

RESEARCH ARTICLE

Neural substrates of neuropsychological profiles in dystrophinopathies: A pilot study of diffusion tractography imaging

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Abstract

Introduction

Cognitive difficulties and neuropsychological alterations in Duchenne and Becker muscular dystrophy (DMD, BMD) boys are not yet sufficiently explored, although this topic could have a relevant impact, finding novel biomarkers of disease both at genetics and neuroimaging point of view. The current study aims to: 1) analyze the neuropsychological profile of a group of DMD and BMD boys without cognitive impairment with an assessment of their executive functions; 2) explore the structural connectivity in DMD, BMD, and age-matched controls focusing on cortico-subcortical tracts that connect frontal cortex, basal ganglia, and cerebellum via the thalamus; 3) explore possible correlations between altered structural connectivity and clinical neuropsychological measures.

Materials and methods

This pilot study included 15 boys (5 DMD subjects, 5 BMD subjects, and 5 age-matched typically developing, TD). They were assessed using a neuropsychological assessment protocol including cognitive and executive functioning assessment and performed a 1.5T MRI brain exam including advanced Diffusion Weighted Imaging (DWI) method for tractography. Structural connectivity measurements were extracted along three specific tracts: Cortico-Ponto-Cerebellar Tract (CPCT), Cerebellar-Thalamic Tract (CTT), and Superior Longitudinal Fasciculus (SLF). Cortical-Spinal Tract (CST) was selected for reference, as control tract.

Results

Regarding intellectual functioning, a major impairment in executive functions compared to the general intellectual functioning was observed both for DMD (mean score = 86.20;

SD = 11.54) and for BMD children (mean score = 88; SD = 3.67). Mean FA resulted tendentially always lower in DMD compared to both BMD and TD groups for all the examined tracts. The differences in FA were statistically significant for the right CTT (DMD vs BMD, $p = 0.002$, and DMD vs TD, $p = 0.0015$) and the right CPCT (DMD vs TD, $p = 0.008$). Concerning DMD, significant correlations emerged between FA-R-CTT and intellectual quotients (FIQ, $p = 0.044$; $\rho_s = 0.821$), and executive functions (Denomination Total, $p = 0.044$, $\rho_s = 0.821$; Inhibition Total, $p = 0.019$, $\rho_s = 0.900$). BMD showed a significant correlation between FA-R-CPCT and working memory index ($p = 0.007$; $\rho_s = 0.949$).

Discussion and conclusion

In this pilot study, despite the limitation of sample size, the findings support the hypothesis of the involvement of a cerebellar-thalamo-cortical loop for the neuropsychological profile of DMD, as the CTT and the CPCT are involved in the network and the related brain structures are known to be implied in executive functions. Our results suggest that altered WM connectivity and reduced fibre organization in cerebellar tracts, probably due to the lack of dystrophin in the brain, may render less efficient some neuropsychological functions in children affected by dystrophinopathies. The wider multicentric study could help to better establish the role of cerebellar connectivity in neuropsychological profile for dystrophinopathies, identifying possible novel diagnostic and prognostic biomarkers.

Introduction

Dystrophinopathies are the most common single gene disorders leading to muscle wasting, due to mutations in *DMD* gene. In-frame mutations generally produce abnormal but partly functional gene product *dystrophin*, which is part of a protein complex located in the muscle cell membrane, and are associated with Becker muscular dystrophy (BMD). The variable expression of the protein reflects the widely variable clinical phenotype observed in BMD children, in terms of motor and cardiorespiratory outcome [1]. On the contrary, mutations that disrupt the reading frame commonly result in the lack of dystrophin, leading to the most severe disease Duchenne muscular dystrophy (DMD) [2].

In addition to the well known peripheral muscular involvement, several studies have also reported a central nervous system involvement, resulting in cognitive difficulties and neuropsychological alterations in DMD boys [3–13], while less is known about possible neuropsychological impairments in BMD [14–16].

From a neuropathological point of view, the cognitive and neuropsychological involvement in DMD may be due to the lack of specific dystrophin isoforms in the brain. Two alternative full-length isoforms (Dp427B and Dp427P) are indeed expressed in the cerebral neocortex and two carboxy-terminal dystrophin proteins, Dp71 and Dp140, in the Purkinje cells of the cerebellar cortex [17–20]. Dp140 is expressed mainly in fetal tissue and in low quantity in adult brain and probably plays a role in the regulation of neuroglial specific gene expression. Dp71 expression gradually increases from the embryo until adult stage, becoming the major product of dystrophin in the brain, particularly in the hippocampus and in some layers of the cerebral cortex. The Dp71 function remains unknown but a role in the formation and/or stabilization of the dystrophin-associated complex and in glutaminergic synaptic maturation and function is supported by studies [21, 22].

In view of the localization of dystrophin isoforms in normal brain and of the neurocognitive involvement of DMD subjects, a possible role of cerebellum and of a complex cerebro-cerebellar network has been hypothesized in DMD [23].

Recently, an impairment of multitasking, problem solving, inhibition, working memory [24] and an implicit learning deficit [25] emerged in a group of 40 DMD boys without intellectual disability during school age. To examine this a selected DMD sample has allowed a targeted neurocognitive detection free from potential bias. Tasks requiring executive functions that have been explored in these studies are believed to activate a cortico-subcortical circuits which connect the prefrontal cortex, the basal ganglia and the cerebellum via the thalamus [26], thus strengthening the hypothesis of a possible involvement of the cerebellum as part of a more general involvement of the cerebellar-thalamo-cortical network.

Neuroimaging studies, although few in number, also corroborate the hypothesis of the involvement of cerebro-cerebellar loops in DMD: for example, positron emission topography (PET) analysis demonstrated in DMD patients glucose hypometabolism in brain areas that are typically rich in dystrophin (as cerebral cortex and cerebellum) [27, 28]. Moreover, the application of Magnetic Resonance Spectroscopy (MRS) in DMD patients revealed alterations in choline levels in both cerebellar white matter among others [29, 30]. A brain involvement in DMD has been also supported by functional MR studies that showed a reduced local synchronization of spontaneous activity of the neural networks in the motor cortex in patients with DMD [31] and a reduction of cerebral perfusion in DMD, regardless of the reduced grey matter volume [32].

Despite growing evidences of a brain dysfunction in DMD involving cerebellar networks for executive functions, to date few studies have explored white matter connectivity by Diffusion Weighted Imaging (DWI).

Brain tractography has been widely applied in developmental age, both in neurodevelopmental disorders, as ADHD [33] and autism spectrum disorders [34–36], and in neurological disorders, as leukodystrophies, cerebral palsy or cerebellar diseases [37–42].

Examples exist of the possible double involvement of brain and muscle in neuromuscular diseases, demonstrated as white matter abnormalities by DWI tractography. In particular, alterations of white matter projection, association and commissural fibres have been described in myotonic dystrophy type 1 [43–45] and alterations of diffusion coefficient of white matter have been revealed for merosin-deficient congenital muscular dystrophy [46, 47].

In DMD patients, some evidence of microstructural differences in scalar measures (Fractional Anisotropy, FA, and Mean Diffusivity, MD) has been shown in the occipital regions [48] and in the splenium of corpus callosum, with a correlation with intellectual quotients [49]. Alterations in diffusion in the prefrontal cortex and hippocampus emerged also in *mdx* mice, the animal model of DMD [50].

The purposes of this explorative study are threefold:

1. to analyze the neuropsychological profile of a group of DMD and BMD boys without cognitive impairment with an assessment of their executive functions. Because of the variable expression of the protein in BMD, we might speculate that the BMD neuropsychological phenotype could be different compared to DMD;
2. to explore the structural connectivity in DMD, BMD and age-matched controls focusing on cortico-subcortical tracts which connect frontal cortex, basal ganglia and cerebellum via the thalamus. Mean FA values along each of the selected tracts were calculated, used as a measure of altered structural connectivity and compared among groups. We hypothesized that DMD boys had lower FA in the tracts compared to BMD and controls;

- to explore possible correlations between altered FA values and clinical measures of neuropsychological function in DMD and BMD groups, even if in a pilot study, in order to identify possible functional neuroimaging biomarkers of the neuropsychological profile for further investigations on wider sample. We hypothesized that reduced FA corresponded to higher functional impairment.

