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**Citation:** Abdoli A, Ghaffarifar F, Sharifi Z, Taghipour A (2024) *Toxoplasma gondii* infection and testosterone alteration: A systematic review and meta-analyses. PLoS ONE 19(4): e0297362. https://doi.org/10.1371/journal.pone.0297362

**Editor:** Rajakumar Anbazhagan, National Institute of Child Health and Human Development (NICHD), NIH, UNITED STATES

Received: July 15, 2023

Accepted: January 3, 2024

Published: April 3, 2024

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**Data Availability Statement:** All relevant data are within the paper.

**Funding:** The author(s) received no specific funding for this work.

**Competing interests:** The authors have declared that no competing interests exist.

Abbreviations: ASD, autism spectrum disorder; CNS, central nervous system; DHEA, dehydroepiandrosterone; FSH, follicle-stimulating hormone; GSH, Glutathione; T. gondii:, RESEARCH ARTICLE

# *Toxoplasma gondii* infection and testosterone alteration: A systematic review and metaanalyses

#### Amir Abdoli<sup>1,2</sup>\*, Fatemeh Ghaffarifar<sup>3</sup>\*, Zohreh Sharifi<sup>4</sup>, Ali Taghipour<sup>1,2</sup>

1 Zoonoses Research Center, Jahrom University of Medical Sciences, Jahrom, Iran, 2 Department of Parasitology and Mycology, Jahrom University of Medical Sciences, Jahrom, Iran, 3 Department of Parasitology, Faculty of Medical Sciences, Tarbiat Modares University, Tehran, Iran, 4 Blood Transfusion Research Center, High Institute for Research and Education in Transfusion Medicine, Tehran, Iran

\* ghafarif@modares.ac.ir (FG); a.abdoli25@gmail.com (AA)

# Abstract

# Background

*Toxoplasma gondii* (*T. gondii*) is a worldwide distributed protozoan parasite which has infected a wide range of warm-blooded animals and humans. The most common form of *T. gondii* infection is asymptomatic (latent); nevertheless, latent toxoplasmosis can induce various alterations of sex hormones, especially testosterone, in infected humans and animals. On the other hand, testosterone is involved in behavioral traits and reproductive functions in both sexes. Hence, the purpose of this systematic review is to summarize the available evidence regarding the association between *T. gondii* infection and testosterone alteration.

# Methods

In the setting of a systematic review, an electronic search (any date to 10 January 2023) without language restrictions was performed using Science Direct, Web of Science, PubMed, Scopus, and Google Scholar. The PRISMA guidelines were followed. Following the initial search, a total of 12,306 titles and abstracts were screened initially; 12,281 were excluded due to the lack of eligibility criteria or duplication. Finally, 24 articles met the included criteria. A mean±standard deviation (SD) was calculated to assess the difference of testosterone between *T. gondii* positive and *T. gondii* negative humans. The possibility of publication bias was assessed using Egger's regression. *P*-value < 0.05 was considered statistically significant.

# Results

This systematic review identified 24 articles (18 studies in humans and six studies in animals). Most human studies (13 out of 19) reported an increased level of testosterone following latent toxoplasmosis in males, while three studies reported decreased levels and two studies reported an insignificant change. Eleven articles (seven datasets in males and seven datasets in females) were eligible to be included in the data synthesis. Based on the random-effects model, the pooled mean± SD of testosterone in *T. gondii* positive than *T*. *Toxoplasma gondii*; LH, Luteinizing Hormone; MeSH, Medical Subject Heading; OCD, obsessive compulsive disorder; PRISMA, The Preferred Reporting Items for Systematic reviews and Meta-Analyses; 2D:4D ratio, second to fourth digit ratio. gondii negative was increased by 0.73 and 0.55 units in males and females, respectively. The Egger's regression did not detect a statistically significant publication bias in males and females (p = value = 0.95 and 0.71), respectively. Three studies in male animals (rats, mice, and spotted hyenas) and two studies in female animals (mice and spotted hyenas) reported a decline in testosterone in infected compared with non-infected animals. While, one study in female rats reported no significant changes of testosterone in infected than non-infected animals. Moreover, two studies in male rats reported an increased level of testosterone in infected than non-infected animals.

#### Conclusions

This study provides new insights about the association between *T. gondii* infection and testosterone alteration and identifies relevant data gaps that can inform and encourage further studies. The consequence of increased testosterone levels following *T. gondii* infection could partly be associated with increased sexual behavior and sexual transmission of the parasite. On the other hand, declining testosterone levels following *T. gondii* infection may be associated with male reproductive impairments, which were observed in *T. gondii*-infected humans and animals. Furthermore, these findings suggest the great need for more epidemiological and experimental investigations in depth to understand the relationship between *T. gondii* infection and testosterone alteration alongside with future consequences of testosterone alteration.

# 1. Introduction

*Toxoplasma gondii* (*T. gondii*) is a worldwide prevalent intracellular protozoan parasite which infects about one-third of human and animal populations [1, 2]. The cat family (Felidae) as the definitive hosts and a wide spectrum of warm-blooded vertebrates including humans serve as intermediate hosts [1, 2]. Humans get the infection through ingestion of contaminated foods and water containing oocytes which shed in the cat feces, or by consumption of raw/under-cooked meat containing parasite tissue cysts [2]. Other routes of transmission include organ transplantation and blood transfusion from infected to uninfected individuals, as well as congenital transmission from infected mothers to their fetus [1, 3]. Recent studies also suggested that the parasite could transmit via sexual intercourse in humans [4] and rats [5].

According to estimations, more than one-third of the human population has been infected with the parasite worldwide [2]. Nevertheless, most human infections are asymptomatic in immunocompetent individuals [6]. In immunocompromised individuals, the infection could have life-threatening sequels, such as toxoplasmic encephalitis, myocarditis, or disseminated infections [7, 8]. Congenital toxoplasmosis is also a life-threatening condition which may lead to abortion, stillbirth, and preterm birth [9–12]. The intensity of *T. gondii* infection depends on several factors, including genetic background [13], immunity status [14], and the parasite virulence [15, 16]. *T. gondii* consists of three main strains (Types I, II, and III), which have some differences in virulence factors and epidemiological patterns [17–19]. While type I strains (such as RH and GT-1) are highly virulent, type II strains (e.g., ME49 and PRU) and type III (e.g., VEG, NED, and CEP) have lower virulence than type I strains [19, 20].

Testosterone is the primary male hormone that is responsible for male sex characteristics and reproductive functions, such as spermatogenesis and fertility. Females also need certain levels of testosterone. In females, most testosterone converts into the sex hormone estradiol [21]. Testosterone is primarily produced in the testes and ovaries in males and females, respectively. A small amount of testosterone is produced in the adrenal glands in both sexes [21].

Several studies in humans and animal models revealed that *T. gondii* infection influenced testosterone levels. While some studies reported an increased level of testosterone, others reported a decline level following *T. gondii* infection [22]. It seems that several factors, such as the parasite strain and intensity of infection could influence this variation [22]. Inasmuch as testosterone is important in different physiological processes (e.g., reproductive function and sexual behavior), this systematic review is aimed to summarize data regarding the effects of *T. gondii* infection on testosterone levels in humans and animals and discusses their influential factors.

#### 2. Materials and methods

The present study was conducted following the guideline of the Preferred Reporting Items for Systematic reviews and Meta-Analyses (PRISMA) statement [23] (S1 Checklist).

#### 2.1 Strategy search

The search was performed in international databases (Science Direct, Web of Science, PubMed, and Scopus) and the search engine, Google Scholar, published from any date to 10 January 2023. The following search terms were selected using Medical Subject Heading (MeSH) terms alone or in combination: (*"Toxoplasma gondii"* OR *"T. gondii"* OR "toxoplasmosis") AND ("testosterone" OR "hormone" OR "androgen"). Additionally, to avoid ignoring the reference lists of all included studies were reviewed. As such, the citations of all selected articles were hand-searched in Google Scholar for potentially eligible articles.

#### 2.2 Eligibility criteria, study selection, and data extraction

Two independent reviewers (AA and AT) selected the articles. After the initial search, all selected articles were screened by title and abstract, then the relevant articles were imported into the EndNote X8 software (Thomson Reuters, New York, USA). Duplicated articles were checked and removed in the next step. Then, if the articles met the following criteria, they were included in the systematic review: (1) papers with full-text or abstract in English, and (2) original research articles, short reports or letters to the editors that studied the association between *T. gondii* infection and testosterone. Articles were included if they fulfilled the following Population, Intervention, Comparison and Outcomes (PICO) criteria [24]: Participants/ Population: animals or humans, Interventions/exposure: *T. gondii* infection, Comparison or control: compared with uninfected human or animals, Outcomes: levels of testosterone.

The data extracted and tabulated from each study, including: (1) First Author, (2) Publication Year, (3) Country, (4) Study Design, (5) Type of Population (case and control), and (6) Findings. All extracted data was entered into the respective tables (for humans and animals) by the primary researcher and verified by another researcher. Any discrepancies were reviewed and resolved by consensus.

#### 2.3 Quality assessment

Quality assessment of the included articles was done by The Joanna Briggs Institute (JBI) Critical Appraisal Checklist [25], which contains eight questions with four options including Yes, No, Unclear, and Not applicable. For including and excluding papers, each paper takes a maximum of one star for each numbered item and the total score of 4–6 and 7–10 points were specified as moderate and high quality, respectively. Based on the obtained score, the authors have decided to include (4–10 points) and exclude ( $\leq$ 3 points) the papers.

#### 2.4. Data synthesis and statistical analysis

Data was analyzed using comprehensive meta-analysis software version 2. To assess the association between *T. gondii* with testosterone in humans, a mean $\pm$ standard deviation (SD) using the random effects model and corresponding 95% confidence intervals (CI) were calculated for each study. Egger's regression (Qualitative method) was applied to assess the possibility of publication bias during the analysis. *P*-value < 0.05 was considered statistically significant.

#### 3. Results

#### 3.1 Study selection

As shown in the PRISMA flowchart (Fig 1), a total of 12,306 titles and abstracts were screened initially; 12,281 were excluded due to the lack of eligibility criteria or duplication. Finally, 24 articles (18 studies in human and six animal studies) met the included criteria. Tables 1 and 2 summarize the information of the included articles regarding the association between *T. gondii* infection and testosterone in humans and animals, respectively.

#### 3.2 Quality assessment

The results of quality assessment according to JBI for eligible studies are depicted in Tables 1 and 2. The included articles in the present meta-analysis showed an acceptable quality.



