

RESEARCH ARTICLE

Significance of eggshell morphology as an additional tool to distinguish species of sand flies (Diptera: Psychodidae: Phlebotominae)

Narissara Jariyapan^{1,2*}, Pongsri Tippawangkosol³, Sriwatapron Sor-Suwan^{1,2}, Chonlada Mano³, Thippawan Yasanga⁴, Pradya Somboon^{3*}, Jérôme Depaquit⁵, Padet Siriyasatien^{1,2}

1 Department of Parasitology, Faculty of Medicine, Chulalongkorn University, Bangkok, Thailand, **2** Vector Biology and Vector Borne Diseases Research Unit, Department of Parasitology, Faculty of Medicine, Chulalongkorn University, Bangkok, Thailand, **3** Department of Parasitology, Faculty of Medicine, Chiang Mai University, Chiang Mai, Thailand, **4** Medical Science Research Equipment Center, Faculty of Medicine, Chiang Mai University, Chiang Mai, Thailand, **5** Faculté de Pharmacie, Université de Reims Champagne-Ardenne, SFR Cap Santé, EA7510 ESCAPE–USC ANSES VECPAR, Reims, France

* narissara.j@chula.ac.th (NJ); pradya.somboon@cmu.ac.th (PS)



OPEN ACCESS

Citation: Jariyapan N, Tippawangkosol P, Sor-Suwan S, Mano C, Yasanga T, Somboon P, et al. (2022) Significance of eggshell morphology as an additional tool to distinguish species of sand flies (Diptera: Psychodidae: Phlebotominae). PLoS ONE 17(2): e0263268. <https://doi.org/10.1371/journal.pone.0263268>

Editor: Albert Schriefer, Universidade Federal da Bahia, BRAZIL

Received: October 21, 2021

Accepted: January 14, 2022

Published: February 25, 2022

Copyright: © 2022 Jariyapan et al. This is an open access article distributed under the terms of the [Creative Commons Attribution License](https://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Data Availability Statement: All nucleotide sequences are available from the GenBank database (accession numbers OK398062–OK398083).

Funding: This work was supported by the Program Management Unit for Human Resources & Institutional Development, Research and Innovation - Chulalongkorn University [grant number B16F630071] received by NJ; the Thailand Science Research and Innovation (TSRI) Fund

Abstract

Morphological characteristics of eggshells are important in sand fly ootaxonomy. In this study, eggshells from *Phlebotomus stantoni* Newstead, *Sergentomyia khawi* (Raynal), and *Grassomyia indica* (Theodor) sand flies collected in Chiang Mai province, Thailand were examined and characterized using light microscopy (LM) and scanning electron microscopy (SEM). Then, eggshell morphology of these three species was described for the first time. Each gravid female was forced to lay eggs by decapitation and the eggs were collected for SEM analysis. Egg laying females were identified by morphological examination and molecular typing using *cytochrome b* (*Cytb*) as a molecular marker. The chorionic sculpturing of *Ph. stantoni* eggs combines two patterns on the same egg: unconnected parallel ridges and reticular patterns. *Sergentomyia khawi* and *Gr. indica* have similar chorionic polygonal patterns, but their exochorionic morphology and aeropylar area are different. Results indicate that eggshell morphological characteristics such as chorionic pattern, exochorionic morphology, inter-ridge/boundary area, aeropylar area (including the number of aeropyles) and basal layer, can be useful to develop morphological identification keys of eggs. These can serve as an additional tool to distinguish species of sand flies. In addition, the chorionic sculpturing of the eggs of the three species of sand flies observed by LM is useful for species identification in gravid females with spermathecae obscured by eggs.

Introduction

Phlebotomine sand flies (Diptera: Psychodidae: Phlebotominae) are hematophagous insects. Some species are vectors of several viral, bacterial, and protozoal diseases, especially leishmaniasis. In the last few decades, new proposals for classification and identification of Phlebotominae sand flies have been based primarily on adult morphology [1]. However, morphological

[grant number CU_FRB640001_01_30_1] received by NJ, <https://www.chula.ac.th/en/>, and <https://www.tsri.or.th/>; and the Faculty of Medicine, Chiang Mai University [grant number 021-2563] received by PT, <https://www.med.cmu.ac.th>. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing interests: The authors have declared that no competing interests exist.

identification of the adult stage alone may be insufficient to differentiate closely related species. Hence, an additional approach which uses the characteristics of immature stages, including eggs, has proved useful for identification [2]. In mosquitoes, analyzing morphological characteristics of the chorionic sculpturing of eggs by means of scanning electron microscopy (SEM) can help to differentiate sibling species of the *Anopheles culicifacies* complex [3].

The chorionic sculpturing of sand fly eggs was first studied and described by Ward and Ready in thirteen species of New World sand flies [4]. The ultrastructure of sand fly eggshells was investigated in many New World species (approximately 50 species) [4–19] but only in a limited number of species of Old World sand flies [20–25]. Chorionic patterns reflect phylogenetic relationships among certain *Lutzomyia* sand fly species in the New World [6–8, 10]. The comparison of eggshell morphology of *Nyssomyia intermedia* (Lutz and Neiva) and *Ny. whitmani* (Antunes and Coutinho) revealed that chorionic patterns of both species are similar. However, details in the microanatomy of the external surface of the eggs observed by SEM can be used to differentiate the two species [14]. Alencar and Scarpassa [19] examined the chorionic sculpturing of ten Brazilian sand fly species using SEM: *Ny. antunesi* (Coutinho), *Ny. whitmani* (Antunes and Coutinho), *Bichromomyia flaviscutellata* (Mangabeira), *Bicholmea nociva* (Young and Arias), *Evandromyia walkeri* (Newstead), *Ev. williamsi* (Antunes and Coutinho), *Deanemyia maruaga* (Alves, Freitas and Barrett), *De. samueli* (Deane), *Viannamyia furcata* (Mangabeira) and *Lutzomyia dispar* (Martins and Silva). The authors demonstrated that eggshell morphology can be used in phlebotomine taxonomy, both at generic and specific levels.

So far, several chorionic patterns of sand fly eggs have been proposed, *i.e.*, “polygonal”, “volcano-like” [4], “connected parallel ridges”, “unconnected parallel ridges” [6], “elliptical” [7], “reticular”, “verrucose”, “disperse” [8], and “placoid” [19]. The ultrastructural morphology of eggs of Old World sand flies in the genera *Sergentomyia* and *Phlebotomus* was studied in Italy, Kenya, Ethiopia, Egypt, and India [20–25]. Ward and Ready suggested that chorionic sculpturing differences are adaptations to the different types of environments of oviposition sites [4].

In Thailand, surveillance studies of sand fly species were carried out in different regions of the country over time. At least 31 species of sand flies have been identified [26–29]. Several studies were carried out to detect the DNA/RNA of various pathogens in sand flies [30–35]. DNA of *Leishmania martiniquensis*, *L. orientalis*, and *L. donovani*/*L. infantum* was detected in some species of *Sergentomyia* [30, 32] probably misidentified as *Se. gemmea* (Lewis and Jeffery) [28]. Moreover, DNA of *L. martiniquensis* was detected in *Se. barraudi* (Sinton), *Se. khawi*, and *Ph. stantoni* [31, 32]. However, the role of sand flies as natural vectors of *L. martiniquensis* remain unclear [28]. The DNA of *Trypanosoma* spp. was detected in *Se. khawi*, and *Ph. stantoni* [33, 34]. Recently, *Orbivirus* genus RNA was detected in *Idiophlebotomus* spp., *Ph. papatasi* (Scopoli) and *Se. khawi* [35]. In addition, the RNA of *Rhabdoviridae* family virus was detected in *Ph. papatasi* [36].

Most researchers have used adult morphology (cibarium, pigment patch, and spermathecae) and/or molecular methods to identify sand fly species in Thailand [26–37]. However, it is difficult to identify spermathecae in gravid females. Moreover, the systematic use of molecular tools/techniques to identify all sand flies collected from the field remains expensive. Therefore, morphological characteristics of sand fly eggs are of interest but there is currently no data on eggshell morphology for sand flies in Thailand. The objective of this study was to examine and characterize the ultrastructural morphology of eggs obtained from individual gravid female sand flies collected from Chiang Mai province, Thailand, using light microscopy (LM) and SEM.

Materials and methods

Sand fly collection and experimental design

Sand flies were collected on a weekly basis in August 2020 (four nights in total) in Doi Saket, Chiang Mai province, Thailand (E 18.892554, S 99.120376). Three CDC light-traps were set up in a cowshed from 6 PM to 6 AM. Collected sand flies were transferred to the laboratory of the Department of Parasitology, Faculty of Medicine, Chiang Mai University, for species identification and examination of eggs.

The experimental design used for species identification of female sand flies and morphological examination and characterization of sand fly eggs is shown in Fig 1. Gravid females with similar morphology were separated into two groups. In Group 1, females were dissected individually for morphological identification using LM. Their carcasses were molecularly identified through DNA sequencing of the *cytochrome b* (*Cytb*) gene. LM analysis of the chorionic sculpturing pattern of eggs in each gravid female was also performed. In Group 2, each gravid female was forced to lay eggs by decapitation on wet Whatman filter papers under a dissecting microscope. After laying eggs, DNA was extracted from the female carcass. DNA sequencing of the *Cytb* gene for molecular identification of species was carried out through polymerase chain reaction (PCR). The eggs were left on the wet papers in a container for 24 h and subjected to SEM for measurements and morphological characterization including chorionic pattern, exochorionic morphology, inter-ridge/boundary area, aeropylar area, and basal layer.

