (Observed Richness = 12 species; Chao1 value = 13.5; SE ± 2.6) (Table 1). The Pielou Index indicated that the high abundance of *Lu. longipalpis*, *Ny. neivai*, and *Ny. whitmani* had a significant impact on the homogeneity of the fauna throughout all months of collection, ranging from 0.51 (August 2018) to 0.63 (May 2019) (Table 1).

Of the total of 12 species, only five (41.6%) were collected in all months and were classified as highly abundant. These included the main vector species *Lu. longipalpis*, *Ny. neivai*, and *Ny. whitmani*. Two species, *Pi. christenseni* and *Sc. sordellii*, were considered frequent, while the remaining five species were categorized as occasional (Table 1).

A total of 224 female specimens, comprising 163 non-engorged (72.8%) and 61 engorged (27.2%), were individually analyzed for the presence of *Leishmania* DNA (Table 2). The 120bp kDNA fragment was amplified in 28 samples (12.5%), including 26 non-engorged and two engorged females. Sanger sequencing identified the presence of *Le. infantum* in 25 of the samples (89.3%), which were obtained from *Ny. neivai* (13), *Lu. longipalpis* (9), *Ev. cortelezzii* (2), and *Ev. lenti* (1). The remaining three samples tested positive for *Leishmania* (*Viannia*) sp.; however, due to the low quality of the sequences, species confirmation could not be obtained. These positive samples were obtained from *Lu. longipalpis* (2) and *Ny. neivai* (1). The kDNA

Species	Sand fly collections							Total	Constancy Index		
	Aug/18		Jan/19		May/19		Sep/19				
	ď	Ŷ	ď	Ŷ	ď	Ŷ	ď	Ŷ		Status	
Brumptomyia brumpti	-	-	2	1	-	-	-	-	3 (0.4)	Occasional	
Evandromyia cortelezzii	7	15	14	17	4	3	-	1	61 (8.0)	Very abundant	
Evandromyia evandroi	-	-	-	-	-	1	-	-	1 (0.1)	Occasional	
Evandromyia lenti	-	2	8	3	2	1	-	2	18 (2.4)	Very abundant	
Evandromyia termitophila	-	-	-	-	-	1	-	-	1 (0.1)	Occasional	
Lutzomyia longipalpis	129	30	135	30	72	10	25	8	439 (57.6)	Very abundant	
Nyssomyia neivai	16	17	52	26	24	12	-	2	149 (19.6)	Very abundant	
Nyssomyia whitmani	4	3	9	7	27	23	6	1	80 (10.5)	Very abundant	
Pintomyia christenseni	-	-	-	2	-	1	-	-	3 (0.4)	Frequent	
Pintomyia pessoai	-	-	2	-	-	-	-	-	2 (0.3)	Occasional	
Psathyromyia lutziana	-	-	-	1	-	-	-	-	1(0.1)	Occasional	
Scyopemyia sordellii	-	1	-	3	-	-	-	-	4 (0.5)	Frequent	
Total (%)	156	68	222	90	129	52	31	14	762	-	
	224 (29.4)		312 (40.9)		181 (23.8)		45 (5.9)		(100)		
Species richness	6		10		8		5		12	-	
Chao 1 estimator (± SD)	8.3 (1.1)		10.6 (2.1)		12.8 (3.0)		13.5 (2.6)		13.5 (2.6)	-	
Shannon diversity (H)	0.92		1.35		1.31		0.87		1.27	-	
Simpson diversity (D)	0.53		0.35		0.31		0.55		0.38	-	
Pielou evenness (J)	0.51		0.58		0.63		0.54		0.51	-	

Table 1. Number of sand flies collected in the municipality of Iguatama, Minas Gerais, Brazil, by sex in the months of August 2018, January 2019, May 2019, and September 2019, and ecological indices.

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sequences were deposited in the GenBank database under accession numbers PP430066-PP430091.