Materials and methods

Subjects

This study involved patients enrolled by IRCCS Stella Maris Foundation, University of Pisa, one of the Centers members of DMD Italian Network. In total, 15 subjects participated in the study: 5 DMD subjects, 5 BMD subjects and 5 age-matched typically developing (TD) boys.

Different reasons made recruiting difficult in DMD children: a) the rarity of the DMD disease; b) the difficulty for the patients families to move from long distances; c) the participation of many DMD patients in experimental trials which already involve several clinical monitoring; d) the refusal of patients to participate in a study involving a brain Magnetic Resonance Imaging (MRI) protocol which is not part of the routine follow up. Therefore, only 5 DMD boys (mean age: 10.1 years old; range: 7–13 years) were enrolled in the study, two coming from Bologna.

Because of the involvement of dystrophin in all dystrophinopathies, 5 BMD boys (mean age: 13.1 years old; range: 11–15 years) were recruited.

The inclusion criteria for the study subjects were the following: i) DMD and BMD boys with proven mutation in the dystrophin gene; ii) the availability of a neuropsychological evaluation assessing cognitive and neuropsychological functioning, according to the protocol already discussed [24]; iii) no cognitive impairment ($IQ < 70$) or any associated neuropsychiatric disorders (drug-resistant epilepsy, autism spectrum, attention deficit and hyperactivity) or any additional neurosensory deficits; iv) steroid treatment and/or other experimental drug stable for at least six months.

Moreover, 5 age-matched typically developing (TD) boys (mean age: 9.5 years old; range: 7–12 years) were enrolled among the children performing brain MRI for headache without other neurological signs and with normal brain MRI.

All the children were recruited only if they have not MRI contraindications and if they were able to collaborate to MRI exam, lying supine in the scanner for at least 30/40 minutes.

The study was approved by Tuscany Pediatric Ethics Committee and a specific informed consent form has been signed by all parents and subjects included in the study.

Methods

Neuropsychological assessment. Both the DMD and the BMD children were assessed using a neuropsychological assessment protocol including cognitive and executive functioning assessment, a simplified format compared to the one previously published [24]. Concerning cognitive evaluation, we performed Wechsler Intelligence Scale for children (WISC-IV) for Full Intellectual Quotients (FIQ) and Working Memory Index (WMI) for all the dystrophinopathy-subjects [51]. With regard to executive functions, Inhibition test of NEPSY-II [52] and Tower Of London (TOL) tests [53] were administered.

MRI acquisition. MRI data were acquired by using a 1.5T MRI scanner (1.5T GE HDx) at MRI Laboratory of IRCCS Stella Maris Foundation. The acquisition protocol consisted of: (1) Isotropic high-resolution T1-weighted sequence (3D BRAVO) with slice thickness = 1 mm,

Field of view (FOV) = 256 mm X 256 mm, matrix = 256 X 256; Time of repetition (TR)/ Time of echo (TE) = 450/5.18 ms, flip angle (fa) = 13°; (2) isotropic diffusion weighted sequence using a 2D single-shot EPI; including 30 non-collinear encoding directions with b value of 1000 and one additional volume without diffusion gradients (b₀), slice thickness = 3 mm; FOV = 240 mm X 240 mm; matrix = 80 x 80; TR/TE = 13000/115.8 ms.

MRI analysis. Brain tissue segmentation was performed using FreeSurfer based on 3D T1-weighted images as in [54]. FreeSurfer is used as pre-processing workflow for structural MRI data to perform volumetric segmentation and cortical reconstruction through 31 processing steps [55]. The processing steps included skull stripping, motion correction, removal of non-brain tissue, spherical surface registration, tissue segmentation and parcellation of the cortex into anatomical regions, obtaining white matter (WM), gray matter (GM), cerebrospinal fluid (CSF) and subcortical gray matter structures for each subject. FreeSurfer provides several descriptive features of mentioned structures as well, such as volume that was considered in our analysis.

DWI data were preprocessed to correct image artifacts caused by involuntary head motion, cardiac pulsation, and intensity inhomogeneities by using FSL tools (<https://fsl.fmrib.ox.ac.uk/fsl>). After preprocessing, a color-encoded track-density image was generated to support the identification of Regions of Interest (ROI) for tract reconstruction. Fiber tractography (MRtrix package) was performed using constrained spherical deconvolution (CSD) with a maximum number of streamlines of ten thousand, by using the iFOD2 algorithm that facilitates more accurate fiber reconstruction in heavily curved regions [56]. Anatomically-Constrained Tractography (ACT) option was applied by using the 5-types-tissue (5TT) segmented images to correct tractography and to increase anatomical plausibility of the reconstructed fibers based on prior information [57], discarding streamlines that are anatomically unfeasible.

Three tracts of interest were identified considering cortico-subcortical tracts, which connect frontal cortex, basal ganglia and cerebellum via the thalamus. In particular, Cortico-Ponto-Cerebellar tract (CPCT), Cerebellar-Thalamic Tract (CTT) and Superior Longitudinal Fasciculus (SLF) were selected in each hemisphere of all subjects. Cortico-Spinal Tract (CST) was identified as “control” tract, since it is not expected to be associated with the cognitive measure of interest. Mean FA value along the tracts were extracted.

A single rater, blinded to the participant status (DMD/BMD/TD), performed tractography, drawing manually appropriate seeding and inclusion ROIs for each tract.

Tracts were checked by two experienced raters on all subjects to verify trajectory and anatomic landmarks described in atlases of human WM and to check false-positive streamlines within the pathways.

The CPCT of left hemisphere was selected setting a seeding ROI in the right middle cerebellar peduncle and an inclusion ROI in the left posterior limb of the internal capsule, as previously [41]. An add inclusion ROI was positioned in a frontal WM area of left hemisphere [58, 59]. The CPCT of right hemisphere was obtained using the same procedure, selecting manually the homologous ROIs of the contralateral hemispheres.

To define the left CTT, a seed in the left superior cerebellar peduncle and a spherical inclusion ROI on the left thalamic WM were chosen [41]. For the right CCT, homologue ROIs were selected in the contralateral hemispheres.

As in Kamali et al. [59] to identify SLF, and for each hemisphere, the first ROI was placed over the green association bundles just superolateral to the cingulum on the color-encoded track-density at the most posterior part of the corpus callosum. The second ROI was placed over the fibers generated on the superolateral aspect of the cingulum at the coronal plane passing through the mid thalamus.

For the delineation of left CST, a spherical seeding ROI was placed in the left white matter in correspondence of the precentral gyrus, at the level of hand omega. An inclusion ROI was selected between the transverse pontine fibers and the middle cerebellar peduncle [60]. The CST of right hemisphere was obtained, drawing the homologous ROIs in the contralateral hemispheres.

The number of streamlines and FA was calculated for each tract examined; FA is an invariant scalar measure that describes the degree of diffusion anisotropy, reflecting fibre density, axonal diameter and myelination, and varies between 0 (equal diffusion in all directions) and 1 (highly directional diffusion).

To improve the specificity of quantitative DTI metrics (i.e. FA), and to take in account Partial Volume Effects (PVE), we calculate the tract volume, to use it as a covariate not-of-interest [61]. In fact, Vos et al. (2011) have shown that tract volume, orientation, and curvature are PVE-modulating factors that can affect the estimation of diffusion metrics when sampled along the tract. The tract volume, in particular, contributes to the explanation of the observed differences in DTI measures between populations. The volume of the tracts was obtained by: i) generating a binary mask starting from the tract; ii) calculating the number of voxels contained within the mask; iii) converting the number of voxels in mm^3 , multiplying it by the voxel volume (image resolution).

Statistical analysis. For each subject, a complete set of neuropsychological clinical measures was included in the analyses. The mean FIQ and WMI were considered in the normal range according to the Diagnostic Mental Index if the mean value was 100 and the SD was 15. The Inhibition test of NEPSY-II was measured as mean 10 and SD 3. TOL results were expressed with T Score (mean = 50; SD = 10).

