

RESEARCH ARTICLE

Light Levels Affect Carbon Utilisation in Tropical Seagrass under Ocean Acidification

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Abstract

Under future ocean acidification (OA), increased availability of dissolved inorganic carbon (DIC) in seawater may enhance seagrass productivity. However, the ability to utilise additional DIC could be regulated by light availability, often reduced through land runoff. To test this, two tropical seagrass species, *Cymodocea serrulata* and *Halodule uninervis* were exposed to two DIC concentrations (447 μatm and 1077 μatm $p\text{CO}_2$), and three light treatments (35, 100, 380 $\mu\text{mol m}^{-2} \text{s}^{-1}$) for two weeks. DIC uptake mechanisms were separately examined by measuring net photosynthetic rates while subjecting *C. serrulata* and *H. uninervis* to changes in light and addition of bicarbonate (HCO_3^-) use inhibitors (carbonic anhydrase inhibitor, acetazolamide) and TRIS buffer (pH 8.0). We observed a strong dependence on energy driven $\text{H}^+ - \text{HCO}_3^-$ co-transport (TRIS, which disrupts H^+ extrusion) in *C. serrulata* under all light levels, indicating greater CO_2 dependence in low light. This was confirmed when, after two weeks exposure, DIC enrichment stimulated maximum photosynthetic rates (P_{max}) and efficiency (α) more in *C. serrulata* grown under lower light levels (36–60% increase) than for those in high light (4% increase). However, *C. serrulata* growth increased with both DIC enrichment and light levels. Growth, NPP and photosynthetic responses in *H. uninervis* increased with higher light treatments and were independent of DIC availability. Furthermore, *H. uninervis* was found to be more flexible in HCO_3^- uptake pathways. Here, light availability influenced productivity responses to DIC enrichment, via both carbon fixation and acquisition processes, highlighting the role of water quality in future responses to OA.

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Introduction

Seagrass meadows are highly productive habitats that offer a wide range of ecologically and economically valuable ecosystem services [1]. Especially important is their ability to capture and convert light energy into organic matter, which then become available to other trophic levels [2, 3]. This primary production is strongly determined by the amount of light available in the aquatic environment [4]. When coastal water quality declines, eutrophication and high turbidity in the water column reduce the light available to these primary producers [5]. Chronic

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light limitation had been shown to lower seagrass productivity and contribute to the accelerating areal loss of these habitats [6–8].

Seagrass productivity may also be limited by the present day seawater composition of dissolved inorganic carbon (DIC) [9, 10]. However, with the rising emissions of carbon dioxide (CO_2) from anthropogenic activities being continually absorbed by the ocean, there is a shift in the relative proportion of each DIC species (HCO_3^- , CO_3^{2-} , CO_2) in seawater [11, 12] as the pH is reduced (Ocean acidification, or OA). Importantly, the proportion of CO_2 will have the greatest percent increase (>250%, compared to 24% for HCO_3^-) among the DIC constituents under the projected pH decrease by 2100 [13]. Studies had indicated that greater availability of DIC under future OA conditions may benefit seagrasses [13–15]. An increase in photosynthesis and growth following exposure to increased DIC in both tropical and temperate species was previously demonstrated [16–22]. Moreover, since tropical and temperate seagrasses displayed a higher photosynthetic affinity for CO_2 than HCO_3^- , an increased availability of CO_2 can further boost productivity [23, 24]. At natural CO_2 vent sites, growth and areal cover of seagrasses were observed to be much greater than at adjacent non- CO_2 enriched sites [15, 25]. Hence, given sufficient light and nutrient availability, seagrasses could utilise the extra provisioning of DIC to enhance productivity [9, 22, 23, 26].