Fig 1. PRISMA flow diagram describing included/excluded studies.

https://doi.org/10.1371/journal.pone.0297362.g001

# PLOS ONE

NO	First Author, Publication Year, and Country	Study aims and design	Studied groups	Findings	QA
1	Flegr et al. [40] 2008 Czech Republic	•A case-control study to assess the relationship between <i>T. gondii</i> seropositivity with testosterone levels.	<ul> <li>A group of 174 female and 91 male students.</li> <li>29 (16.7%) females and 23 (25.3%) males were <i>T. gondii</i> seropositive.</li> </ul>	► <i>Toxoplasma</i> -infected <b>men</b> have a <u>higher</u> concentration of <b>testosterone</b> and <i>Toxoplasma</i> -infected <b>women</b> have a <u>lower</u> concentration of <b>testosterone</b> than <i>Toxoplasma</i> -free controls.	9
2	Flegr et al. [ <u>41</u> ] 2008 Czech Republic	•A case-control study to test the relationship between <i>T. gondii</i> seropositivity with testosterone levels and 2D:4D ratio.	<ul> <li>A group of 194 females and 106 males.</li> <li>31 (15.9%) females and 25 (23.6%) males were <i>T. gondii</i> seropositive.</li> </ul>	<ul> <li>Infected males had higher and infected females had lower testosterone levels than <i>Toxoplasma</i>-free males and females, respectively (<i>P</i> = 0.007).</li> <li><i>Toxoplasma</i>-infected males had a lower left hand 2D:4D ratio than Toxoplasma-free males (<i>P</i> = 0.008).</li> </ul>	9
3	Shirbazou et al. [26] 2011 Iran	•A case-control study to investigate the relationship between <i>T. gondii</i> seropositivity with serum cortisol and testosterone levels, as well as with depression, anxiety, and stress index.	<ul> <li>180 people (73 females and 107 males) healthy individuals.</li> <li>24 females (13/33%) and 39 males (21/66%) had anti-<i>Toxoplasma</i> IgG antibody.</li> <li>12 females and 19 males without <i>Toxoplasma</i> gondii IgG antibody</li> </ul>	<ul> <li>Serum cortisol and testosterone concentrations were significantly increased in seropositive women and men compared with seronegative counterparts.</li> <li>Stress and anxiety index were also increased in seropositive men and women, whereas depression index increased only in seropositive men.</li> </ul>	9
4	Abdul-Lateef et al. [32] 2012 Iraq	• A case-control study to investigate the relationship between <i>T. gondii</i> seropositivity with testosterone, IFN- and IL-12 serum levels.	•77 healthy individuals with anti- <i>Toxoplasma</i> antibodies (40 females and 37 males) and 30 (15 females and 15 males) seronegative control	► <i>T. gondii</i> seropositive women and men had significantly <u>higher</u> levels of IL-12, IFN- $\gamma$ , and testosterone than seronegative control group.	8
5	Eslamirad et al. [28] 2013 Iran	•A case-control study to investigate the relationship between chronic <i>T. gondii</i> infection with serum testosterone in men	<ul> <li>1026 healthy men who referred to Arak Post Marriage Center.</li> <li>365 men with <i>T. gondii</i> IgG antibody were selected as case group and 365 seronegative men was selected as control group</li> </ul>	► <b>Testosterone</b> concentration in case group (seropositive <b>men</b> ) was <u>decreased</u> than control group and this difference was statistically significant ( <i>P</i> <0.05).	10
6	Eslamirad et al. [27] 2014 Iran	•A case-control study to examine the relationship between <i>T. gondii</i> infection with serum testosterone and lipid profile in healthy men	•100 men with <i>T. gondii</i> IgG antibody and equal number of men without <i>T. gondii</i> antibodies	<ul> <li>Testosterone levels level was significantly lower among seropositive men</li> <li>No significant difference was found between <i>T. gondii</i> seropositivity and serum lipid levels.</li> </ul>	9
7	Mahbodfar et al. [29] 2015 Iran	•A case-control study to examine the relationship between <i>T. gondii</i> infection with serum testosterone, DHEA, cortisol and prolactin among young persons •The prevalence of hirsutism, acne and alopecia were also investigated	<ul> <li>•215 (106 women and 107 men) blood samples (age 18–35 years).</li> <li>•61 men and 58 women were seropositive for (IgG) <i>T. gondii</i> antibodies</li> <li>•47 men and 49 women were seronegative for (IgG) <i>T. gondii</i> antibodies</li> </ul>	<ul> <li>A significant <u>increase</u> in testosterone and cortisol was found in <i>T. gondii</i> seropositive individuals, but not for DHEA.</li> <li>A significantly <u>increased</u> in the rates of alopecia and acne in seropositive men than seronegative men.</li> <li>A significantly <u>increased</u> in the rates of hirsutism in seropositive women than seronegative women.</li> </ul>	10
8	Colosi et al. [42] 2015 Romania	•A case-control study to examine the relationship between <i>T. gondii</i> infection with male fertility in human	<ul> <li>60 immunocompetent males.</li> <li>15 men with <i>T. gondii</i> IgG antibody</li> <li>45 men without <i>T. gondii</i> IgG antibody</li> </ul>	<ul> <li>Serum testosterone concentration in seropositive men was lower than seronegative individuals, but it was not statistically significant (P = 0.62).</li> <li>Serum FSH concentration in seropositive men was increased than seronegative individuals, but it was not statistically significant (P = 0.97).</li> <li>Sperm characteristics (ejaculate quantity, sperm count, motility, morphology) was not statistically significant changed (P&gt;0.05) in <i>T. gondii</i> infected than noninfected individuals.</li> </ul>	10

#### Table 1. Association of *T. gondii* infection with testosterone alteration in human in the case-control studies.

(Continued)

NO	First Author, Publication Year, and Country	Study aims and design	Studied groups	Findings	QA
9	Zghair et al. [33] 2015 Iraq	•A case-control study to investigate the relationship between chronic <i>T. gondii</i> infection with total testosterone, free testosterone and FSH levels healthy men	<ul> <li>400 apparently healthy blood donor males were evaluated for <i>T. gondii</i> antibodies.</li> <li>10 <i>T. gondii</i>-IgM positive and 121 <i>T. gondii</i>-IgG positive were enrolled as case group.</li> <li>30 seronegative individuals were enrolled as control group</li> </ul>	<ul> <li>Concentrations of total and free</li> <li>testosterone were significantly higher in</li> <li>both IgM and IgG seropositive men</li> <li>compared with seronegative control group</li> <li>Concentration of FSH was not significant</li> <li>differences in both IgM and IgG</li> <li>seropositive men compared with</li> <li>seronegative control group</li> </ul>	10
10	Zouei et al. [30] 2018 Iran	•A case-control study to investigate the relationship between <i>T. gondii</i> seropositivity with serum testosterone in men and women	•76 positive sera were selected as case group (38 from men and 38 from women) and a same number of negative sera as control group	► The mean concentration of serum testosterone was statistically <u>higher</u> in <i>T</i> . <i>gondii</i> - seropositive <b>men and women</b> compared to non-infected men and women.	9
11	Borráz-León et al. [43] 2021 Mexico	•A case-control study to examine the relationship between <i>T. gondii</i> infection with serum testosterone, Interpersonal Sensitivity and Psychoticism symptoms among men and women	<ul> <li>•213 healthy subjects (males = 108, females = 105) were enrolled.</li> <li>•22 and 13 men and women had <i>T. gondii</i> IgG antibody</li> <li>86 and 92 women were seronegative for <i>T. gondii</i> IgG antibody</li> </ul>	<ul> <li><i>Toxoplasma</i>-seropositive men had higher testosterone levels (<i>P</i>&lt;0.001), Interpersonal Sensitivity (<i>P</i>&lt;0.03) and Psychoticism symptoms (<i>P</i>&lt;0.037) than non-infected men.</li> <li><i>Toxoplasma</i>-infected women did not differ from control women.</li> </ul>	10
12	Kadhim and AL- awadi. [ <u>34</u> ] 2013 Iraq	•A case-control study to examine the relationship between <i>T. gondii</i> infection with serum testosterone, progesterone and prolactin levels in pregnant women with chronic toxoplasmosis	•A total number of 55 <i>T. gondii</i> -IgG seropositive pregnant women and 51 seronegative pregnant women were enrolled as case and control groups, respectively	► Toxoplasma-seropositive women had a significant <u>higher</u> level of testosterone, <u>but</u> not progesterone and prolactin than Toxoplasma-seronegative women.	8
13	Al-Masoudi et al. [35] 2018 Iraq	•A case-control study to examine the relationship between toxoplasmosis and, testosterone and LH hormones	<ul> <li>66 healthy subjects were enrolled</li> <li>20 (male = 15 and female = 5) seropositive individuals were enrolled as case and the same number of healthy subjects were enrolled as control group</li> </ul>	► A <u>decreased</u> level of <b>testosterone</b> and an <u>increased</u> level of <b>LH</b> were found in <i>T</i> . <i>gondii</i> seropositive individuals compared to control.	8
14	Al-Kurdy et al. [36] 2020 Iraq	•A case-control study to examine the relationship between toxoplasmosis and testosterone in healthy <b>men</b>	•38 <i>T. gondii</i> seropositive <b>men</b> and the same number of seronegative men were enrolled as the case and control groups, respectively	► No statistical differences were found in concentration of testosterone among the case than the control group.	8
15	AL-Asady [37] 2017 Iraq	•A case-control study to examine the relationship between toxoplasmosis with testosterone, FSH and LH in healthy pregnant women	•59 <i>T. gondii</i> -IgG positive <b>pregnant</b> <b>women</b> and 28 <i>T. gondii</i> -IgG negative pregnant women were enrolled as case and control groups, respectively	► The result showed very slightly higher serum levels of testosterone and LH and an insignificant lower level of FSH were detected in seropositive women compared to controls.	9
16	El-Gebaly et al. [38] 2019 Egypt	•A case-control study to assess seroprevalence/serointensity of toxoplasmosis in schizophrenic patients in relation to the levels of testosterone, cortisol and GSH activity	•120 schizophrenic inpatients were compared with 120 individuals attending the outpatients' clinics	<ul> <li>In <i>T. gondii</i>-seropositive patients, testosterone was higher in both genders and glutathione was lower, while no significant difference was documented in relation to PANSS, treatment with electroconvulsive-therapy (ECT) or cortisol level.</li> <li>Schizophrenic patients showed higher <i>Toxoplasma</i> antibody titer, cortisol, and free testosterone levels in both genders and lower GSH than control.</li> </ul>	9

(Continued)