For all subjects, the number of streamlines and FA were calculated for each reconstructed tract (CTT, CPCT, SLF and CST). Within each subject, a paired sample t test was used to compare mean FA and number of streamlines between the right and left side of each tract.

A general linear model was used to determine the difference among groups for FA and clinical measures, and post hoc pair-wise comparisons were performed. Effect size was calculated for significant differences, by using the Hedges's g parameter, that corrects the standard Cohen's d effect size for potential bias due to small samples [62].

For tracts that showed altered connectivity among groups, ANCOVA analyses were performed using age and the tract volume as covariate. Moreover, for these tracts, the relationship with clinical measures was explored.

Statistical analyses were performed by using SPSS, Version 2.0 (IBM, Armonk, New York).

To correct for multiple comparisons, we used the method of Benjamini and Hochberg [63], which recalculates the level of significance in terms of p-values, limiting the false discovery rate (FDR) to 5%.

Results

Neuropsychological profile in DMD and BMD children

The neuropsychological profile of the small group of BMD children resulted almost comparable to that already described in DMD [24]. In fact, regarding intellectual functioning, a major impairment in WMI compared to the general intellectual functioning was observed both for DMD (mean score = 86.20; SD = 11.54) and for BMD children (mean score = 88; SD = 3.67). Worse performances were confirmed in the Switching task of the Inhibition test (NEPSY-II) (mean score for DMD boys = 5.60; SD = 2.70; mean score for BMD boys = 6; SD = 1.87) compared to the other tasks of that test. In TOL test, the BMD boys showed worse impairments in

Table 1. Intellectual and executive functioning in DMD and BMD cohorts.

	DMD	BMD
	5 subjects	5 subjects
	Mean (SD)	Mean (SD)
<i>WISC-IV Index</i>		
Working Memory Index	86.2 (11.5)	88 (3.7)
Full Intellectual Quotient	100.2 (7.0)	95.2 (14.2)
<i>Inhibition test (NEPSY-II) (standard score)</i>		
Denomination Total	7.2 (1.6)*	6.4 (0.6)*
Inhibition Total	7.6 (2.1)	7.4 (1.3)
Switching Total	5.6 (2.7)	6.0 (1.9)
<i>TOL test (T score)</i>		
Total corrected	45.4 (11.0)	36.6 (13.4)

* = statistically significant differences

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“total correct answers” than DMD (mean score for DMD = 45.40; SD = 11.01; mean score for BMD = 36.60; SD = 13.35).

No significant difference emerged between the BMD and DMD groups enrolled in the study in neuropsychological measures, except for the Denomination task of the Inhibition test (NEPSY-II), where BMD obtained worse performances ($p = 0.014$).

Table 1 shows mean scores and SDs of cognitive and executive functioning tests in BMD and DMD groups.

Fiber tracts reconstruction

All fiber tracts were successfully extracted on each hemisphere of each subject (Fig 1).

The number of streamlines was >10 in each of the examined tracts. No statistically significant differences for number of streamlines of all the examined tracts were found between the left and the right side within each group of children (complete dataset in S1 Table), neither between the three groups of subjects.

Moreover, within each group of subjects, no significant differences are found between the mean FA of the right and the left representations of all examined tracts.

Mean FA resulted tendentially always lower in DMD compared to both BMD and TD groups for all the examined tracts, with the exception of the right SLF (FA-R-SLF), which showed the same values of FA in DMD and TD.

A statistically significant difference emerged between DMD and BMD in the mean FA value of the right CTT (FA-R-CTT) ($p = 0.002$), with a “huge” effect size ($g = 3.3$). Moreover, a statistically significant difference emerged between DMD and TD in the mean FA-R-CTT ($p = 0.0015$) and in the FA-R-CPCT ($p = 0.008$). The effect size was “huge” in both cases ($g = 2.8$ for FA-R-CTT and $g = 1.8$ for FA-R-CPCT).

No statistically significant differences emerged among groups for FA values of bilateral SLF and bilateral CST. The results are shown in Table 2.

Adding age and tract volume as covariates in the statistical analyses, the significances remain valid, according to S2 Table.

Interestingly, with regard to FA-R-CTT, a possibly “gradient effect” was observed, with mean FA of BMD children resulting in the middle between mean FA of DMD and mean FA of controls (Fig 2).

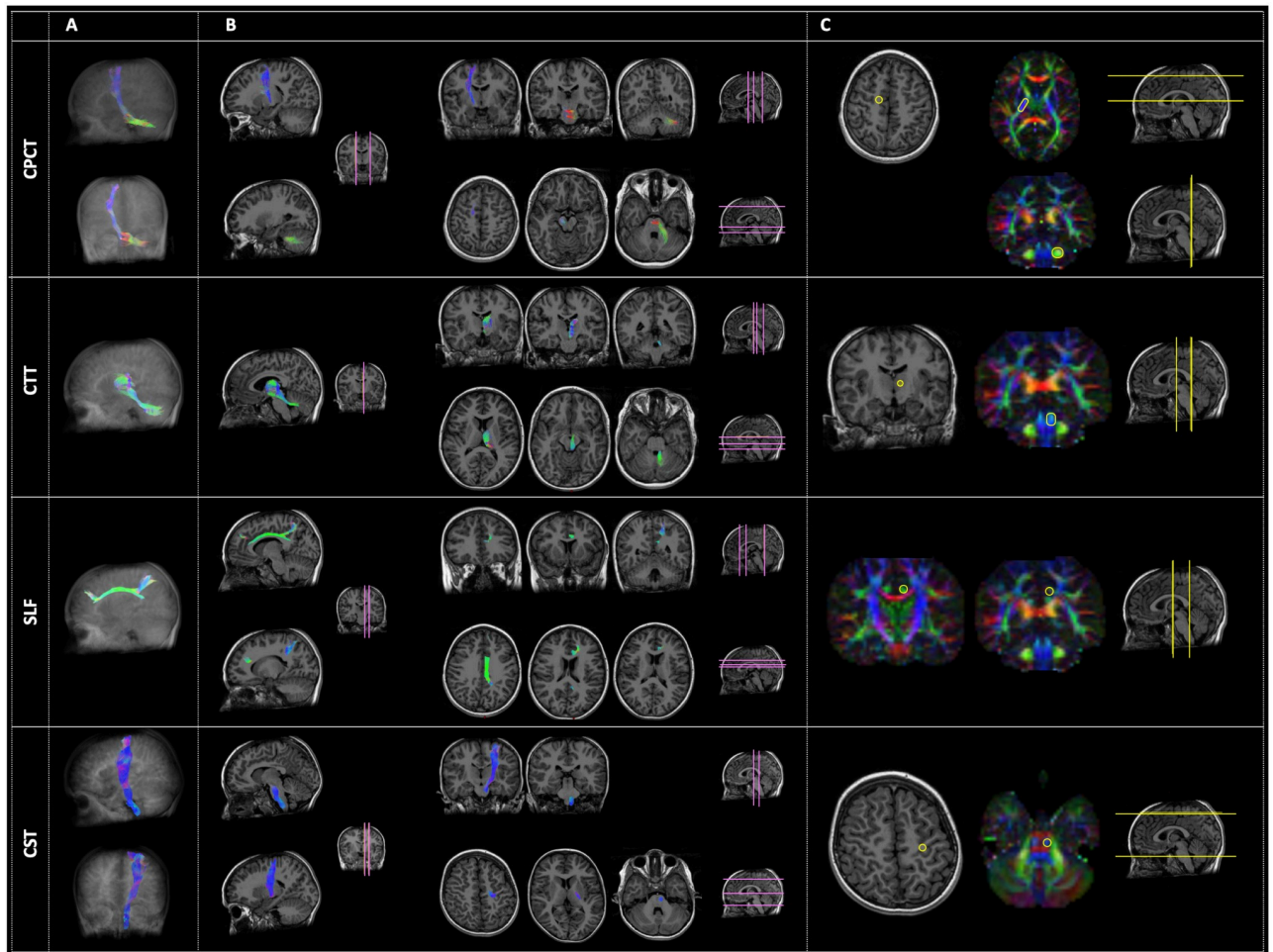


Fig 1. Examples of the four tracts of interest: From the top, the Cortico-Ponto-Cerebellar Tract (CPCT), the Cerebellar-Thalamic Tract (CTT), the Superior Longitudinal Fasciculus (SLF) and the Cortico-Spinal Tract (CST). Panel A represents the three-dimensional reconstruction of each tract in a glass brain. In panel B, the pathway of tractography is depicted in some representative slices with different orientation: Sagittal (right side of the panel), coronal (left side of the panel, upper rows), and axial (left side of the panel, lower rows). The location of these slices is reported in pink color on a reference brain. Panel C reports the ROIs (yellow circles) used for the tractography of each tract, both in anatomical images or in the diffusion-encoded-color (DEC) maps. Analogously to panel B, the location of these images is represented in yellow color on a reference brain. The order of representation of images corresponds to the Right-Left (R-L) direction for sagittal images, to the Anterior-Posterior (A-P) direction for coronal images, and to the Superior-Inferior (S-I) direction for axial ones. The color code of tractography pathways and DEC maps is the standard red-green-blue (RGB) code for diffusion: red for R-L direction, blue for S-I, and green for A-P.