Seagrasses can use bicarbonate (HCO_3^-), the dominant DIC species, for photosynthesis [27]. Bicarbonate utilization involves the dehydration and transport of HCO_3^- into the plant cells [27, 28]. Various HCO_3^- utilization pathways that have been proposed involved the enzyme carbonic anhydrase (CA) and the presence of acidic zones maintained by proton (H^+) gradients [26, 28, 29]. Extracellular CA dehydrates HCO_3^- to CO_2 , allowing CO_2 to diffuse into the cell. Another uptake pathway involves H^+ extrusion-driven co-transport of H^+ and HCO_3^- [30]. In general, active extrusion of H^+ into localized regions of the leaf boundary layer (acidic zones) for HCO_3^- utilisation is energetically costly compared to passive CO_2 uptake, and thus could be limited by low light levels [22].

Light availability can influence the ability of seagrasses to exploit enriched DIC conditions for photosynthesis. Fluctuating light availability [31–33] could vary the carbon demand for the downstream carbon fixation cycle. On the other hand, elevated DIC (with CO_2 as substrate) increased photosynthetic efficiency and reduced light requirements of seagrasses [19, 22]. So far, the interaction of light and DIC availability on seagrass photosynthesis has only been explored in a few studies [16, 19, 34–36]. Productivity responses in seagrasses to DIC enrichment under low light conditions had been variable, ranging from no change in growth rates in *Zostera marina* [16], to increased growth in *Amphibolis antarctica* [35] and enhanced photosynthesis in *Halophila ovalis*, *Cymodocea serrulata* [34] and *Thalassia hemprichii* [36]. Studies demonstrating positive effects of DIC enrichment under low light [34, 35] suggested that seagrass growing under low light may benefit more from DIC enrichment, through a lowered epiphyte load and/or reduced energetic demand from HCO_3^- use with an increase in CO_2 supply. However, it has to be noted that some of these studies did not directly measure light levels, instead using water depth [34] and epiphyte cover [36] as proxies for light reduction. It is unclear if this DIC limitation stemmed directly from a reduced HCO_3^- utilization due to low light availability, or was compounded with reduced mixing or competitive DIC uptake by epiphytic algae.

This study aimed to examine the effects of light availability on carbon utilisation in two tropical seagrasses, *C. serrulata* and *Halodule uninervis*. Growth, net primary productivity and photosynthetic responses to DIC enrichment under different light treatment levels were characterised in a two-week aquaria experiment. DIC levels were chosen to represent present day and end-of-century ($\text{CO}_2 \sim 1000\text{ppm}$) emission scenarios (RCP8.5) [37]. It was hypothesized that while DIC enrichment (7.5% increase relative to ambient seawater) will increase photosynthesis and growth of seagrasses, the extent of increase will be greater under lower light levels.

To further test the influence of light availability on HCO_3^- use in tropical seagrasses, HCO_3^- utilization mechanisms were investigated using CA and H^+ gradient inhibitors (CA-mediated and/or H^+ co-transport mediated) under contrasting light levels [23, 38]. Energetic demand from using HCO_3^- may make it unfavourable as a photosynthetic substrate under limiting light levels. Hence the hypothesis was that under limiting light availability, the ability to use of HCO_3^- as a carbon substrate would be reduced [39, 40].