Light microscopy

The female samples in Group 1 were dissected in Phosphate Buffer Saline (PBS; 10 mM sodium phosphate, 145 mM sodium chloride, pH 7.2) under a dissecting binocular microscope. Only the female heads and abdominal segments 8–11 were detached and mounted in Hoyer's medium. The morphological characteristics of the sand flies (cibarium, including armature and pigment patch; spermathecae) and eggs (chorionic sculpturing pattern) were examined and photographed using the LEICA DM1000 microscopy camera (Leica microsystems, Wetzlar, Germany). Species were identified using the adult morphological keys of Lewis [38, 39] as well as original descriptions and a recent taxonomical study by Depaquit and colleagues [28].

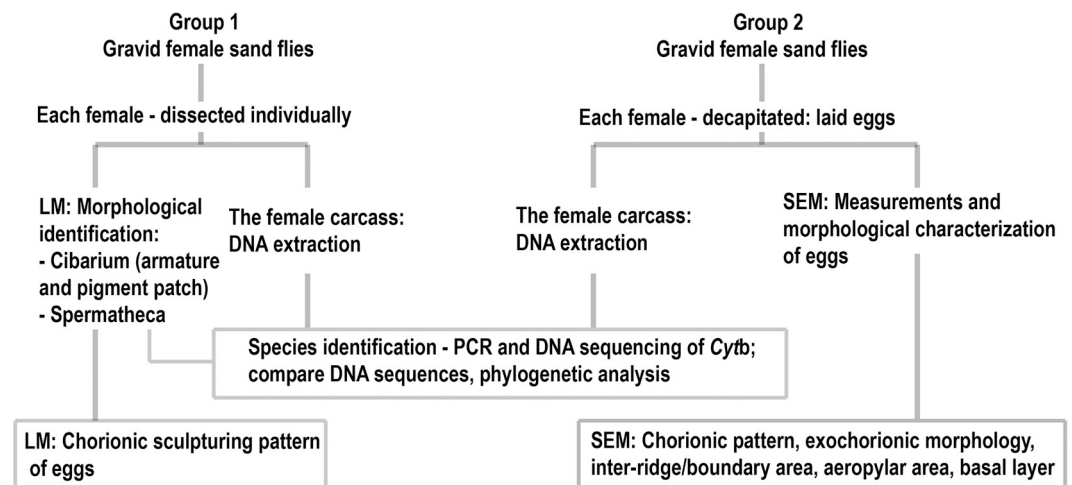


Fig 1. Experimental design for species identification of female sand flies and morphological examination and characterization of sand fly eggs.

<https://doi.org/10.1371/journal.pone.0263268.g001>

Scanning electron microscopy and measurements of eggs

Eggs from individual females were fixed with 2.5% glutaraldehyde in 0.1 cacodylate buffer (pH 7.2) for 12 h at 4°C and washed twice for 10 min with PBS (pH 7.2). Next, samples were fixed with 1% osmium tetroxide in PBS for 1 h, and then dehydrated in a graded series of ethanol (50%, 70%, 90%, 95% for 10 min each and then twice with 100% ethanol for 30 min each), followed by critical point drying in liquid CO₂ and coating with gold particles in a sputter-coating apparatus. The gold-coated preparations were examined under a scanning electron microscope, JEOL JSM-6610LV (JEOL, Tokyo, Japan), at 25–30 kV. SEM images of 30 eggs from at least three females in each species were used for egg measurements, including: length and width of egg, width and height of ridge, diameter and height of palisade unit, and diameter of tubercle. The values were presented as mean ± SD.

DNA extraction, PCR, DNA sequencing and phylogenetic analysis

Each female carcass was transferred into 1.5 ml microcentrifuge tubes containing 200 µl of lysis buffer for gDNA extraction using a tissue DNA extraction kit (GeneJet Genomic DNA Purification kit, Thermo scientific, USA) following the manufacturer's instructions. Genomic DNA was stored at 4°C until use. PCR of the *Cytb* gene was performed using the N1N-PDR (5'–CAY–ATT–CAA–CCW–GAA–TGA–TA –3') and C3B-PDR (5'–GGT–AYW–TTG–CCT–CGA–WTT–CGW–TAT–GA –3') primers [40, 41]. The reaction was carried out in a final volume of 25 µl including 2.5 µl PCR buffer, 2.5 µl MgCl₂, 2.5 µl dNTPs, 0.2 µl Taq polymerase, 0.4 µl forward, 0.4 µl reverse primers, 10.5 µl distilled water and 6 µl DNA template. PCR condition was performed with an initial denaturation step at 94°C for 3 min for 1 cycle, followed by 5 cycles: denaturation at 94°C for 30 sec, annealing at 40°C for 60 sec, extension at 68°C for 60 sec; and then 35 cycles of denaturation at 94°C for 60 sec, annealing at 45°C for 60 sec, extension at 68°C for 60 sec and a final extension step at 68°C for 10 min. PCR products were analyzed using electrophoresis on 1.5% agarose/1X TAE buffer gels, stained with ethidium bromide, and visualized under a UV transilluminator. PCRs were purified using the GeneJet PCR purification kit (Thermo Fisher scientific, USA) following the manufacturer's instructions. The purified PCRs were sent to Macrogen Inc., Seoul, South Korea, for sequencing. A phylogenetic tree was built using the maximum likelihood method with the Tamura 3-parameter distance model in MEGA X [42].

Results

Species identification

Twenty-two (22) gravid female sand flies were collected from the study area. They were examined and molecularly identified according to species. The *Cytb* sequences of each sample were deposited in GenBank as listed in Table 1. Phylogenetic analysis revealed three species of sand flies, *i.e.*, *Ph. stantoni*, *Se. khawi*, and *Gr. indica* (Fig 2).

LM and SEM analyses

The eggshell structures of the three sand fly species, *Ph. stantoni*, *Se. khawi*, and *Gr. indica*, were examined and described. In general, all eggs were elongated, ellipsoidal in shape with one side slightly flattened and rounded polar regions. In this study, only an aeropyle of *Ph. stantoni* was observed at the anterior pole of eggs.

Two chorionic patterns of eggs were found: (1) an unconnected parallel ridges pattern combined with a reticular pattern in *Ph. stantoni* and (2) a polygonal pattern in *Se. khawi* and *Gr. indica*. The details of the eggshell structures of the three sand fly species are described below.

Table 1. GenBank accession numbers of *Cytb* sequences used to identify the species of sand flies in this study.

Species	Group 1 (No.)	GenBank accession number
<i>Ph. stantoni</i>	3	OK398078, OK398080, OK398081
<i>Se. khawi</i>	4	OK398070, OK398071, OK398072, OK398073
<i>Gr. indica</i>	3	OK398062, OK398063, OK398069
	Group 2 (No.)	
<i>Ph. stantoni</i>	3	OK398079, OK398082, OK398083
<i>Se. khawi</i>	4	OK398074, OK398075, OK398076, OK398077
<i>Gr. indica</i>	5	OK398064, OK398065, OK398066, OK398067, OK398068

<https://doi.org/10.1371/journal.pone.0263268.t001>

Phlebotomus stantoni. *Phlebotomus stantoni* has a cibarium with approximately 15 pointed denticles of various lengths, which are irregularly arranged. The two or three median denticles are longer and stouter than the others and without a pigment patch (Fig 3A). The species has a fusiform spermatheca with 15 or 16 rings. The neck is thick and short, the head more or less oblong. The duct is striated and the common duct is very long, about 1.5 times the length of the individual ones, with a thick wall sclerotized at its basal part (Fig 3B). LM analysis of eggs in *Ph. stantoni* gravid females showed two patterns: unconnected parallel ridges and a reticular pattern on the same egg (Fig 3C).

Egg size. length = 301.20 ± 2.11 μm ; width = 83.65 ± 1.63 μm ; N = 30.

Chorionic pattern. The chorionic pattern of *Ph. stantoni* combines two patterns on the same egg, unconnected parallel ridges and a reticular pattern (Fig 4A–4D).

Exochorionic morphology. Ridges are formed by double non-columnar palisades that project from the surface of the eggshell. In some areas, the ridges are fused, forming an unconnected parallel ridges pattern. In other areas, the ridges exist separately thereby forming a reticular pattern. Each ridge is 3.37 ± 0.79 μm in width and 5.88 ± 0.35 μm in height (Fig 4C–4F).

Inter-ridge/boundary area. Fibrous material in the inter-ridge area is coarsely arranged with minute protuberances covering the basal layer (Fig 4E and 4F).

Aeropylar area. One aeropyle opening is present. The aeropyle is surrounded by randomly distributed protuberances and covered by very small dots (Fig 4G and 4H). The aeropylar area is delimited by a single non-columnar circular ridge.

Basal layer. A wide reticular mesh conformation of the basal layer near the aeropyle opening is present (Fig 4I).