NO	First Author, Publication Year, and Country	Study aims and design	Studied groups	Findings	QA
117	Bayani et al. [31] 2022 Iran	•A case-control study to examine the relationship between toxoplasmosis with testosterone, prolactin, DHEA, FSH, LH, and TSH among <i>T. gondii</i> infected and uninfected infertile couples	<ul> <li>•376 (188 males and 188 females) were enrolled.</li> <li>•<i>T. gondii</i>-IgG and IgM seropositivity were detected in 56.9% (107/188) and 6.5% (7/107) of <b>females</b>, respectively.</li> <li>•<i>T. gondii</i>-IgG and IgM seropositivity were detected in 111/188 (59.0%) and 9/111 (8.1%) of males, respectively.</li> </ul>	<ul> <li>In females, DHEA was lower and the mean level of prolactin, LH, FSH and TSH were higher among seropositive cases compared with seronegative cases, but there were no statistically significant differences. Testosterone was unchanged among seropositive and seronegative females.</li> <li>A positive correlation was seen between toxoplasmosis and the upper and lower ranges of the normal value of prolactin in females (x 2 = 6.5, p = 0.039) but not in male cases (x 2 = 1.06, p = 0.59).</li> <li>In males, the mean level of testosterone and TSH were higher and the mean level of prolactin and DHEA were lower among seropositive cases, but there were mostatistically significant differences.</li> <li>A positive cases, but there were mostatistically significant differences.</li> <li>A positive sasociation was observed between <i>T. gondii</i> infection and the upper and lower ranges of the normal value of testosterone in males (x 2 = 6.8, p = 0.033) but not in females (x 2 = 0.62, p = 0.99).</li> </ul>	10
18	Hagag et al. [39] 2022 Egypt	•A case-control study to examine the association of latent toxoplasmosis with testosterone levels among androgenic alopecia and acne vulgaris patients	•30 androgenic and alopecia and 30 acne vulgaris patients	<ul> <li>There was a statistical significance relationship between <i>T. gondii</i> seropositivity with androgenic alopecia severity (<i>P</i> = 0.001).</li> <li>There was a significant elevation of free testosterone in seropositive subgroups of androgenic alopecia compared with seronegative group.</li> <li>There was a statistical significance between <i>T. gondii</i> seropositivity with acne vulgaris severity (<i>P</i> = 0.019)</li> <li>There was a significant elevation of free testosterone in seropositive subgroups of acne vulgaris compared with seronegative group.</li> </ul>	10

2D:4D ratio: second to fourth digit ratio, DHEA: dehydroepiandrosterone, FSH: follicle-stimulating hormone, LH: Luteinizing Hormone, GSH: Glutathione, TSH: thyroid stimulating hormone, QA: Quality Assessment.

https://doi.org/10.1371/journal.pone.0297362.t001

#### 3.3 Description of included studies

**3.3.1 Human studies.** In human studies, 18 articles were included (Table 1). The studies were reported from six countries, including Iran (six studies [26–31]), Iraq (six studies [32–37]), Egypt (two studies [38, 39]), and each of Czech Republic [40, 41], Romania [42], and Mexico [43] with one study (Table 1).

3.3.1.1 Evidence for increased testosterone in human infected with T. gondii. Fig 2 summarizes the included studies and Table 1 represents the details of each study. In males, 9 studies reported an increased level of testosterone in *T. gondii* seropositive individuals compared to seronegative counterparts [29, 30, 32, 33, 38–41, 43]. As such, in females, seven studies reported elevated levels of testosterone in *T. gondii* seropositive than seronegative counterparts [29, 30, 32, 34, 37–39].

NO	First Author, Publication Year, and Country	Animal type	<i>T. gondii</i> strain	Type of inoculation	Findings	QA
1	Kan <sup>*</sup> ková et al. [49] 2011 Czech Republic	Cross-breeds of BALB/c female mice and C57 80 Black male mice of the F1 generation	Virulent strain T38 isolated from oocysts released by a stray cat	<ul> <li>Oral cysts</li> <li>12 female mice and 12 male mice were orally infected with brain homogenate from mice infected with cystogenic but relatively virulent strain T38 of <i>T. gondii.</i></li> <li>21 female mice and 20 male mice were given the same amount of isotonic saline (0.8% NaCl)</li> </ul>	<ul> <li>► Infected mice had significantly <u>lower</u> concentration of testosterone than controls (Tau = 0.271, P = 0.001).</li> <li>► Both female and male mice with latent toxoplasmosis had significantly lower levels of testosterone (females: Z = 2.32, P = 0.020; males: Z = 2.76, P = 0.005)</li> </ul>	10
2	Abdoli et al. [48] 2012 Iran	Male rats (Albino Wistar type)	RH strain (type 1 T. gondii strain)	<ul> <li>Intraperitoneal injection of tachyzoites</li> <li>Case: 35 infected male rats</li> <li>Control: 21 uninfected male rats</li> </ul>	<ul> <li>► A temporary decline in serum and intratesticular testosterone, and fructose in seminal vesicle were observed.</li> <li>► The rates of sperm motility (%), viability (%), and concentration were significantly decreased and sperm abnormality (%) was significantly increased after infection, but it reverts to the normal level on day 60 and 70 post infection.</li> </ul>	9
3	Lim et al. [46] 2013 Singapore	Male Wistar rats	Prugniaud strain (type 2 <i>T. gondii</i> strain)	•Intraperitoneal injection of tachyzoites	► <i>T. gondii</i> infection <u>enhances testicular</u> expression of genes involved in facilitating synthesis of testosterone (LHR, StAR and P450scc), resulting in greater testicular testosterone production.	8
4	Afshari et al. [47] 2013 Iran	Male Wistar rats	<i>RH</i> strain (type 1 <i>T. gondii</i> strain)	<ul> <li>Intraperitoneal injection of tachyzoites</li> <li>Case: 10 infected male rats</li> <li>Control: 10 uninfected male rats</li> </ul>	► Serum alkaline phosphatase and testosterone were significantly <u>increased</u> in case than control group	9
5	Abdulai-Saiku and Vyas. [51] 2017 Singapore	Female Wistar rats	Prugniaud strain (type 2 <i>T. gondii</i> strain)	•Intraperitoneal injection of tachyzoites	<ul> <li><i>T. gondii</i> infection <u>did not change</u> circulating levels of testosterone in the blood</li> <li><i>T. gondii</i> infection <u>did not affect</u> levels of serum estrogen and progesterone in gonadally intact females</li> </ul>	9
6	Laubach et al. [50] 2022 USA	spotted hyenas (Crocuta crocuta)	-	• The relationship between <i>T. gondii</i> infection and plasma testosterone and cortisol levels were investigated among 109 spotted hyenas	<ul> <li>A negative association was found between <i>T. gondii</i> infection and plasma testosterone among female (cubs and subadults) and adult male hyenas, which means that the infected animals have <u>lower testosterone</u> levels than uninfected animals.</li> <li>No associations were found between <i>T. gondii</i> infection and cortisol in any age class or sex group of hyenas.</li> </ul>	9

LHR: Luteinizing hormone receptor, StAR: Steroidogenic acute regulatory, QA: Quality Assessment.

https://doi.org/10.1371/journal.pone.0297362.t002

3.3.1.2 Evidence for a decreased or unchanged level of testosterone in human infected with *T*. gondii. Three studies in males [27, 28, 35] and three studies in females [35, 40, 41] reported a declined level of testosterone in *T. gondii* seropositive than seronegative counterparts. Moreover, three studies in males [31, 36, 42] and two studies in females [31] reported no significant change in testosterone levels in *T. gondii* seropositive than seronegative counterparts (Table 1 and Fig 2).

3.3.1.3 Evidence for increased cortisol levels in human infected with *T. gondii*. Three studies in males [26, 29, 38] and two studies in females [26, 38] reported an increased level of cortisol in *T. gondii* seropositive than seronegative counterparts.

Species	Testoste	rone	Cortiso	ol 🛛
	Male	Female	Male	Female
Humans (Homo sapiens)	▲ Flegr et al. (2008) [1], Flegr et al. (2008) [2], Abdul-Lateef et al. (2012) [3], Mahbodfar et al. (2015) [4], Zghair et al. (2015) [5], Zouei et al. (2018) [6], Borráz-León et al. (2021) [7], El-Gebaly et al. (2019) [8], Hagag et al. (2022) [9] ↔ (Colosi et al. (2015) [10], Al-Kurdy et al. (2020) [11], Bayani et al. (2022) [12] ↓ Eslamirad et al. (2013) [14], Al-Masoudi et al. (2- 18) [15]	A Abdul-Lateef et al. (2012) [3], Mahbodfar et al. (2015) [4], Zouei et al. (2018) [6], Kadhim and AL- awadi (2013) [16], AL-Asady (2017) [17], El-Gebaly et al. (2019) [8], Hagag et al. (2022) [9] ↔ Borráz-León et al. (2021) [7], Bayani et al. (2022) [12] ♥ Flegr et al. (2008) [1], Flegr et al.	A Shirbazou et al. (2011) [18], Mahbodfar et al. (2015) [4], El-Gebaly et al. (2019) [8]	Shirbazou et al. (2011) [18], El-Gebaly et al. (2019) [8]
		(2008) [2], Al-Masoudi et al. (2018) [15]		
Rat (Rattus norvegicus)	Lim et al. [19], Afshari et al. [20] Abdoli et al. [21]	↔ Abdulai-Saiku and Vyas (2017) [22]	▼ Mitra et al. (2013) [23]	
Mice (Mus musculus)				
C	▼ Kaňková et al. (2011) [24]	▼Kaňková et al. (2011) [24]		
Spotted hyenas (Crocuta crocuta)	▼ Laubach et al. (2022) [25]	▼ Laubach et al. (2022) [25]	↔ Laubach et al. (2022) [25]	↔ Laubach et al. (2022) [25]

This table is adopted from Laubach et al. (*Int J Parasitol: Parasit Wildlife. 2022;17:53-9*) with some modification and update.

▲ Increase, ▼Decrease, ↔No significant change.