Materials and Methods

Experimental species

Two common tropical seagrass species, *Cymodocea serrulata* and *Halodule uninervis*, were collected from an intertidal meadow at Cockle Bay, Magnetic Island, northern Great Barrier Reef (19°10.88'S, 146°50.63'E). Photosynthetically active radiation (PAR) was measured at Cockle Bay using planar irradiance collectors (Odyssey Photosynthetic Irradiance Recording System, Dataflow Systems Pty Ltd, New Zealand) installed at seagrass canopy height from 2009 to 2014. The mean integrated daily irradiance at Cockle Bay from September 2009 to July 2014 was $14.9 \text{ mol m}^{-2} \text{ d}^{-1}$. Calculated over the light period, the average PAR at this site was $385 \text{ } \mu\text{mol m}^{-2} \text{ s}^{-1}$, the average maximum PAR was $961 \text{ } \mu\text{mol m}^{-2} \text{ s}^{-1}$, while the median PAR was $196 \text{ } \mu\text{mol m}^{-2} \text{ s}^{-1}$ [41]. Variation in light levels between the wet and dry season was low (S1 Fig). Average water temperature at this site (2005 to 2012) was around 26°C [41]. Cores of *H. uninervis* and sediment were collected as intact plugs and *C. serrulata* as intact horizontal rhizomes [21], first in July 2012 for the DIC and light aquaria study, and later in May 2014 for the inhibitors study (detailed below). On both occasions, seagrasses were collected under a limited impact research permit (MTB41), which was assessed and issued by the Department of Employment, Economic Development and Innovation (Fisheries Queensland Code MP05) for the removal of marine plants from the Great Barrier Reef Marine Park. The potted seagrasses were kept in an outdoor flow-through aquarium (1000 L) with filtered seawater ($5 \text{ } \mu\text{m}$) for a three-week acclimation period. Maximum light level in the outdoor aquarium was $350 \text{ } \mu\text{mol m}^{-2} \text{ s}^{-1}$ with seawater temperature of $23\text{--}25^\circ\text{C}$ and salinity at $35\text{--}36$ (PSS-78).

Light and DIC effects on photosynthetic and growth response

Experimental set-up. The seagrasses were exposed to three different light levels (35 , 100 and $380 \text{ } \mu\text{mol m}^{-2} \text{ s}^{-1}$) and two seawater DIC concentrations (high vs control; Table 1) in an indoor flow-through system over two weeks. Two weeks exposure were shown to be sufficient for inducing photosynthetic changes in the two species in a previous experiment [21]. The three light levels chosen for the experiment (35 , 100 and $380 \text{ } \mu\text{mol m}^{-2} \text{ s}^{-1}$) provided 1.5 , 4.3

Table 1. Measured and calculated carbonate system parameters for high DIC and control treatments.

DIC treatment	Measured parameters					Calculated parameters					Nutrient concentrations		
	DIC ($\mu\text{mol kg}^{-1}\text{SW}$)	pH [NBS]	AT ($\mu\text{mol kg}^{-1}\text{SW}$)	Temp ($^\circ\text{C}$)	Salinity (psu)	pH [NBS]	pCO ₂ (μatm)	CO ₂ ($\mu\text{mol kg}^{-1}\text{SW}$)	HCO ₃ ⁻ ($\mu\text{mol kg}^{-1}\text{SW}$)	CO ₃ ²⁻ ($\mu\text{mol kg}^{-1}\text{SW}$)	NH ₄ ⁺ ($\mu\text{mol/L}$)	NO ₃ ⁻ ($\mu\text{mol/L}$)	NO ₂ ⁻ ($\mu\text{mol/L}$)
High	2215 (16.1)	7.85 (0.1)	2327 (8.4)	23.8 (0.3)	36.4 (0.7)	7.82 (0.04)	1077 (104.1)	31.7 (3.1)	2083 (20.6)	101 (8.2)	0.12 (0.01)	1.35 (0.11)	0.07 (0.01)
Control	2063 (9.8)	8.26 (0.1)	2327 (10.0)	23.9 (0.3)	36.4 (0.8)	8.15 (0.02)	447 (23.1)	13.1 (0.7)	1858 (14.1)	192 (7.1)	0.13 (0.01)	1.26 (0.11)	0.07 (0.01)

Temperature and pH readings in the aquaria tanks were measured with a hand held pH probe calibrated on the NBS scale. Dissolved inorganic carbon (DIC) and total alkalinity (AT) concentrations were measured from water samples taken every four days from each aquaria tank. Carbonate system parameters were calculated using USGS CO2calc software [43]. Average values and S.D. (in brackets) were given.