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Correlations between FA fiber tracts and neuropsychological profile

Concerning DMD, significant correlations emerged between FA-R-CTT and FIQ ($p = 0.044$; $\rho_s = 0.821$), Denomination Total ($p = 0.044$; $\rho_s = 0.821$) and Inhibition Total ($p = 0.019$; $\rho_s = 0.900$) of the Inhibition test (NEPSY-II) (Fig 3). No other significant correlation emerged between FA-R-CTT and neuropsychological tests.

We also found a slight negative correlation between FA-R-CPCT and WMI ($p = 0.027$; $\rho_s = -0.872$), while, regarding BMD, a significant correlation emerged between FA-R-CPCT and WMI ($p = 0.007$; $\rho_s = 0.949$) (Fig 4). No other significant correlation emerged between FA-R-CPCT nor FA-R-CTT and neuropsychological tests in BMD.

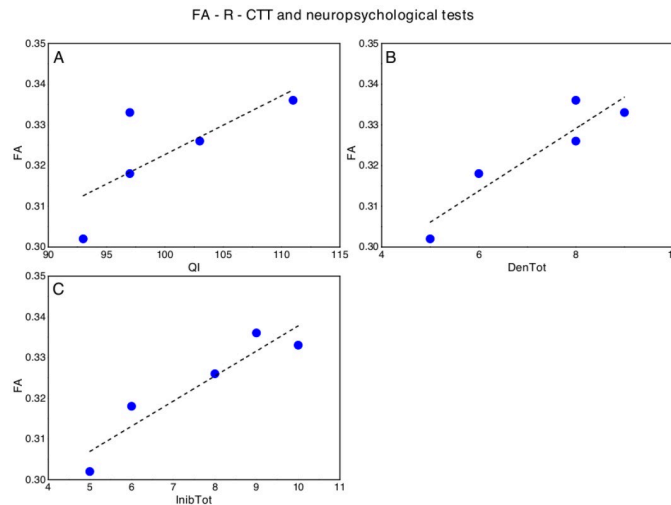


Fig 3. Significant correlations between FA-R-CTT and neuropsychological tests in DMD, in particular with respect to (A) Full Intellectual Quotient (FIQ, WISC-IV), (B) Denomination Total (DenTot, Inhibition test-NEPSY II) and (C) Inhibition Total (InhibTot, Inhibition test-NEPSY II). The dotted lines represent the linear fit of the data with a function $y = a + b \cdot x$. A) FA-R-CTT versus FIQ: $a = 0.18 \pm 0.08$, $b = 0.001 \pm 0.0008$, Pearson's $R = 0.74$, Adj $R^2 = 0.40$; B) FA-R-CTT versus DenTot: $a = 0.27 \pm 0.01$, $b = 0.008 \pm 0.0002$, Pearson's $R = 0.93$, Adj $R^2 = 0.81$; C) FA-R-CTT versus InhibTot: $a = 0.28 \pm 0.01$, $b = 0.006 \pm 0.0001$, Pearson's $R = 0.94$, Adj $R^2 = 0.84$.

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Discussion

In this study, we explored the neuropsychological profile in a group of DMD and BMD boys without intellectual impairment and we investigate possible correlations with the obtained DWI data comparing DMD, BMD and controls. We explored WM microstructure in tracts that are known or supposed to be part of executive functions networks by investigating FA, as the measure of disrupted connectivity. In particular, we explored two cerebellar tracts, separately on the right and left sides: the CTT, which is the main efferent pathway from cerebellum, and the CPCT, which is the major input of the cerebellum from the cerebral cortex, focusing specifically on the fibers originating from the frontal area. Moreover, the SLF, involved in fronto-striatal connections and in executive functions, as already described in literature [64], has been included, separately on the right and left sides. The CST was included as a control tract not expected to be associated with cognitive functions.

Our neuropsychological results suggest that the BMD neuropsychological phenotype may be similar to DMD one, due to the involvement of specific dystrophin isoforms in the brain. The variable expression of the dystrophin in BMD can account for the variability of neurocognitive profile. Our BMD sample showed a major impairment of working memory compared to the general intellectual functioning and a failure in executive functions like problem solving, inhibition and shifting abilities, as already described in literature for DMD [7, 11, 24].

Regarding DWI data, the overall mean FA resulted lower in DMD children than in the other groups of subjects for the examined tracts, suggesting a possible alteration in WM microstructural integrity. More specifically, the cerebellar connectivity seemed to be more compromised, compared to SLF and CST in the DMD group.

Even if with few and explorative contributions, possible altered WM connectivity has been already described in DMD boys [48, 49]. We suppose that the lack of cerebral dystrophin protein in DMD children, already during neural development, may be responsible for reduced fiber coherence, and altered myelination and axonal density in the explored tracts. This speculation may be supported also by the results obtained in BMD children, that variably express