Sergentomyia khawi. *Sergentomyia khawi* has a cibarium containing 10–12 hind teeth with two or three rows of fore teeth with a light pigment patch (Fig 5A). It has a narrow shaped spermatheca with a few wrinkles, with a knob in a deep narrow pit (Fig 5C). LM analysis of eggs in *Se. khawi* gravid females reveals polygonal (rectangular, pentagonal, or hexagonal) patterns of the chorion (Fig 5B–5D).

Egg size. length = 289.88 ± 1.78 μm ; width = 68.85 ± 1.20 μm ; N = 30.

Chorionic pattern. The ornamentation of the chorionic sculpturing pattern is polygonal (rectangular, pentagonal, and hexagonal). The polygonal pattern is due to ridges that are continuous, forming alternating transversal rows of polygonal cells (Fig 6A–6E).

Exochorionic morphology. Ridges are formed by the intersection of a single or double parallel series of rounded palisade units that project from the surface of eggshell. The diameter and height of each palisade unit are 1.26 ± 0.21 μm and 0.8 ± 0.14 μm , respectively. Almost all palisade units are linked or united at the top (Fig 6B and 6C).

Inter-ridge/boundary area. Finely arranged fibrous material as a compact coat covering the basal layer of the egg is present in the inter-ridge area (Fig 6D).

Aeropylar area. A wide reticular mesh conformation is present at the pole (Fig 6E).

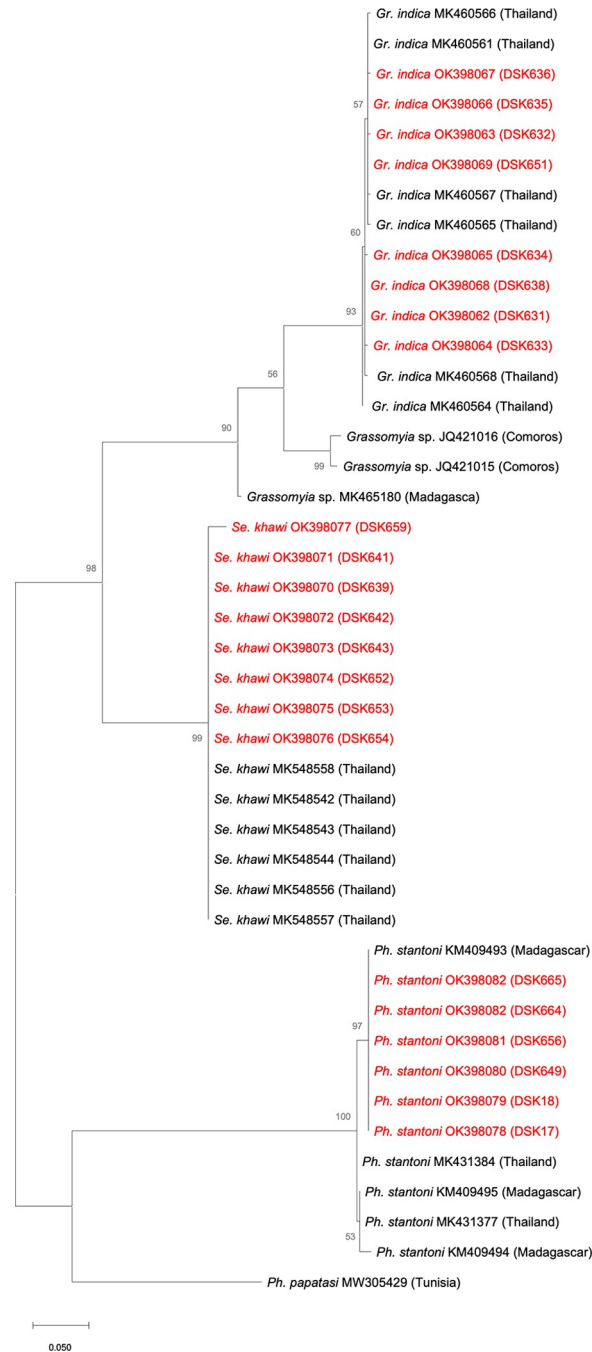


Fig 2. Maximum Likelihood tree based on the *Cytb* sequences using the Tamura 3-parameter (T92) distance in MEGA X. The sequences obtained in the present study are written in red.

<https://doi.org/10.1371/journal.pone.0263268.g002>

Grassyomyia indica. *Grassyomyia indica* has a cibarium with 29–30 teeth along a convex curved line and a dark brown pigment patch with a straight anterior margin (Fig 7A). It has a round finely speculate capsule spermatheca with minute projections at its tip (Fig 7C). LM analysis of eggs inside *Gr. indica* females reveals polygonal patterns (rectangular, pentagonal, or hexagonal) of the chorionic sculpturing (Fig 7B–7D).

Egg size. length = $265.63 \pm 0.99 \mu\text{m}$; width = $55.61 \pm 0.91 \mu\text{m}$; N = 30.

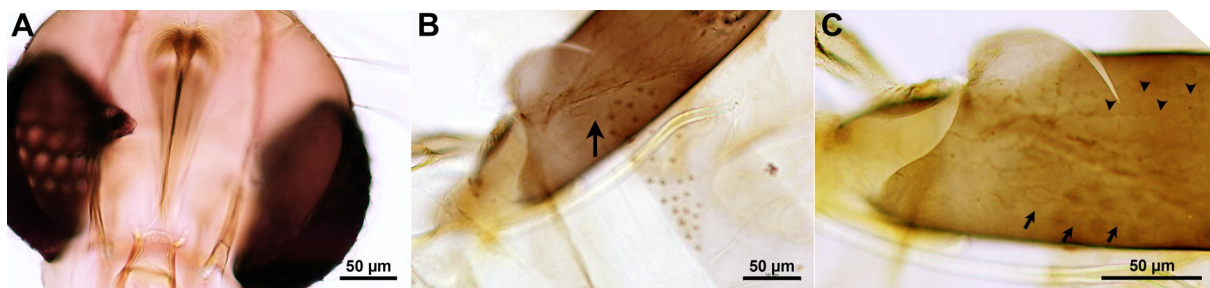


Fig 3. *Phlebotomus stantoni*. A. LM of head showing ciberial teeth; B. LM showing a spermatheca (arrow); C. LM showing a chorionic pattern of the egg with unconnected parallel ridges (arrowheads) and reticular pattern (arrows).

<https://doi.org/10.1371/journal.pone.0263268.g003>

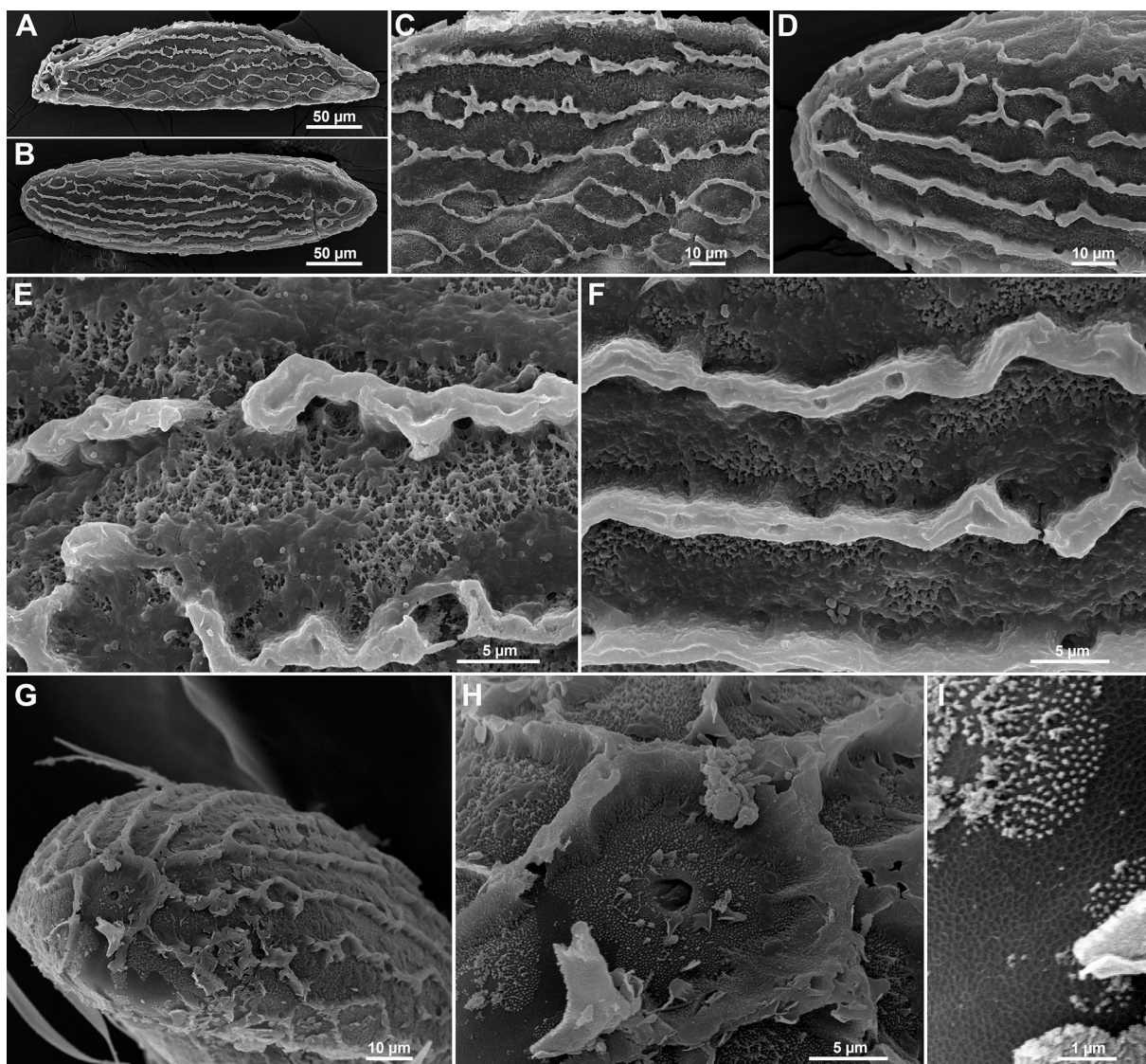


Fig 4. Ultrastructural morphology of *Ph. stantoni* eggs. A, B. SEM of whole eggs showing chorionic sculpturing with unconnected parallel ridges and a reticular pattern on the same egg; C, D. Details of the chorionic pattern; E, F. Coarsely arranged fibrous material which formed minute protuberances between ridges; G. Anterior pole showing the aeropylar area with one aeropyle opening; H. Higher magnification of the aeropylar area delimited by a single non-columnar circular ridge; I. A reticular basal layer near the aeropylar area.