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and $16.4 \text{ mol m}^{-2} \text{ d}^{-1}$ of light per day over a 12:12 h light:dark cycle. The two lower light levels represented low light conditions that were less common, but still ecologically relevant at the source meadow. The highest light level ($380 \mu\text{mol m}^{-2} \text{ s}^{-1}$) provided an integrated daily irradiance which was similar to that most commonly encountered at Cockle Bay ($12\text{--}14 \text{ mol m}^{-2} \text{ d}^{-1}$). The experiment was conducted at the Australian Institute of Marine Sciences, Townsville. Eighteen glass aquaria (working volume 18 litres) with three replicates for each treatment were supplied with fresh filtered ($5 \mu\text{m}$) seawater from header tanks. One pot of each seagrass species was placed into each aquarium. Leaves were cleaned of epiphytes every two days by gentle rubbing between fingers. Light and CO_2 levels were randomly assigned to each aquarium. LED lights mounted over the top of each aquarium provided illumination to cover the full sunlight spectrum (Aqua Illumination LED, USA). The LED lights were set to provide $380 \mu\text{mol m}^{-2} \text{ s}^{-1}$. Irradiance was determined using a planar irradiance meter (LICOR, USA). Light reduction was achieved by placing neutral density shade-cloth over individual aquaria. DIC concentrations were manipulated by means of feedback-regulated CO_2 input (AquaMedic, Germany) into the header tanks, as described in [42]. In all the header and aquarium tanks, diffusers and pumps were installed to ensure thorough mixing of DIC enriched water. Additional pH and temperature measurements were taken manually (pH probe: Eutech, USA; console: Oakton, USA) and pH levels calibrated to TRIS seawater standards (Batch 10, Supplied by A. Dixon, Scripps Institute of Oceanography). Salinity was measured with a handheld refractometer. Every four days, water samples were taken from each aquarium and analysed for dissolved inorganic carbon (DIC) and total alkalinity (AT) concentrations using a Vindta 3C analyser. Measured values of DIC, AT, temperature and salinity were used to calculate carbonate system parameters in USGS CO2calc software [43]. Water samples for inorganic nutrient (NH_4^+ and NO_3^-) measurements were collected in duplicate, every five days, from each aquarium, filtered ($0.45 \mu\text{m}$) and analysed [44].

Growth measurements. Growth rates of seagrass shoots in the experiment were measured following [45]. At the start, all shoots were marked at the top of the bundle sheath with a needle. After two weeks of growth, the shoots were harvested; new tissue growth was excised and dried at 60°C for 48 hours before weighing. Biomass of new tissue growth was normalised to the total above-ground biomass in each pot to calculate relative growth rates (RGR, $\text{g g}^{-1} \text{ DW day}^{-1}$).

Photosynthesis vs irradiance curves. To characterise photosynthetic parameters, dark respiration and photosynthetic rates over a light range were measured to construct photosynthesis vs irradiance (P-E) curves two weeks after the initiation of the experiment. Oxygen consumption and evolution rates of seagrass leaves in seawater from their respective treatment aquaria (DIC enriched: $1077 \pm 104 \mu\text{atm}$; control: $447 \pm 23 \mu\text{atm}$) were monitored using optical oxygen sensors (Sensor spots-PSt3, PreSens) and a fibre-optic oxygen meter (PreSens Oxy 4) in 200 mL volume incubation chambers [21]. The chambers were incubated at 25°C water temperature in a flow-through water bath system (Lauda, Ecoline RE 106). Magnetic stirrers ($2 \text{ cm} \times 0.8 \text{ cm}$, 128 revolutions per minute) provided even mixing in each chamber. One mature epiphyte-free (rank 2) leaf was held upright in the chamber to mimic natural orientation. Respiration was measured over a 20-min period in the dark. Photosynthetic rates were measured on the same leaf over a series of light steps (10, 20, 35, 70, 100, 200, 380, 520, $600 \mu\text{mol m}^{-2} \text{ s}^{-1}$), with each light step lasting 25 min. Adjustable LED lights were used to provide the different light intensities (Aqua Illumination LED). Light spectra of LED lights are provided in S2 Fig. Incubation media in chambers were replaced with fresh filtered seawater from respective aquaria prior to measurements at these light steps: 35, 100, 380 and $600 \mu\text{mol m}^{-2} \text{ s}^{-1}$. Initial trials showed that low photosynthetic rates at low light levels, and short incubation times between replacements of incubation media at higher light levels reduced the possibility of DIC limitation during incubations (S1 Table). Dark respiration and photosynthetic rates were derived by