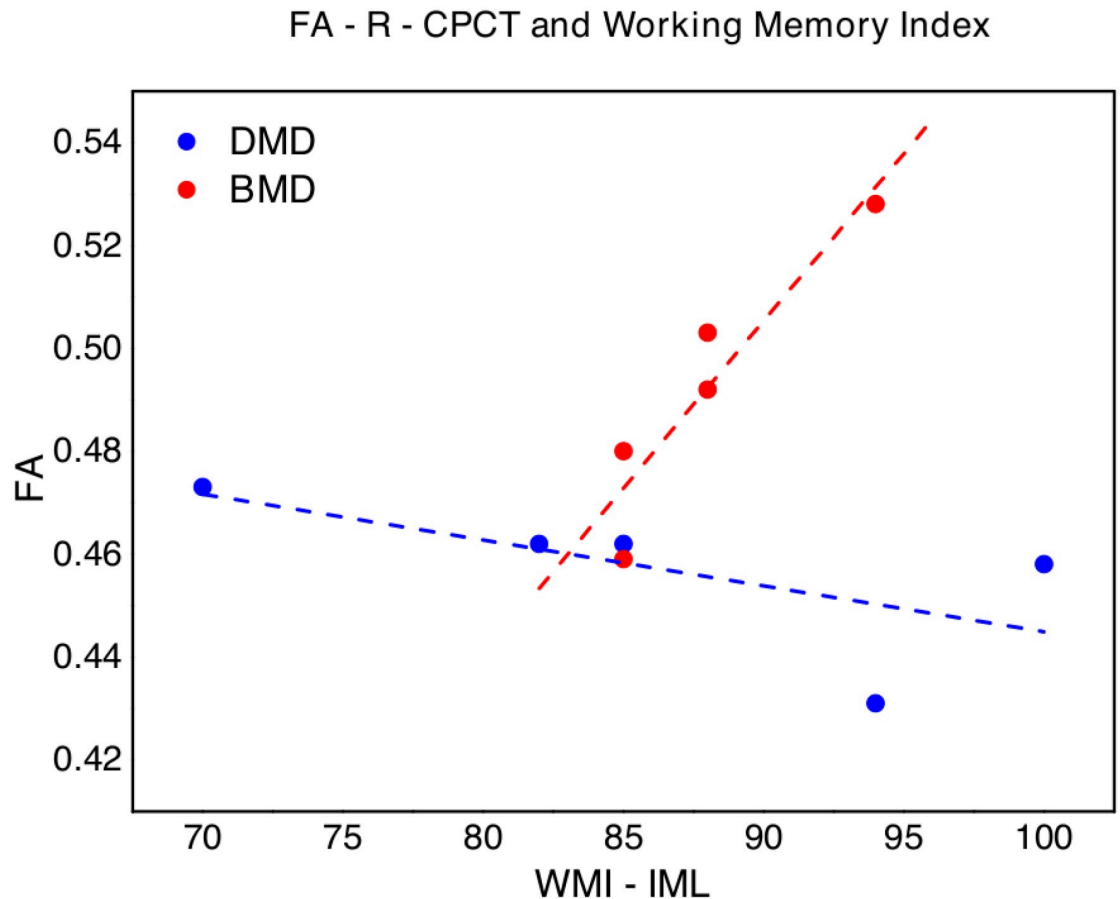


Fig 4. FA of the right corticopontocerebellar tract (FA-R-CPCT) values versus Working Memory Index (WMI, WISC-IV) in DMD (blue points) and BMD (red points). The dotted lines (blue for DMD and red for BMD respectively) represent the linear fit of the data with a function $y = a + b \cdot x$. DMD: $a = 0.53 \pm 0.05$, $b = -0.001 \pm 0.0005$, Pearson's $R = -0.66$, $\text{Adj } R^2 = 0.24$; BMD: -0.08 ± 0.13 , $b = 0.007 \pm 0.0001$, Pearson's $R = 0.93$, $\text{Adj } R^2 = 0.81$.

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dystrophin protein, who showed an intermediate mean FA in some of the examined tracts, with a sort of “gradient effect” between DMD and controls, at least in tracts that might be responsible for specific cerebral symptoms (i.e. neuropsychological dysfunction). A greater effect regarding differences between DMD and BMD groups seems to be observed in CTT than in CPCT, but the results must be confirmed in wider samples, considering also the slight difference in age between the two groups.

In fact, some studies have shown a development and maturation of executive functioning, using neuropsychological or neuroimaging approaches, both in normotypical children and adolescents [65–67] and in clinical populations, such as children with autism spectrum disorders and ADHD, in which emerged that not until 7 to 9 years of age switch flexibility begin operating [68, 69].

Our recent results about neuropsychological profile confirmed a similar neuropsychological maturation also in DMD [70].

In order to explore possible functional correlates of altered connectivity for specific tracts, we thus studied the relationship between mean FA values in the cerebellar tracts that showed reduced FA and neuropsychological measures. Indeed, we demonstrated a less widespread involvement in BMD compared to DMD. In detail, regarding DMD children, we observed

significantly lower FA-R-CTT in boys with lower FIQ and a major impairment in inhibition abilities.

FA-R-CPCT seems to show a different behavior with respect to the working memory abilities for DMD and BMD boys. This result may be due to the small sample size and must be verified in a next wider analysis, but could also underline unexpected mechanisms of maladaptive neural plasticity for DMD, which may produce abilities globally slightly reduced in DMD compared to BMD children.

Overall, these findings support the hypothesis of the involvement of a cerebellar-thalamo-cortical loop for the neuropsychological profile of DMD, as the CTT and the CPCT are involved in the network and the related brain structures are known to be implied in executive functions.

It has long been known that frontal cortex, in particular prefrontal area, plays a central role in global aspect of general intelligence [71] and executive functions [69] across the lifespan; thanks to connections between the prefrontal cortex and other brain regions, the neural substrates of executive functions include also the parietal cortex, the anterior cingulate cortex, and subcortical regions as the striatum and the cerebellum [72]. Moreover, the superior cerebellar peduncle, involved in CTT, and the posterior limb of internal capsule, involved in CPCT, are thought to be key components of the circuit [73]. Recent data have also emphasized a role of the thalamus, in particular mediodorsal nucleus, in cognition and executive functions because of its significant interconnectivity with prefrontal cortex [74, 75].

More in detail, a bilateral contribution of cerebellum in DMD seem to be suggested by our findings. In fact, both CTT which originates from the right cerebellum and CPCT fibers which, originating from the right frontal cortex, project to the pontine nuclei and cross the midline, thus terminating in the contralateral half of cerebellum [76], resulted more damaged in DMD than in BMD and controls. In literature, a bilateral contribution of cerebellum in executive functions is reported. For example, a cross cerebral-cerebellar circuitry with left prefrontal cortex predominantly involved and strong right cerebellum activation has been shown for verbal working memory [77]. However, a fMRI study has demonstrated a bilateral cerebellar activation for working memory paradigms, while other executive function tasks showed converging activation in lobules VI, Crus I and left VIIB of cerebellum [78]. Moreover, left and right cerebellum involvement in switching attention has been demonstrated [79]. To verify if differences in FA values were related to smaller brain size of DMD, we performed post hoc ANCOVA analyses to compare brain size between groups, using age as covariate. We found that DMD has significant smaller volumes of the cerebellar Gray Matter (Cereb-GM) both at a global level ($p = 0.006$), both for each hemisphere separately (L Cereb-GM $p = 0.006$, R Cereb-GM $p = 0.007$), than TD, confirming the involvement of cerebellum in the dystrophiaopathies.

The great limitation of the study is due to the small sample size that limits the possibility to correlate the neuroimaging data with the neuropsychological findings in a robust manner and suggest caveats in the interpretation of results. The enlargement of the sample could help us to better establish the role of cerebellar connectivity in neuropsychological profile for dystrophiaopathies.

The slight unexpected negative correlation between FA-R-CPCT and WMI in DMD children needs to be further explored, also in consideration of the general difficulty of tracking CPCT, that is a long tract that sharply turns between cerebellum and cerebrum, and consequently of calculating the FA along it. This is evident also in the high values of standard deviation for the number of streamlines of CPCT, in particular in DMD, suggesting higher variability in fiber tracking among subjects within the same group.

Moreover, to increase the robustness and reliability of the data, it could be useful to replicate the study with state-of-art DWI acquisition techniques. In particular, 3T MR scanners, with new generation gradients, could allow acquisitions at higher spatial resolutions, in a multi-shell approach, in reasonable times of acquisition, allowing to implement multiple compartment models of diffusion, maximizing the accuracy of diffusion measurements and reducing PVE contributions.

Furthermore, DMD and BMD children enrolled in the study underwent only a brief neuropsychological assessment, and a more detailed evaluation could help not only to better describe the role of WM abnormalities but also, for BMD children, to define their neuropsychological profile.

In conclusion, to our knowledge, this is the first study to specifically explore the role of the cerebellar-thalamo-cortical network in DMD boys using DWI-based connectivity measures. Our results suggest that altered WM connectivity and reduced fibre organization in cerebellar tracts, probably due to the lack of dystrophin in the brain, may render less efficient some neuropsychological functions in children affected by dystrophinopathies. The findings lead us to identify possible functional neuroimaging biomarkers for the neuropsychological profile of DMD without intellectual disability.