# Fig 2. A summary of studies on the relationship between *T. gondii* infection, testosterone, and steroid hormone levels in males and females.

https://doi.org/10.1371/journal.pone.0297362.g002

3.3.1.4 Description of human studies. The first studies regarding *T. gondii* and testosterone in humans were conducted by Flegr et al in 2008. In the Czech Republic [40, 41]. They conducted case-control studies among *T. gondii* IgG seropositive female and male students. The results showed that *T. gondii*- seropositive men have a higher concentration of testosterone and *T. gondii*- seropositive women have a lower concentration of testosterone compared with *Toxoplasma*-free subjects [40, 41]. An article was published by a group of researchers in Iran

[26]. They found that *T. gondii* seropositive women and men had a higher concentration of serum cortisol and testosterone than seronegative individuals. As such, a significant association was found between T. gondii seropositivity with hair loss in women, hirsutism in women, and height increase in women and men. Stress and anxiety indices were also increased in T. gondii seropositive men and women, whereas the depression index increased only in seropositive men compared with the control group [26]. Abdul-Lateef et al. [32] found a significant correlation between T. gondii IgG seropositivity with an increase in serum testosterone, IL-12, and IFN- $\gamma$  among an Iraqi population [32]. Eslamirad et al. [28] found an association between T. gondii IgG seropositivity with decreased testosterone levels in healthy men than the seropositivity control group [28], but they did not find an association between T. gondii seropositivity and serum lipid levels [27]. Mahbodfar et al. [29] found that T. gondii seropositive individuals had significantly higher levels of testosterone and cortisol than seronegative individuals. As such, the rates of alopecia and acne were significantly increased in seropositive men than seronegative men, and the rate of hirsutism was significantly increased in seropositive women than seronegative women [29]. Colosi et al. [42] found no statistically significant difference in serum testosterone, follicle-stimulating hormone (FSH), and sperm characteristics among T. gondii seropositive men compared with seronegative individuals. Zghair et al. [33] demonstrated that the levels of total and free testosterone, but not FSH, were significantly higher in T. gondii-seropositive men compared with the seronegative control group. Zouei et al. [30] found a statistically significant increase in the level of serum testosterone among T. gondii- seropositive men and women compared to non-infected men and women in an Iranian population. Borráz-León et al. [43] showed a significantly positive relationship between T. gondii IgG seropositivity with higher testosterone levels, interpersonal sensitivity, and psychoticism symptoms in seropositive men, but not women, than non-infected control groups [34]. A study among T. gondii seropositive and seronegative women revealed an increased level of testosterone, but not progesterone and prolactin, in seropositive women compared with seronegative control groups. Al-Masoudi et al. [35] found a decreased level of testosterone and an increased level of luteinizing hormone (LH) in T. gondii seropositive individuals compared to controls in a healthy Iraqi population. Al-Kurdy et al. [36] found no statistical differences in the concentration of testosterone among the T. gondii seropositive men than seronegative controls. AL-Asady et. Al. [37] found a very slightly higher serum level of testosterone and LH and insignificant lower levels of FSH in seropositive women compared to controls. El-Gebaly et al. [38] demonstrated that schizophrenic patients showed higher *T. gondii* antibody titer, cortisol, and free testosterone levels in both genders and lower Glutathione (GSH) than controls. As such, T. gondii seropositive schizophrenic patients had higher testosterone levels and lower glutathione levels than seronegative patients. Bayani et al. [31] investigated the relationship between toxoplasmosis with testosterone, prolactin, dehydroepiandrosterone (DHEA), FSH, LH, and thyroid stimulating hormone (TSH) among T. gondii seropositive and seronegative infertile couples. Although some alterations were observed, no statistically significant differences were detected in these hormones among T. gondii seropositive and seronegative groups [31]. In an interesting report, Hagag et al. [39] found a positive association between T. gondii seropositivity and a significant elevation of free testosterone levels among patients with androgenic alopecia and acne vulgaris compared with the seronegative group. There are also some case reports regarding the association of acute toxoplasmosis with lower testosterone levels in males with hypogonadotrophic hypogonadism [44] as well as a case with intracranial toxoplasmosis presenting as panhypopituitarism [45].

3.3.1.5 Meta-analysis of human studies. As shown in Table 3, eleven papers (seven datasets in males and seven datasets in females) on the association between *T. gondii* and testosterone were eligible to include in the data synthesis. Based on the random-effects model, the pooled

	-	1			1			
First author         Abdul-Lateef et al., 2012 [32]         Abdul-Lateef et al., 2012 [32]         Bayani et al., 2022 [31]         Hagag et al., 2022 [39]         Hagag et al., 2022 [39]         Kadhim and AL-awadi., 2013 [34]         Colosi et al., 2015 [42]         El-Gebaly et al., 2019 [38]         El-Gebaly et al., 2019 [38]         Al-Masoudi et al., 2018 [35]	Gender	Toxoplasma positi	ve		Toxoplasma negati	P-value		
		Total sample size	Mean (ng/ml)	St.Deviation	Total sample size	Mean (ng/ml)	St.Deviation	
Abdul-Lateef et al., 2012 [32]	Male	37	8.0601	3.04751	15	4.1123	3.17078	0
Abdul-Lateef et al., 2012 [32]	Female	40	0.7213	0.35507	15	0.5249	0.18708	0.011
Bayani et al., 2022 [31]	Mixed †	99	0.6	0.5	71	0.6	0.5	0.9
Hagag et al., 2022 [39]	Male	14	28.01	12.95	16	13.62	6.86	0.001
Hagag et al., 2022 [39]	Female	14	11.98	14.26	16	2.3	1.04	0.001
Kadhim and AL-awadi., 2013 [34]	Female	55	1.95	1.37	51	0.94	0.84	1.80E-05
Colosi et al., 2015 [42]	Male	15	399.07	185.18	45	425.96	170.05	0.62
El-Gebaly et al., 2019 [38]	Male	42	10.8	6.23	39	7	6.59	0.01
El-Gebaly et al., 2019 [38]	Female	12	8.5	9.62	27	2.2	1.65	0.003
Al-Masoudi et al., 2018 [35]	Male	4	0.85	6.25	4	0.73	5.95	NR
Al-Masoudi et al., 2018 [35]	Female	8	0.3	0.87	8	0.31	0.54	NR
Mahbodfar et al., 2015 [29]	Mixed †	119	5.83	5.39	96	3.38	3.92	0
Zouei et al., 2018 [30]	Male	38	5.6	1.99	38	4.56	1.96	NR
Zouei et al., 2018 [30]	Female	38	0.41	0.22	38	0.31	0.17	NR
Zghair et al., 2015 [33]	Mixed †	121	6.515	0.51	30	6.78	0.61	NR
Borráz-León et al., 2021 [43]	Male	22	7.78	2.66	86	4.32	2.82	<0.001
Borráz-León et al., 2021 [43]	Female	13	0.63	0.37	92	1.18	1.69	0.49

Table 3. Included studies on the association between T. gondii positive and T. gondii negative with testosterone.

† Not included in Meta-analysis.

https://doi.org/10.1371/journal.pone.0297362.t003

mean± SD of testosterone in *T. gondii* positive than *T. gondii* negative were calculated to be 0.73 and 0.55 in males and females, respectively (Figs 3 and 4). It means that, testosterone increased by 0.73 and 0.55 units in *T. gondii* positive compared to *T. gondii* negative males and females, respectively. The publication bias was not statistically significant in males (p = 0.95) and females (p = 0.71), respectively.

**3.3.2 Animal studies.** *3.3.2.1 Evidence for increased levels of testosterone in animals infected with T. gondii.* Two studies in rats [46, 47] reported an increased level of testosterone in infected than non-infected animals (Fig 2 and Table 2).

Study name		Statistics	for each st	tudy		Std diff in means and 95% CI				
	Std diff in means	Standard error	Variance	Lower limit	Upper limit					
Abdul-Lateef et al., 2012	1.281	0.331	0.109	0.632	1.929					$\rightarrow$
Hagag et al., 2022	1.417	0.409	0.167	0.615	2.219					
Colosi et al., 2015	-0.155	0.298	0.089	-0.740	0.430				_	
El-Gebaly et al., 2019	0.593	0.227	0.052	0.148	1.039			-		
Al-Masoudi et al., 2018	0.020	0.707	0.500	-1.366	1.406	<del>(</del>		-	_	
Zouei et al., 2018	0.527	0.233	0.054	0.069	0.984			_	_	_
Borraz-Leon et al., 2021	1.241	0.253	0.064	0.744	1.737					$\rightarrow$
	0.737	0.217	0.047	0.312	1.162					
						1 00	0.50	0.00	0.50	1 00

Fig 3. Forest plot of the pooled mean± SD of testosterone in *T. gondii* positive than *T. gondii* negative in males, estimated with random-effects model.

https://doi.org/10.1371/journal.pone.0297362.g003

Study name		Statistics for each study					Std diff in means and 95% CI			
	Std diff in means	Standard error	Variance	Lower limit	Upper limit					
Abdul-Lateef et al., 2012	0.615	0.308	0.095	0.010	1.219			- I		$\rightarrow$
Hagag et al., 2022	0.993	0.388	0.150	0.233	1.753					
Kadhim and AL-awadi, 2013	0.881	0.204	0.041	0.482	1.280					-
El-Gebaly et al., 2019	1.161	0.371	0.138	0.434	1.889				-	$\rightarrow$
Al-Masoudi et al., 2018	-0.014	0.500	0.250	-0.994	0.966			-		—
Zouei et al., 2018	0.509	0.233	0.054	0.052	0.966				_	
Borraz-Leon et al., 2021	-0.345	0.297	0.088	-0.928	0.237	_		_		
	0.558	0.196	0.038	0.174	0.941			-		
						-1.00	-0.50	0.00	0.50	1.00

Fig 4. Forest plot of the pooled mean± SD of testosterone in *T. gondii* positive than *T. gondii* negative in females, estimated with random-effects model.

https://doi.org/10.1371/journal.pone.0297362.g004

3.3.2.2 Evidence for decreased or unchanged levels of testosterone in animals infected with *T*. gondii. Three studies in male animals (rats [48], mice [49], and spotted hyenas [50]) and two studies in female animals (mice [49], and spotted hyenas [50]) reported a decline level of testosterone in infected animals compared with non-infected animals. While, one study in female rats [51] reported no significant changes of testosterone in infected than non-infected animals (Fig 2 and Table 2).

*3.3.2.3 Evidence for alteration of cortisol levels in animals infected with T. gondii.* One study reported a declined level of cortisol in *T. gondii*-infected male rats [52], while one study [50] reported no significant change of cortisol levels in *T. gondii*-infected male and female spotted hyenas.

3.3.2.4 Description of animal studies. Kan ková et al. [49], reported that *T. gondii*-infected mice (both females and males) had significantly lower concentration of testosterone. Abdoli et al. [48] reported that male rats with *T. gondii* infection had a temporary decline in serum and intratesticular testosterone and fructose in seminal vesicles. As such, the percentage rates of sperm motility, viability, and concentration were significantly decreased and sperm abnormality was significantly increased after infection, but it reverts to the normal level on days 60 and 70 post infection [48]. Lim et al. [46] observed that *T. gondii* infection in male rats enhances testicular expression of genes involved in the synthesis of testosterone (LHR, StAR, and P450scc), resulting in greater testicular testosterone production. Afshari et al. [47] showed significantly increased levels of serum alkaline phosphatase and testosterone in *T. gondii*-infected male rats compared with the uninfected control group. Laubach et al. [50] found a negative association between *T. gondii* infection and plasma testosterone among female (cubs and subadults) and adult male hyenas, which means that the infected animals have lower testosterone levels than uninfected animals. Indeed, no associations were found between *T. gondii* infection and cortisol in any age class or sex group of hyenas [50].

