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RESEARCH ARTICLE

Effects of flower thrips (Thysanoptera: Thripidae) on nutritional quality of banana (Zingiberales: Musaceae) buds

Deyi Yu^{1,2©}*, Peng Huang^{1,2©}, Yong Chen^{1,2}, Yongwen Lin^{1,3}, Komivi Senyo Akutse⁴, Yanyang Lan⁵, Hui Wei^{1,2}*

- 1 Institute of Plant Protection, Fujian Academy of Agricultural Sciences, Fuzhou, China, 2 Fujian Key Laboratory for Monitoring and Integrated Management of Crop Pests, Fuzhou, China, 3 Plant Protection College, Fujian Agriculture and Forestry University, Fuzhou, China, 4 International Centre of Insect Physiology and Ecology, Nairobi, Kenya, 5 Research and Development Centre of Zhangzhou National Agricultural Science and Technology Zone, Zhangzhou, China
- These authors contributed equally to this work.
- * yudy_2004@126.com (DY); weihui@faas.cn (HW)

Abstract

The abundance of banana flower thrips (*Thrips hawaiiensis* Morgan) in a banana (*Musa acuminata* Colla "Williams" cultivar) plantation was investigated using yellow sticky traps (29.70 cm × 21.00 cm) in 2015. Banana flower thrips occurred throughout the year with monthly variation, and the maximum occurrence was observed in October and November during the bud burst (73.80 ± 6.32 adults/trap) and young fruit (70.06 ± 5.69 adults/trap) periods. The damage rates were as follows: interior flowers >3rd-layer flowers > 2nd-layer flowers > 1st-layer flowers > young fruits. This result indicates that thrips migrated to lower bracts, young fruits, and other flower buds as bracts gradually opened. Results also showed that the reducing sugar, vitamin C, protein and ash contents in thrips-damaged flowers were all significantly lower than those in undamaged flowers, while there was no significant difference between damaged and undamaged young banana fruit. Our results indicated that the abundances of banana flower thrips were closely associated with the growing stage of banana. Thrips mainly infested flower buds and caused a reduction in nutrients for the host plant, especially the reducing sugar and vitamin C contents, which reduced the nutritional quality of banana fruits and the quality of flower bud by-products of banana.

Introduction

Banana (*Musa acuminata*) is the world's most important fruit crop and one of the top 10 crops by production [1]. It is widely grown in the tropics and subtropics, where it acts as an important dietary component, both raw (as a dessert fruit) and cooked (often as the major source of carbohydrates) [2]. Banana produces a large number of flower buds, which have been developed for edible and pharmaceutical functions due to their abundance of nutrients and medicinal contents [3, 4]. In recent years, banana flower thrips (*Thrips hawaiiensis* Morgan) that damage flower buds have become major pests of banana crops [4–6]. The thrips cause dark-



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brown and black bumps on the surface of flower buds and young fruit through their rasping-sucking and oviposition behaviors [6]. Theoretically, *T. hawaiiensis* not only impacts flower nutritional quality, edibility, and pharmaceutical value but also affects the development and utilization of banana bioproducts. It is necessary to establish reasonable management methods to control banana flower thrips [7].

The strategy for controlling banana flower thrips in the field is the use of chemical insecticides that not only kill targets but also result in "3R" (residue, resistance and resurgence) [8] and phytotoxicity problems [9, 10]. Some researchers tried sticky traps and attraction based on the susceptibility of thrips to particular colors or volatile compounds [11, 12]. However, this approach is more likely to monitor pests' occurrence than reduce their populations. Using resistant host species is one of the most effective approaches to control pests and improve crop quality, as it reduces the infestation by pests at the source [13, 14]. For using this method, it is necessary to investigate the characteristics of damage caused by target pests to the host plant and the effect of pest feeding on the nutritional quality of the host plant [12, 15]. Thrips intake nutrition for growth, reproduction, and physiological metabolism by piercing the host plant and sucking up protein, carbohydrates, lipids, vitamins, water, inorganic salts, and other nutrients. This process not only changes the nutrient contents of the host plants but also causes compensation, eventually affecting the resistance and nutritional quality of the host plant [16, 17].