<https://doi.org/10.1371/journal.pone.0263268.g004>

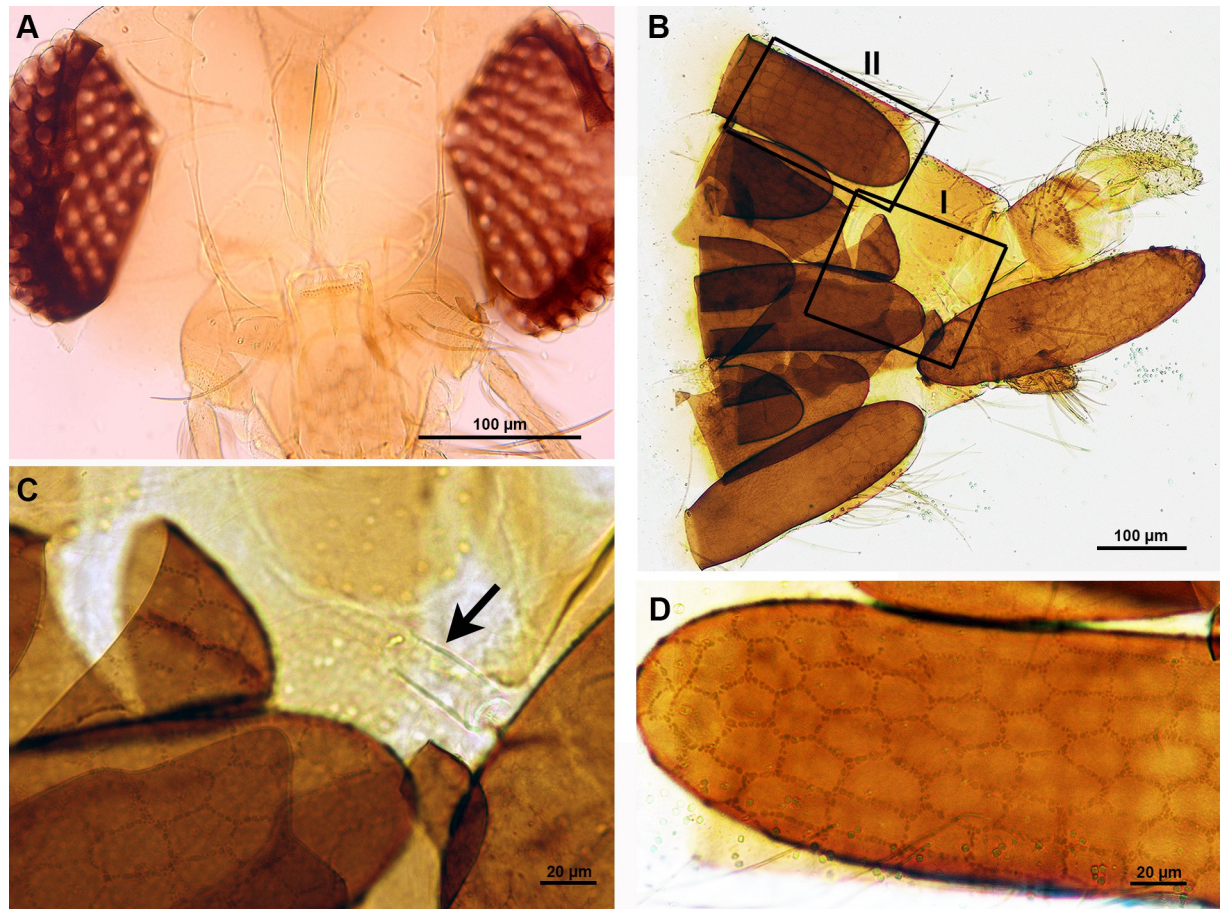


Fig 5. *Sergentomyia khawi*. A. LM of a head showing the cibarial teeth; B. Posterior abdominal region of a gravid female sand fly with eggs inside; C. Higher magnification of the area of box I in B showing a spermatheca (arrow); D. Higher magnification of the area of box II in B showing the polygonal pattern of the sand fly egg.

<https://doi.org/10.1371/journal.pone.0263268.g005>

Chorionic pattern. The ornamentation of the chorionic pattern is polygonal, mostly rectangular and pentagonal shapes, occasionally hexagonal shapes (Fig 8A–8D).

Exochorionic morphology. Boundaries of the polygonal shapes are formed by the intersection of 2–5 lines of small randomly arranged tubercles. Some tubercles project from the surface of the eggshell. The average of diameter of the tubercles is $0.68 \pm 0.14 \mu\text{m}$. No tubercles are linked or united at the top (Fig 8C and 8D).

Inter-ridge/boundary area. Fibrous material in the inter-ridge area is finely arranged as a compact coat covering the basal layer (Fig 8D).

Aeropylar area. A thick coat material is present at the pole (Fig 8C).

Table 2 summarizes the important details of ultrastructural morphology of Old World sand fly eggs examined by SEM.

Discussion

Eggs of approximately ten species of the genus *Phlebotomus* belonging to four subgenera, i.e., *Euphlebotomus*, *Larrousius*, *Phlebotomus*, and *Synphlebotomus* [20–25] and five species in the genus *Sergentomyia* belonging to two subgenera, i.e., *Rondanomyia* and *Sergentomyia* [20, 25]

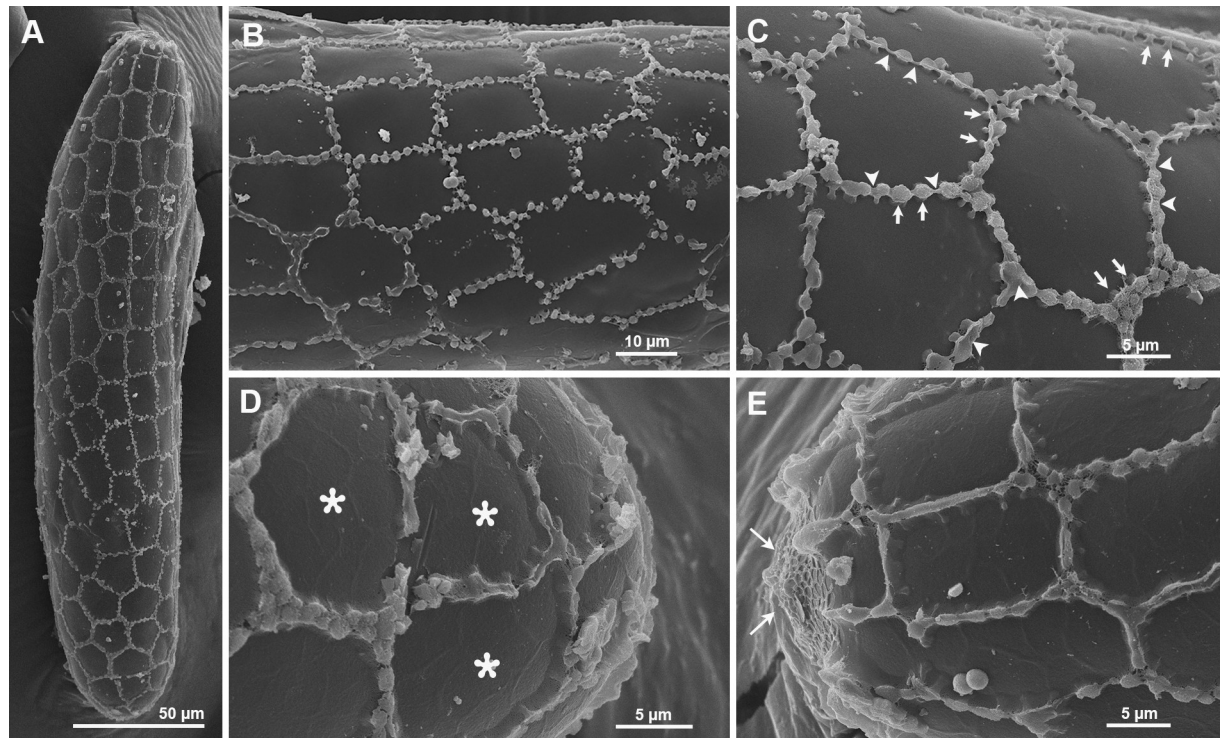


Fig 6. Ultrastructural morphology of *Se. khawi* eggs. A. SEM of a whole egg showing chorionic sculpturing with a polygonal pattern; B. Details of the general appearance of the chorionic sculpturing; C. High magnification image showing ridges formed by the intersection of single or double parallel series of rounded palisade units (arrows). Palisade units are linked or united at the top (arrowheads); D. Details of finely arranged fibrous material as a compact coat in inter-ridge areas (asterisks); E. Chorionic pattern near the pole and a thick reticular mesh (arrows).