A total of 61 engorged females were tested for blood source identification. Among them, 24 samples (39.3%), representing six species, tested positive by *cytb*-PCR for blood meal identification. Sanger sequencing revealed that the females had fed on four species of vertebrates: *Homo sapiens* (83.3%), *Rattus rattus* (8.3%), *Canis familiaris*, and *Gallus gallus* (both 4.2%). Females of *Ev. lenti* (1), *Lu. longipalpis* (9), and *Ny. whitmani* (1) had fed exclusively on *H. sapiens*, while the single female of *Sc. sordellii* had fed on *R. rattus*. *Nyssomyia neivai* females had fed on *H. sapiens* (9) and *G. gallus*, whereas *Ev. cortelezzii* females had fed on *H. sapiens*, *C. familiaris*, and *R. rattus* (Table 3). No instances of mixed blood meal (two or more blood sources simultaneously detected in the same sand fly) was observed in the engorged females. The *cytb* sequences were deposited in the GenBank database under accession numbers PP430092- PP430114.

## Discussion

The findings of this study shed light on the sand fly fauna of the municipality of Iguatama, Minas Gerais (MG), Brazil, including *Leishmania* infection rates and blood meal sources. The predominant sand fly species identified were *Lu. longipalpis*, *Ny. neivai*, and *Ny. whitmani*, which together constituted the majority (88%) of the collected specimens. These species have been previously implicated as important vectors in the transmission of VL and CL. The detection of *Leishmania* DNA in a considerable proportion of sand fly specimens, with *Le. infantum* being the predominant species identified, highlights the potential role of these sand fly species



**Fig 2. Species accumulation curve for the sand fly fauna of the municipality of Iguatama, Minas Gerais, Brazil.** The numbers 1 to 4 on the x-axis refer to collections performed from August 2018 to September 2019, while numbers 5 and 6 refer to extrapolated hypothetical collections to produce the trend curve by non-linear logarithmic regression analysis. The y-axis represents species richness. The black line indicates the number of collected species while the dotted line indicates the number of species estimated by regression analysis.

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in the transmission cycle of leishmaniasis. Furthermore, the identification of diverse blood meal sources suggests the opportunistic feeding behavior of the sampled sand flies. These results provide important insights into the ecology and potential transmission dynamics of leishmaniasis in the study area.

The sand fly fauna in the municipality of Iguatama was found to consist of 12 species of seven genera. Although the species accumulation curve reached its peak after the third collection, the non-linear logarithmic regression analysis indicated that the sampling effort may not have been sufficient to capture all sand fly species in the study area. This result was further

Species	Total number of females	Positive samples (%)	Leishmania infantum	Leishmania (Viannia) sp.	Positivity rate	
Ny. neivai	57	14 (24.5)	13	1	24.6%	
Lu. longipalpis	78	11 (14.1)	9	2	14.1%	
Ev. lenti	8	1 (12.5)	1	-	12.5%	
Ev. cortelezzii	36	2 (5.5)	2	-	5.6%	
Br. brumpti	1	-	-	-	-	
Ev. evandroi	1	-	-	-	-	
Ev. termitophila	1	-	-	-	-	
Ny. whitmani	34	-	-	-	-	
Pi. christenseni	3	-	-	-	-	
Pa. lutziana	1	-	-	-	-	
Sc. sordellii	4	-	-	-	-	
Total (%)	224 (100)	28 (12.5)	25 (89.2)	3 (10.8)	-	

Table 2. Molecular detection of *Leishmania* in female sand flies collected with CDC light traps in the municipality of Iguatama, Minas Gerais, Brazil, from August 2018 to September 2019.

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Sand fly species	Total number of engorged females	Cytb-PCR positive	Blood meal source				
			Homo sapiens	Canis familiaris	Gallus gallus	Rattus rattus	
Ev. cortelezzii	8 (13.1)	3 (12.5)	1	1	-	1	
Ev. lenti	1 (1.6)	1 (4.2)	1	-	-	-	
Lu. longipalpis	22 (36.3)	8 (33.3)	8	-	-	-	
Ny. neivai	26 (42.6)	10 (41.6)	9	-	1	-	
Ny. whitmani	2 (3.2)	1 (4.2)	1	-	-	-	
Pa. lutziana	1 (1.6)	0 (0)	-	-	-	-	
Sc. sordellii	1 (1.6)	1 (4.2)	-	-	-	1	
Total (%)	61 (100)	24 (39.3)	20 (83.3)	1 (4.2)	1 (4.2)	2 (8.3)	

Table 3. Total number of engorged female sand flies and vertebrate species identified from their blood meals, for collections from the municipality of Iguatama, Minas Gerais, Brazil.

