



updates



Citation: Simmons EE, Bergeron ER, Florian JP (2017) The impact of repetitive long-duration water immersion on vascular function. PLoS ONE 12(7): e0181673. https://doi.org/10.1371/journal.pone.0181673

Editor: Christopher Torrens, University of Southampton, UNITED KINGDOM

Received: December 28, 2016

Accepted: July 4, 2017

Published: July 27, 2017

Copyright: This is an open access article, free of all copyright, and may be freely reproduced, distributed, transmitted, modified, built upon, or otherwise used by anyone for any lawful purpose. The work is made available under the Creative Commons CCO public domain dedication.

Data Availability Statement: The Navy
Experimental Diving Unit has restricted distribution
of the data to allow access upon request. To
request access to vascular function data from the
RIP Study, please contact the Commanding Officer
of the Navy Experimental Diving Unit at 850-2303100 or at http://www.navsea.navy.mil/Home/
SUPSALV/NEDU/.

Funding: This research was supported by NAVSEA DSBDP N0002416WX02277 and Office of Naval Research Grant N0001416WX00313. The funders had no role in study design, data collection and

RESEARCH ARTICLE

The impact of repetitive long-duration water immersion on vascular function

Erin E. Simmons, Elizabeth R. Bergeron, John P. Florian*

Navy Experimental Diving Unit, Panama City, Florida, United States of America

* iohn.florian@navv.mil

Abstract

While physiological responses to water immersion (WI) are well-studied, the vascular responses after WI are less understood. Fifteen male subjects performed six-hour resting thermoneutral water immersions (WI) at 1.35 atmospheres absolute for four consecutive days, with follow-up on the fifth day. Measurements included peripheral endothelial function and augmentation index (PAT, peripheral arterial tonometry), beat-to-beat blood pressure (BP, photoplethysmography), heart rate (HR), and plasma volume (PV) calculated from changes in hemoglobin and hematocrit. The reactive hyperemia index (RHI), a marker of peripheral endothelial function, increased with repeated immersions (p = 0.008). By WI2 and WI3, RHI increased 12% and 16%, respectively, compared to WI1 values, but no significant differences were detected between WI4 and WI1 for either measure. Absolute augmentation index (AI) increased by an average of 33% (p<0.001) and AI normalized for HR (Al@75) by 11% (p = 0.12) following each WI. PV decreased significantly by 13.2% following WI and remained 6.8% lower at follow-up compared to pre-WI. Systolic blood pressure significantly decreased by an average of 2.5% following each WI (p = 0.012). Compared to pre-WI HR, average post-WI HR decreased 4.3% lower (p<0.001), but increased overall by 8.2% over the course of repeated WI (p<0.001). Total peripheral resistance increased by an average of 13.1% following WI (p = 0.003). Thus, peripheral endothelial function increases after two days of WI, and PAT-derived measures of arterial stiffness increase transiently post-WI. Additionally, BP and PAT-derived endothelial function diverge from their usual associations with arterial stiffness (i.e. augmentation index) in the context of WI.

Introduction

A number of physiological effects of diving have been studied in an attempt to better understand acute and long-term risks associated with both pressure and water immersion. Complete characterization of systemic effects of pressure and immersion is especially important for individuals who are concerned with safety and performance in association with diving (i.e., working, rescue, or military divers). Autonomic and hemodynamic responses to water immersion (WI) have been documented [1,2,3,4,5,6,7], as well as changes in fluid distribution and excretion [5,8,9,10,11]. Water immersion increases central blood volume and plasma volume (PV),



analysis, decision to publish, or preparation of the manuscript.

Competing interests: The authors have declared that no competing interests exist.

resulting in immersion diuresis [12,13]. The removal of hydrostatic pressure following WI leads to hypovolemia [11], increased calf vascular resistance (CVR), and decreased stroke volume (SV), cardiac output (CO), systolic blood pressure (SBP), and leg blood flow [13,14]. Vascular responses following long-duration WI are not well-characterized, although it is plausible that endothelial function and vascular stiffness may be affected.

While the endothelial cells lining blood vessels are responsible for many functions, including secretion of regulatory factors and inhibition of cellular adhesion, the primary role of the endothelium is facilitation of flow-mediated dilation [15,16]. A healthy endothelium is able to respond to increases in shear stress by causing vasorelaxation, a relationship first tested and observed by Furchgott and Zawadzki [17]. Nitric oxide (NO) has been identified as the endothelium-derived relaxation factor (EDRF) that causes vasodilation as well as inhibits the vasoconstrictor endothelin[16,18]. Changes in shear stress affect endothelial nitric oxide synthase (eNOS) activation and subsequently, NO production; it is therefore plausible that the changes in blood volume associated with hypovolemia following water immersion may affect endothelial function by altering vascular shear stress, but this pathway has not been studied.

Endothelial regulation of vessel tone in response to the balance between vasodilating and vasoconstricting substances may be related to measures of arterial stiffness. A reduction in nitric oxide may increase the stiffness of smooth muscle underlying the arterial endothelial cells, reducing the endothelial responses to shear stress [16]. Structural and functional changes in vessel walls that increase stiffness result in an increase in pulse wave velocity [19]. The increase in velocity causes the reflected wave to reach the heart during systole, thus increasing systolic blood pressure. While carotid-femoral pulse wave velocity is the current clinical gold standard for assessing arterial stiffness, analysis of pressure waveforms can also estimate arterial stiffness. The augmentation index (AI) is obtained by dividing the difference between the first and second systolic peaks by the pulse pressure. Utilizing this measure, a negative AI indicates normal, healthy vasculature while a positive AI is indicative of vessels with increased stiffness. There is some evidence that diving and water immersion may present cardiovascular challenges that affect arterial compliance, pulse pressures, and wave reflections[20]; however, such a relationship has yet to be determined.