<https://doi.org/10.1371/journal.pone.0263268.g006>

have been previously examined by SEM. In these past studies, the sand fly eggs were obtained from females reared in laboratories (laboratory strains) [22, 23, 25] or collected in the field. The females were allowed to feed on animal blood in the laboratory to favor egg development [20, 24, 25] but no molecular methods for species identification were mentioned. In the case of field-caught female sand flies, sibling species which are morphologically indistinguishable but reproductively isolated may be obtained together. Rogo and colleagues [24] observed that eggs laid by an individual female of *Ph. martini* were identical but those laid by different females of the same species (identified by morphology) were not. In addition, eggs of *Ph. martini* females collected from different origins, such as Kenya and Ethiopia, showed different chorionic patterns [20, 21].

Therefore, in this study, both morphological identification keys of female sand flies and DNA sequences of the *Cytb* gene were used for species identification and confirmation. In addition, a systematic collection of egg batches from individual females was performed by decapitating the gravid females. It is the first time this method was used in sand flies from Thailand and it allowed us to obtain synchronized eggs for SEM analysis. The species of the females and eggs were reliable and consistent by using these different approaches.

Our results show that the egg morphology of the three confirmed species, *Ph. stantoni*, *Se. khawi* and *Gr. indica*, followed the shape of eggs of all the previously described species: they have a long and oval shape with one side slightly flattened and rounded polar regions. Five details in the ultrastructural morphology of sand fly eggs including chorionic pattern, exochorionic morphology, inter-ridge/boundary area, aeropylar area and basal layer were examined

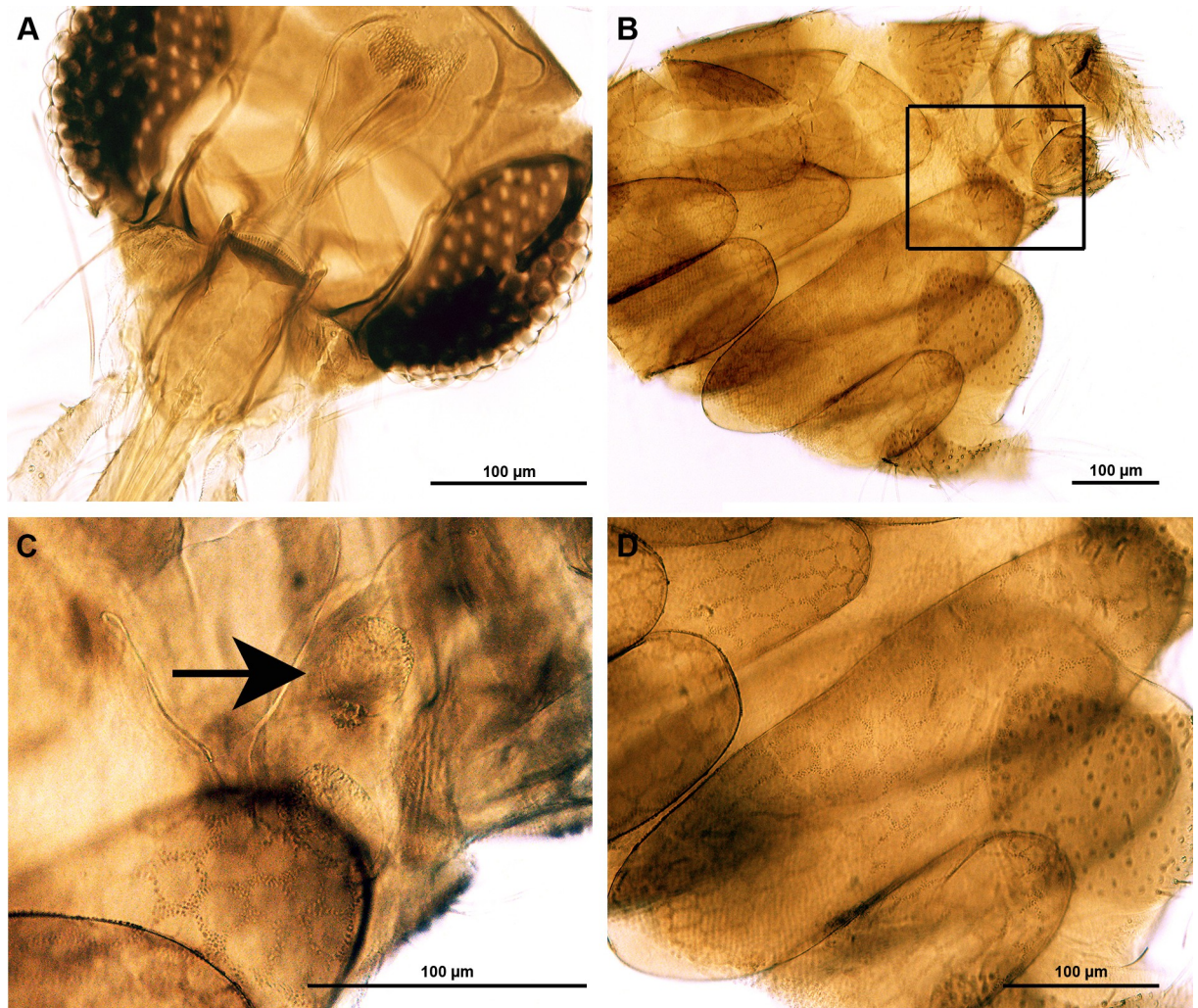


Fig 7. *Grassomyia indica*. A. LM of head showing cibarial teeth; B. Posterior abdominal region of a gravid female sand fly with eggs inside; C. Higher magnification from the area of box in B showing spermathecae (arrows); D. Polygonal pattern of the sand fly eggs.

<https://doi.org/10.1371/journal.pone.0263268.g007>

and characterized by SEM. However, only complete details of *Ph. stantoni* eggs were obtained. The chorionic sculpturing of *Ph. stantoni* (subgenus *Anaphlebotomus*) eggs differs from that of sand flies in other subgenera. We demonstrated for the first time that the chorionic pattern of *Ph. stantoni* eggs combines “unconnected parallel ridges” and a “reticular” pattern on the same egg. In *Ph. argentipes* (subgenus *Euphlebotomus*) and *Ph. papatasi* (subgenus *Phlebotomus*), although the chorionic pattern of eggs has been grouped under “connected parallel ridges”, a reticular pattern was observed in some areas on the eggs [25]. A combination of double parallel connected ridges and reticular pattern has also been reported in *Lu. verrucarum* [8]. Moreover, a variation of the chorionic sculpturing pattern among sand flies in the genus *Phlebotomus* has been reported. For example, in the subgenus *Larroussius*, the pattern of *Ph. perniciosus* and *Ph. perfiliewi* eggs is “unconnected parallel ridges” [23] whereas various patterns (polygonal, beehive cell, and parallel ridges) have been reported in *Ph. aculeatus* [24]. Furthermore, in the subgenus *Synphlebotomus*, the chorionic pattern of *Ph. celiae* eggs is “reticular” while *Ph. martini* sand flies from different countries (Kenya and Ethiopia) have different chorionic patterns