A previous study showed that the damage characteristics of banana flower thrips and its preference for "Williams" banana, "red" banana, and "pink" banana flower buds and young fruits[18]. However, the influences of flower thrips on the biology of banana flowers and young fruits are still unknown. Here, the abundance of banana flower thrips was observed in a "Williams" banana plantation in Tianbao Town, Zhangzhou City. Before the buds were cut off, six nutritional indicators, including protein, reducing sugar (easily absorbed by insects), crude fat, ash (consisting of inorganic salts and oxides), vitamin C (improves plant resistance), and water were compared among different flower layers and young fruits. The effects of banana flower thrips on the nutritional quality of banana flower buds were clarified to provide technical support for the management of thrips and banana flower buds.

Materials and methods

Crop culture

The *M. acuminata* "Williams" cultivar was selected for this study. Banana crops were planted in the field in an area of approximately 2.3 hm² located in Tianbao Town, Zhangzhou City (23°34′53″ N, 117°32′14″ E). Banana plants were grown in a deep-furrow border-check system (4.0–4.5 m in border-check width, 0.5–0.7 m in furrow width, 0.4–0.6 m in depth). The spacing of individual plants was 2.0–2.5 m, and the height of plants varied from 2.0–2.5 m.

Classification of developmental stages

The developmental stages of banana in Tianbao Town were classified as seedling stage (fifty days from planting in the field), vigorous growth stage (sixty days), flower bud burst stage (sixty days), formation of fruits stage (forty days), fruit development stage (sixty days) and harvest stage (thirty days) (Fig 1).

Sampling method

Twenty 40-day-old seedling plants were selected for tissue culture, randomly sampled in a "Z" pattern and labeled for observation at 10-day intervals. The damage rate of plants was recorded in all the plant tissues (stems, leaves, flower buds, and fruits) during the whole growth period.



Population dynamics of thrips

The twenty selected banana plants mentioned above were investigated, and the population dynamics of banana flower thrips were analyzed as described by [19]. Bamboo poles (1.7 m high) used to hang yellow sticky traps (29.7 cm \times 21 cm) were stuck into the earth, 50 cm away from the sampled plants. The sticky traps were 1.5 m above the ground. The number of thrips on each sticky trap was recorded every 10 days using a 40 \times magnifier. The sticky traps were replaced every 10 days. This experiment was repeated 3 times.

Preference of thrips for different tissues

Ten thirty-day-old thrips-free inflorescences (3 layers of young fruit) were covered with insect net (50 mesh) bags, and one hundred banana flower thrips were then transferred into each bag for twenty days. One fruit and flower bud from each layer of inflorescence on the selected banana trees were marked and collected in black transparent bags. All samples were transported to the lab immediately to assess the damage rate. All the dark spots (S1 Fig) on the surface of each sample (young fruit and flower bud) were recorded under a 20X binocular microscope (SZ760, Chongqing Optec Instrument Co., Ltd., Chongqing, China). The severity of damage to fruits and flowers was divided into four levels: L0, undamaged; L1, < 10 spots; L2, < 30 spots; and L3, > 30 spots. Thrips-free inflorescences were used as a control, and this experiment was repeated five times.

Nutritional quality of flower buds and young fruit

The protein, reducing sugar, crude fat, ash, vitamin C, and water contents in each sample were measured in the same layers for samples with different damage rates. The L0 samples were

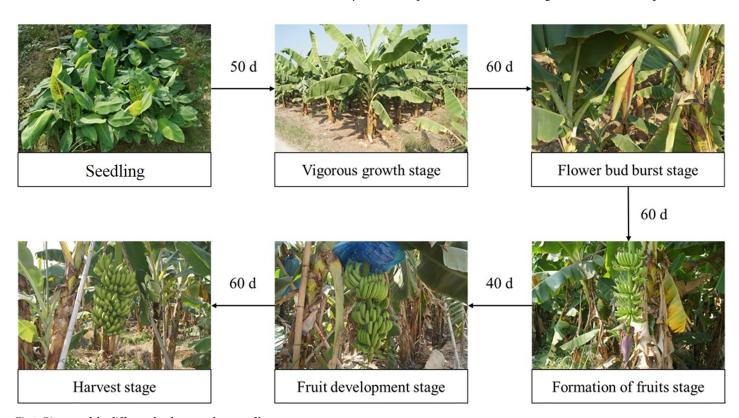


Fig 1. Pictures of the different developmental stages of banana.

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regarded as the control (CK) and the experiment was repeat for 5 times. The protein content was determined with the method described by Kjeldah [20], reducing sugar content was determined using direct titration [21], crude fat content was extracted using Soxhlet extraction [22], ash content was determined with the method described by Dinuzzo et al. [23], vitamin C content was determined using 2,6-dichloroindophenol titration [24], and water content was determined using direct drying [25].