Vascular responses to diving, as characterized by increased pressure with or without WI, while breathing compressed gases have been examined. The depth and duration of a dive are known to affect the uptake of inhaled gas into tissue, and subsequent reduction of pressure can cause bubbles to form as excess gas comes out of solution. Breathing compressed gas and the resulting decompression stress may thus decrease endothelial function. This is evidenced by the significant increase in the number of endothelial microparticles (a marker of endothelial damage) observed following compressed gas dives at 283 kPa[21]. This notion is supported by research showing that the production of reactive oxygen species (ROS) causes a reduction in endothelial function by inhibition of NO-mediated vasoregulation [22,23,24,25], and both ROS and intravascular bubble formation can result in physical endothelial cell damage [21,23,26,27,28,29].

Although previous diving studies have shown a decrease in endothelial function after diving with associated decompression stress [21,27,30,31], it is possible that head-out or shallow head-in WI alone will have a different effect on endothelial function [32], especially over the course of repeated immersions. Likewise, the effects of WI and resulting hypovolemia on measures of vascular stiffness are also unclear. It is reasonable to hypothesize that following resting immersion, the combination of inactivity together with post-immersion hypovolemia could increase arterial stiffness. Therefore, the objective of this study was to quantify the changes in endothelial function and vascular stiffness parameters following repeated six-hour WI with surface-supplied air. Endothelial function was tested by reactive hyperemia peripheral artery



tonometry (RH-PAT), which is a non-invasive measure of the ratio of the pulse wave amplitude signal during reactive hyperemia compared to baseline. We also measured blood pressure and resistance measures as these are related to vascular function responses.

Methods

Subjects

Fifteen healthy male military divers with an average of 7 ± 3 years of diving experience participated in the study. All subject characteristics can be found in Table 1. Subjects were not taking any medications pertinent to the outcome of the study, had not used nicotine in any form for the previous six months, and were asked to refrain from taking any elective medications, herbal supplements, or training supplements during the experimental period. Subjects were prohibited from alcohol for two days prior, and caffeine and exercise one day prior to the experimental period, and subjects were prohibited from diving for at least 10 days prior to the start of the experiment. A medical screening was performed, including complete blood count, complete metabolic panel, lipid profile evaluation, urinalysis, physical examination, and determination of maximal oxygen consumption ($\dot{V}O_{2_{\rm max}}$). This protocol was approved by the Navy Experimental Diving Unit Institutional Review Board. Subjects provided written, informed consent, and all procedures conformed to the Declaration of Helsinki.

Study design

The timeline of the experimental period is illustrated in Fig 1. Subjects were asked to abstain from food and drink (except water) for 2 hours prior to reporting to the laboratory. All physiological testing was performed before and after the 6-hour WI, and was conducted in a laboratory with air temperature maintained at $21-24^{\circ}$ C. Subjects wore running shorts and t-shirts for each laboratory visit and during the dive. Pre-immersion measurements included the EndoPAT test and a blood draw to measure hemoglobin (Hb) and hematocrit (Hct) for Δ PV calculation. Subjects were then administered a standardized snack, emptied their bladder, and were weighed immediately prior to being immersed in the tank. They surfaced after hour 3 for a 10-min standardized lunch and returned to full immersion until hour 6. Post-WI weight was taken after the subject had emptied his bladder and was completely dried. Weight loss was calculated from the difference in pre- and postimmersion weight. Post-WI physiological measurements commenced after drying and weighing. This procedure was repeated for four consecutive days, and a follow-up (FU) test was conducted the morning of the fifth day.

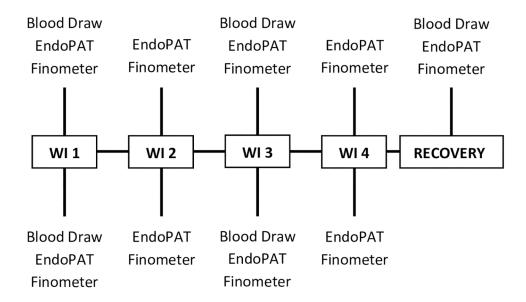
Table 1. Subject characteristics.

30 ± 6 (21–40)
88 ± 10
178±9
28 ± 2
52 ± 7
123±7
75 ± 11
4.27 ± 0.75
7±3

Values are means \pm SD. n = 15 subjects. BMI, body mass index; SBP, systolic blood pressure; DBP, diastolic blood pressure.

https://doi.org/10.1371/journal.pone.0181673.t001

Pre-WI



Post-WI

Fig 1. Water immersion and experimental timeline. Four consecutive water immersions (WI) were performed, immediately preceded and followed by blood draws (WI1, WI3, and Recovery), EndoPAT testing (WI1-4 and Recovery), and Finometer measurement (WI1-4 and Recovery).

https://doi.org/10.1371/journal.pone.0181673.g001

Water immersion

Each WI was conducted for 6 hours at the bottom of a 15 foot test pool with a thermoneutral water temperature of 32–33°C. Subjects were provided weights if necessary to maintain negative buoyancy, and were instructed to remain still and seated in a chair for the duration of WI. Subjects sat reclined so that their chest remained at a depth of 12–13 feet. They breathed surface-supplied air through a MK20 breathing apparatus (Aga mask, Interspiro), which delivers breathing gas at a pressure slightly greater than ambient pressure to minimize breathing resistance. At hour 3 of WI, participants were instructed to surface and stand on a platform with only head and shoulders out while eating a 2.2 MJ lunch containing 24% fat, 64% carbohydrate, 12% protein, and 500 ml of liquids, which included a supplementation shake (Ensure) and sports drink (Gatorade). After 10 minutes, participants returned to the bottom of the test pool until surfacing at hour 6.