#### 4. Discussion

Testosterone is involved in a variety of physiological functions, such as behavioral traits and reproductive functions in both sexes [21]. In this study, we reviewed data regarding *T. gondii* infection and testosterone variations in human studies and animal models (Fig 2 and Tables 1 and 2). We observed that most of the included studies in humans reported an increased level of testosterone [26, 29–34, 37–41, 43], while some studies reported a decreased level [27, 28,

35] or insignificant changes [36, 42] (Fig 1). As such, these variations were different in males and females in some studies [40, 41]. In animal models, some studies reported a declining level of testosterone [48–50], while others reported an increased level [46, 47] or insignificant changes [51]. Notably, variations in testosterone levels are most probably due to infection with different parasite strains, or a difference in host variations, which consequently influence the intensity of infection [15, 16, 22, 53]. Host variations also influence the intensity of *T. gondii* infection [54]. Among animals, mice and New and Old-World monkeys are highly sensitive to *T. gondii* infection; while sheep are intermediately sensitive, and goats, cattle, deer, horses, and pigs are resistant to the infection [54]. In humans, immunocompromised patients and pregnant women are at high risk of severe *T. gondii* infection, while *T. gondii* infection is usually asymptomatic (latent) among immunocompetent individuals, [7, 12]. Like humans, the laboratory rat (*Rattus norvegicus*) is resistant to *T. gondii* infection and is a suitable model for the study of chronic *T. gondii* infection [55, 56].

Testosterone plays an important role in sexual behavior and mating success [57-60]. On the other hand, recent evidence revealed that T. gondii infection augments sexual behavior and attractiveness in humans [61] and experimentally infected rodents [5]. In this regard, Borráz-León et al. [61] assessed several factors related to attractiveness among T. gondii-infected and non -infected individuals. They found that both T. gondii-infected men and women had lower facial fluctuating asymmetry, while infected women had lower body mass index, higher number of sexual partners, and a higher self-perceived attractiveness than non-infected control groups. They also assessed the attractiveness and perceived health of facial pictures of T. gondii-infected and non-infected subjects by an independent group of raters and found that both infected women and men were rated as more attractive and healthier than non-infected individuals [61]. Increased testosterone could enhance sexual behavior and attractiveness in infected subjects and could increase mating opportunity and transmission of T. gondii through sexual intercourse. In this regard, Lim et al. [46] reported that T. gondii infection (induced by Prugniaud strain) enhances testicular expression of genes that are involved in the synthesis of testosterone in experimentally infected male rats. Dass et al. [5] demonstrated that T. gondiiinfected male rats had higher sexual attractiveness to non-infected females, resulting in increased mating of infected males with non-infected females. They also confirmed sexual transmission of T. gondii through intercourse, whereas T. gondii cysts were detected in the epididymis of infected males, vaginal lavage of naïve females that mated with infected males, as well as in brains of pups which born from these matings [5]. As such, secretion of *T. gondii* in semen and sexual transmission of the parasite have been reported in dogs [62], goats [63-65], sheep [66, 67], cattle [68], and pigs [69]. Notably, T. gondii transmission in sheep was reported by artificial insemination of contaminated frozen semen [70]. There is also indirect evidence that suggests sexual transmission of T. gondii in humans. In this regard, a recent study by Tong et al. [4] confirmed the presence of *T. gondii* tissue cysts in human semen by immunofluorescence staining and molecular methods. Furthermore, it is proposed that unprotected sex and oral sex could be an important route of T. gondii transmission in humans [71, 72]. Hlaváčová et al. [73] performed a two-year study to compare the seropositivity to T. gondii in couples and analyzed the serological status of sexual partners. The results indicated that the prevalence of T. gondii infection was higher in women who had infected male partners than in women with uninfected male partners (25.6% vs 18.2%, respectively; P = 0.045). This study also suggests that a partner's seropositivity may be a risk factor for infection in women (prevalence ratio = 1.418; P = 0.045) but not in men (prevalence ratio = 1.058; P = 0.816) [73]. This evidence was also supported by studies among female sex workers [74] and individuals with a history of sexual promiscuity [75] in Mexico. In this regard, Alvarado-Esquivel et al. [74] found a significantly higher incidence of latent toxoplasmosis among female sex workers

compared with age- and sex-matched control groups (15.44% *vs* 3.67% in case and control groups, respectively, P = 0.0001). As such, female sex workers had significantly higher anti-*T. gondii* IgG titers (>150 IU/mL) than the control group (9.6% *vs* 2.9%, respectively P = 0.007) [74]. Another study by the same group of researchers in Mexico [75] revealed a significantly higher prevalence of anti-*T. gondii* IgG antibodies among individuals with sexual promiscuity than individuals without this practice (18.1% *vs* 10.3%, respectively; OR: 1.91; 95% CI: 1.41– 2.60; P < 0.0001). Indeed, higher titers of anti-*T. gondii* IgG antibodies (>150 IU/mL) were significantly increased in participants with sexual promiscuity than participants without this history (9.2% *vs* 4.6% respectively; OR: 2.09; 95% CI: 1.38–3.16; P = 0.0003). Additionally, the association of *T. gondii* seropositivity and serointensity with sexual promiscuity was observed in men but not in women [75]. Collectively, it seems that *T. gondii* infection could manipulate the mate choice of their host to increase their transmission rates. This phenomenon could be mediated partly by enhancing testosterone levels, which consequently increase sexual behavior and mating success [22, 76].

Testosterone has a pivotal role in spermatogenesis and male reproductive functions. A declined level of testosterone was reported following T. gondii infection in mice [49] and rats following infection with a T. gondii type I strain [48], as well as male and female spotted hyenas (Crocuta crocuta) which were naturally infected with T. gondii [50]. On the other hand, T. gon*dii* infection could induce male reproduction impairment by interfering in spermatogenesis and testicular damage [44, 48, 77–80], which may be partly mediated by declining testosterone levels. In this regard, Abdoli et al. [48] showed that T. gondii infection (induced by RH strain) induced a temporary decline in serum and intratesticular testosterone levels, fructose in seminal vesicles, as well as declining of sperm motility, viability, concentration, and increased of sperm abnormality in male rats. Hlaváčová et al. [81] compared the prevalence of latent toxoplasmosis in men with and without semen abnormalities and found that T. gondii-infected men had significantly lower sperm concentration and motility compared with T. gondii-negative men. Although another human study did not find a significant association between latent toxoplasmosis and semen abnormalities [42]. Considering the possible role of T. gondii in male reproductive impairment, it is recommended that populations with high prevalence of male infertility be examined for T. gondii infection.

Testosterone has also a pivotal role on behavioral traits in males and females, such as aggressive behavior [22, 82–84]. On the other hand, latent toxoplasmosis is also involved in the etiopathogenesis of different behavioral alterations (e.g., psychoticism [43], aggressive behavior [85, 86], and violent behavior [87]) and neuropsychiatric diseases, such as schizophrenia [88, 89], depression [90, 91] and anxiety disorders [90, 92, 93], obsessive compulsive disorder (OCD) [94], and autism spectrum disorder (ASD) [95–98]. Different mechanisms have been proposed to be involved in the etiopathogenesis of these disorders following *T. gon-dii* infection, including CNS Inflammation [99, 100], neurotransmitter alterations (alterations in dopamine [101–106] and serotonin synthesis [91]) and testosterone alteration [22, 107]. On the other hand, *in vitro* experiments revealed that testosterone [108] and dopamine [109] stimulate the propagation of *T. gondii* tachyzoites *in vitro*. Increasing fetal testosterone is also involved in autistic traits [110–113]. It is an important point because toxoplasmosis is a worldwide prevalent infection [114]. It is plausible an increased risk of ASD among infants of mothers with latent toxoplasmosis, and this phenomenon may partly be mediated via maternal testosterone alteration in mothers with latent toxoplasmosis [95].

There are some limitations to this systematic review. The lack of published articles from many countries were infertility is common is a major limitation. The observed association should be interpreted with caution, because the timeline of *T. gondii* infection and disease process could not be evaluated from the available data. Importantly, *T. gondii* seroprevalence has

been associated with many different risk factors, which were not evaluated in this work. As such, such confounding factors, including environmental toxins [115–117] and coinfections with other pathogens [100] may also affect the levels of sex hormones.

The results of this work can provide useful guidance for planning future studies. It would be important to focus on those parts of the world in which there is a lack of data on this subject. Moreover, including all pertinent risk factors would allow to better clarify the epidemiological aspects of *T. gondii* infection in infertility individuals and testosterone alterations. Optimally, prospective cohort studies and using more comprehensive serology panels (e.g., including IgG avidity testing) for estimating the timing of *T. gondii* infection could elucidate the timeline of risk factors of infection.

# 5. Conclusion

This study indicated that latent toxoplasmosis is associated with increased testosterone levels in most studies in humans and some studies in non-human animals. This change could be associated with increased sexual attractiveness in infected subjects which lead to sexual transmission of the parasite. On the other hand, some studies demonstrated a decreased level of testosterone in *T. gondii*-infected animals and humans. This change could partly be associated with male reproductive impairments, which were observed in *T. gondii*-infected human and non-human animals. These findings suggest the great need for more epidemiological and experimental studies in depth understanding the relationship between *T. gondii* infection, testosterone alteration, and further consequences.

# Supporting information

**S1 Checklist. PRISMA 2020 checklist.** (DOCX)

#### Acknowledgments

This study was supported by Jahrom University of Medical Sciences, Iran National Science Foundation (INSF) (95007218), and Tarbiat Modares University, Tehran, Iran. The Ethics Committee of Jahrom University of Medical Sciences (IR.JUMS.REC.1402.078) was approved the study protocol.

# **Author Contributions**

Conceptualization: Amir Abdoli.

Data curation: Amir Abdoli.

Formal analysis: Amir Abdoli, Ali Taghipour.

Investigation: Amir Abdoli.

Methodology: Amir Abdoli, Fatemeh Ghaffarifar, Ali Taghipour.

Project administration: Amir Abdoli.

Supervision: Fatemeh Ghaffarifar, Zohreh Sharifi.

Validation: Amir Abdoli, Zohreh Sharifi.

Writing - original draft: Amir Abdoli.

Writing - review & editing: Amir Abdoli.