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supported by the *Chao*1 Index, which estimated the presence of 13 species (range 11–15 species). However, the highest species richness was observed in January and May 2019, with ten and eight species collected, respectively. Overall, despite the significant presence of *Lu. longipalpis*, the Shannon Index indicated a high level of diversity (H = 1.27), while the Pielou Evenness Index (J = 0.51) suggested reduced evenness due to the dominance of certain species. The presence of *Lu. longipalpis* in the urban area of Iguatama, as well as in other cities in the Midwest Region of MG [25–27], highlights its role in the transmission of *Le. infantum* in this endemic region. The year-round presence of *Ny. neivai* and *Ny. whitmani*, important vectors of *Le. braziliensis* in Southeast Brazil [28], is also of epidemiological significance, considering the reported cases of human CL from 2007 to 2021 [29]. *Evandromyia lenti* and *Ev. cortelezzii* were consistently collected throughout all sampling periods. These species exhibit similar behavior and adaptability to urban areas of MG [25,27,30].

The detection of *Leishmania* DNA in *Lu. longipalpis* (14.1%) reinforces its significance in the parasite life cycle, as it has been shown to support late-stage infections of various *Leishmania* species both experimentally [31,32] and naturally [33–35]. Natural infections and molecular detections of *Leishmania*, primarily *Le. infantum*, in *Lu. longipalpis* are commonly reported in Brazilian areas endemic for VL, highlighting the significance of this species in the parasite life cycle [5,25,27,35]. While *Lu. longipalpis* is considered a permissive species, it has yet to be confirmed as a proven vector of *Le. (Viannia)* parasites. However, the molecular findings of parasites belonging to this subgenus draw attention to the possibility of its involvement in the transmission cycle of CL in Iguatama.

*Nyssomyia neivai* is a confirmed vector of *Le. braziliensis*, primarily in South Brazil. This sand fly has been reported carrying *Le. braziliensis* DNA in the states of Paraná [36] and Rio Grande do Sul [37]. Moreover, its role in transmitting *Le. infantum* is also suspected, as molecular detections have identified this parasite in the states of Paraná [38], Santa Catarina [39], and MG in the Southeast Region of the country [40].

Several species of the genus *Evandromyia*, particularly *Ev. lenti* and *Ev. cortelezzii*, have been suggested as potential vectors of *Leishmania* in Brazil. *Evandromyia lenti* appears to be widely distributed, at least in the Southeast Region of the country and shares similar ecological preferences with *Lu. longipalpis* [41], being predominantly found in peridomestic areas [42] and domestic animal shelters in rural areas [43], as observed in Iguatama where this species was collected throughout all study periods. Similarly, *Ev. cortelezzii* seems to be adapting to urban and peri-urban areas in the state of MG, such as in the state capital Belo Horizonte [30]. Molecular detections of *Le. infantum* in *Ev. lenti* [22,44,45] and *Ev. cortelezzii* [27,44,46–48] raise concerns about their potential role in transmitting this parasite, although further studies on vector capacity are needed.

The primary blood source for female sand flies in the municipality of Iguatama was humans, which raises epidemiological concerns, particularly with regards to the vectors Lu. longipalpis and Ny. whitmani, which exclusively fed on humans. Evandromyia cortelezzii exhibited a more eclectic feeding habit, having fed on dogs, rats, and humans, as previously reported [49]. This feeding versatility, coupled with the presence of *Le. infantum*, reinforces the potential role of sand flies in transmitting this parasite to multiple hosts. Nyssomyia neivai was the only species that fed on chickens. Although chickens are refractory to Leishmania infection, their presence in peridomestic sites provides a valuable blood source for maintaining the sand fly population [50]. Moreover, high molecular prevalence of Leishmania has been reported in sand flies that fed on chicken blood, indicating that the quality of chicken blood supports the development of *Leishmania* in sequential blood meals [51,52]. A single engorged female of *Sc. sordellii*, a species known to feed on cold-blooded animals such as frogs [53], was found, and DNA sequencing revealed rats as the blood source. This finding warrants further investigation, as it represents the first report of this sand fly feeding on potential vertebrate hosts of Leishmania. Interestingly, Sc. sordellii has been found carrying Leishmania DNA in several regions of Brazil [54-57].