Statistical analysis

The mean values of preference damage rate and nutritional quality were compared by Tukey's HSD test ($\alpha = 0.05$) following analysis of variance using SPSS 20.0 (Microsoft, USA).

Results

Abundance of banana flower thrips

Thrips appeared sporadically in young leaves during the seedling period, while stems, leaves, and other tissues did not show any sign of damage during this growth stage. Thrips started rasping-sucking and oviposition behavior on the flower buds from the bud burst to young fruit stage. This damage became gradually worse before the flower clusters were cut off. Thrips invaded tissues before buds opened and then migrated to the lower bracts, young fruits, and other flower buds as the bracts gradually opened. The abundance of thrips decreased rapidly when plants were covered with fruit bags during the fruit developmental stage. No additional damage was observed at the end of the harvest. Thrips occurred throughout the year, and the population abundance fluctuated widely from July to December (Fig 2). The abundance of thrips in the bud burst and young fruit stages, which occur in October-November, was higher than that in the other stages ($F_{2,5} = 121.78$, P = 0.0001). The abundance observed on seedlings and during the vigorous growth stage (June-August) was significantly lower than that of the other stages ($F_{2.5} = 28.37$, P = 0.017). The abundance rapidly increased to 73.80 adults/trap in October-November (flower bud burst and young fruit stages) and was significantly higher than in any other month ($F_{2,5} = 18.52$, P = 0.012). The abundance sharply declined (0.80–7.60 adults/trap) in December (fruit development stage) and then maintained a smooth downward trend in the following months (January-May).

Damage rates in different tissues

The damage rates were ranked as follows: interior flowers $> 3^{\rm rd}$ -layer flowers $> 2^{\rm nd}$ -layer flowers $> 1^{\rm st}$ -layer flowers $> 2^{\rm nd}$ -layer flowers $> 1^{\rm st}$ -layer flowers $> 2^{\rm nd}$ -layer flowers $> 2^{\rm nd}$ -layer flowers $> 2^{\rm nd}$ -layer flowers (87.56%) than in the other 3 tissues ($F_{4,4} = 44.75$, P = 0.0001). A significant difference in damage rate was also found between the first-layer flowers (23.69%) and $2^{\rm nd}$ -layer flowers (45.13%) ($F_{4,4} = 13.57$, P = 0.024). The lowest damage rate (5.17%) was observed in the young fruit. There was no significant difference in damage rate between the $3^{\rm rd}$ -layer flowers and interior flowers ($F_{4,4} = 0.89$, $F_{4,4} = 0.89$).

For the severity of damage, we also found the following rank: interior flowers $> 3^{\rm rd}$ -layer flowers $> 2^{\rm nd}$ -layer flowers $> 1^{\rm st}$ -layer flowers > 9 young fruits (Fig 3-II). The severity of damage to interior flowers (38.69 spots) was significantly higher than that to the other tissues ($F_{4,4} = 54.76$, P = 0.003), while the severity of damage to young fruits (2.58%) was significantly lower than that to the other tissues ($F_{4,4} = 33.89$, P = 0.011).



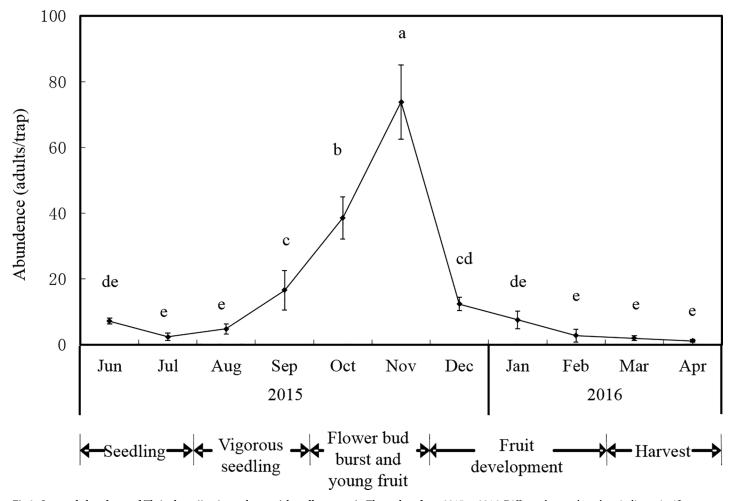


Fig 2. Seasonal abundance of *Thrips hawaiiensis* caught on sticky yellow traps in Zhangzhou from 2015 to 2016. Different latters above bars indicate significant differences at P < 0.05.