fitting a linear regression to the logged oxygen concentration data in the each chamber. Only steady-state measurements of respiration and photosynthesis (obtained after ~5 min) were used for regressions. After incubation, leaves were dried (60°C, 48 h) and weighed. Rates were normalised to the dry weight of the leaf. Each optical oxygen sensor was calibrated prior to use according to method described in [46].

Net primary productivity (NPP) was taken to be the photosynthetic rate measured at the respective experimental light levels (35, 100 and 380 $\mu\text{mol m}^{-2} \text{s}^{-1}$). To derive photosynthetic parameters, photosynthetic rates were fitted to the adapted hyperbolic tangent model of [47]. The model describes the linear increase (photosynthetic efficiency, α , $\text{mg O}_2 \mu\text{mol}^{-1} \text{photons}$) in photosynthetic rates with irradiance, up until the saturating irradiance (E_k , $\mu\text{mol m}^{-2} \text{s}^{-1}$) where photosynthesis plateaus at the maximum rate (P_{max} , $\text{mg O}_2 \text{g}^{-1} \text{DW h}^{-1}$). Compensation irradiance (E_c , $\mu\text{mol m}^{-2} \text{s}^{-1}$) is the light level when photosynthetic rate is equal to respiration rate.

Effect of light levels on HCO_3^- uptake

To assess the effect of light on HCO_3^- utilization, net photosynthesis under specific inhibition of HCO_3^- uptake mechanisms was measured under two contrasting light levels. The light levels represented limiting (40 $\mu\text{mol m}^{-2} \text{s}^{-1}$) and saturating (600 $\mu\text{mol m}^{-2} \text{s}^{-1}$) light as determined from the previous experiment where the derived saturating light (E_k) was 100 and 98 $\mu\text{mol m}^{-2} \text{s}^{-1}$ for *C. serrulata* and *H. uninervis* respectively. Therefore, the limiting light level was well below E_k and the saturating light level was above E_k for both species.

Bicarbonate utilization pathways can be elucidated by the effect of specific inhibitors on photosynthetic rates, as described in [28]. One pathway involves the membrane-bound extracellular CA dehydrating HCO_3^- into CO_2 , which then diffuses into the cell. Addition of acetazolamide (AZ) inhibits extracellular CA activity [28]. Another possible pathway for HCO_3^- utilization consists of localised active H^+ extrusion to create H^+ gradients that facilitate the inward co-transport of H^+ and HCO_3^- . Addition of a buffer, e.g. Tris (hydroxymethyl) aminomethane (TRIS), dissipates the H^+ gradient as the buffer reacts with the extruded H^+ , thus altering photosynthetic rates [27]. A third HCO_3^- utilization pathway consists of extracellular CA-mediated HCO_3^- conversion to CO_2 in acidic zones created at H^+ extrusion sites. The acidic zones help concentrate CO_2 and facilitate diffusion into the cells. Strong inhibition of photosynthetic rates by the combined addition of buffer and AZ indicates a strong dependence on this pathway [29, 48]. Thus, we used AZ, TRIS and their combination in the experiments described below.