In conclusion, this study provides valuable insights into the sand fly fauna of the municipality of Iguatama, Minas Gerais, Brazil, including *Leishmania* detection rates and blood meal sources. The presence of diverse sand fly species, including the predominant vectors, *Lu. longipalpis, Ny. neivai*, and *Ny. whitmani*, highlights the potential for VL and CL occurrence in this region. The detection of *Leishmania* DNA, primarily *Le. infantum*, in sand fly specimens emphasizes their potential role in the parasite's life cycle and raises concerns about their involvement in disease transmission. Additionally, the identification of humans as the predominant blood source for sand flies, along with the feeding habits observed in other species, underscores the risk of human exposure to *Leishmania* parasites. These findings contribute to understanding the epidemiology of leishmaniasis in the municipality of Iguatama, underscoring the importance of vector surveillance, especially in areas where sand flies positive for *Leishmania* were found and where dogs seropositive for VL were previously detected [13]. Geospatial analysis of human and canine cases of leishmaniasis can help identify critical points where the disease occurs. Altogether, these measures, associated with health education, should assist local authorities in mitigating disease transmission.

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#### References

- Bruschi F, Gradoni L. The leishmaniases: Old neglected tropical diseases. 1st ed. The Leishmaniases: Old Neglected Tropical Diseases. Berlim, Germany: Springer International Publishing; 2018. https:// doi.org/10.1007/978-3-319-72386-0
- 2. PAHO. Leishmaniases. Epidemiological Report of the Americas, December 2020. 2020.
- Brasil. Ministério da Saúde. Secretaria de Vigilância em Saúde. Departamento de Vigilância Epidemiológica. Manual de vigilância e controle da leishmaniose visceral. Ministério da Saúde. Secretaria de Vigilância em Saúde. Departamento de Vigilância Epidemiológica., editor. Brasília: Ministério da Saúde; 2014. p. 120.
- 4. SINAN. Brazilian Notifiable Diseases Information System. In: Ministério da Saúde do Brasil [Internet]. 2020 [cited 30 Apr 2020]. Available: http://portalsinan.saude.gov.br/dados-epidemiologicos-sinan.
- Cardoso DT, de Souza DC, de Castro VN, Geiger SM, Barbosa DS. Identification of priority areas for surveillance of cutaneous leishmaniasis using spatial analysis approaches in Southeastern Brazil. BMC Infect Dis. 2019; 19: 1–11.
- Lamounier VV, Teixeira-Neto RG, Lopes VV, Barbosa JR, Fontes G. Leishmaniose visceral canina autóctone em área indene no Centro Oeste de Minas Gerais, Brasil. Medicina Veterinária (UFRPE). 2017; 11: 179–184.
- 7. Faria MT de, Barbosa FS, Teixeira-Neto RG, Pinheiro GRG, Manhani MN, Marcelino AP, et al. Autochthonous case of Canine Visceral Leishmaniasis in a non-endemic area in Minas Gerais, Brazil. Pesquisa Veterinária Brasileira. 2017; 37: 1505–1508.
- Faria MT de, Barbosa FS, Teixeira-Neto RG, Pinheiro GRG, Manhani MN, Marcelino AP, et al. Autochthonous case of Canine Visceral Leishmaniasis in a non-endemic area in Minas Gerais, Brazil. Pesquisa Veterinária Brasileira. 2017; 37: 1505–1508.
- Vaz TP, Gama-Melo MO, Quaresma PF, Gontijo CMF, Santos G, Barbosa FS, et al. Evaluation of the euthanasia of seropositive dogs for canine visceral leishmaniasis as the only method of controling the disease in the enzootic area in the Midwestern Minas Gerais. Pesquisa Veterinária Brasileira. 2020; 40: 107–112.
- 10. Brasil M da S. Banco de dados do Sistema Único de Saúde-DATASUS. In: Banco de dados do Sistema Único de Saúde-DATASUS [Internet]. 2022 [cited 11 Feb 2023]. Available: http://www.datasus.gov.br.
- Beck HE, Zimmermann NE, McVicar TR, Vergopolan N, Berg A, Wood EF. Present and future Köppen-Geiger climate classification maps at 1-km resolution. Sci Data. 2018; 5: 1–12.
- 12. Brasil. Nota técnica N° 11/2016/CPV/DFIP/SDA/GM/MAPA. Brasília, DF; 2016.
- Vaz TP, Quaresma PF, Rêgo FD, Souza CB, Fontes G, Gontijo CMF. Clinical and Laboratory Response of Domiciled Dogs with Visceral Leishmaniasis Treated with Miltefosine and Allopurinol. Trop Med Infect Dis. 2023; 8: 472. https://doi.org/10.3390/tropicalmed8100472 PMID: 37888600