Vascular function measurement

Peripheral endothelial function and augmentation index were measured before and approximately 30 min after WI with the EndoPAT 2000 device (Itamar Medical, Caesarea, Israel) as previously described and reviewed [18,33,34,35]. RH-PAT measurements using the EndoPAT 2000 (Itamar Medical, Caesarea, Israel) have been shown to be highly reproducible [33,36,37], correlate well with brachial artery ultrasound measurement of flow-mediated dilation (FMD) [38,39,40], and detect the effects of acute interventions [21,41,42,43,44,45]. It is also recognized



as an operator-independent method for both dynamic and diagnostic measures of vascular function [15,46,47,48]. The use of EndoPAT for finger tonometry is appropriate to measure endogenous NO-mediated vasodilation regulation since distal regions of limbs are major sites of sympathetic alpha adrenergic vasoconstrictor activity [15]. EndoPAT measures the change in peripheral arterial tone (PAT) signal at the fingertips in response to reactive hyperemia (RH-PAT) following upper arm occlusion. Endothelial function was measured using the reactive hyperemia index (RHI), which is determined from the ratio of post- to pre-occlusion PAT signal in the occluded arm relative to the same ratio in the non-occluded arm, and corrected for vascular tone of the occluded arm at baseline [37]. Augmentation index (AI) is a measure of wave reflections influenced by arterial stiffness, which is calculated by averaging multiple valid pulses during baseline in the occluded arm to determine the augmentation of the forward propagated wave by the returning reflective wave. This is accomplished by finding the systolic peak (P_1) and the backward reflected peak (P_2) and then using the formula: $(P_2-P_1)/P_1$. Since the pulse waveform may be influenced by heart rate, AI can be corrected for heart rate by standardizing the measure to 75 bpm (AI@75). Study participants rested supine for 10 minutes before the start of the measurement period. Probes were placed on the index finger of both hands and a blood pressure cuff was applied to the non-dominant arm. Each recording consisted of a six-minute baseline period, a five-minute period of upper arm occlusion, and a fiveminute post-occlusion measurement period. During occlusion, the blood pressure cuff was inflated to a suprasystolic pressure that did not exceed 300 mmHg. The RHI and AI values are automatically calculated by the EndoPAT system.

Hemodynamic measurements

Beat-to-beat arterial pressure was measured during the EndoPAT protocol using photo-plethysmography (Finometer, Finapres Medical Systems) on the ring finger of the dominant hand, and heart rate was measured using the EndoPAT. Total peripheral resistance (TPR), calculated by the Finometer as MAP / Finometer CO, was also recorded. Calibration of finger pressure to brachial artery pressure was performed using the manufacturer's return-to-flow system. Beat-to-beat values of SBP, DBP, MAP, and TPR were averaged for approximately five minutes of the resting baseline period prior to occlusion of the non-dominant arm.

Blood analysis

Blood was collected before and after WI and was immediately analyzed for Hb and Hct levels using the conductimetric method (Rapidpoint 405, Siemens). Blood draws were performed after ten minutes of seated rest, after which subjects lay supine during blood collection to prevent possible presyncopal symptoms which may occur after WI. The relative change in PV following WI was calculated from changes in Hb and Hct concentrations according to the Harrison modification of the Dill and Costill equation [49,50].

Statistical analysis

A repeated measures analysis of variance (ANOVA) was conducted to determine the effect of WI on the dependent variables. SAS (Version 9.2) was used to run a 2 [Pre-WI, Post-WI] x 4 [Day1, Day2, Day3, Day4] mixed linear model ANOVA. When appropriate, a post-hoc Bonferroni-Holm correction was applied to identify differences between factors. Differences between FU and WI1 pre values were compared using a paired t-test.



Results

Weight loss and plasma volume

Mean weight loss after WI, adjusted for food and fluid intake was 1.80 ± 0.14 kg (p<0.001). Plasma volume significantly decreased by 13.2% on average following each WI (p<0.001) and remained decreased by 6.8% at FU. There was no significant effect of WI day on the change in PV (Table 2).

Endothelial function

The effect of WI day on RHI was significant (p = 0.008) and post-hoc analysis revealed a significant increase of 12.3% (p < 0.01) on WI2 and 16.1% (p < 0.01) on WI3 compared to WI1; WI4 was not significantly different from baseline values on Day 1 (Fig 2). There were no significant differences in RHI between pre- and post-WI measurements, but FU RHI was significantly higher than that at baseline (WI1 pre, p < 0.005).

Arterial stiffness parameters

AI was significantly increased by an average of 33.3% following WI (p<0.001). There were no cumulative effects from consecutive days of WI (Fig 3A). When normalized for heart rate (AI@75), AI still showed a significant 10.7% increase post-WI (p = 0.012, Fig 3B). AI was not significantly different at FU compared to baseline (WI1) measures.

All interaction effects of the dependent variables were non-significant (Table 2).

Table 2. Responses to 6-hr water immersion.

Variable	WI 1	WI 2	WI 3	WI 4	FU	Main Pre/Post	Main Day	Interaction Pre/Post* Time	
Weight (kg)									
Pre	88.3±2.8	88.7±2.9	87.9±2.7	87.7±2.8	87.5±2.8	<0.001*	<0.001*	0.572	
Post	86.8±2.8	86.7±2.8	86.7±2.8	87.3±2.8					
dPV (%)									
Pre	-	-	-4.5±2.2	-	-6.8±1.9	<0.001*	0.397	0.312	
Post	-13.0±1.8	-	-13.3±2.0	-					
SBP (mmHg	3)	·							
Pre	131±3	130±2	131±3	132±2	131±2	0.014*	0.169	0.425	
Post	129±3	124±4	128±3	130±3					
DBP (mmHg	3)								
Pre	71±2	73±2	72±2	74±2	73±1	0.071	0.617	0.104	
Post	77±2	73±3	72±3	74±2					
MAP (mmH	g)	·							
Pre	92±3	92±2	93±2	94±2	92±2	0.580	0.455	0.291	
Post	96±2	91±3	92±2	94±2					
TPR (dynes	· s · cm ⁻⁵)								
Pre	912±76	892±41	856±65	894±60	950±69	950±69	0.002*	0.064	0.253
Post	1110±80	1062±70	919±60	933±34					

Values are means ± SE for pre- and post-immersion variables. FU = follow-up, dPV = change in plasma volume, SBP = systolic blood pressure, DBP = diastolic blood pressure, MAP = mean arterial pressure, TPR = total peripheral resistance. Main, P-value of main effects of pre- and post-immersion and day of water immersion. Interaction, P-value of a two-way (pre/post * day) repeated-measures ANOVA interaction.