Net photosynthesis was determined by measuring rates of oxygen evolution in 60mL incubation chambers, similar to the procedure described above. During measurements, incubation media (fresh filtered seawater (pH 8.0) with or without inhibitor) in each chamber were stirred and maintained at 25°C. Mature and non-epiphytised leaves from non-connected shoots collected fresh from the field, were used for the measurements. Prior to the addition of the inhibitor(s), control rates of dark respiration and photosynthesis for each leaf were measured in fresh filtered seawater (pH 8.0). Dark respiration rates of the leaves were measured over 20 min, before the lights were switched on to provide 40 $\mu\text{mol m}^{-2} \text{s}^{-1}$ of light. Net photosynthesis was then measured for 30 min. Subsequently, light levels were increased to 600 $\mu\text{mol m}^{-2} \text{s}^{-1}$ and photosynthetic rates of the same leaves were measured. After the control measurements, incubation media for all chambers were replaced with fresh filtered seawater mixed with inhibitor solutions (AZ, TRIS or AZ+TRIS). The second set of incubations used the same leaves and followed the same sequence of measurements (in dark, at limiting light, then at saturating light). Oxygen production and consumption rates were calculated and normalised to leaf dry weight as above. For this experiment, net photosynthetic rates to TRIS and/or AZ were expressed as percentages of the control net photosynthetic rates.

Three inhibitor solutions were used: 1) 0.1 mM AZ, 2) 45 mM TRIS, and 3) combination of 0.1 mM AZ and 45 mM TRIS [38]. A stock solution of 20 mM AZ was prepared by dissolving the powder in 50 mM sodium hydroxide (NaOH). An aliquot of 350 μ l AZ stock solution was added to each individual chamber to achieve a final concentration of 0.1 mM. TRIS was prepared as 1 M stock solution and adjusted to pH 8.0. The buffer yielded pH 8.0 when mixed with seawater to a final concentration of 45 mM. The final TRIS concentration used here, although was lower than that commonly found in literature [22, 38], was sufficient in eliciting a reduction in net photosynthetic rates.

Statistical analyses

To evaluate effects of DIC and light for each species, growth, net primary productivity and photosynthetic parameters were analysed using univariate two-factor ANOVA to test for significance in fixed effects of light (three levels) and DIC (two levels) and their interaction. Data were checked for homogeneity of variance using Levene's test and for normality using Q-Q plots. Where the assumptions of ANOVA were violated, data were square root transformed. However when the assumptions were still not met, as observed in the E_c and α data for *H. uninervis*, the alpha-value was lowered to 0.01 to minimize the risk of a Type I error [49]. In case of significant light effects, Tukey's HSD was carried out to test which of the treatment levels were significantly different from each other. Similarly, Tukey's HSD was used to test for significance between selected groups in case of significant light and DIC interactions.

To determine if the addition of AZ and/or TRIS significantly affects net productivity of each seagrass species in fresh seawater (control), raw net productivity data were compared using paired t-tests. Further, to test if light levels affect HCO_3^- utilization, paired t-tests were used to compare the change in percentage net photosynthesis between the two light levels for each inhibitor type. Arcsine transformation was not performed on the the percentage net photosynthesis data, since the data exceeded the range of 0 to 100. Instead, percentage net photosynthesis responses were square root transformed prior to analysis to meet the assumption of normality for the test. All statistical analyses were conducted using R statistical software [50].

Results

Light and DIC effects on photosynthetic and growth response

Experimental parameters. Water temperature and salinity in the experimental aquaria did not vary considerably throughout the experiment (average \pm S.D.: $23.9 \pm 0.3^\circ\text{C}$ and 35.9 ± 0.5 (PSS-78)). Manipulation of the CO_2 system resulted in a 140% increase in CO_2 concentration, a 10% increase in HCO_3^- concentration, and a corresponding decrease of ~ 0.3 units in pH, compared to present day control conditions (Table 1). Carbonate system parameters of the DIC treatments remained in the target range. Nutrient concentrations were similar between aquarium tanks; with an average (\pm S.D.) water column ammonium concentration of $0.12 \pm 0.03 \mu\text{M}$ and nitrate concentration of $1.30 \pm 0.55 \mu\text{M}$.