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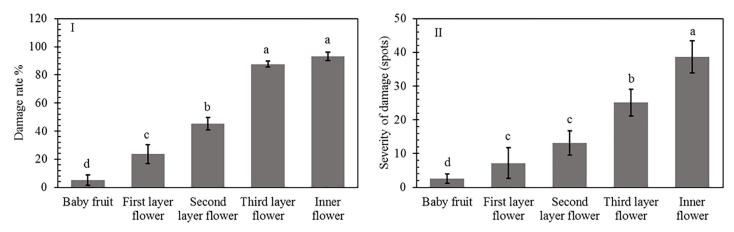


Fig 3. Damage rates in different tissues attacked by banana flower thrips. I, damage rate of banana flowers and young fruits caused by thrips; II, severity of damage to banana flowers and young fruits caused by thrips. Different letters above bars indicate significant differences at P < 0.05.

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Table 1. Effects of banana flower thrips on the nutritional quality of different flower layers.

| Nutritional content | 1st-layer flowers | 2 nd -layer flowers | 3 rd -layer flowers | Interior flowers | Average | Control (CK) |
|--|-------------------|--------------------------------|--------------------------------|------------------|-------------|--------------|
| Protein content (g/100 g) | 13.10±0.02d | 15.00±0.04c | 13.30±0.09d | 15.60±0.17b | 14.25±0.07e | 17.33±0.48a |
| Reducing sugar content (g/100 g) | 4.00±0.07d | 6.30±0.13c | 6.80±0.14b | 6.90±0.05b | 6.00±0.05e | 7.24±0.30a |
| Crude fat content (g/100 g) | 6.00±0.13bc | 6.20±0.07b | 6.30±0.12b | 6.30±0.03b | 6.20±0.07b | 7.50±0.37a |
| Ash content (inorganic salt) (g/100 g) | 12.20±0.30d | 12.30±0.20cd | 12.60±0.40c | 14.10±0.10a | 12.80±0.20b | 13.10±0.35b |
| Vitamin C content (mg/100 g) | 9.60±0.27e | 13.80±0.18d | 18.80±0.27a | 16.40±0.18c | 14.65±0.07f | 17.20±0.60b |
| Water content (g/100 g) | 90.32±0.47a | 90.57±1.50a | 91.04±0.14a | 91.61±0.48a | 90.89±0.58a | 90.84±0.43a |

Note: Means \pm SE followed by different alphabets represent significant differences at P < 0.05.

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Effects of thrips on nutritional quality

As shown in Table 1, the protein, reducing sugar, crude fat and vitamin C contents in thrips-damaged flower were 14.25%, 6.00%, 6.20% and 17.20%, all significantly lower than in undamaged flower (control)($F_{1,4} = 22.17$, P = 0.010; $F_{1,4} = 10.41$, P = 0.036; $F_{1,4} = 17.99$, P = 0.021; $F_{1,4} = 93.41$, P = 0.001). Thereafter, the protein content of the 2nd flower layer rebounded due to host plant compensation, but this did not counter the loss caused by thrips damage. Hence, the protein content of the 1st flower layer showed a significant decrease and then recovered to the level of the 3rd flower layer ($F_{1,4} = 18.65$, P = 0.018). Similar effects of plant compensation were observed for vitamin C in the 3rd flower layer. The vitamin C content even significantly higher than that of the control ($F_{1,4} = 38.41$, P = 0.002). A continuous, significant decline in vitamin C was observed in the 1st and 2nd flower layers ($F_{1,4} = 187.35$, P = 0.0001; $F_{1,4} = 105.33$, P = 0.0001). There was no significant difference in water content among the different flower layers, although it decreased in the range of 90.30 to 91.61 g/100 g ($F_{4,16} = 1.25$, P > 0.05).

The effect of thrip damage on the nutritional quality of young fruit was showed in Table 2. The content of reducing sugar and vitamin C in thrips-damaged young fruit were 4.91% and 9.60, both significantly lower than in undamaged one (control) ($F_{1,4} = 9.82$, P = 0.030; $F_{1,4} = 53.87$, P = 0.0001). The changes in protein, crude fat, ash, and water contents were not significant (P > 0.05). Protein and water contents decreased slightly. Conversely, crude fat and ash contents increased slightly and fluctuated in the range of 0.90% to 1.64% ($F_{1,4} = 1.98$, P = 0.121; $F_{1,4} = 1.13$, P = 0.213).