#### References

- Montoya JG, Liesenfeld O. Toxoplasmosis. Lancet (London, England). 2004; 363(9425):1965–76. https://doi.org/10.1016/S0140-6736(04)16412-X PMID: 15194258
- Tenter AM, Heckeroth AR, Weiss LM. *Toxoplasma gondii*: from animals to humans. Int J Parasitol. 2000; 30(12):1217–58. https://doi.org/10.1016/S0020-7519(00)00124-7.
- Robert-Gangneux F, Dardé M-L. Epidemiology of and Diagnostic Strategies for Toxoplasmosis. Clin Microbiol Rev. 2012; 25(2):264–96. https://doi.org/10.1128/CMR.05013-11 PMID: 22491772
- Tong WH, Hlaváčová J, Abdulai-Saiku S, Kaňková Š, Flegr J, Vyas A. Presence of *Toxoplasma gondii* tissue cysts in human semen: Toxoplasmosis as a potential sexually transmissible infection. J Infect. 2023; 86(1):60–5. https://doi.org/10.1016/j.jinf.2022.10.034.
- 5. Dass SAH, Vasudevan A, Dutta D, Soh LJT, Sapolsky RM, Vyas A. Protozoan parasite *Toxoplasma gondii* manipulates mate choice in rats by enhancing attractiveness of males. PloS One. 2011; 6(11): e27229.
- Dalimi A, Abdoli A. Latent toxoplasmosis and human. Iran J Parasitol. 2012; 7(1): 1–17. <u>https://doi.org/10.1111/j.1439-0272.2011.01249.x PMID: 23133466</u>
- 7. Abdoli A, Barati M, Dalimi A, Pirestani M, Shokouh H, Javad S. Toxoplasmosis among patients with immunocompromising conditions: A snapshot. J Arch Milit Med. 2016; 4(4).
- Rasti S, Hassanzadeh M, Soliemani A, Hooshyar H, Mousavi SGA, Nikoueinejad H, et al. Serological and molecular survey of toxoplasmosis in renal transplant recipients and hemodialysis patients in Kashan and Qom regions, central Iran. Ren Fail. 2016; 38(6):970–3. <u>https://doi.org/10.3109/ 0886022X.2016.1172940</u> PMID: 27097530
- Bollani L, Auriti C, Achille C, Garofoli F, De Rose DU, Meroni V, et al. Congenital Toxoplasmosis: The State of the Art. Front Pediatr. 2022; 10:894573. <u>https://doi.org/10.3389/fped.2022.894573</u> PMID: 35874584
- Garweg JG, Kieffer F, Mandelbrot L, Peyron F, Wallon M. Long-Term Outcomes in Children with Congenital Toxoplasmosis—A Systematic Review. Pathogens [Internet]. 2022; 11(10):[1187 p.]. <u>https://</u> doi.org/10.3390/pathogens11101187 PMID: 36297244
- Ghasemi FS, Rasti S, Piroozmand A, Bandehpour M, Kazemi B, Mousavi SGA, et al. Toxoplasmosisassociated abortion and stillbirth in Tehran, Iran. The Journal of Maternal-Fetal & Neonatal Medicine. 2016; 29(2):248–51. https://doi.org/10.3109/14767058.2014.996127 PMID: 25564725
- Khademi SZ, Ghaffarifar F, Dalimi A, Davoodian P, Abdoli A. Spontaneous abortion among *Toxo-plasma gondii* IgG seropositive women: Molecular detection, genotype identification, and serological assessment with conventional ELISA and avidity ELISA. J Obstet Gynaecol Res. 2022; 48(10):2479–85. https://doi.org/10.1111/jog.15349 PMID: 35793814
- **13.** Fernández C, Jaimes J, Ortiz MC, Ramírez JD. Host and *Toxoplasma gondii* genetic and non-genetic factors influencing the development of ocular toxoplasmosis: A systematic review. Infect Genet Evol. 2016; 44:199–209. https://doi.org/10.1016/j.meegid.2016.06.053 PMID: 27389360
- 14. Yarovinsky F. Innate immunity to *Toxoplasma gondii* infection. Nat Rev Immunol. 2014; 14(2):109–21. https://doi.org/10.1038/nri3598 PMID: 24457485
- Xiao J, Yolken RH. Strain hypothesis of *Toxoplasma gondii* infection on the outcome of human diseases. Acta Physiol. 2015; 213(4):828–45. https://doi.org/10.1111/apha.12458 PMID: 25600911
- Abdoli A. Toxoplasma gondii and neuropsychiatric diseases: strain hypothesis. Neurol Sci. 2013; 34 (9):1697–8. https://doi.org/10.1007/s10072-012-1264-x PMID: 23224584
- Galal L, Hamidović A, Dardé ML, Mercier M. Diversity of *Toxoplasma gondii* strains at the global level and its determinants. Food Waterborne Parasitol. 2019; 15:e00052. https://doi.org/10.1016/j.fawpar. 2019.e00052.
- Saeij JPJ, Boyle JP, Boothroyd JC. Differences among the three major strains of *Toxoplasma gondii* and their specific interactions with the infected host. Trend Parasitol. 2005; 21(10):476–81. <u>https://doi.org/10.1016/j.pt.2005.08.001</u>.
- Boothroyd JC, Grigg ME. Population biology of *Toxoplasma gondii* and its relevance to human infection: do different strains cause different disease? Curr Opin Microbiol. 2002; 5(4):438–42. https://doi.org/10.1016/S1369-5274(02)00349-1.
- 20. Dardé M. Toxoplasma gondii, "new" genotypes and virulence. Parasite. 2008; 15(3):366-71.
- 21. Nelson DL, Lehninger AL, Cox MM. Lehninger principles of biochemistry: Macmillan; 2008.
- Abdoli A. Toxoplasma, testosterone, and behavior manipulation: the role of parasite strain, host variations, and intensity of infection. Front Biol. 2014; 9(2):151–60.
- Moher D, Liberati A, Tetzlaff J, Altman DG, Group\* P. Preferred reporting items for systematic reviews and meta-analyses: the PRISMA statement. Ann Intern Med. 2009; 151(4):264–9.

- Eriksen MB, Frandsen TF. The impact of patient, intervention, comparison, outcome (PICO) as a search strategy tool on literature search quality: a systematic review. Journal of the Medical Library Association: JMLA. 2018; 106(4):420–31. Epub 2018/10/03. https://doi.org/10.5195/jmla.2018.345 PMID: 30271283; PubMed Central PMCID: PMC6148624.
- Porritt K, Gomersall J, Lockwood C. JBI's systematic reviews: study selection and critical appraisal. AJN The American Journal of Nursing. 2014; 114(6):47–52. https://doi.org/10.1097/01.NAJ. 0000450430.97383.64 PMID: 24869584
- Shirbazou S, Abasian L, Talebi Meymand F. Effects of *Toxoplasma gondii* infection on plasma testosterone and cortisol level and stress index on patients referred to Sina hospital, Tehran. Jundishapur J Microbiol. 2011; 4(3):167–74.
- 27. Eslamirad Z, Hajihossein R, Ghorbanzadeh B. Relationship between blood testosterone level and lipid profile among a group of men with and without *Toxoplasma* IgG antibody referred to pre-marital clinics in Arak city, 2012. Arak Med Uni J. 2014; 16(12):1–8.
- Eslamirad Z, Hajihossein R, Ghorbanzadeh B, Alimohammadi M, Mosayebi M, Didehdar M. Effects of *Toxoplasma gondii* Infection in Level of Serum Testosterone in Males with Chronic Toxoplasmosis. Iran J Parasitol. 2013; 8(4):622–6. Epub 2013/10/01. PMID: 25516745; PubMed Central PMCID: PMC4266128.
- Mahbodfar HR, Yousefi-Razin E, Saki J, Rafiei A, Khademvatan S. Study of latent *Toxoplasma gondii* role in level of testosterone, DHEA, cortisol and prolactin hormones of young persons. Asian J Epidemiol. 2015; 8(3):64–71.
- Zouei N, Shojaee S, Mohebali M, Keshavarz H. The association of latent toxoplasmosis and level of serum testosterone in humans. BMC Res Notes. 2018; 11(1):365. https://doi.org/10.1186/s13104-018-3468-5 PMID: 29884208
- Bayani M, Kalantari N, Esmailzadeh S, Ghaffari S, Mahjoub S, Ghofrani F, et al. An evaluation of the level of testosterone, DHEA and prolactin among *Toxoplasma gondii* infected and uninfected infertile couples attending to Fatima Al-Zahra infertility treatment center, Babol, Northern Iran. Acta facultatis medicae Naissensis. 2022; 39(1):37–47.
- **32.** Abdul-Lateef HI, AL-Najar SA, Majeed NGA. The levels of IFN-, IL-12 and testosterone hormone in persons with asymptomatic toxoplasmosis. J Facul Med Baghdad. 2012; 54(1):79–82.
- Zghair KH, Al-Qadhi BN, Mahmood SH. The effect of toxoplasmosis on the level of some sex hormones in males blood donors in Baghdad. J Parasitic Dis. 2015; 39(3):393–400. https://doi.org/10. 1007/s12639-013-0382-6 PMID: 26345040
- Kadhim RA, AL-awadi HM. Changes in testosterone, progesterone and prolactin levels in pregnant women with chronic toxoplasmosis. Med J Babylon. 2013; 10(3):699–708.
- Al-Masoudi HK, Al-Khafaji MS, Noori RS. Molecular and Hormonal Study (Testosterone and Luteinizing hormone) among Applicants For Marriage and Blood Donors Peoples who Infected With Toxoplasma gondii in Babylon Province, Iraq. J Pharmaceutical Sci Res. 2018; 10(9):2391–5.
- Al-Kurdy MJ, A'aiz NN, Jawad TI. Study the Effect of Chronic Toxoplasmosis on Serum Testosterone Level in Men. Indian J Forensic Med Toxicol. 2020; 14(4):3133.
- AL-Asady RA. The Levels of Testosterone, FSH and LH in Pregnant Women with Chronic Toxoplasmosis in Najaf Province. Al-Qadisiyah Medical Journal. 2017; 13(23):34–41.
- El-Gebaly N, Abd-Eltawab M, Hamed A, Mahfouz N, Adel S, Mahfoz A, et al. Insights into the interplay of latent toxoplasmosis, testosterone, cortisol and oxidative stress in screened schizophrenic patients in Egypt. Parasitologists United J. 2019; 12(2):102–9.
- Hagag MM, Al Shaimaa M, Saafan MA. Association of latent toxoplasmosis with hormonal disturbance of androgenic alopecia and acne vulgaris. Menoufia Med J. 2022; 35(1):72–7.
- Flegr J, Lindová J, Kodym P. Sex-dependent toxoplasmosis-associated differences in testosterone concentration in humans. Parasitology. 2008; 135(4):427–31. <u>https://doi.org/10.1017/</u> S0031182007004064 PMID: 18205984
- Flegr J, Lindová J, Pivoñková V, Havlíček J. Brief Communication: Latent toxoplasmosis and salivary testosterone concentration—Important confounding factors in second to fourth digit ratio studies. Ame J Physical Anthropol. 2008; 137(4):479–84. https://doi.org/10.1002/ajpa.20888 PMID: 18615572
- Colosi HA, Jalali-Zadeh B, Colosi IA, Simon LM, Costache CA. Influence of *Toxoplasma gondii* infection on male fertility: a pilot study on immunocompetent human volunteers. Iran J Parasitol. 2015; 10 (3):402–9.
- 43. Borráz-León JI, Rantala MJ, Luoto S, Krams I, Contreras-Garduño J, Cerda-Molina AL, et al. *Toxoplasma gondii* and Psychopathology: Latent Infection Is Associated with Interpersonal Sensitivity, Psychoticism, and Higher Testosterone Levels in Men, but Not in Women. Adapt Hum Behav Physiol. 2021; 7(1):28–42. https://doi.org/10.1007/s40750-020-00160-2