*P < 0.05.

https://doi.org/10.1371/journal.pone.0181673.t002

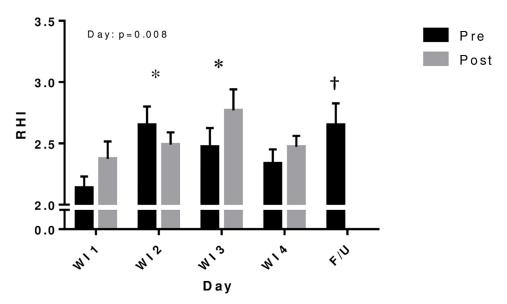


Fig 2. Peripheral endothelial function before and after repeated 6-h water immersions. Values are mean ± SE. *P<0.05 compared to water immersion 1 (WI1). †P<0.05 compared to WI1 pre. Immersion resulted in increased reactive hyperemia index (RHI) by the second and third days of WI. RHI during follow-up (F/U) remained elevated compared to baseline (WI1 pre).

https://doi.org/10.1371/journal.pone.0181673.g002

Blood pressure, heart rate, and resistance

On average, SBP significantly decreased by 2.5% following WI (p = 0.012), but no significant differences in diastolic or mean arterial pressures were observed (Table 2). As shown in Fig 4, heart rate was 4.3% lower after WI on average (p<0.001), but showed a significant increase of 5.9% (p<0.002) by WI3 and 8.2% (p<0.001) by WI4 compared to WI1 (p<0.001), and remained elevated during FU compared to baseline (WI 1 pre). Total peripheral resistance (TPR) increased an average of 13.1% following WI (p = 0.003) with no significant effect of WI day (Table 2).

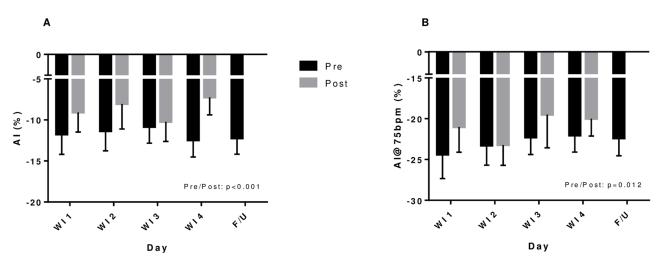


Fig 3. Arterial stiffness parameters before and after repeated 6-h water immersions. Values are mean ± SE. *P<0.05 compared to water immersion 1 (WI1). Increases in augmentation index (AI—A) and AI corrected for heart rate (AI@75—B) were seen following WI but there was no cumulative effect of repeated WI.

https://doi.org/10.1371/journal.pone.0181673.g003

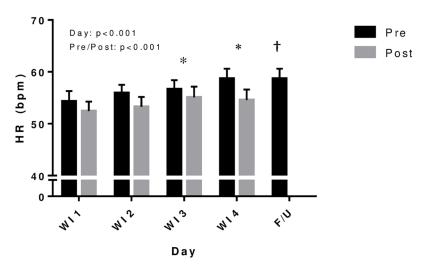


Fig 4. Heart rate before and after repeated 6-h water immersions. Values are mean ± SE. *P<0.05 compared to water immersion 1 (WI1). †P<0.05 compared to WI1 pre. Heart rate was lower post-WI but increased over the course of repeated immersions. Heart rate during follow-up (F/U) remained elevated compared to baseline (WI1 pre).

https://doi.org/10.1371/journal.pone.0181673.g004

Discussion

This study is the first to our knowledge to examine the effect of long, repeated water immersions on post-immersion RHI and AI, measures of endothelial function and arterial stiffness. RHI-PAT significantly increased by the third day of WI, but returned toward baseline levels on the fourth day. On the other hand, WI acutely increased AI after each WI, though this effect seemed to be transient as there was no change in AI detected pre-immersion for any WI days. Interestingly, blood pressure and endothelial function diverged from their usual associations with measures of arterial stiffness. These findings thus reveal atypical effects of WI on post-immersion vascular function.

This study contributes novel information related to diving and immersion physiology by separating out immersion effects from those caused by diving exposures with decompression stress. Previous studies have found that breathing air at depth resulted in decreased endothelial function, likely due to production of reactive oxygen species as well as bubbles in the circulation. Decreases in endothelial function measured by FMD were found following both dry and open-sea single dives for 80 minutes at 280 kPa and 30 minutes at 400 kPa, respectively, but these decreases in function were not correlated with bubble formation [27,31]. Differences may also exist depending on which breathing gases are utilized. In a previous study by Madden et al. [21], post-dive measures of endothelial function were significantly lower after dry, 60-minute air dives at 283 kPa compared to the same dive breathing 100% oxygen. Future studies should investigate whether RHI exhibits the same pattern with repeated oxygen WI as it does for WI while breathing air.

While endothelial function might be expected to decrease in response to the repetitive water immersions based on such previous diving studies, we recognized that there could be distinctive results for immersion without decompression. Indeed, our study showed that peripheral endothelial function actually improved from baseline by the second day of WI. We hypothesize that chronic exposure to increased arterial stiffness post-WI may lead to increased shear stress following each immersion. This could potentially increase NO production and cause an increase in RHI over the course of repeated immersions. An investigation of



rehabilitation programs utilizing WI did in fact detect an increase in resting plasma concentration of the NO metabolite, nitrate, following five, 50-minute immersions per week for 3 weeks, suggesting upregulated endothelial function with chronic WI [32]. This may indeed be a chronic exposure effect, considering that a study measuring endothelial function in response to reactive hyperemia during WI found no change in plasma NO metabolites [51]. Future studies should further investigate the mechanisms behind the observed increase in RHI after repeated immersions, since a previous study found that FMD decreased on the third dive day in a series of repetitive dives [52]. While there is apparently a very different effect on endothelial function with diving compared to WI, it seems that there may be a consistent timing threshold (~2–3 days) for changes in endothelial function to take place. This observation should be tested further, and mechanisms by which an adaptive effect might occur should be explored.