Supporting information

S1 Table. Mean number of streamlines in the examined tracts in DMD, BMD and TD.
(DOCX)

S2 Table. Statistical significance (p-value) in the analyses without and with covariates.
(DOCX)

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References

1. Flanigan KM. Duchenne and Becker muscular dystrophies. *Neurol Clin.* 2014; 32(3):671–viii. <https://doi.org/10.1016/j.ncl.2014.05.002> PMID: 25037084
2. Muntoni F, Torelli S, Ferlini A. Dystrophin and mutations: one gene, several proteins, multiple phenotypes. *Lancet Neurol* 2003 Dec; 2(12):731–40. [https://doi.org/10.1016/s1474-4422\(03\)00585-4](https://doi.org/10.1016/s1474-4422(03)00585-4) PMID: 14636778
3. Ogasawara A. Downward shift in IQ in persons with Duchenne muscular dystrophy compared to those with spinal muscular atrophy. *Am. J. Ment. Retard.* 1989; 93, 544–547. PMID: 2706122
4. Billard C, Gillet P, Barthez M, Hommet C, Bertrand P. Reading ability and processing in Duchenne muscular dystrophy and spinal muscular atrophy. *Dev Med Child Neurol.* 1998 Jan; 40(1):12–20. <https://doi.org/10.1111/j.1469-8749.1998.tb15351.x> PMID: 9459212
5. Moizard MP, Billard C, Toutain A, Berret F, Marmin N, Moraine C. Are Dp71 and Dp140 brain dystrophin isoforms related to cognitive impairment in Duchenne muscular dystrophy? *Am J Med Genet* 1998; 80: 32–41. PMID: 9800909
6. Felisari G, Martinelli Boneschi F, Bardoni A, Sironi M, Comi GP, Robotti M, et al. Loss of Dp140 dystrophin and intellectual impairment in Duchenne dystrophy. *Neurology.* 2000 Aug 22; 55(4):559–64. <https://doi.org/10.1212/wnl.55.4.559> PMID: 10953192
7. Cotton S, Voudouris NJ, Greenwood KM. Intelligence and Duchenne muscular dystrophy: full-scale, verbal, and performance intelligence quotients. *Dev Med Child Neurol.* 2001 Jul; 43(7):497–501. <https://doi.org/10.1017/s0012162201000913> PMID: 11463183
8. Marini A, Lorusso ML, D'Angelo MG, Civati F, Turconi AC, Fabbro F, et al. Evaluation of narrative abilities in patients suffering from Duchenne Muscular Dystrophy. *Brain Lang.* 2007 Jul; 102(1):1–12. <https://doi.org/10.1016/j.bandl.2007.02.003> PMID: 17428527
9. Daoud F, Angeard N, Demerre B, Martie I, Benyaou R, Leturcq F, et al. Analysis of Dp71 contribution in the severity of mental retardation through comparison of Duchenne and Becker patients differing by mutation consequences on Dp71 expression. *Hum Mol Genet.* 2009; 18, 3779–94. <https://doi.org/10.1093/hmg/ddp320> PMID: 19602481
10. Taylor PJ, Betts GA, Maroulis S, Gilissen C, Pedersen RL, Mowat DR, et al. Dystrophin gene mutation location and the risk of cognitive impairment in Duchenne muscular dystrophy. *PLoS One* 2010; 5, e8803. <https://doi.org/10.1371/journal.pone.0008803> PMID: 20098710
11. Mento G, Tarantino V, Bisiacchi PS. The neuropsychological profile of infantile Duchenne muscular dystrophy. *Clin Neuropsychol.* 2011 Nov; 25(8):1359–77. <https://doi.org/10.1080/13854046.2011.617782> PMID: 21999586
12. Lorusso ML, Civati F, Molteni M, Turconi AC, Bresolin N, D'Angelo MG. Specific profiles of neurocognitive and reading functions in a sample of 42 Italian boys with Duchenne Muscular Dystrophy. *Child Neuropsychol.* 2013; 19(4):350–69. <https://doi.org/10.1080/09297049.2012.660912> PMID: 22385039
13. Astrea G, Pecini C, Gasperini F, Brisca G, Scutifero M, Bruno C, et al. Reading impairment in Duchenne muscular dystrophy: A pilot study to investigate similarities and differences with developmental dyslexia. *Res Dev Disabil.* 2015 Oct-Nov; 45–46:168–77. <https://doi.org/10.1016/j.ridd.2015.07.025> PMID: 26255617
14. Young HK, Barton BA, Waisbren S, Portales Dale L, Ryan MM, Webster RI, et al. Cognitive and psychological profile of males with Becker muscular dystrophy. *J Child Neurol.* 2008 Feb; 23(2):155–62. <https://doi.org/10.1177/0883073807307975> PMID: 18056690
15. Banihani R, Baskin B, Halliday W, Kobayashi J, Kawamura A, McAdam L, et al. A Novel Mutation in DMD (c.10797+5G>A) Causes Becker Muscular Dystrophy Associated with Intellectual Disability *J Dev Behav Pediatr.* 2016 Apr; 37(3):239–44. <https://doi.org/10.1097/DBP.0000000000000262> PMID: 26836830
16. Cuisset JM, Rivier F. Manifestations centrales des dystrophinopathies [Central manifestations of dystrophinopathies]. *Arch Pediatr.* 2015; 22(12 Suppl 1):12S58–12S62. [https://doi.org/10.1016/S0929-693X\(16\)30010-0](https://doi.org/10.1016/S0929-693X(16)30010-0) PMID: 26773588
17. Huard J, Tremblay JP. Localization of dystrophin in the Purkinje cells of normal mice. *Neuroscience Letters* 1992; 137, 105–8. [https://doi.org/10.1016/0304-3940\(92\)90309-u](https://doi.org/10.1016/0304-3940(92)90309-u) PMID: 1625810
18. Tinsley JM, Blake DJ, Pearce M, Knight AE, Kendrick-Jones J, Davies KE. Dystrophin and related proteins. *Curr Opin Genet Dev.* 1993; 3(3):484–90. [https://doi.org/10.1016/0959-437x\(93\)90124-8](https://doi.org/10.1016/0959-437x(93)90124-8) PMID: 8353425
19. Lidov HG, Selig S, Kunkel LM. Dp140: a novel 140 kDa CNS transcript from the dystrophin locus. *Hum Mol Genet.* 1995; 4(3):329–35. <https://doi.org/10.1093/hmg/4.3.329> PMID: 7795584
20. Anderson JL, Head SI, Rae C, Morley JW. Brain function in Duchenne muscular dystrophy. *Brain* 2002; 125(Pt 1):4–13. <https://doi.org/10.1093/brain/awf012> PMID: 11834588