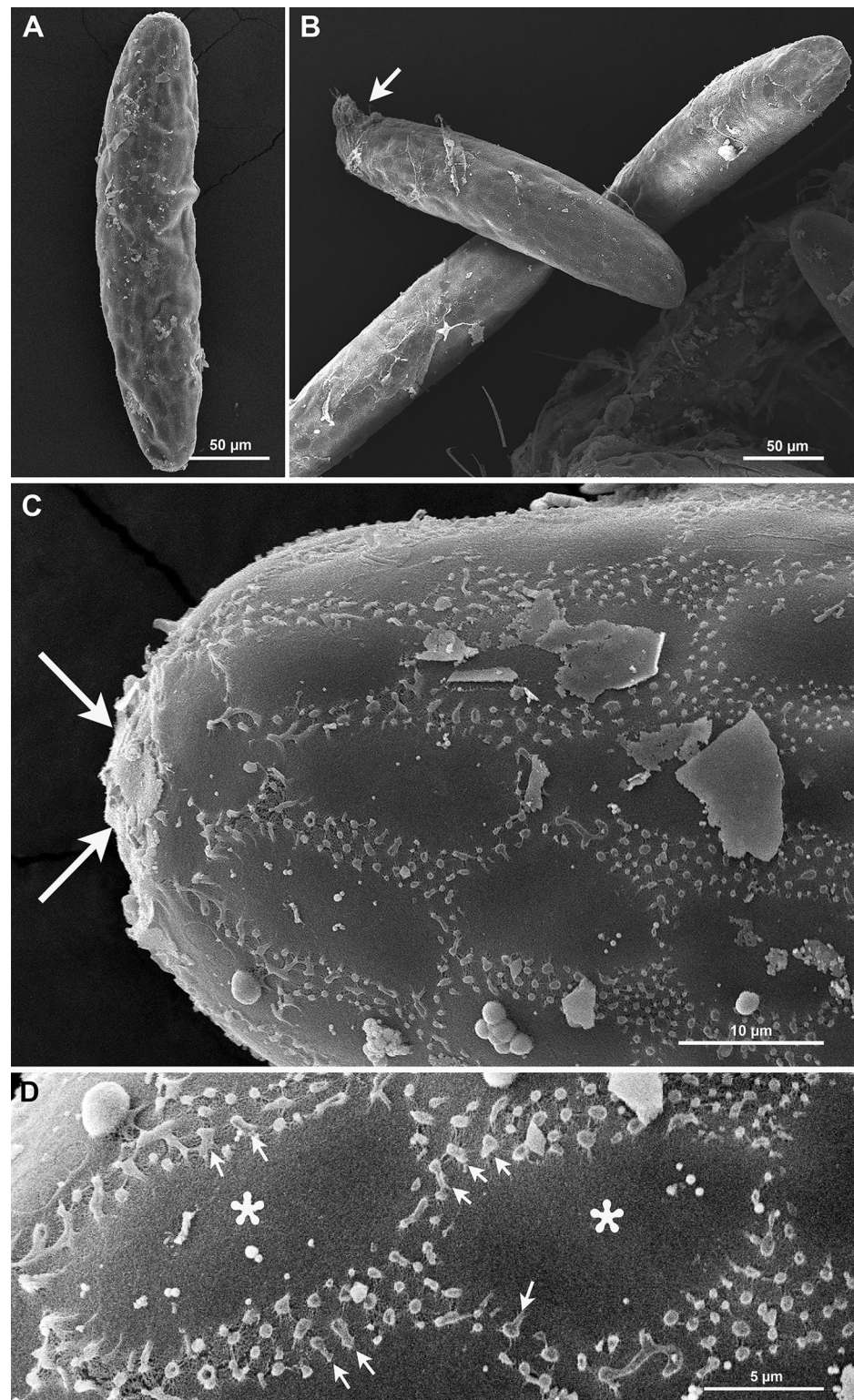


Fig 8. Ultrastructural morphology of *Gr. indica* eggs. A. SEM of whole eggs showing chorionic sculpturing with a polygonal pattern; B. Larval head protruding from the anterior pole (arrow); C. Details of the general appearance of the chorionic pattern and a thick coat material at the pole (arrows); D. High magnification image showing boundaries of polygonal shapes formed by intersection of 2–5 lines of small randomly arranged tubercles. Some tubercles project from the surface of the eggshell (arrows). Details of finely arranged fibrous material as a compact coat in inter-boundary areas (asterisks).

<https://doi.org/10.1371/journal.pone.0263268.g008>

Table 2. Important details of Old World sand fly eggs examined by SEM.

Species	Chorionic pattern	Exochorionic morphology	Inter-ridge/boundary area	Number of aeropyle, Aeropylar area	Basal layer	Reference (s)
Genus <i>Phlebotomus</i>						
Subgenus <i>Anaphlebotomus</i>						
<i>Ph. stantoni</i>	Unconnected parallel ridges and reticular	Ridges formed by double non-columnar palisades, Width = 2.1–4.2 μm and Height = 5.5–6.3 μm	Coarse minute protuberances	1, aeropyle surrounded by randomly distributed protuberances and covered by very small dots, single non-columnar circular ridge	Reticular	This study
Subgenus <i>Euphlebotomus</i>						
<i>Ph. argentipes</i> Annandale & Brunetti in Annandale	Connected parallel ridges (hexagonal in some areas)	Ridges formed by single column palisade series	Rough, having intercalated networks	ND ^a	ND	[25]
Subgenus <i>Phlebotomus</i>						
<i>Ph. papatasi</i> Scopoli	Connected parallel ridges (reticular in some areas)	Ridges formed by double column palisade series, Height = 4–4.5 μm	Coarse minute protuberances, rough, having uniform small granular protuberances	2, double non-columnar circular ridge, sub-divided by a transverse ridge into two semi-circular areas	Reticular	[22, 23, 25]
<i>Ph. duboscqi</i> Neveu-Lemaire	Reticular	Ridges formed by double column palisade series, Height = 3.5–4 μm	Coarse minute protuberances	2, single non-columnar circular ridge, sub-divided by transverse ridge into two semi-circular areas	Reticular	[23]
Subgenus <i>Larrousius</i>						
<i>Ph. perniciosus</i> Newstead	Unconnected parallel ridges	Ridges formed by double column palisade series, Height = 2–2.5 μm	Fine, as a compact coat	2, single non-columnar circular ridge, sub-divided by a transverse ridge into two semi-circular areas	Rough	[23]
<i>Ph. perfiliewi</i> Parrot	Unconnected parallel ridges	Ridges formed by single thin column palisade series, Height = 1.5–2 μm	Fine, as a compact coat	2, single non-columnar circular ridge, sub-divided by a transverse ridge into two semi-circular areas	Rough	[23]
<i>Ph. pedifer</i> Lewis, Mutinga & Ashford	Parallel ridges	ND	ND	ND	ND	[24]
<i>Ph. aculeatus</i> Lewis, McMillan & Ashford	Variations: polygonal, beehive cell, parallel ridges	ND	ND	ND	ND	[24]
<i>Ph. major</i> Annandale	Polygonal	Ridges formed by columns of granular cells arranged to form a palisade	ND	ND	ND	[25]
Subgenus <i>Synphlebotomus</i>						
<i>Ph. celiae</i> Minter	Reticular	ND	ND	ND	ND	[21]
<i>Ph. martini</i> Parrot	Kenya: Connected parallel ridges	ND	ND	ND	ND	[21]
	Ethiopia: Reticular	ND	ND	ND	ND	[20]
Genus <i>Sergentomyia</i>						
Subgenus <i>Rondanomyia</i>						
<i>Se. khawi</i>	Polygonal	Ridges formed by single or double parallel series of rounded palisade units, Diameter = 0.7–1.5 μm and Height = 0.6–1.0 μm	Fine, as a compact coat	ND, a wide reticular mesh conformation	ND	This study

(Continued)

Table 2. (Continued)

Species	Chorionic pattern	Exochorionic morphology	Inter-ridge/boundary area	Number of aeropyle, Aeropylar area	Basal layer	Reference (s)
<i>Se. zeylanica</i> (Annandale)	Polygonal	Solid granules of various sizes, arranged in a column to form a palisade, ridges appear to be wide and solid, uniform cord like structures	ND	ND	ND	[25]
<i>Se. kirki</i> (Parrot)	Volcano-like	ND	ND	ND	ND	[20]
Subgenus <i>Sergentomyia</i>						
<i>Se. garnhami</i> (Heisch, Guggisberg & Teesdale)	Parallel ridges	ND	ND	ND	ND	[20]
<i>Se. bedfordi</i> (Newstead)	Polygonal	ND	ND	ND	ND	[20]
Genus <i>Grassomyia</i>						
<i>Gr. indica</i>	Polygonal	Boundaries formed by 2–5 lines of small randomly arranged tubercles, Diameter = 0.3–0.8 μ m	Fine, as a compact coat	ND, covered by a thick coat material	ND	This study

^aND = not described

<https://doi.org/10.1371/journal.pone.0263268.t002>

of eggs, such as “connected parallel ridges”, “reticular”, “mountain-like”, irregular patterns [20, 21, 24].

In the genus *Sergentomyia*, a polygonal pattern is quite common, however, a “volcano-like” pattern and “parallel ridges” have been reported in *Se. kirki* (subgenus *Rondanomyia*) and *Se. garnhami* (subgenus *Sergentomyia*), respectively [20]. The chorionic pattern of *Se. khawi* and *Gr. indica* from Thailand is polygonal but their exochorionic morphology and aeropylar area are different. The exochorionic morphology of *Gr. indica* is unique. Boundaries of the polygonal pattern formed by 2–5 lines of small randomly arranged tubercles in *Gr. indica* were noted and described for the first time. The aeropylar area of *Se. khawi* is covered by a wide reticular mesh conformation whereas the aeropylar area of *Gr. indica* is covered by a thick coat material. Differences in exochorionic morphology, the inter-ridge/boundary area, the aeropylar area (including the number of aeryples) and basal layer of some sand fly species in the genus *Phlebotomus* in the New World have been reported [10, 12, 15–19, 22, 23, 25]. Therefore, the chorionic pattern together with other important details of the ultrastructural morphology may be useful for developing morphological identification keys of eggs as an additional tool to distinguish species of sand flies. More information on egg ultrastructural morphology of sand flies in the Old World and the New World with correct molecular genetic information are required for further analysis to ascertain their taxonomic values before any conclusion can be drawn.