Endothelial cell signaling has the potential to affect arterial stiffness via eNOS stimulation and NO production, and increased stiffness may promote a decline in eNOS activity and worsen endothelial cell functioning [53]. Though they have the potential to exert effects on each other, endothelial function and arterial stiffness are not necessarily linked [54]. Similarly, while chronic arterial stiffness is typically related to hypertension, there is evidence that measures of arterial stiffness and blood pressure can diverge acutely. A study of head-out WI found that blood pressure was significantly reduced while arterial stiffness increased [55]. Lazar et al. [55] proposed several factors that could potentially cause decreased BP with WI, including increased NO release and atrial natriuretic peptide, and decreased renin activity and plasma norepinephrine levels [50]. It is possible that the observed transient effects of WI on measures of arterial stiffness are not strong enough to affect blood pressure or endothelial function in ways previously determined in clinical studies of chronic cardiovascular states. Chronic increased stiffness generally contributes to increases in systolic blood pressure since the reflected wave reaches the heart faster and at systole, driving up systolic pressures. The acute increase in stiffness during immersion may not be enough to counteract the drop in systolic blood pressure caused by immersion-induced hypovolemia.

Other extrinsic influences on arterial stiffness include shear stress, angiotensin, and sympathetic neurohormones [53]. It is unlikely that traditional factors affecting arterial stiffness, such as increased collagen, decreased elastin, or deposition of sulfates, proteoglycans, or fibronectins, would act over the short time periods in this study. The lack of a significant effect of the WI day provides evidence that chronic structural changes did not occur over the course of the four days of WI, an observation supported by a previous study that found no tolerance effect of recent or repetitive head-out WI on changes in AI during immersion[50]. The transient increase in AI could be in response to reduced blood flow and inactivity during immersion, as well as post-immersion hypovolemia. Since previous studies have found that exercise has an acutely positive effect on compliance and stiffness, it is possible that inactivity and hypovolemia could have the opposite effect. Previous studies have shown that short-term inactivity results in greater arterial stiffness [56], and that blood volume and blood flow are reduced to extremities following immersion [14]. Additionally, increased extremity vascular resistance seen in our previous study may be linked to post-immersion peripheral arterial stiffness [14].

Previous studies have documented changes in arterial stiffness following both acute and chronic interventions, and acute changes have been detected in response to many stimuli other than diving or immersion [57,58,59,60,61]. Acute aerobic exercise (rowing) caused stiffness to decrease[62], and acute sprint interval exercise caused increased compliance but no change in brachial artery stiffness [63]. In both studies, however, chronic training resulted in decreased compliance and increased arterial stiffness. In another study [64], trained endurance athletes with higher $\dot{V}O_{2\,\text{peak}}$ were found to have decreased arterial stiffness (AIx@75)



compared to age-matched controls. Intensity of exercise may also affect arterial stiffness, which was shown to decrease significantly after high intensity interval exercise but not moderate continuous exercise [65]. Conversely, short-term bed rest (five days) resulted in decreased FMD and increased arterial stiffness [56].

Future work should identify the cause and time course of the transient increase in arterial stiffness parameters following WI. Although exercise and immersion appear to have opposite effects on arterial stiffness, the observation that some forms of chronic exercise tend to increase arterial stiffness begs the question of whether or not chronic diving or WI responds in a similar fashion. A recent study of female pearl divers suggests that chronic free-diving decreases arterial stiffness, but only compared to sedentary age-matched females; no difference was observed in arterial stiffness when compared to active age-matched females [20]. Taken with the observation that inactivity promotes increased arterial stiffness[56], there may be a greater effect of inactivity on arterial stiffness during our study than the immersion itself. Studies employing exercise during WI or an inactive time control condition on the surface would be able to determine whether it is the immersion or inactivity that contributes to the transient increase in arterial stiffness parameters that we observed.

While this study characterized the vascular response to repeated, long-duration WI, more research is needed to outline the mechanisms by which these responses occurred. The results of this study are limited to endothelial function and arterial stiffness, as measured by Endo-PAT, in the context of resting WI while breathing surface-supplied air at 130 kPa. The effects of breathing oxygen or exercising during WI have yet to be determined. It is also important to note that AI has been questioned as a measure of arterial stiffness, as the characteristics of pulse waves can be determined by a variety of factors including small and large artery compliance, left ventricular function, artery elasticity, wave velocity, and heart rate (though a measure of AI normalized to heart rate is also presented here). Despite these limitations, we conclude that four consecutive days of six-hour WI while breathing air caused a transient increase in arterial stiffness parameters following each WI as well as an increase in endothelial function on the second day. Changes in systolic blood pressure also diverged from changes in arterial stiffness, which is a deviation from typical clinical findings. The mechanisms contributing to our results can only be speculated, and future studies should seek to determine the mechanisms by which our observations occurred.

Acknowledgments

We thank Dr. Barbara Shykoff and the NEDU corpsmen, electronics technicians, and technical support staff who helped with experimentation. We also appreciate the participation by the research subjects. The opinions and assertions contained herein are those of the authors, not to be construed as the official views of the Department of the Navy or the US Government.

Author Contributions

Conceptualization: John P. Florian.

Data curation: Erin E. Simmons, Elizabeth R. Bergeron, John P. Florian.

Formal analysis: Erin E. Simmons, Elizabeth R. Bergeron.

Funding acquisition: John P. Florian.

Investigation: Erin E. Simmons, Elizabeth R. Bergeron, John P. Florian.

Methodology: John P. Florian.

Project administration: John P. Florian.



Resources: John P. Florian.

Software: Erin E. Simmons, Elizabeth R. Bergeron, John P. Florian.

Supervision: John P. Florian.Validation: John P. Florian.

Visualization: Erin E. Simmons.

Writing - original draft: Erin E. Simmons.

Writing - review & editing: Elizabeth R. Bergeron, John P. Florian.