Discussion

Understanding the host preference of target pests, their abundance and their influence on nutritional quality are prerequisites for the application of resistant cultivars, which are one of the most effective strategies of pest control [26, 27]. Thrips damaged flower buds by rasping-sucking and ovipositing throughout the year. The order of damage rates was as follows:

Table 2. Effects of banana flower thrips on the nutritional quality of young fruits.

| Nutritional content | Damaged | Undamaged (control) | P |
|--|-------------------|---------------------|--------|
| Protein content (g/100 g) | $12.80 \pm 0.02a$ | 13.01 ± 0.03a | 0.1508 |
| Reducing sugar content (g/100 g) | $4.91 \pm 0.02b$ | $5.31 \pm 0.05a$ | 0.0308 |
| Crude fat content (g/100 g) | $6.69 \pm 0.01a$ | $6.63 \pm 0.03a$ | 0.1004 |
| Ash content (inorganic salt) (g/100 g) | 19.18 ± 0.03a | 18.89 ± 0.02a | 0.3203 |
| Vitamin C content (mg/100 g) | $9.60 \pm 0.02b$ | 12.51 ± 0.04a | 0.0001 |
| Water content (g/100 g) | 92.88 ± 0.28a | 94.12 ± 0.21a | 0.0740 |

Note: Means \pm SE followed by different alphabets represent significant differences at P < 0.05.

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interior flowers $> 3^{\rm rd}$ -layer flowers $> 2^{\rm nd}$ -layer flowers $> 1^{\rm st}$ -layer flowers > young fruit. Thrips abundance was closely related to banana growth stage and varied widely each month. The highest abundance was observed during the bud burst and young fruit stages (October-November). This finding indicated that flower buds are a defining characteristic of determination materials used for breeding resistant cultivars.

Plants have evolved a series of complicated defense mechanisms against insect attacks. Plants supply nutrients, including protein, carbohydrates, lipids, vitamins, water, and inorganic salts, that are necessary to maintain insect life, but they are also induced to produce a series of physiological and biochemical reactions to compensate for the damage caused by insects accessing these resources [28, 29]. Many researchers have focused on the interspecific relationship between plants and herbivores, such as the photophysiological responses of alfalfa Medicago sativa to thrips damage (Odontothrips) [30], the effects of rasping-sucking by thrips (Gynaikothrips uzeli) on the chemical constitution of surface wax and amino acid content in Ficus benjamina [31], the relationship between soybean aphid (Aphis glycines) damage and physiological indexes of soybean (Glycine max) leaves [32], and the response of secondary metabolism of plane tree (Platanus acerifolia) leaves to tree bug (Corythucha ciliata) damage [33]. However, there are few studies that identify the resistance and nutritional changes of banana attacked by banana flower thrips. In our study, the nutritional contents (protein, reducing sugar, crude fat, ash, vitamin C, and water) were compared between damaged and undamaged flowers and young fruits. The results showed that all indexes fluctuated with damage level, except for water content. Plant compensation appeared in the 2nd and 3rd flower layers with higher protein and vitamin C contents but did not equal the loss caused by thrips damage. Overall, the nutritional values decreased with increasing damage, and the same results were found in previous studies [31–34]. There were significant decreases in reducing sugar and vitamin C contents in young fruit but not in the other nutritional contents. These results not only confirmed the complicated and subtle interspecific relationship between banana and thrips but also provided evidence to quantitatively evaluate the benefits and costs to buds that have been attacked by thrips.

Our evaluation of the nutritional quality of buds also showed the benefits and costs to physiological indexes caused by thrips attacks. This could also explain how plants and herbivorous insects coevolved, but it is not clear how plant metabolism, such as ultramicrostructural tissues, secondary substances (phenolic compounds, terpenoids), and enzyme activities, could change. Therefore, the effects of thrips attacks on morphological structures, secondary substances, and enzyme activities should be included in future studies to clarify the function of flower buds and establish suitable management methods for thrips.

In conclusion, the degree of damage to banana plants depended on *T. hawaiiensis* abundance. Thrips mainly invaded flower buds and caused poor nutritional quality of fruits and their bud by-products, particularly in reducing sugar and vitamin C contents.