- 44. Oktenli C, Doganci L, Ozgurtas T, Araz RE, Tanyuksel M, Musabak U, et al. Transient hypogonadotrophic hypogonadism in males with acute toxoplasmosis: suppressive effect of interleukin-1β on the secretion of GnRH. Hum Reprod. 2004; 19(4):859–66.
- Hamdeh S, Abbas A, Fraker J, Lambrecht J. Intracranial toxoplasmosis presenting as panhypopituitarism in an immunocompromised patient. Am J Emerg Med. 2015; 33(12):1848. e1–. e2. <u>https://doi.org/ 10.1016/j.ajem.2015.04.071 PMID: 26027887</u>
- 46. Lim A, Kumar V, Hari Dass SA, Vyas A. *Toxoplasma gondii* infection enhances testicular steroidogenesis in rats. Mol Ecol. 2013; 22(1):102–10. https://doi.org/10.1111/mec.12042 PMID: 23190313
- 47. Afshari F, Imani AM, Najjari Asl S, Farhang H, Ghasempour K, Ezzatzadeh A, et al. Evaluation of testosterone and alkaline phosphatase activity changes in epidydimis of *Toxoplasma gondii* infected rats. Int J Women's Health Reproduction Sci. 2013; 1(2):64–71.
- Abdoli A, Dalimi A, Movahedin M. Impaired reproductive function of male rats infected with *Toxoplasma gondii*. Andrologia. 2012; 44(s1):679–87. <u>https://doi.org/10.1111/j.1439-0272.2011.01249.x</u> PMID: 22098674
- Kaňková Š, Kodym P, Flegr J. Direct evidence of *Toxoplasma*-induced changes in serum testosterone in mice. Exp Parasitol. 2011; 128(3):181–3.
- Laubach ZM, Gering E, Yang E, Montgomery TM, Getty T, Holekamp KE. Associations between *Toxoplasma gondii* infection and steroid hormone levels in spotted hyenas. Int J Parasitol: Parasit Wildlife. 2022; 17:53–9. https://doi.org/10.1016/j.ijppaw.2021.11.007.
- Abdulai-Saiku S, Vyas A. Loss of predator aversion in female rats after *Toxoplasma gondii* infection is not dependent on ovarian steroids. Brain Behav Immun. 2017; 65:95–8. <u>https://doi.org/10.1016/j.bbi</u>. 2017.04.005 PMID: 28400143
- Mitra R, Sapolsky RM, Vyas A. *Toxoplasma gondii* infection induces dendritic retraction in basolateral amygdala accompanied by reduced corticosterone secretion. Dis Model Mech. 2013; 6(2):516–20.
- 53. Dubey JP, Ferreira LR, Martins J, McLeod R. Oral oocyst-induced mouse model of toxoplasmosis: effect of infection with Toxoplasma gondii strains of different genotypes, dose, and mouse strains (transgenic, out-bred, in-bred) on pathogenesis and mortality. Parasitology. 2012; 139(1):1–13. Epub 2011/11/14. https://doi.org/10.1017/S0031182011001673 PMID: 22078010
- Innes EA. Toxoplasmosis: Comparative species susceptibility and host immune response. Comp Immunol Microbiol Infect Dis. 1997; 20(2):131–8. <u>https://doi.org/10.1016/s0147-9571(96)00038-0</u> PMID: 9208198
- Sullivan WJ Jr., Jeffers V. Mechanisms of *Toxoplasma gondii* persistence and latency. FEMS Microbiol Rev. 2012; 36(3):717–33. https://doi.org/10.1111/j.1574-6976.2011.00305.x PMID: 22091606
- 56. Dubey J, Frenkel J. Toxoplasmosis of rats: a review, with considerations of their value as an animal model and their possible role in epidemiology. Vet Parasitol. 1998; 77(1):1–32. <u>https://doi.org/10.1016/s0304-4017(97)00227-6 PMID: 9652380</u>
- Martel KL, Baum MJ. Adult testosterone treatment but not surgical disruption of vomeronasal function augments male-typical sexual behavior in female mice. J Neurosci. 2009; 29(24):7658–66. https://doi. org/10.1523/JNEUROSCI.1311-09.2009 PMID: 19535577
- Peters M, Simmons LW, Rhodes G. Testosterone is associated with mating success but not attractiveness or masculinity in human males. Anim Behav. 2008; 76(2):297–303.
- 59. Muller MN. Testosterone and reproductive effort in male primates. Hormon Behav. 2017; 91:36–51. https://doi.org/10.1016/j.yhbeh.2016.09.001 PMID: 27616559
- Roney JR, Gettler LT. The role of testosterone in human romantic relationships. Curr Opin Psychol. 2015; 1:81–6. https://doi.org/10.1016/j.copsyc.2014.11.003.
- Borráz-León JI, Rantala MJ, Krams IA, Cerda-Molina AL, Contreras-Garduño J. Are *Toxoplasma*infected subjects more attractive, symmetrical, or healthier than non-infected ones? Evidence from subjective and objective measurements. PeerJ. 2022; 10:e13122.
- Arantes TP, Lopes WDZ, Ferreira RM, Pieroni JSP, Pinto VMR, Sakamoto CA, et al. *Toxoplasma gondii*: Evidence for the transmission by semen in dogs. Exp Parasitol. 2009; 123(2):190–4. <u>https://doi.org/10.1016/j.exppara.2009.07.003</u>.
- **63.** Dubey J, Sharma S. Prolonged excretion of *Toxoplasma gondii* in semen of goats. Am J Vet Res. 1980; 41(5):794–5.
- Santana LF, Rossi GAM, Gaspar RC, Pinto VMR, Oliveira GPd, Costa AJd. Evidence of sexual transmission of *Toxoplasma gondii* in goats. Small Rumin Res. 2013; 115(1):130–3. <u>https://doi.org/10.1016/j.smallrumres.2013.08.008</u>.
- **65.** Wanderley FS, Porto WJN, Câmara DR, de Oliveira VVG, Garcia JL, de Albuquerque PPF, et al. Venereal transmission of *Toxoplasma gondii* in goats after a buck was experimentally infected. Small Rumin Res. 2015; 123(2):301–5. https://doi.org/10.1016/j.smallrumres.2014.11.017.

- De Moraes EPBX, Batista AM, Faria EB, Freire RL, Freitas AC, Silva MAR, et al. Experimental infection by *Toxoplasma gondii* using contaminated semen containing different doses of tachyzoites in sheep. Vet Parasitol. 2010; 170(3–4):318–22.
- Lopes WDZ, Rodriguez JDA, Souza FA, dos Santos TR, dos Santos RS, Rosanese WM, et al. Sexual transmission of *Toxoplasma gondii* in sheep. Vet Parasitol. 2013; 195(1):47–56. <u>https://doi.org/10. 1016/j.vetpar.2012.12.056.</u>
- Scarpelli L, Lopes WDZ, Migani M, Bresciani KDS, Costa AJd. *Toxoplasma gondii* in experimentally infected Bos taurus and Bos indicus semen and tissues. Pesq Vet Bras. 2009; 29:59–64.
- 69. Moura AB, Costa AJ, Jordão Filho S, Paim BB, Pinto FR, Di Mauro DC. *Toxoplasma gondii* in semen of experimentally infected swine. Pesq Vet Bras. 2007; 27:430–4.
- Consalter A, Silva AF, Frazão-Teixeira E, Matos LF, de Oliveira FCR, Leite JS, et al. *Toxoplasma gondii* transmission by artificial insemination in sheep with experimentally contaminated frozen semen. Theriogenology. 2017; 90:169–74. https://doi.org/10.1016/j.theriogenology.2016.12.004.
- Flegr J, Klapilová K, Kaňková Š. Toxoplasmosis can be a sexually transmitted infection with serious clinical consequences. Not all routes of infection are created equal. Med Hypotheses. 2014; 83 (3):286–9. https://doi.org/10.1016/j.mehy.2014.05.019 PMID: 24986706
- Kaňková Š, Hlaváčová J, Flegr J. Oral sex: A new, and possibly the most dangerous, route of toxoplasmosis transmission. Med Hypotheses. 2020; 141:109725. https://doi.org/10.1016/j.mehy.2020. 109725 PMID: 32315924
- 73. Hlaváčová J, Flegr J, Řežábek K, Calda P, Kaňková Š. Male-to-Female Presumed Transmission of Toxoplasmosis Between Sexual Partners. Am J Epidemiol. 2021; 190(3):386–92. <u>https://doi.org/10.1093/aje/kwaa198 PMID</u>: 32929444
- 74. Alvarado-Esquivel C, Sánchez-Anguiano LF, Hernández-Tinoco J, Arreola-Cháidez E, López J, Salcido-Meraz KI, et al. High seroprevalence of *Toxoplasma gondii* infection in female sex workers: a case-control study. Eur J Microbiol Immunol. 2015; 5(4):285–92.
- 75. Alvarado-Esquivel C, Estrada-Martínez S, Ramos-Nevárez A, Pérez-Álamos AR, Beristain-Garcia I, Alvarado-Félix ÁO, et al. Is *Toxoplasma gondii* Infection Associated with Sexual Promiscuity? A Cross-Sectional Study. Pathogens [Internet]. 2021; 10(11):[1393 p.].
- 76. Vyas A. Parasite-augmented mate choice and reduction in innate fear in rats infected by *Toxoplasma gondii*. J Exp Biol. 2013; 216(1):120–6. https://doi.org/10.1242/jeb.072983 PMID: 23225874
- 77. Dalimi A, Abdoli A. *Toxoplasma gondii* and male reproduction impairment: a new aspect of toxoplasmosis research. Jundishapur J Microbiol. 2013; 6(8):e7184.
- 78. Terpsidis KI, Papazahariadou MG, Taitzoglou IA, Papaioannou NG, Georgiadis MP, Theodoridis IT. *Toxoplasma gondii*: Reproductive parameters in experimentally infected male rats. Exp Parasitol. 2009; 121(3):238–41. https://doi.org/10.1016/j.exppara.2008.11.006.
- Lopes WD, Santos TR, Luvizotto MCR, Sakamoto C, Oliveira G, Costa A. Histopathology of the reproductive system of male sheep experimentally infected with *Toxoplasma gondii*. Parasitol Res. 2011; 109:405–9.
- Dvorakova-Hortova K, Sidlova A, Ded L, Hladovcova D, Vieweg M, Weidner W, et al. *Toxoplasma gondii* Decreases the Reproductive Fitness in Mice. PLoS One. 2014; 9(6):e96770. https://doi.org/10. 1371/journal.pone.0096770 PMID: 24940596
- Hlaváčová J, Flegr J, Řežábek K, Calda P, Kaňková Š. Association between latent toxoplasmosis and fertility parameters of men. Andrology. 2021; 9(3):854–62. https://doi.org/10.1111/andr.12969 PMID: 33420759
- Abdulai-Saiku S, Tong WH, Vyas A. Behavioral Manipulation by Toxoplasma gondii: Does Brain Residence Matter? Trend Parasitol. 2021; 37(5):381–90. <u>https://doi.org/10.1016/j.pt.2020.12.006</u> PMID: 33461902
- Webster JP. The effect of Toxoplasma gondii on animal behavior: playing cat and mouse. Schizophrenia Bulletin. 2007; 33(3):752–6. https://doi.org/10.1093/schbul/sbl073 PMID: 17218613
- Batrinos ML. Testosterone and aggressive behavior in man. International journal of endocrinology and metabolism. 2012; 10(3):563–8. Epub 2013/07/12. <u>https://doi.org/10.5812/ijem.3661</u> PMID: 23843821; PubMed Central PMCID: PMC3693622.
- Coccaro EF, Lee R, Groer MW, Can A, Coussons-Read M, Postolache TT. *Toxoplasma gondii* infection: relationship with aggression in psychiatric subjects. The Journal of clinical psychiatry. 2016; 77 (3):334–41. Epub 2016/04/06. https://doi.org/10.4088/JCP.14m09621 PMID: 27046307.
- Cook TB, Brenner LA, Cloninger CR, Langenberg P, Igbide A, Giegling I, et al. "Latent" infection with *Toxoplasma gondii*: Association with trait aggression and impulsivity in healthy adults. J Psychiatr Res. 2015; 60:87–94. https://doi.org/10.1016/j.jpsychires.2014.09.019.