21. Lidov HGW, Byers TJ, Watkins SC, Kunkel LM. Localization of dystrophin to postsynaptic regions of central nervous system cortical neurons. *Nature*. 1990 Dec 20–27; 348(6303):725–8. <https://doi.org/10.1038/348725a0> PMID: 2259381
22. Daoud F, Candelario-Martínez A, Billard JM, Avital A, Khelifaoui M, Rozenvald Y, et al. Role of mental retardation-associated dystrophin-gene product Dp71 in excitatory synapse organization, synaptic plasticity and behavioral functions. *PLoS One* 2008; 4, e6574. <https://doi.org/10.1371/journal.pone.0006574> PMID: 19649270
23. Cyrulnik SE, Hinton VJ. Duchenne muscular dystrophy: a cerebellar disorder? *Neurosci Biobehav Rev*. 2008; 32(3):486–96. <https://doi.org/10.1016/j.neubiorev.2007.09.001> PMID: 18022230
24. Battini R, Chieffo D, Bulgheroni S, Piccini G, Pecini C, Lucibello S, et al. Cognitive profile in Duchenne muscular dystrophy boys without intellectual disability: The role of executive functions. *Neuromuscul Disord*. 2018 Feb; 28(2):122–128. <https://doi.org/10.1016/j.nmd.2017.11.018> PMID: 29305139
25. Vicari S, Piccini G, Mercuri E, Battini R, Chieffo D, Bulgheroni S, et al. Implicit learning deficit in children with Duchenne muscular dystrophy: Evidence for a cerebellar cognitive impairment? *PLoS One*. 2018 Jan 6; 13(1):e0191164. <https://doi.org/10.1371/journal.pone.0191164> PMID: 29338029
26. Heyder K, Suchan B, Daum I. Cortico-subcortical contributions to executive control. *Acta Psychol (Amst)*. 2004 Feb-Mar; 115(2–3):271–89. <https://doi.org/10.1016/j.actpsy.2003.12.010> PMID: 14962404
27. Bresolin N, Castelli E, Comi GP, Felisari G, Bardoni A, Perani D, et al. Cognitive impairment in Duchenne muscular dystrophy. *Neuromuscul Disord* 1994; 4:359–369. [https://doi.org/10.1016/0960-8966\(94\)90072-8](https://doi.org/10.1016/0960-8966(94)90072-8) PMID: 7981593
28. Lee JS, Pfund Z, Juhász C, Behen ME, Muzik O, Chugani DC, et al. Altered regional brain glucose metabolism in Duchenne muscular dystrophy: a pet study. *Muscle Nerve*. 2002 Oct; 26(4):506–12. <https://doi.org/10.1002/mus.10238> PMID: 12362416
29. Rae C, Scott RB, Thompson CH, Dixon RM, Dumughn I, Kemp GJ, et al. Brain biochemistry in Duchenne muscular dystrophy: a H1 magnetic resonance and neuropsychological study. *J Neurol Sci*. 1998 Oct 8; 160(2):148–57. [https://doi.org/10.1016/s0022-510x\(98\)00190-7](https://doi.org/10.1016/s0022-510x(98)00190-7) PMID: 9849797
30. Kreis R, Wingeier K, Vermathen P, Giger E, Joncourt F, Zwygart K, et al. Brain metabolite composition in relation to cognitive function and dystrophin mutations in boys with Duchenne muscular dystrophy. *NMR Biomed*. 2011 Apr; 24(3):253–62. <https://doi.org/10.1002/nbm.1582> PMID: 21404337
31. Lv SY, Zou QH, Cui JL, Zhao N, Hu J, Long XY, et al. Decreased gray matter concentration and local synchronization of spontaneous activity in the motor cortex in Duchenne muscular dystrophy. *Am. J. Neuroradiol* 2011; 32, 2196–200. <https://doi.org/10.3174/ajnr.A2718> PMID: 21960496
32. Doorenweerd N, Dumas EM, Ghariq E, Schmid S, Straathof CS, Roest AA, et al. Decreased cerebral perfusion in Duchenne muscular dystrophy patients. *Neuromuscul Disord*. 2017 Jan; 27(1):29–37. <https://doi.org/10.1016/j.nmd.2016.10.005> PMID: 27927595
33. Sudre G, Szekely E, Sharp W, Kasperek S, Shaw P. Multimodal Mapping of the Brain's Functional Connectivity and the Adult Outcome of Attention Deficit Hyperactivity Disorder. *Proc Natl Acad Sci U S A*. 2017 Oct 31 114 (44), 11787–11792. <https://doi.org/10.1073/pnas.1705229114> PMID: 29078281
34. Conti E, Calderoni S, Gaglianese A, Pannek K, Mazzotti S, Rose S, et al. Lateralization of Brain Networks and Clinical Severity in Toddlers With Autism Spectrum Disorder: A HARDI Diffusion MRI Study. *Autism Res*. 2016 Mar; 9(3):382–92. <https://doi.org/10.1002/aur.1533> PMID: 26280255
35. Conti E, Mitra J, Calderoni S, Pannek K, Shen KK, Pagnozzi A, et al. Network Over-Connectivity Differentiates Autism Spectrum Disorder From Other Developmental Disorders in Toddlers: A Diffusion MRI Study. *Hum Brain Mapp*. 2017 May; 38(5):2333–2344. <https://doi.org/10.1002/hbm.23520> PMID: 28094463
36. Qin B, Wang L, Zhang Y, Cai J, Chen J, Li T. Enhanced Topological Network Efficiency in Preschool Autism Spectrum Disorder: A Diffusion Tensor Imaging Study. *Front Psychiatry*. 2018 Jun 27; 9:278. <https://doi.org/10.3389/fpsy.2018.00278> PMID: 29997534
37. Poretti A, Boltshauser E, Loenneker T, Valente EM, Brancati F, Il'yasov K, et al. Diffusion tensor imaging in Joubert syndrome. *AJNR Am J Neuroradiol*. 2007 Nov-Dec; 28(10):1929–33. <https://doi.org/10.3174/ajnr.A0703> PMID: 17898198
38. Escolar ML, Poe MD, Smith JK, Gilmore JH, Kurtzberg J, Lin W, et al. Diffusion tensor imaging detects abnormalities in the corticospinal tracts of neonates with infantile Krabbe disease. *AJNR Am J Neuroradiol*. 2009 May; 30(5):1017–21. <https://doi.org/10.3174/ajnr.A1476> PMID: 19386732
39. Poretti A, Meoded A, Bunge M, Fatemi A, Barrette P, Huisman TA, et al. Novel diffusion tensor imaging findings in Krabbe disease. *Eur J Paediatr Neurol*. 2014 Mar; 18(2):150–6. <https://doi.org/10.1016/j.ejpn.2013.09.008> PMID: 24149099

40. Pannek K, Boyd RN, Fiori S, Guzzetta A, Rose SE. Assessment of the structural brain network reveals altered connectivity in children with unilateral cerebral palsy due to periventricular white matter lesions. *Neuroimage Clin*. 2014 Jun 4; 5:84–92. <https://doi.org/10.1016/j.nicl.2014.05.018> PMID: 25003031
41. Fiori S, Poretti A, Pannek K, Del Punta R, Pasquariello R, Tosetti M, et al. Diffusion Tractography Biomarkers of Pediatric Cerebellar Hypoplasia/Atrophy: Preliminary Results Using Constrained Spherical Deconvolution. *AJNR Am J Neuroradiol*. 2016 May; 37(5):917–23. <https://doi.org/10.3174/ajnr.A4607> PMID: 26659337
42. Kimiskidis VK, Papaliagkas V, Papagiannopoulos S, Zafeiriou D, Kazis D, Tsatsali-Foroglou E, et al. Investigation of the motor system in two siblings with Canavan's disease: a combined transcranial magnetic stimulation (TMS)—diffusion tensor imaging (DTI) study. *Metab Brain Dis*. 2017 Apr; 32(2):307–310. <https://doi.org/10.1007/s11011-017-9955-x> PMID: 28130616
43. Okkersen K, Monckton DG, Le N, Tuladhar AM, Raaphorst J, van Engelen BGM. Brain Imaging in Myotonic Dystrophy Type 1: A Systematic Review. *Neurology*. 2017; 89(9):960–969. <https://doi.org/10.1212/WNL.0000000000004300> PMID: 28768849
44. van Dorst M, Okkersen K, Kessels RPC, Meijer FJA, Monckton DG, van Engelen BGM, et al. Structural White Matter Networks in Myotonic Dystrophy Type 1. 2019; 21:101615.
45. Labayru G, Diez I, Sepulcre J, Fernández E, Zulaica M, Cortés JM, et al. Regional Brain Atrophy in Gray and White Matter Is Associated With Cognitive Impairment in Myotonic Dystrophy Type 1. *Neuroimage Clin*. 2019; 24:102078. <https://doi.org/10.1016/j.nicl.2019.102078> PMID: 31795042
46. Sijens PE, Fock JM, Meiners LC, Potze JH, Irwan R, Oudkerk M. MR Spectroscopy and Diffusion Tensor Imaging of the Brain in Congenital Muscular Dystrophy With Merosin Deficiency: Metabolite Level Decreases, Fractional Anisotropy Decreases, and Apparent Diffusion Coefficient Increases in the White Matter. *Brain Dev*. 2007; 29(5):317–321. <https://doi.org/10.1016/j.braindev.2006.10.004> PMID: 17113260
47. Ip JJK, Hui PKT, Chau MT, Lam WWM. Merosin-deficient Congenital Muscular Dystrophy (MDCMD): A Case Report With MRI, MRS and DTI Findings. *J Radiol Case Rep*. 2012; 6(8):1–7. <https://doi.org/10.3941/jrcr.v6i8.997> PMID: 23365711
48. Doorenweerd N, Straathof CS, Dumas EM, Spitali P, Ginjaar IB, Wokke BH, et al. Reduced cerebral gray matter and altered white matter in boys with Duchenne muscular dystrophy. *Ann Neurol*. 2014 Sep; 76(3):403–11. <https://doi.org/10.1002/ana.24222> PMID: 25043804
49. Fu Y, Dong Y, Zhang C, Sun Y, Zhang S, Mu X, et al. Diffusion tensor imaging study in Duchenne muscular dystrophy. *Ann Transl Med*. 2016 Mar; 4(6):109. <https://doi.org/10.21037/atm.2016.03.19> PMID: 27127762
50. Xu S, Shi D, Pratt SJP, Zhu W, Marshall A, Lovering RM. Abnormalities in brain structure and biochemistry associated with mdx mice measured by in vivo MRI and high resolution localized (1)H MRS. *Neuromuscul Disord*. 2015 Oct; 25(10):764–72. <https://doi.org/10.1016/j.nmd.2015.07.003> PMID: 26236031
51. Orsini, A., Pezzuti, L., Picone, L. WISC-IV. Contributo alla taratura italiana. Firenze: Giunti O.S. Organizzazioni Speciali; 2012.
52. Urgesi C, Campanella F, Fabbro F. NEPSY-II. Second edition. Contributo alla taratura italiana. Firenze: Giunti O.S. Organizzazioni Speciali; 2011.
53. Sannio Fancello G., Vio C., Cianchetti C. TOL Torre di Londra: test di valutazione delle funzioni esecutive (pianificazione e problem solving). Erickson; 2006.
54. Conti E, Retico A, Palumbo L, Spera G, Bosco P, Biagi L, et al. Autism Spectrum Disorder and Childhood Apraxia of Speech: Early Language-Related Hallmarks across Structural MRI Study. *J Pers Med*. 2020 Dec 12; 10(4):275. <https://doi.org/10.3390/jpm10040275> PMID: 33322765
55. Fischl B. Freesurfer. *Neuroimage*. 2012; 62(2): 774–781. <https://doi.org/10.1016/j.neuroimage.2012.01.021> PMID: 22248573
56. Tournier JD, Calamante F, Connelly A. Improved probabilistic streamlines tractography by 2 nd order integration over fibre orientation distributions. *Proceedings of the international society for magnetic resonance in medicine ISMRM*; 2010;1670.
57. Horbuegger M, Loewe K, Kaufmann J, Wagner M, Schippling S, Pawlitzki M, et al. Anatomically constrained tractography facilitates biologically plausible fiber reconstruction of the optic radiation in multiple sclerosis. *Neuroimage Clin*. 2019; 22:101740. <https://doi.org/10.1016/j.nicl.2019.101740> PMID: 30870736
58. Kamali A, Kramer LA, Frye RE, Butler IJ, Hasan KM. Diffusion tensor tractography of the human brain cortico-ponto-cerebellar pathways: a quantitative preliminary study. *J Magn Reson Imaging* 2010; 32:809–17. <https://doi.org/10.1002/jmri.22330> PMID: 20882611
59. Kamali A, Flanders AE, Brody J, Hunter JV, Hasan KM. Tracing Superior Longitudinal Fasciculus Connectivity in the Human Brain using High Resolution Diffusion Tensor Tractography. *Brain Struct Funct* 2014 Jan; 219(1):269–81. <https://doi.org/10.1007/s00429-012-0498-y> PMID: 23288254