- Galati EAB. Phlebotominae (Diptera, Psychodidae): classification, morphology and terminology of adults and identification of American Taxa. 1st ed. In: Rangel E, Shaw J, editors. Brazilian Sand Flies: Biology, Taxonomy, Medical Importance and Control. 1st ed. Springer International Publishing; 2018. pp. 9–212.
- Galati EAB, Nunes VLB, Oshiro ET, Rego F de A Jr. Nova espécie de Phlebotominae, Lutzomiya corumbaensis, sp. N. (Diptera, Psychodidae) do complexo Lutzomiya cortelezzii. Balint G, Antala B, Carty C, Mabieme J-MA, Amar IB, Kaplanova A, editors. Rev Bras Entomol. 1989; 33: 465–75. https://doi. org/10.2/JQUERY.MIN.JS
- **16.** Marcondes CB. A proposal of generic and subgeneric abbreviations for phlebtomine sandflies (Diptera: Psychodidae: Phelbotomina) of the world. Entomol News. 2007; 118: 351–356.
- 17. Peet RK. The Measurement of Species Diversity. 1974; 5: 285–307. https://doi.org/10.1146/ ANNUREV.ES.05.110174.001441
- Buckland ST, Magurran AE, Green RE, Fewster RM. Monitoring change in biodiversity through composite indices. Philosophical Transactions of the Royal Society B: Biological Sciences. 2005; 360: 243– 254. https://doi.org/10.1098/rstb.2004.1589 PMID: 15814343
- Linsdale JM. A method of showing relative frequency of occurrence of birds. Condor. 1928; 30: 180– 184.
- Degrave W, Fernandes O, Campbell D, Bozza M, Lopes U. Use of molecular probes and PCR for detection and typing of Leishmania-a mini-review. Mem Inst Oswaldo Cruz. 1994; 89: 463–469. https://doi. org/10.1590/s0074-02761994000300032 PMID: 7476234
- Passos V, Lasmar EB, Gontijo CMF, Fernandes O, Degrave W. Natural infection of a domestic cat (Felis domesticus) with Leishmania (Viannia) in the metropolitan region of Belo Horizonte, State of Minas Gerais, Brazil. Mem Inst Oswaldo Cruz. 1996; 91: 19–20. <u>https://doi.org/10.1590/s0074-02761996000100003</u> PMID: 8734945
- Rêgo FD, Rugani JMN, Shimabukuro PHF, Tonelli GB, Quaresma PF, Gontijo CMF. Molecular detection of Leishmania in phlebotomine sand flies (Diptera: Psychodidae) from a cutaneous leishmaniasis focus at Xakriabá Indigenous Reserve, Brazil. PLoS One. 2015; 10: e0122038. <u>https://doi.org/10.1371/journal.pone.0122038</u> PMID: 25853254
- 23. Carvalho GM de L, Rêgo FD, Tanure A, Silva ACP, Dias TA, Paz GF, et al. Bloodmeal Identification in Field-Collected Sand Flies From Casa Branca, Brazil, Using the Cytochrome b PCR Method. J Med Entomol. 2017; 54: 1049–1054. https://doi.org/10.1093/jme/tjx051 PMID: 28399200
- Steuber S, Abdel-Rady A, Clausen PH. PCR-RFLP analysis: A promising technique for host species identification of blood meals from tsetse flies (Diptera: Glossinidae). Parasitol Res. 2005; 97: 247–254. https://doi.org/10.1007/s00436-005-1410-y PMID: 15999278
- 25. Carvalho GM de L, Silva DF, Xavier L do A, Soares JVR, Ramos VDV, Madureira AP, et al. Sand fly bioecological aspects and risk mapping of leishmaniasis by geographical information systems approach in a mineral exploration area of Brazil. Acta Trop. 2022; 232: 106491. <u>https://doi.org/10.1016/j.actatropica.2022.106491</u> PMID: 35504313
- Nascimento BWL, Saraiva L, Neto RGT, Meira PCLS e., Sanguinette C de C, Tonelli GB, et al. Study of sand flies (Diptera: Psychodidade) in visceral and cutaneous leishmaniasis areas in central western of Minas Gerais state–Brazil. Acta Trop. 2013; 125: 262–268. https://doi.org/10.1016/J.ACTATROPICA. 2012.11.005 PMID: 23178219
- Capucci DC, Campos AM, Soares JVR, Ramos VDV, Binder C, Lima MA, et al. Ecology and natural infection of phlebotomine sand flies in different ecotopes and environments in the municipality of Pains, Minas Gerais, Brazil. Acta Trop. 2023; 238: 106789. https://doi.org/10.1016/j.actatropica.2022.106789 PMID: 36463952
- 28. Brazil RP, Rodrigues AAF, Andrade-Filho JD. Sand Fly Vectors of Leishmania in the Americas—A Mini Review. Entomology, Ornithology & Herpetology. 2015; 4: 1–4.
- 29. SINAN. Brazilian Notifiable Diseases Information System. In: Ministério da Saúde do Brasil [Internet]. 2020 [cited 29 Apr 2020]. Available: http://portalsinan.saude.gov.br/dados-epidemiologicos-sinan.
- Saraiva L, Leite CG, Lima ACVM da R, Carvalho LOA de, Pereira AAS, Rugani JMN, et al. Seasonality of sand flies (Diptera: Psychodidae) and Leishmania DNA detection in vector species in an area with endemic visceral leishmaniasis. Mem Inst Oswaldo Cruz. 2017; 112: 309–318. <u>https://doi.org/10.1590/ 0074-02760160438</u> PMID: 28327794
- da Silva ALFF, Williams P, Melo MN, Mayrink W. Susceptibility of laboratory-reared female Lutzomyia longipalpis (Lutz & Neiva, 1912) to infection by different species and strains of Leishmania Ross, 1903. Mem Inst Oswaldo Cruz. 1990; 85: 453–458.
- 32. Barbosa AF, Oliveira SMP, Bertho ÁL, Franco AMR, Rangel EF. Single and concomitant experimental infectionsby Endotrypanum spp. and Leishmania (Viannia) guyanensis (Kinetoplastida:

Trypanosomatidae) in the Neotropical sand fly Lutzomyia longipalpis (Diptera: Psychodidae). Mem Inst Oswaldo Cruz. 2006; 101: 851–856. <u>https://doi.org/10.1590/s0074-02762006000800005</u> PMID: 17293978

- 33. Paiva BR de Secundino NFC, Nascimento JC do Pimenta PFP, Galati EAB Junior HFA, et al. Detection and identification of Leishmania species in field-captured phlebotomine sandflies based on mini-exon gene PCR. Acta Trop. 2006; 99: 252–259. <u>https://doi.org/10.1016/j.actatropica.2006.08.009</u> PMID: 17055444
- Paiva BR, Oliveira AG, Dorval M, Galati EAB, Malafronte R dos S. Species-specific identification of Leishmania in naturally infected sand flies captured in Mato Grosso do Sul State, Brazil. Acta Trop. 2010; 115: 126–130. https://doi.org/10.1016/j.actatropica.2010.02.013 PMID: 20219438
- 35. Silva MD da, Galvis-Ovallos F, Casanova C, Silva VG da, Leonel JAF, Oliveira TMF de S, et al. Natural infection of Lutzomyia longipalpis (Cembrene-1 population) with Leishmania infantum in a new visceral leishmaniasis focus in the eastern region of São Paulo State, Brazil. Rev Soc Bras Med Trop. 2021; 54.
- 36. Pita-Pereira D de, Souza GD, Zwetsch A, Alves CR, Britto C, Rangel EF. First Report of Lutzomyia (Nyssomyia) neivai (Diptera: Psychodidae: Phlebotominae) Naturally Infected by Leishmania (Viannia) braziliensis in a Periurban Area of South Brazil Using a Multiplex Polymerase Chain Reaction Assay. Am J Trop Med Hyg. 2009; 80: 593–595. https://doi.org/10.4269/ajtmh.2009.80.593 PMID: 19346382
- Thomaz-Soccol V, Goncalves AL, Piechnik CA, Baggio RA, Boeger WA, Buchman TL, et al. Hidden danger: Unexpected scenario in the vector-parasite dynamics of leishmaniases in the Brazil side of triple border (Argentina, Brazil and Paraguay). PLoS Negl Trop Dis. 2018; 12: e0006336. https://doi.org/ 10.1371/journal.pntd.0006336 PMID: 29624586
- Thomaz-Soccol V, Gonçalves AL, Baggio RA, Bisetto A Jr, Celestino A, Hospinal-Santiani M, et al. One piece of the puzzle: Modeling vector presence and environment reveals seasonality, distribution, and prevalence of sandflies and Leishmania in an expansion area. One Health. 2023; 17: 100581. https:// doi.org/10.1016/j.onehlt.2023.100581 PMID: 37332885
- 39. Dias ES, Michalsky ÉM, do Nascimento JC, de Castro Ferreira E, Lopes JV, Fortes-Dias CL. Detection of Leishmania infantum, the etiological agent of visceral leishmaniasis, in Lutzomyia neivai, a putative vector of cutaneous leishmaniasis. Journal of Vector Ecology. 2013; 38: 193–196. <u>https://doi.org/10.1111/j.1948-7134.2013.12028.x PMID: 23701627</u>