References

- Mano T, Iwase S, Saito M, Koga K, Abe H, Inamura K, et al. Neural and humoral controlling mechanisms of cardiovascular functions in man under weightlessness simulated by water immersion. Acta Astronaut. 1991; 23: 31–33. PMID: 11537141
- Miwa C, Sugiyama Y, Mano T, Iwase S, Matsukawa T. Sympatho-vagal responses in humans to thermoneutral head-out water immersion. Aviat Space Environ Med. 1997; 68: 1109–1114. PMID: 9408561
- Schipke JD, Pelzer M. Effect of immersion, submersion, and scuba diving on heart rate variability. Br J Sports Med. 2001; 35: 174–180. https://doi.org/10.1136/bjsm.35.3.174 PMID: 11375876
- Tripathi KK. Very low frequency oscillations in the power spectra of heart rate variability during dry supine immersion and exposure to non-hypoxic hypobaria. Physiol Meas. 2011; 32: 717–729. https://doi.org/10.1088/0967-3334/32/6/008 PMID: 21606562
- Wilcock IM, Cronin JB, Hing WA. Physiological response to water immersion: a method for sport recovery? Sports Med. 2006; 36: 747–765. PMID: 16937951
- Mourot L, Bouhaddi M, Gandelin E, Cappelle S, Dumoulin G, Wolf JP, et al. Cardiovascular autonomic control during short-term thermoneutral and cool head-out immersion. Aviat Space Environ Med. 2008; 79: 14–20. PMID: 18225773
- Mourot L, Bouhaddi M, Gandelin E, Cappelle S, Nguyen NU, Wolf JP, et al. Conditions of autonomic reciprocal interplay versus autonomic co-activation: effects on non-linear heart rate dynamics. Auton Neurosci. 2007; 137: 27–36. https://doi.org/10.1016/j.autneu.2007.06.284 PMID: 17662671
- Epstein M. Renal effects of head-out water immersion in humans: a 15-year update. Physiol Rev. 1992; 72: 563–621. PMID: 1626032
- Greenleaf JE, Morse JT, Barnes PR, Silver J, Keil LC. Hypervolemia and plasma vasopressin response during water immersion in men. J Appl Physiol Respir Environ Exerc Physiol. 1983; 55: 1688–1693. PMID: 6363363
- Harrison MH, Keil LC, Wade CA, Silver JE, Geelen G, Greenleaf JE. Effect of hydration on plasma volume and endocrine responses to water immersion. J Appl Physiol (1985). 1986; 61: 1410–1417.
- Norsk P, Bonde-Petersen F, Warberg J. Central venous pressure and plasma arginine vasopressin during water immersion in man. Eur J Appl Physiol Occup Physiol. 1985; 54: 71–78. PMID: 4018059
- Greenleaf JE, Shvartz E, Kravik S, Keil IC. Fluid shifts and endocrine responses during chair rest and water immersion in man. J Appl Physiol Respir Environ Exerc Physiol. 1980; 48: 79–88. PMID: 6986349
- Boussuges A, Gole Y, Mourot L, Jammes Y, Melin B, Regnard J, et al. Haemodynamic changes after prolonged water immersion. J Sports Sci. 2009; 27: 641–649. https://doi.org/10.1080/ 02640410902717001 PMID: 19296363
- **14.** Florian JP, Simmons EE, Chon KH, Faes L, Shykoff BE. Cardiovascular and autonomic responses to physiological stressors before and after six hours of water immersion. J Appl Physiol (1985). 2013; 115: 1275–1289.
- Celermajer DS. Reliable endothelial function testing: at our fingertips? Circulation. 2008; 117: 2428–2430. https://doi.org/10.1161/CIRCULATIONAHA.108.775155 PMID: 18474821
- Vanhoutte PM, Shimokawa H, Feletou M, Tang EH. Endothelial dysfunction and vascular disease—a 30th anniversary update. Acta Physiol (Oxf). 2017; 219: 22–96.
- Furchgott RF, Zawadzki JV. The obligatory role of endothelial cells in the relaxation of arterial smooth muscle by acetylcholine. Nature. 1980; 288: 373–376. PMID: 6253831