Supporting information

S1 Fig. Damage symptoms of banana flower thrips against banana flowers and baby fruit. The tiny black points in the red circles were damage symptoms of banana flower thrips. (DOCX)

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Author Contributions

Conceptualization: Deyi Yu, Hui Wei.

Data curation: Deyi Yu, Peng Huang, Yong Chen, Yongwen Lin.

Formal analysis: Deyi Yu, Peng Huang, Yong Chen, Yongwen Lin, Hui Wei.

Funding acquisition: Deyi Yu, Peng Huang.

Investigation: Deyi Yu, Yanyang Lan.

Methodology: Deyi Yu, Peng Huang, Yanyang Lan.

Project administration: Deyi Yu.

Supervision: Hui Wei.

Writing - original draft: Deyi Yu, Peng Huang, Yong Chen.

Writing – review & editing: Yongwen Lin, Komivi Senyo Akutse, Hui Wei.

References

- 1. Hu W, Zuo J, Hou X, Yan Y, Wei Y, Liu J, et al. The auxin response factor gene family in banana: genome-wide identification and expression analyses during development, ripening, and abiotic stress. Frontiers in Plant Science. 2015; 6:742. https://doi.org/10.3389/fpls.2015.00742 PMID: 26442055
- Sreedharan S, Shekhawat UKS, Ganapathi TR. Transgenic banana plants overexpressing a native plasma membrane aquaporin MusaPIP1;2 display high tolerance levels to different abiotic stresses. Plant Biotechnology Journal. 2013; 11(8):942–52. https://doi.org/10.1111/pbi.12086 PMID: 23745761
- 3. Zaffari GR, Kerbauy GB, Kraus JE, Romano EC. Hormonal and histological studies related to in vitro banana bud formation. Plant Cell Tissue & Organ Culture. 2000; 63(3):187–92.
- Jamuna JB, Nandini CD. Feeding of banana flower and pseudostem to diabetic rats results in modulation of renal GLUTs, TGFβ, PKC and extracellular matrix components. Nutrition Metabolism & Cardiovascular Diseases Nmcd. 2014; 24(6):623–31.
- Roditakis E, Mound LA, Roditakis NE. First record in crete of Hercinothrips femoralis in greenhouse banana plantations. Phytoparasitica. 2006; 34(5):488–90.
- Lu H, Zhong Y, Liu K, Liang X, Peng S. The taxis of the banana flower thrips *Thrips hawaiiensis* to different colors and field trapping effect of sticky cards. Plant Protection. 2011; 55(12):21–30.
- Murai T. Development and reproductive capacity of *Thrips hawaiiensis* (Thysanoptera: Thripidae) and its potential as a major pest. Bulletin of Entomological Research. 2001; 91(3):193–8. PMID: 11415473
- Boyden S. The environment and human health. Medical Journal of Australia. 1972; 1(24):1229–34.
 PMID: 5038902
- Peterson HG, Boutin C, Martin PA, Freemark KE, Ruecker NJ, Moody MJ. Aquatic phyto-toxicity of 23
 pesticides applied at expected environmental concentrations. Aquatic Toxicology. 1994; 28(3–4):275
 92.
- Harms HH. In-vitro systems for studying phytotoxicity and metabolic fate of pesticides and xenobiotics in plants. Pest Management Science. 2010; 35(3):277–81.
- Loomans AJM, Lenteren JCV. Biological control of thrips pests: a review on thrips parasitoids. Developments in Plant Pathology. 1995; 14(1995):244–53.
- Harman JA, Mao CX, Morse JG. Selection of colour of sticky trap for monitoring adult bean thrips, Caliothrips fasciatus (Thysanoptera: Thripidae). Pest Management Science. 2007; 63(2):210–6. https://doi.org/10.1002/ps.1322 PMID: 17125151
- Gatehouse JA. Biotechnological prospects for engineering insect-resistant plants. Plant Physiology. 2008; 146(3):881–7. https://doi.org/10.1104/pp.107.111096 PMID: 18316644
- 14. Russell GE. Resistant plants. Nature. 1974; 250(5463):272-.
- Chen Y, Ruberson JR, Ni X. Influence of host plant nitrogen fertilization on hemolymph protein profiles of herbivore *Spodoptera exigua* and development of its endoparasitoid *Cotesia marginiventris*. Biological Control. 2014; 70(1):9–16.
- **16.** Haile FJ, Higley LG. Changes in soybean gas-exchange after moisture stress and spider mite injury. Environmental Entomology. 2003; 32(3):433–40.