- 40. Saraiva L, Carvalho GM de L, Gontijo CMF, Quaresma PF, Lima ACVM da R, Falcão AL, et al. Natural infection of Lutzomyia neivai and Lutzomyia sallesi (Diptera: Psychodidae) by Leishmania infantum chagasi in Brazil. J Med Entomol. 2009; 46: 1159–1163. <u>https://doi.org/10.1603/033.046.0525</u> PMID: 19769049
- Pinto I de S, Ferreira AL, Valim V, Carvalho F dos S, da Silva GM, Falcão AL, et al. Sand fly vectors (Diptera, Psychodidae) of American visceral leishmaniasis areas in the Atlantic Forest, State of Espírito Santo, southeastern Brazil. Journal of Vector Ecology. 2012; 37: 90–96.
- Oliveira AG de, Andrade Filho JD, Falcão AL, Brazil RP. Estudo de flebotomíneos (Diptera, Psychodidae, Phlebotominae) na zona urbana da cidade de Campo Grande, Mato Grosso do Sul, Brasil, 1999– 2000. Cad Saude Publica. 2003; 19: 933–944.
- 43. Galati EAB, Nunes VLB, Dorval MEC, Oshiro ET, Cristaldo G, Espíndola MA, et al. Estudo dos flebotomíneos (Diptera, Pychodidae), em área de leishmaniose tegumentar, no Estado de Mato Grosso do Sul, Brasil. Rev Saude Publica. 1996; 30: 115–128.
- 44. Lopes J V., Michalsky EM, Pereira NCL, De Paula AJV, Lara-Silva FO, Silva-Lana R, et al. Entomological studies in Itaúna, Brazil, an area with visceral leishmaniasis transmission: fauna survey, natural Leishmania Infection, and molecular characterization of the species circulating in phlebotomine sand flies (Diptera: Psychodidae). J Med Entomol. 2019; 56: 1368–1376. https://doi.org/10.1093/jme/tjz061 PMID: 31121044
- **45.** Lana RS, Michalsky ÉM, Fortes-Dias CL, França-Silva JC, Lara-Silva F de O, Lima ACVM da R, et al. Phlebotomine sand fly fauna and Leishmania infection in the vicinity of the Serra do Cipó National Park, a natural Brazilian heritage site. Biomed Res Int. 2015; 2015.
- 46. Carvalho GM de L, Andrade-Filho JD, Falcão AL, Lima ACVM da R, Gontijo CMF. Naturally infected Lutzomyia sand flies in a Leishmania-endemic area of Brazil. Vector Borne Zoonotic Dis. 2008; 8: 407– 414. https://doi.org/10.1089/vbz.2007.0180 PMID: 18429695
- 47. Carvalho GM de L, Silva DF, Xavier L do A, Soares JVR, Ramos VDV, Madureira AP, et al. Sand fly bioecological aspects and risk mapping of leishmaniasis by geographical information systems approach in a mineral exploration area of Brazil. Acta Trop. 2022; 232: 106491. https://doi.org/10.1016/j. actatropica.2022.106491 PMID: 35504313
- **48.** Lana RS, Michalsky EM, Lopes LO, Lara-Silva FO, Nascimento JL, Pinheiro LC, et al. Ecoepidemiological aspects of visceral leishmaniasis in an endemic area in the Steel Valley in Brazil: An ecological