- Kandhai-Ragunath JJ, Jorstad HT, de Man FH, Peters RJ, von Birgelen C. Approaches for non-invasive assessment of endothelial function: focus on peripheral arterial tonometry. Neth Heart J. 2013; 21: 214–218. https://doi.org/10.1007/s12471-011-0202-5 PMID: 22311175
- Najjar SS, Scuteri A, Lakatta EG. Arterial aging: is it an immutable cardiovascular risk factor? Hypertension. 2005; 46: 454–462. https://doi.org/10.1161/01.HYP.0000177474.06749.98 PMID: 16103272
- Tanaka H, Tomoto T, Kosaki K, Sugawara J. Arterial stiffness of lifelong Japanese female pearl divers. Am J Physiol Regul Integr Comp Physiol. 2016; 310: R975–978. https://doi.org/10.1152/ajpregu. 00048.2016 PMID: 26984889
- Madden LA, Chrismas BC, Mellor D, Vince RV, Midgley AW, McNaughton LR, et al. Endothelial function and stress response after simulated dives to 18 msw breathing air or oxygen. Aviat Space Environ Med. 2010; 81: 41–45. PMID: 20058736
- Giordano FJ. Oxygen, oxidative stress, hypoxia, and heart failure. J Clin Invest. 2005; 115: 500–508. https://doi.org/10.1172/JCI200524408 PMID: 15765131
- Berenji Ardestani S, Buzzacott P, Eftedal I. The aging diver: endothelial biochemistry and its potential implications for cardiovascular health. Diving Hyperb Med. 2015; 45: 235–239. PMID: 26687310
- Brubakk AO, Ross JA, Thom SR. Saturation diving; physiology and pathophysiology. Compr Physiol. 2014; 4: 1229–1272. https://doi.org/10.1002/cphy.c130048 PMID: 24944036
- 25. Theunissen S, Balestra C, Boutros A, De Bels D, Guerrero F, Germonpre P. The effect of pre-dive ingestion of dark chocolate on endothelial function after a scuba dive. Diving Hyperb Med. 2015; 45: 4–9. PMID: 25964032
- Wang Q, Guerrero F, Mazur A, Lambrechts K, Buzzacott P, Belhomme M, et al. Reactive Oxygen Species, Mitochondria, and Endothelial Cell Death during In Vitro Simulated Dives. Med Sci Sports Exerc. 2015; 47: 1362–1371. https://doi.org/10.1249/MSS.0000000000000563 PMID: 25380471
- Brubakk AO, Duplancic D, Valic Z, Palada I, Obad A, Bakovic D, et al. A single air dive reduces arterial endothelial function in man. J Physiol. 2005; 566: 901–906. https://doi.org/10.1113/jphysiol.2005. 089862 PMID: 15961424
- Sureda A, Batle JM, Ferrer MD, Mestre-Alfaro A, Tur JA, Pons A. Scuba diving activates vascular antioxidant system. Int J Sports Med. 2012; 33: 531–536. https://doi.org/10.1055/s-0031-1297957 PMID: 22562744
- Jamieson D, Chance B, Cadenas E, Boveris A. The relation of free radical production to hyperoxia.
 Annu Rev Physiol. 1986; 48: 703–719. https://doi.org/10.1146/annurev.ph.48.030186.003415 PMID: 3010832
- Theunissen S, Guerrero F, Sponsiello N, Cialoni D, Pieri M, Germonpre P, et al. Nitric oxide-related endothelial changes in breath-hold and scuba divers. Undersea Hyperb Med. 2013; 40: 135–144. PMID: 23682545
- Lambrechts K, Pontier JM, Balestra C, Mazur A, Wang Q, Buzzacott P, et al. Effect of a single, opensea, air scuba dive on human micro- and macrovascular function. Eur J Appl Physiol. 2013; 113: 2637– 2645. https://doi.org/10.1007/s00421-013-2676-x PMID: 23949788
- Mourot L, Teffaha D, Bouhaddi M, Ounissi F, Vernochet P, Dugue B, et al. Training-induced increase in nitric oxide metabolites in chronic heart failure and coronary artery disease: an extra benefit of waterbased exercises? Eur J Cardiovasc Prev Rehabil. 2009; 16: 215–221. https://doi.org/10.1097/HJR.0b013e3283292fcf PMID: 19276981
- 33. Xu Y, Arora RC, Hiebert BM, Lerner B, Szwajcer A, McDonald K, et al. Non-invasive endothelial function testing and the risk of adverse outcomes: a systematic review and meta-analysis. Eur Heart J Cardiovasc Imaging. 2014; 15: 736–746. https://doi.org/10.1093/ehjci/jet256 PMID: 24399339
- Axtell AL, Gomari FA, Cooke JP. Assessing endothelial vasodilator function with the Endo-PAT 2000. J Vis Exp. 2010.
- Faizi AK, Kornmo DW, Agewall S. Evaluation of endothelial function using finger plethysmography. Clin Physiol Funct Imaging. 2009; 29: 372–375. https://doi.org/10.1111/j.1475-097X.2009.00882.x PMID: 19552735
- Brant LC, Barreto SM, Passos VM, Ribeiro AL. Reproducibility of peripheral arterial tonometry for the assessment of endothelial function in adults. J Hypertens. 2013; 31: 1984–1990. https://doi.org/10. 1097/HJH.0b013e328362d913 PMID: 23751970
- McCrea CE, Skulas-Ray AC, Chow M, West SG. Test-retest reliability of pulse amplitude tonometry measures of vascular endothelial function: implications for clinical trial design. Vasc Med. 2012; 17: 29–36. https://doi.org/10.1177/1358863X11433188 PMID: 22363016
- Dhindsa M, Sommerlad SM, DeVan AE, Barnes JN, Sugawara J, Ley O, et al. Interrelationships among noninvasive measures of postischemic macro- and microvascular reactivity. J Appl Physiol (1985). 2008; 105: 427–432.