- Zangerl AR, Hamilton JG, Miller TJ, Crofts AR, Oxborough K, Berenbaum MR, et al. Impact of folivory on photosynthesis is greater than the sum of its holes. Proceedings of the National Academy of Sciences of the United States of America. 2002; 99(2):1088–91. https://doi.org/10.1073/pnas.022647099 PMID: 11792866
- Yu D, Hu S, Huang P, Chen H, Yao J. Characteristic damages caused by and preference on three varieties of bananas of pest insect, *Thrips hamaiiensis*. Fujian Journal of Agricultural Sciences. 2015; 30 (10):997–1000.
- Pearsall IA, Myers JH. Population dynamics of western flower thrips (Thysanoptera: Thripidae) in nectarine orchards in British Columbia. Journal of Economic Entomology. 2000; 93(2):264–75. PMID: 10826171
- Bradstreet RB, Chem A. Kjeldahl method for organic nitrogen. Analytical Chemistry. 1965; 26(1):169– 234
- Zhao K, Xue PJ, Gu GY. Study on determination of reducing sugar content using 3, 5-dinitrosalicylic acid method. Food Science. 2008; 71(25):36–7.
- Castro LD, Priegocapote F. Soxhlet extraction: Past and present panacea. Journal of Chromatography A. 2010; 1217(16):2383–9. https://doi.org/10.1016/j.chroma.2009.11.027 PMID: 19945707
- Dinuzzo P, Gravitis V, Toop A, Watt J, Zastawny H. In-stream determination of solids weight fraction and ash content of coal in flotation circuit slurries of a coal preparation plant. Nuclear Geophysics. 1987; 1(4):329–43.
- **24.** Wu W, Sun Z, Zhang W. Simple and rapid determination of Vitamin C in vegetables and fruits by a commercial electrochemical reader. Food Analytical Methods. 2016; 9(11):3187–92.
- Sezgintürk M, Dinçkaya E. Direct determination of sulfite in food samples by a biosensor based on plant tissue homogenate. Talanta. 2005; 65(4):998–1002. https://doi.org/10.1016/j.talanta.2004.08.037
 PMID: 18969901
- Nangju D, Flinn JC, Singh SR. Control of cowpea pests by utilization of insect-resistant cultivars and minimum insecticide application. Field Crops Research. 1979; 2(4):373–85.
- Daryono BS, Natsuaki KT. Application of random amplified polymorphic DNA markers for detection of resistant cultivars of melon (*Cucumis melo*) against *Cucurbitaceae viruses*. Acta Horticulturae. 2002; 588(588):321–9.
- Lou Y, Cheng J. Induced plant resistance to phytophagous insects. Acta Entomologica Sinica. 1997; 40
 (3):320–31.
- Zhang Y, Yan F. Herbivore-induced volatiles and their roles in plant defence. Acta Entomologica Sinica. 1998; 41(2):204–14.
- Kou J, Shi S, Guixin HU, Zhou W. Photosynthetic physiological response of Medicago sativa to Odontothrips loti damage. Acta Ecologica Sinica. 2014; 34(20):2459–70.
- 31. Yu DY, Hu JF, Huang P, Yao JA, Lan YY, Fang DL. Influence of rasping-sucking feeding of thrips on surface ultrastructure and the contents of amino acids and proteins in *Ficus benjamina* leaves. Acta Entomologica Sinica. 2013; 56(1):60–8.
- Butler CD, O'Neil RJ. Life history characteristics of Orius insidiosus (Say) fed diets of soybean aphid, Aphis glycines Matsumura and soybean thrips, Neohydatothrips variabilis (Beach). Biological Control. 2007; 40(3):339–46.
- Ju RT, Wang F, Li B. Leaf physiological responses in the london plane tree (Platanus × acerifolia) (platanaceae) to injury by the sycamore lace bug, *Corythucha ciliata* (Hemiptera: Tingidae). Acta Entomologica Sinica. 2010; 53(9):1009–14.
- Kou JT, Shi SL, Gui-Xin HU, Zhou WH, Yao T. Photosynthetic physiology of odontothrips damaged Medicago sativa. Scientia Agricultura Sinica. 2013; 46(12):2459–70.