Although the use of SEM has significantly improved the characterization of the ultrastructural morphology of sand fly eggs, in this study, the chorionic sculpturing of the eggs of *Ph. stantoni*, *Se. khawi*, and *Gr. indica*, were observed by LM. We demonstrate that in the case of gravid females (whose spermathecae is obscured by eggs), the chorionic sculpturing of eggs is useful to identify these three species. In this study, the developmental stages of eggs in the gravid female sand flies could not be estimated since these samples were taken from females caught in the wild. Our results showed that in LM, the chorionic pattern of eggs inside the females of each species was the same as shown in SEM. The egg samples for SEM were allowed to fully melanise for 24 h before SEM processing. This suggests that the chorionic patterns of fully mature eggs were present in the abdomens of gravid females of these three sand fly

species. In *Lu. longipalpis*, fully melanised eggs have been observed in the abdomens of gravid sand flies. It highlights that sand fly eggs can fully melanise prior to oviposition [43]. Therefore, it is interesting to further investigate chorionic patterns in each developmental stages of sand fly eggs of each species. In addition, further studies on eggshell morphology of sand flies from other areas and other species are needed to compare eggshell patterns as the environments of oviposition sites influence the exochorion of sand fly eggs [4]. At least two species in the same genus in the Old World, i.e., *Phlebotomus*, *Sergentomyia*, *Grassomyia*, *Idiophlebotomus*, *Chinius*, *Parvidens*, *Spelaomyia* and *Spelaephlebotomus* should be investigated to evaluate their informativity for classification of sand flies.

In conclusion, the ultrastructural morphology of the eggshells of *Ph. stantoni*, *Se. khawi*, and *Gr. indica* sand flies was described for the first time. Our findings indicate that the chorionic sculpturing pattern together with other important details of the ultrastructural morphology can be used to develop morphological identification keys of eggs. These can serve as an additional tool to distinguish species of sand flies. The systematic collection of egg batches from individually gravid females by decapitating allowed us to obtain synchronized eggs for SEM analysis. In addition, insect species were confirmed using both *Cytb* sequencing as well as adult morphological identification keys. Together, these methods provided useful information for sand fly ootaxonomy. Furthermore, the chorionic sculpturing of the eggs of these three species observed by LM is useful for species identification in the case of gravid females with spermathecae obscured by eggs.

Acknowledgments

We thank Mr. Sakone Sunantaraporn for his technical help and the house owner of the collection site for giving his permission.

Author Contributions

Conceptualization: Narissara Jariyapan.

Formal analysis: Narissara Jariyapan, Pradya Somboon, Jérôme Depaquit.

Funding acquisition: Narissara Jariyapan, Pongsri Tippawangkosol.

Investigation: Narissara Jariyapan, Sriwatapron Sor-Suwan, Chonlada Mano, Thippawan Yasanga.

Methodology: Narissara Jariyapan, Pongsri Tippawangkosol.

Project administration: Narissara Jariyapan, Pongsri Tippawangkosol.

Resources: Narissara Jariyapan, Pradya Somboon.

Validation: Narissara Jariyapan, Jérôme Depaquit.

Writing – original draft: Narissara Jariyapan.

Writing – review & editing: Narissara Jariyapan, Pradya Somboon, Jérôme Depaquit, Padet Siriyasatien.

References

1. Galati EAB. Phlebotominae (Diptera, Psychodidae): Classification, Morphology and Terminology of Adults and Identification of American Taxa. In: Rangel E., Shaw J. (eds) Brazilian Sand Flies. Springer, Cham; 2018. Pages 9–212.
2. Marialva EF, Secundino NF, Fernandes FF, Araújo HRC, Ríos-Velásquez CM, Pimenta PFP, et al. Morphological aspects of immature stages of *Migonemyia migonei* (Diptera: Psychodidae, Phlebotominae),

- an important vector of Leishmaniasis in South America, described by scanning electron microscopy. PLoS One. 2020; m15(11):e0242163. <https://doi.org/10.1371/journal.pone.0242163> PMID: 33180853; PMCID: PMC7660558.
3. Tyagi V, Sharma AK, Dhiman S, Srivastava AR, Yadav R, Sukumaran D, et al. Malaria vector *Anopheles culicifacies* sibling species differentiation using egg morphometry and morphology. Parasit Vectors. 2016; 9:202. <https://doi.org/10.1186/s13071-016-1478-5> PMID: 27075571; PMCID: PMC4831176.
 4. Ward RD, Ready P. Chorionic sculpturing of some sandfly eggs (Diptera: Psychodidae). J. Entomol. 1975; Ser. A 50:127–134.
 5. Zimmerman JH, Newson HD, Hooper GR, Christensen HA. A comparison of the egg surface structure of six anthropophilic phlebotomine sand flies (*Lutzomyia*) with the scanning electron microscope (Diptera: Psychodidae). J Med Entomol. 1977; 13(4–5):574–579. <https://doi.org/10.1093/jmedent/13.4-5.574> PMID: 557563.
 6. Endris RG, Young DG, Perkins PV. Ultrastructural comparison of egg surface morphology of five *Lutzomyia* species (Diptera: Psychodidae). J. Med. Entomol. 1987; 24:412–415. <https://doi.org/10.1093/jmedent/24.4.412>.
 7. Feliciangeli MD, Castejon OC, Limongi J. Egg surface ultrastructure of eight New World phlebotomine sand fly species (Diptera: Psychodidae). J Med Entomol. 1993; 30(4):651–656. <https://doi.org/10.1093/jmedent/30.4.651> PMID: 8360889.
 8. Enrique Pérez J, Ogusuku E. Chorion patterns on eggs of *Lutzomyia* sandflies from the Peruvian Andes. Med Vet Entomol. 1997; 11(2):127–133. <https://doi.org/10.1111/j.1365-2915.1997.tb00301.x> PMID: 9226640.
 9. Sierra DA, Vélez IDB, Uribe SS. Identificación de *Lutzomyia* spp. (Diptera: Psychodidae) grupo verrucarum por medio de microscopía electrónica de sus huevos. Rev. Biol. Trop. 2000; 48(2–3):615–622. http://www.scielo.sa.cr/scielo.php?script=sci_arttext&pid=S0034-7744200000200033&lng=en. PMID: 11354968
 10. Fausto AM, Feliciangeli MD, Maroli M, Mazzini M. Ootaxonomic investigation of five *Lutzomyia* species (Diptera, Psychodidae) from Venezuela. Mem Inst Oswaldo Cruz. 2001; 96(2):197–204. <https://doi.org/10.1590/s0074-02762001000200011> PMID: 11285497
 11. Noguera P, Chaves LF, Nieves E. Effects of blood ingestion on patterns on the chorion of eggs of *Lutzomyia ovallesi* (Diptera: Psychodidae). Parasitol. Latinoam. 2003; 58:49–53. <https://doi.org/10.4067/S0717-77122003000100008>.
 12. De Almeida DN, Da Silva OR, Brazil BG, Soares MJ. Patterns of exochorion ornaments on eggs of seven South American species of *Lutzomyia* sand flies (Diptera: Psychodidae). J Med Entomol. 2004; 41(5):819–825. <https://doi.org/10.1603/0022-2585-41.5.819> PMID: 15535608.
 13. Pessoa FAC, Feitosa MAC, Bermudez EGC, Ríos-Velásquez CM, Ward RD. Immature stages two species of *Evandromyia* (*Aldamyia*) and the systematic importance of larval mouthparts within Psychodidae (Diptera, Phlebotominae, Psychodinae). Zootaxa. 2008; 1740:1–14. <https://doi.org/10.11646/zootaxa.1740.1.1>.
 14. Bahia AC, Secundino NF, Miranda JC, Prates DB, Souza AP, Fernandes FF, et al. Ultrastructural comparison of external morphology of immature stages of *Lutzomyia* (*Nyssomyia*) *intermedia* and *Lutzomyia* (*Nyssomyia*) *whitmani* (Diptera: Psychodidae), vectors of cutaneous leishmaniasis, by scanning electron microscopy. J Med Entomol. 2007; 44(6):903–914. [https://doi.org/10.1603/0022-2585\(2007\)44\[903:ucoemo\]2.0.co;2](https://doi.org/10.1603/0022-2585(2007)44[903:ucoemo]2.0.co;2) PMID: 18047187.
 15. Costa WA, da Costa SM, Rangel EF, dos Santos-Mallet JR, Serrão JE. Eggshell as a characteristic to identify *Lutzomyia* (*Nyssomyia*) *intermedia* (Lutz & Neiva, 1912) and *Lutzomyia* (*Nyssomyia*) *neivai* (Pinto, 1926) (Diptera: Psychodidae: Phlebotominae), vectors of cutaneous leishmaniasis. Revista Pan-Amazônica de Saúde 2012; 3:19–24. <https://doi.org/10.5123/S2176-62232012000100003>.
 16. Castillo A, Serrano AK, Mikery OF, Pérez J. Life history of the sand fly vector *Lutzomyia cruciata* in laboratory conditions. Med Vet Entomol. 2015; 29(4):393–402. <https://doi.org/10.1111/mve.12127> PMID: 26147368.
 17. Montes de Oca-Aguilar AC, Rebollar-Téllez EA, Piermarini PM, Ibáñez-Bernal S. Descriptions of the immature stages of *Lutzomyia* (*Tricholateralis*) *cruciata* (Coquillett) (Diptera: Psychodidae, Phlebotominae). Neotrop Entomol. 2017; 46(1):66–85. <https://doi.org/10.1007/s13744-016-0439-1> PMID: 27631128.
 18. Alencar RB, Justiniano SCB, Scarpassa VM. Morphological description of the immature stages of *Nyssomyia umbratilis* (Ward & Frahia) (Diptera: Psychodidae: Phlebotominae), the main vector of *Leishmania guyanensis* Floch (Kinetoplastida: Trypanosomatidae) in the Brazilian Amazon Region. Neotrop Entomol. 2018; 47(5):668–680. <https://doi.org/10.1007/s13744-018-0587-6> PMID: 29374388.