- 39. Kuvin JT, Patel AR, Sliney KA, Pandian NG, Sheffy J, Schnall RP, et al. Assessment of peripheral vascular endothelial function with finger arterial pulse wave amplitude. Am Heart J. 2003; 146: 168–174. https://doi.org/10.1016/S0002-8703(03)00094-2 PMID: 12851627
- 40. Hamburg NM, Palmisano J, Larson MG, Sullivan LM, Lehman BT, Vasan RS, et al. Relation of brachial and digital measures of vascular function in the community: the Framingham heart study. Hypertension. 2011; 57: 390–396. https://doi.org/10.1161/HYPERTENSIONAHA.110.160812 PMID: 21263120
- Worthley MI, Prabhu A, De Sciscio P, Schultz C, Sanders P, Willoughby SR. Detrimental effects of energy drink consumption on platelet and endothelial function. Am J Med. 2010; 123: 184–187. https:// doi.org/10.1016/j.amjmed.2009.09.013 PMID: 20103032
- Ochiai R, Sugiura Y, Shioya Y, Otsuka K, Katsuragi Y, Hashiguchi T. Coffee polyphenols improve peripheral endothelial function after glucose loading in healthy male adults. Nutr Res. 2014; 34: 155– 159. https://doi.org/10.1016/j.nutres.2013.11.001 PMID: 24461317
- Bonetti PO, Barsness GW, Keelan PC, Schnell TI, Pumper GM, Kuvin JT, et al. Enhanced external counterpulsation improves endothelial function in patients with symptomatic coronary artery disease. J Am Coll Cardiol. 2003; 41: 1761–1768. PMID: 12767662
- 44. Li Z, Wong A, Henning SM, Zhang Y, Jones A, Zerlin A, et al. Hass avocado modulates postprandial vascular reactivity and postprandial inflammatory responses to a hamburger meal in healthy volunteers. Food Funct. 2013; 4: 384–391. https://doi.org/10.1039/c2fo30226h PMID: 23196671
- Martin EA, Tan SL, MacBride LR, Lavi S, Lerman LO, Lerman A. Sex differences in vascular and endothelial responses to acute mental stress. Clin Auton Res. 2008; 18: 339–345. https://doi.org/10.1007/s10286-008-0497-5 PMID: 18850310
- 46. Urbina EM, Williams RV, Alpert BS, Collins RT, Daniels SR, Hayman L, et al. Noninvasive assessment of subclinical atherosclerosis in children and adolescents: recommendations for standard assessment for clinical research: a scientific statement from the American Heart Association. Hypertension. 2009; 54: 919–950. https://doi.org/10.1161/HYPERTENSIONAHA.109.192639 PMID: 19729599
- Poredos P, Jezovnik MK. Testing endothelial function and its clinical relevance. J Atheroscler Thromb. 2013; 20: 1–8. PMID: 22972428
- 48. Hamburg NM, Benjamin EJ. Assessment of endothelial function using digital pulse amplitude tonometry. Trends Cardiovasc Med. 2009; 19: 6–11. https://doi.org/10.1016/j.tcm.2009.03.001 PMID: 19467447
- **49.** Harrison MH, Graveney MJ, Cochrane LA. Some sources of error in the calculation of relative change in plasma volume. Eur J Appl Physiol. 1982; 50: 13–21.
- Strauss MB, Davis RK, Rosenbaum JD, Rossmeisl EC. "Water diuresis" produced during recumbancy by the intravenous infusion of isotonic saline solution. J Clin Invest. 1951; 30: 862–868. https://doi.org/ 10.1172/JCI102501 PMID: 14861307
- Ayme K, Gavarry O, Rossi P, Desruelle AV, Regnard J, Boussuges A. Effect of head-out water immersion on vascular function in healthy subjects. Appl Physiol Nutr Metab. 2014; 39: 425–431. https://doi.org/10.1139/apnm-2013-0153 PMID: 24669983
- 52. Madden D, Barak O, Thom SR, Yang M, Bhopale VM, Ljubkovic M, et al. The impact of predive exercise on repetitive SCUBA diving. Clin Physiol Funct Imaging. 2016; 36: 197–205. https://doi.org/10.1111/ cpf.12213 PMID: 25371042
- 53. Zieman SJ, Melenovsky V, Kass DA. Mechanisms, pathophysiology, and therapy of arterial stiffness. Arterioscler Thromb Vasc Biol. 2005; 25: 932–943. https://doi.org/10.1161/01.ATV.0000160548. 78317.29 PMID: 15731494
- **54.** Cussons AJ, Watts GF, Stuckey BG. Dissociation of endothelial function and arterial stiffness in nonobese women with polycystic ovary syndrome (PCOS). Clin Endocrinol (Oxf). 2009; 71: 808–814.
- 55. Lazar JM, Morris M, Qureshi G, Jean-Noel G, Nichols W, Qureshi MR, et al. The effects of head-out-of-water immersion on arterial wave reflection in healthy adults. J Am Soc Hypertens. 2008; 2: 455–461. https://doi.org/10.1016/j.jash.2008.04.013 PMID: 20409926
- 56. Nosova EV, Yen P, Chong KC, Alley HF, Stock EO, Quinn A, et al. Short-term physical inactivity impairs vascular function. J Surg Res. 2014; 190: 672–682. https://doi.org/10.1016/j.jss.2014.02.001 PMID: 24630521
- Barnes JN, Trombold JR, Dhindsa M, Lin HF, Tanaka H. Arterial stiffening following eccentric exerciseinduced muscle damage. J Appl Physiol (1985). 2010; 109: 1102–1108.
- 58. Boos CJ, Hodkinson P, Mellor A, Green NP, Woods DR. The effects of acute hypobaric hypoxia on arterial stiffness and endothelial function and its relationship to changes in pulmonary artery pressure and left ventricular diastolic function. High Alt Med Biol. 2012; 13: 105–111. https://doi.org/10.1089/ham. 2012.1009 PMID: 22724613



- Sunbul M, Kanar BG, Durmus E, Kivrak T, Sari I. Acute sleep deprivation is associated with increased arterial stiffness in healthy young adults. Sleep Breath. 2014; 18: 215–220. https://doi.org/10.1007/ s11325-013-0873-9 PMID: 23852444
- 60. Vlachopoulos C, Aznaouridis K, Bratsas A, Ioakeimidis N, Dima I, Xaplanteris P, et al. Arterial stiffening and systemic endothelial activation induced by smoking: The role of COX-1 and COX-2. Int J Cardiol. 2015; 189: 293–298. PMID: 25919966
- 61. West SG, McIntyre MD, Piotrowski MJ, Poupin N, Miller DL, Preston AG, et al. Effects of dark chocolate and cocoa consumption on endothelial function and arterial stiffness in overweight adults. Br J Nutr. 2014; 111: 653–661. https://doi.org/10.1017/S0007114513002912 PMID: 24274771
- 62. Franzen K, Reppel M, Koster J, Mortensen K. Acute and chronic effects on central hemodynamics and arterial stiffness in professional rowers. Physiol Meas. 2016; 37: 544–553. https://doi.org/10.1088/0967-3334/37/4/544 PMID: 26999470
- **63.** Olver TD, Reid SM, Smith AR, Zamir M, Lemon PW, Laughlin MH, et al. Effects of acute and chronic interval sprint exercise performed on a manually propelled treadmill on upper limb vascular mechanics in healthy young men. Physiol Rep. 2016; 4.
- Denham J, Brown NJ, Tomaszewski M, Williams B, O'Brien BJ, Charchar FJ. Aortic augmentation index in endurance athletes: a role for cardiorespiratory fitness. Eur J Appl Physiol. 2016; 116: 1537– 1544. https://doi.org/10.1007/s00421-016-3407-x PMID: 27278522
- 65. Hanssen H, Nussbaumer M, Moor C, Cordes M, Schindler C, Schmidt-Trucksass A. Acute effects of interval versus continuous endurance training on pulse wave reflection in healthy young men. Atherosclerosis. 2015; 238: 399–406. https://doi.org/10.1016/j.atherosclerosis.2014.12.038 PMID: 25558034