60. Liégeois F, Tournier JD, Pigdon L, Connelly A, Morgan AT. Corticobulbar tract changes as predictors of dysarthria in childhood brain injury. *Neurology*. 2013 Mar 5; 80(10):926–32. <https://doi.org/10.1212/WNL.0b013e3182840c6d> PMID: 23390172
61. Vos SB, Jones DK, Viergever MA, Leemans A. Partial volume effect as a hidden covariate in DTI analyses. *NeuroImage*. 2011 Apr 15; 55(4): 1566–1576. <https://doi.org/10.1016/j.neuroimage.2011.01.048> PMID: 21262366
62. Hedges LV, Olkin I. *Statistical methods for meta-analysis*. San Diego, CA: Academic Press; 1985.
63. Benjamini Y, Hochberg Y. Controlling the false discovery rate—a practical and powerful approach to multiple testing. *J R Stat Soc Series B Stat Methodol* 1995; 57:289–300.
64. Catani M, Dell’acqua F, Bizzi A, Forkel SJ, Williams SC, Simmons A, et al. Beyond Cortical Localization in Clinico-Anatomical Correlation. *Cortex*. Nov-Dec 2012; 48(10):1262–87. <https://doi.org/10.1016/j.cortex.2012.07.001> PMID: 22995574
65. Igazság B, Demetrovics Z, Cserjési R. The developmental trajectory of executive functions and their stress sensitivity in adolescence. *Psychiatr Hung*. 2019; 34(3):300–310. PMID: 31570661
66. Baron Nelson M, O’Neil SH, Wisnowski JL, Hart D, Sawardekar S, Rauh V, et al. Maturation of Brain Microstructure and Metabolism Associates with Increased Capacity for Self-Regulation during the Transition from Childhood to Adolescence. *J Neurosci*. 2019 Oct 16; 39(42):8362–8375. <https://doi.org/10.1523/JNEUROSCI.2422-18.2019> PMID: 31444243
67. Wierenga LM, Bos MGN, van Rossenberg F, Crone EA. Sex Effects on Development of Brain Structure and Executive Functions: Greater Variance than Mean Effects. *J Cogn Neurosci*. 2019 May; 31(5):730–753. https://doi.org/10.1162/jocn_a_01375 PMID: 30726177
68. Richardson C, Anderson M, Reid CL, Fox AM. Development of inhibition and switching: A longitudinal study of the maturation of interference suppression and reversal processes during childhood. *Dev Cogn Neurosci*. 2018 Nov; 34:92–100. <https://doi.org/10.1016/j.dcn.2018.03.002> PMID: 30114552
69. Diamond A. Executive functions. *Annu Rev Psychol*. 2013; 64:135–68. <https://doi.org/10.1146/annurev-psych-113011-143750> PMID: 23020641
70. Battini R, Lenzi S, Lucibello S, Chieffo D, Moriconi F, Cristofani P, et al. Longitudinal data of neuropsychological profile in a cohort of Duchenne muscular dystrophy boys without cognitive impairment. *Neuromuscul Disord*. 2021 Feb 4: <https://doi.org/10.1016/j.nmd.2021.01.011> Epub ahead of print.
71. Barbey AK, Colom R, Grafman J. Dorsolateral prefrontal contributions to human intelligence. *Neuropsychologia*. 2013 Jun; 51(7):1361–9. <https://doi.org/10.1016/j.neuropsychologia.2012.05.017> PMID: 22634247
72. Fiske A, Holmboe K. Neural substrates of early executive function development. *Dev Rev*. 2019; 52:42–62. <https://doi.org/10.1016/j.dr.2019.100866> PMID: 31417205
73. Schmahmann JD, Pandya DN. Prefrontal cortex projections to the basilar pons in rhesus monkey: implications for the cerebellar contribution to higher function. *Neurosci Lett*. 1995; 199(3):175–178. [https://doi.org/10.1016/0304-3940\(95\)12056-a](https://doi.org/10.1016/0304-3940(95)12056-a) PMID: 8577391
74. Ouhaz Z, Fleming H, Mitchell AS. Cognitive Functions and Neurodevelopmental Disorders Involving the Prefrontal Cortex and Mediodorsal Thalamus. *Front Neurosci*. 2018 Feb 6; 12:33. <https://doi.org/10.3389/fnins.2018.00033> PMID: 29467603
75. Wolff M, Vann SD. The Cognitive Thalamus as a Gateway to Mental Representations. *J Neurosci*. 2019; 39(1):3–14. <https://doi.org/10.1523/JNEUROSCI.0479-18.2018> PMID: 30389839
76. Nolte J. *The human brain. An introduction to its functional anatomy*, 5th edition: St. Louis, MO: Mosby; 2002. p 650.
77. Emch M, von Bastian CC, Koch K. Neural Correlates of Verbal Working Memory: An fMRI Meta-Analysis. *Front Hum Neurosci*. 2019 Jun 12; 13:180. <https://doi.org/10.3389/fnhum.2019.00180> PMID: 31244625
78. Stoodley CJ, Valera EM, Schmahmann JD. Functional topography of the cerebellum for motor and cognitive tasks: an fMRI study. *NeuroImage*. 2012; 59(2):1560–1570. <https://doi.org/10.1016/j.neuroimage.2011.08.065> PMID: 21907811
79. Berninger VW, Richards TL, Abbott RD. Brain and Behavioral Assessment of Executive Functions for Self-Regulating Levels of Language in Reading Brain. *J Nat Sci*. 2017; 3(11):e464. PMID: 29104930