19. Alencar RB, Scarpassa VM. Morphology of the eggs surface of ten Brazilian species of phlebotomine sandflies (Diptera: Psychodidae). *Acta Trop*. 2018; 187:182–189. <https://doi.org/10.1016/j.actatropica.2018.07.023> PMID: 30055173.
20. Irungu LW, Mutinga MJ, Kokward ED. Chorionic sculpturing of eggs of some Kenyan phlebotomine sand flies. *Insect Sci. Appl*. 1985; 7:45–48. <https://doi.org/10.1017/S174275840000309X>.
21. Gebre-Michael T, Lane RP. Scanning electron microscopy of eggs of *Phlebotomus* (*Synphlebotomus*) *martini* and *P.* (*Syn.*) *celiae* (Diptera: Phlebotominae). *Parassitologia*. 1991; 33 Suppl:261–266. PMID: 1841216.
22. Fausto AM, Maroli M, Mazzini M. Ootaxonomy investigation of three sandfly species (Diptera: Psychodidae) from Italy. *Parassitologia*. 1991; 33 Suppl:225–228. PMID: 1841211.
23. Fausto AM, Maroli M, Mazzini M. Ootaxonomy and eggshell ultrastructure of *Phlebotomus* sandflies. *Med Vet Entomol*. 1992; 6(3):201–208. <https://doi.org/10.1111/j.1365-2915.1992.tb00607.x> PMID: 1421499.
24. Rogo LM, Kokwaro ED, Mutinga MJ, Khamala CP. Differentiation of vector species of phlebotominae (Diptera: Psychodidae) in Kenya by chorionic sculpturing of their eggs. *J Med Entomol*. 1992; 29(6):1042–1044. <https://doi.org/10.1093/jmedent/29.6.1042> PMID: 1460620.
25. Ghosh KN, Mukhopadhyay JA. A comparison of chorionic sculpturing of four Indian phlebotomine sand flies (Diptera: Psychodidae) by scanning electron microscopy. *Parasite*. 1996; 3:61–68. <https://doi.org/10.1051/parasite/1996031061>.
26. Polseela R, Vitta A, Apiwathnasorn C. Distribution of phlebotomine sand flies (Diptera: Psychodidae) in limestone caves, Khao Pathawi, Uthai Thani province, Thailand. *Southeast Asian J Trop Med Public Health*. 2015; 46(3):425–433. PMID: 26521515.
27. Phumee A, Tawatsin A, Thavara U, Pongsakul T, Thammapalo S, Depaquit J, et al. Detection of an unknown *Trypanosoma* DNA in a *Phlebotomus stantoni* (Diptera: Psychodidae) collected from Southern Thailand and records of new sand flies with reinstatement of *Sergentomyia hivernus* Raynal & Gaschen, 1935 (Diptera: Psychodidae). *J Med Entomol*. 2017; 54(2):429–434. <https://doi.org/10.1093/jme/tjw161> PMID: 27744363.
28. Depaquit J, Vongphayloth K, Siriyasatien P, Polseela R, Phumee A, Loyer M, et al. On the true identity of *Sergentomyia gemmea* and description of a closely related species: *Se. raynali* n. sp. *Med Vet Entomol*. 2019; 33(4):521–529. <https://doi.org/10.1111/mve.12393> PMID: 31155766.
29. Phuphisut O, Nitatsukprasert C, Pathawong N, Jaichapor B, Pongsiri A, Adisakwattana P, et al. Sand fly identification and screening for *Leishmania* spp. in six provinces of Thailand. *Parasit Vectors*. 2021; 14(1):352. <https://doi.org/10.1186/s13071-021-04856-6> PMID: 34217359; PMCID: PMC8254935.
30. Kanjanopas K, Siripattanapipong S, Ninsaeng U, Hitakarun A, Jitkaew S, Kaewtaphaya P, et al. *Sergentomyia* (*Neophlebotomus*) *gemmea*, a potential vector of *Leishmania siamensis* in southern Thailand. *BMC Infect Dis*. 2013; 13:333. <https://doi.org/10.1186/1471-2334-13-333> PMID: 23870062; PMCID: PMC3725172.
31. Chusri S, Thammapalo S, Chusri S, Thammapalo S, Silpapojakul K, Siriyasatien P. Animal reservoirs and potential vectors of *Leishmania siamensis* in southern Thailand. *Southeast Asian J Trop Med Public Health*. 2014; 45(1):13–9. PMID: 24964648.
32. Sriwongpan P, Nedsuwan S, Manomat J, Charoensakulchai S, Lacharajana K, Sankwan J, et al. Prevalence and associated risk factors of *Leishmania* infection among immunocompetent hosts, a community-based study in Chiang Rai, Thailand. *PLoS Negl Trop Dis*. 2021; 15(7):e0009545. <https://doi.org/10.1371/journal.pntd.0009545> PMID: 34252099; PMCID: PMC8297947.
33. Srisuton P, Phumee A, Sunantaraporn S, Boonserm R, Sor-Suwan S, Brownell N, et al. Detection of *Leishmania* and *Trypanosoma* DNA in field-caught sand flies from endemic and non-endemic areas of Leishmaniasis in southern Thailand. *Insects*. 2019; 10(8):238. <https://doi.org/10.3390/insects10080238> PMID: 31382501; PMCID: PMC6722825.
34. Phumee A, Tawatsin A, Thavara U, Pongsakul T, Thammapalo S, Depaquit J, et al. Detection of an unknown *Trypanosoma* DNA in a *Phlebotomus stantoni* (Diptera: Psychodidae) collected from southern Thailand and records of new sand flies with reinstatement of *Sergentomyia hivernus* Raynal & Gaschen, 1935 (Diptera: Psychodidae). *J Med Entomol*. 2017; 54(2):429–434. <https://doi.org/10.1093/jme/tjw161> PMID: 27744363.
35. Phumee A, Wacharapluesadee S, Petcharat S, Tawatsin A, Thavara U, Siriyasatien P. Detection of Changuinola virus (Reoviridae: Orbivirus) in field-caught sand flies in southern Thailand. *Trans R Soc Trop Med Hyg*. 2021; 115(9):1039–1044. <https://doi.org/10.1093/trstmh/traa203> PMID: 33515044; PMCID: PMC8417081.
36. Phumee A, Wacharapluesadee S, Petcharat S, Siriyasatien P. A new cluster of Rhabdovirus detected in field-caught sand flies (Diptera: Psychodidae: Phlebotominae) collected from southern Thailand. *Parasit Vectors*. 2021; 14:569. <https://doi.org/10.1186/s13071-021-05047-z> PMID: 34749797

37. Sor-Suwan S, Jariyapan N, Mano C, Apiwathnasorn C, Sriwichai P, Samung Y, et al. Species composition and population dynamics of phlebotomine sand flies in a *Leishmania* infected area of Chiang Mai, Thailand. *Trop Biomed*. 2017; 34(4):855–862. PMID: [33592954](#).
38. Lewis D J. The Phlebotomine sandflies (Diptera: Psychodidae) of the Oriental Region. *Bulletin of the British Museum (Natural History). Entomol Ser*. 1978; 37:217–343.
39. Lewis D J. Phlebotomine sandflies (Diptera: Psychodidae) from the Oriental Region. *Syst Entomol*. 1987; 12:163–180. <https://doi.org/10.1111/j.1365-3113.1987.tb00194.x>.
40. Esseghir S, Ready PD, Killick-Kendrick R, Ben-Ismaïl R. Mitochondrial haplotypes and phylogeography of *Phlebotomus* vectors of *Leishmania major*. *Insect Mol Biol*. 1997; 6(3):211–225. <https://doi.org/10.1046/j.1365-2583.1997.00175.x> PMID: [9272439](#).
41. Depaquit J, Leger N, Randrianambinintsoa FJ. Paraphyly of the subgenus *Anaphlebotomus* and creation of *Madaphlebotomus* subg. nov. (Phlebotominae: *Phlebotomus*). *Med Vet Entomol*. 2015; 29(2):159–1570. <https://doi.org/10.1111/mve.12098> PMID: [25613531](#).
42. Kumar S, Stecher G, Li M, Knyaz C, Tamura K. MEGA X: Molecular evolutionary genetics analysis across computing platforms. *Mol Biol Evol*. 2018; 35(6):1547–1549. <https://doi.org/10.1093/molbev/msy096> PMID: [29722887](#); PMCID: [PMC5967553](#).
43. Jeffries CL, Rogers ME, Walker T. Establishment of a method for *Lutzomyia longipalpis* sand fly embryo microinjection: The first step towards potential novel control strategies for leishmaniasis. *Wellcome Open Res*. 2018; 3:55. <https://doi.org/10.12688/wellcomeopenres.14555.2> PMID: [30175240](#); PMCID: [PMC6107984](#).