approach with spatial analysis. PLoS One. 2018; 13: e0206452. <u>https://doi.org/10.1371/journal.pone.</u> 0206452 PMID: 30376577

- 49. Tanure A, Peixoto JC, Afonso MM dos S, Duarte R, Pinheiro A da C, Coelho SVB, et al. Identification of sandflies (Diptera: Psychodidae: Phlebotominae) blood meals in an endemic leishmaniasis area in Brazil. Rev Inst Med Trop Sao Paulo. 2015; 57: 321–324. <u>https://doi.org/10.1590/S0036-46652015000400008 PMID: 26422156</u>
- 50. Sant'Anna MR V, Nascimento A, Alexander B, Dilger E, Cavalcante RR, Diaz-Albiter HM, et al. Chicken blood provides a suitable meal for the sand fly Lutzomyia longipalpis and does not inhibit Leishmania development in the gut. Parasit Vectors. 2010; 3: 1–11.
- Ferreira T de S, Timbo RV, Minuzzi-Souza TTC, de Almeida Rocha D, Neiva M, de Albuquerque Ribeiro J, et al. High molecular prevalence of Leishmania in phlebotomine sand flies fed on chicken blood in Brazil. Vet Parasitol. 2018; 259: 80–84. https://doi.org/10.1016/j.vetpar.2018.07.004 PMID: 30056989
- Serafim TD, Coutinho-Abreu IV, Oliveira F, Meneses C, Kamhawi S, Valenzuela JG. Sequential blood meals promote Leishmania replication and reverse metacyclogenesis augmenting vector infectivity. Nat Microbiol. 2018; 3: 548–555. https://doi.org/10.1038/s41564-018-0125-7 PMID: 29556108
- Costa JCR, Marchi GH, Santos CS, Andrade MCM, Chaves Junior SP, Silva MAN, et al. First molecular evidence of frogs as a food source for sand flies (Diptera: Phlebotominae) in Brazilian caves. Parasitol Res. 2021; 120: 1571–1582. https://doi.org/10.1007/s00436-021-07154-3 PMID: 33852067
- 54. Costa TS da, Ferreira RM dos A, Santos GS, Garcia Júnior MD, Pinto CB, Souto RNP. Ecological aspects and molecular detection of Leishmania DNA (Kinetoplastida: Trypanosomatidae) in phlebotomine sand flies (Diptera: Psychodidae) from a rural settlement in the Eastern Amazon, Brazil. Rev Bras Entomol. 2021; 65.
- 55. Carvalho GM de L, Brazil RP, Rêgo FD, Ramos MCNF, Zenóbio APL de A, Andrade-Filho JD. Molecular detection of Leishmania DNA in wild-caught phlebotomine sand flies (Diptera: Psychodidae) from a cave in the state of Minas Gerais, Brazil. J Med Entomol. 2017; 54: 196–203. https://doi.org/10.1093/jme/tjw137 PMID: 28082647
- 56. Chagas EC da S, Silva AS, Fé NF, Ferreira LS, Sampaio V de S, Terrazas WCM, et al. Composition of sand fly fauna (Diptera: Psychodidae) and detection of Leishmania DNA (Kinetoplastida: Trypanosomatidae) in different ecotopes from a rural settlement in the central Amazon, Brazil. Parasit Vectors. 2018; 11: 1–10.
- 57. Da Silva YY, Sales KGDS, Miranda DEDO, Figueredo LA, Brandão-Filho SP, Dantas-Torres F. Detection of Leishmania DNA in Sand Flies (Diptera: Psychodidae) From a Cutaneous Leishmaniasis Outbreak Area in Northeastern Brazil. J Med Entomol. 2020; 57: 529–533. https://doi.org/10.1093/jme/tjz189 = PMID: 31693145