- Rocha-Salais A, Muñoz-Larreta FY, García-Pérez SI, Serrato-Enríquez AI, Rivas-González MA, Sifuentes-Alvarez A, et al. Survey on the association between *Toxoplasma gondii* infection and violent behavior in inmates. PLoS ONE. 2023; 18(4):e0284202. https://doi.org/10.1371/journal.pone. 0284202 PMID: 37027388
- Contopoulos-Ioannidis DG, Gianniki M, Ai-Nhi Truong A, Montoya JG. *Toxoplasmosis* and Schizophrenia: A Systematic Review and Meta-Analysis of Prevalence and Associations and Future Directions. Psychiatr Res Clin Pract. 2022; 4(2):48–60. https://doi.org/10.1176/appi.prcp.20210041 PMID: 36254187
- Wang HL, Wang GH, Li QY, Shu C, Jiang MS, Guo Y. Prevalence of *Toxoplasma* infection in first-episode schizophrenia and comparison between *Toxoplasma*-seropositive and *Toxoplasma*-seronegative schizophrenia. Acta Psychiatr Scand. 2006; 114(1):40–8. https://doi.org/10.1111/j.1600-0447. 2006.00780.x.
- 90. Groër MW, Yolken RH, Xiao JC, Beckstead JW, Fuchs D, Mohapatra SS, et al. Prenatal depression and anxiety in *Toxoplasma gondii*-positive women. Am J Obstet Gynecol. 2011; 204(5):433.e1-.e7. https://doi.org/10.1016/j.ajog.2011.01.004.
- Mahmoud ME, Ihara F, Fereig RM, Nishimura M, Nishikawa Y. Induction of depression-related behaviors by reactivation of chronic *Toxoplasma gondii* infection in mice. Behav Brain Res. 2016; 298:125–33. https://doi.org/10.1016/j.bbr.2015.11.005.
- 92. Hu M, Richard JE, Maliqueo M, Kokosar M, Fornes R, Benrick A, et al. Maternal testosterone exposure increases anxiety-like behavior and impacts the limbic system in the offspring. Proc Natl Acad Sci USA. 2015; 112(46):14348. https://doi.org/10.1073/pnas.1507514112 PMID: 26578781
- Markovitz AA, Simanek AM, Yolken RH, Galea S, Koenen KC, Chen S, et al. *Toxoplasma gondii* and anxiety disorders in a community-based sample. Brain Behavior Immun. 2015; 43:192–7. https://doi. org/10.1016/j.bbi.2014.08.001.
- Nayeri Chegeni T, Sarvi S, Amouei A, Moosazadeh M, Hosseininejad Z, A. Aghayan S, et al. Relationship between toxoplasmosis and obsessive compulsive disorder: A systematic review and meta-analysis. PLoS Negl Trop Dis. 2019; 13(4):e0007306. https://doi.org/10.1371/journal.pntd.0007306 PMID: 30969961
- Abdoli A, Dalimi A. Are There any Relationships between Latent Toxoplasma gondii Infection, Testosterone Elevation, and Risk of Autism Spectrum Disorder? Front Behav Neurosci. 2014; 8(339). <u>https://</u> doi.org/10.3389/fnbeh.2014.00339 PMID: 25309376
- 96. Spann MN, Sourander A, Surcel H-M, Hinkka-Yli-Salomäki S, Brown AS. Prenatal toxoplasmosis antibody and childhood autism. Autism Res. 2017; 10(5):769–77. <u>https://doi.org/10.1002/aur.1722</u> PMID: 27874276
- 97. Hamid N, Azizy B, Hamidinejad H. *Toxoplasma gondii* Infection and Aggression in Autistic Children. Pediatr Infect Dis J. 2022; 41(6).
- Al Malki JS, Hussien NA, Al Malki F. Maternal toxoplasmosis and the risk of childhood autism: serological and molecular small-scale studies. BMC Pediatr. 2021; 21(1):133. https://doi.org/10.1186/ s12887-021-02604-4 PMID: 33731054
- **99.** Xiao J. Behavioral Changes Induced by Latent Toxoplasmosis Could Arise from CNS Inflammation and Neuropathogenesis. Berlin, Heidelberg: Springer Berlin Heidelberg; 2022. p. 1–11.
- Abdoli A, Taghipour A, Pirestani M, Mofazzal Jahromi MA, Roustazadeh A, Mir H, et al. Infections, inflammation, and risk of neuropsychiatric disorders: the neglected role of "co-infection". Heliyon. 2020; 6(12):e05645. https://doi.org/10.1016/j.heliyon.2020.e05645.
- 101. Cromar GL, Epp JR, Popovic A, Gu Y, Ha V, Walters BJ, et al. *Toxoplasma* infection in male mice alters dopamine-sensitive behaviors and host gene expression patterns associated with neuropsychiatric disease. PLoS Neglect Trop Dis. 2022; 16(7):e0010600. https://doi.org/10.1371/journal.pntd. 0010600 PMID: 35857765
- **102.** Prandovszky E, Gaskell E, Martin H, Dubey J, Webster JP, McConkey GA. The neurotropic parasite *Toxoplasma gondii* increases dopamine metabolism. PloS One. 2011; 6(9):e23866.
- 103. Wang ZT, Harmon S, O'Malley KL, Sibley LD. Reassessment of the role of aromatic amino acid hydroxylases and the effect of infection by *Toxoplasma gondii* on host dopamine. Infect Immun. 2015; 83(3):1039–47.
- 104. SkallovÁ A, Kodym P, Frynta D, Flegr J. The role of dopamine in *Toxoplasma*-induced behavioural alterations in mice: an ethological and ethopharmacological study. Parasitology. 2006; 133(5):525–35. Epub 2006/08/02. https://doi.org/10.1017/S0031182006000886 PMID: 16882355
- 105. Xiao J, Li Y, Prandovszky E, Karuppagounder SS, Talbot CC Jr, Dawson VL, et al. MicroRNA-132 dysregulation in *Toxoplasma gondii* infection has implications for dopamine signaling pathway. Neuroscience. 2014; 268:128–38.

- 106. Wang T, Sun X, Qin W, Zhang X, Wu L, Li Y, et al. From inflammatory reactions to neurotransmitter changes: Implications for understanding the neurobehavioral changes in mice chronically infected with *Toxoplasma gondii*. Behav Brain Res. 2019; 359:737–48. https://doi.org/10.1016/j.bbr.2018.09.011 PMID: 30253194
- 107. Hodková H, Kolbeková P, Skallová A, Lindová J, Flegr J. Higher perceived dominance in *Toxoplasma* infected men—a new evidence for role of increased level of testosterone in toxoplasmosis-associated changes in human behavior. Neuroendocrinol Lett. 2007; 28(2):110–4.
- 108. Abdoli A, Ghaffarifar F, Sharifi Z, Zaki L. Testosterone Augments Propagation of *Toxoplasma gondii* in Glioblastoma Cells In Vitro. Acta Parasitol. 2022; 67(3):1425–31. <u>https://doi.org/10.1007/s11686-022-00571-z PMID</u>: 35616833
- 109. Strobl JS, Goodwin DG, Rzigalinski BA, Lindsay DS. Dopamine Stimulates Propagation of *Toxoplasma gondii* Tachyzoites in Human Fibroblast and Primary Neonatal Rat Astrocyte Cell Cultures. J Parasitol. 2012; 98(6):1296–9. https://doi.org/10.1645/GE-2760.1 PMID: 22512377
- Auyeung B, Baron-Cohen S, Ashwin E, Knickmeyer R, Taylor K, Hackett G. Fetal testosterone and autistic traits. Brit J Psychol. 2009; 100(1):1–22.
- Auyeung B, Taylor K, Hackett G, Baron-Cohen S. Foetal testosterone and autistic traits in 18 to 24month-old children. Mol Autism. 2010; 1:1–8.
- 112. Ingudomnukul E, Baron-Cohen S, Wheelwright S, Knickmeyer R. Elevated rates of testosteronerelated disorders in women with autism spectrum conditions. Horm Behav. 2007; 51(5):597–604. https://doi.org/10.1016/j.yhbeh.2007.02.001 PMID: 17462645
- 113. Baron-Cohen S, Auyeung B, Nørgaard-Pedersen B, Hougaard DM, Abdallah MW, Melgaard L, et al. Elevated fetal steroidogenic activity in autism. Mol Psychiatry. 2015; 20(3):369–76. <u>https://doi.org/10.1038/mp.2014.48 PMID: 24888361</u>
- 114. Flegr J, Prandota J, Sovičková M, Israili ZH. Toxoplasmosis–a global threat. Correlation of latent toxoplasmosis with specific disease burden in a set of 88 countries. PloS One. 2014; 9(3):e90203. https:// doi.org/10.1371/journal.pone.0090203 PMID: 24662942
- Kumar M, Sarma DK, Shubham S, Kumawat M, Verma V, Prakash A, et al. Environmental Endocrine-Disrupting Chemical Exposure: Role in Non-Communicable Diseases. Front Public Health. 2020; 8:553850. https://doi.org/10.3389/fpubh.2020.553850 PMID: 33072697
- 116. Dutta S, Gorain B, Choudhury H, Roychoudhury S, Sengupta P. Environmental and occupational exposure of metals and female reproductive health. Environ Sci Pollut Res. 2022; 29(41):62067–92. https://doi.org/10.1007/s11356-021-16581-9 PMID: 34558053
- 117. Sharara FI, Seifer DB, Flaws JA. Environmental toxicants and female reproduction 44Additional references are available from the authors. Fertil Steril. 1998; 70(4):613–22. https://doi.org/10.1016/S0015-0282(98)00253-2.