Productivity responses to light and DIC enrichment. For *C. serrulata*, DIC enrichment and light availability both increased relative growth rates (RGR) significantly (ANOVA: DIC $p = 0.015$; light $p = 0.002$) (Table 2; Fig 1A). When light levels were at $100 \mu\text{mol m}^{-2} \text{s}^{-1}$ and $380 \mu\text{mol m}^{-2} \text{s}^{-1}$, RGRs increased by 20% and 26% respectively, relative to that at $35 \mu\text{mol m}^{-2} \text{s}^{-1}$ (Tukey HSD: $100 \mu\text{mol m}^{-2} \text{s}^{-1}$ $p = 0.015$; $380 \mu\text{mol m}^{-2} \text{s}^{-1}$: $p = 0.002$). DIC enrichment raised RGR by 13%. There was no significant interaction between light and DIC enrichment on RGR in *C. serrulata*. In *H. uninervis*, effect of DIC enrichment on RGR varied with light levels (ANOVA: $p = 0.009$) (Table 2; Fig 1B). DIC enrichment increased RGR only for seagrass growing at

100 $\mu\text{mol m}^{-2} \text{s}^{-1}$ (Tukey HSD: $p = 0.027$), but not at lower (Tukey HSD: $p = 0.955$) or higher (ANOVA: $p = 0.905$) light levels.

In *C. serrulata*, net primary productivity (NPP) was significantly influenced by light levels (ANOVA: $p < 0.001$) (Table 2; Fig 1C). DIC enrichment did not significantly raise NPP (ANOVA: $p = 0.312$) (Table 2). NPP in *C. serrulata* increased with light, by 12% (at 100 $\mu\text{mol m}^{-2} \text{s}^{-1}$) and 29% (at 380 $\mu\text{mol m}^{-2} \text{s}^{-1}$) (Tukey HSD: 100 $\mu\text{mol m}^{-2} \text{s}^{-1}$: $p < 0.001$; 380 $\mu\text{mol m}^{-2} \text{s}^{-1}$: $p < 0.001$). Similarly for *H. uninervis*, NPP increased significantly with light but not with DIC enrichment (ANOVA: light $p < 0.001$; DIC $p = 0.713$) (Table 2; Fig 1D). NPP increased by 13 to 22% in *H. uninervis* with light (Tukey HSD: 100 $\mu\text{mol m}^{-2} \text{s}^{-1}$: $p = 0.002$; 380 $\mu\text{mol m}^{-2} \text{s}^{-1}$: $p < 0.001$). Dark respiration rates did not vary with light treatments or DIC in *C. serrulata* (Table 2). In *H. uninervis*, dark respiration rates responded to light treatments only (ANOVA: $p = 0.031$). Dark respiration rates increased by 67% at 380 $\mu\text{mol m}^{-2} \text{s}^{-1}$ relative to at 35 $\mu\text{mol m}^{-2} \text{s}^{-1}$ (Tukey HSD: $p = 0.032$).

Photosynthetic-irradiance (P-E) curves. The adapted hyperbolic tangent model provided a good fit for all P-E curves ($R^2 > 0.85$; $p < 0.050$). Photosynthetic rates increased linearly (initial slope, α) with irradiance before plateauing off at the maximum photosynthetic rate (P_{max}) above saturating irradiance (E_k).

The increase in maximal photosynthetic rates (P_{max}) in *C. serrulata* with DIC enrichment depended on light levels (ANOVA: $p = 0.049$) (Table 2; Fig 2A). The observed increase in P_{max} due to DIC enrichment became smaller with increasing light availability (Fig 2). Post-hoc tests

Table 2. Two-way ANOVA results.

		<i>Cymodocea serrulata</i>				<i>Halodule uninervis</i>			
		df	MS	F	P	df	MS	F	p
RGR	DIC	1	1.07×10^{-3}	8.115	0.015	1	8.94×10^{-6}	1.189	0.297
	Light	2	1.43×10^{-3}	10.793	0.002	2	5.63×10^{-5}	7.488	0.008
	DIC x light	2	3.84×10^{-5}	0.290	0.753	2	5.45×10^{-5}	7.238	0.009
NPP	DIC	1	3.400	1.116	0.312	1	1.600	0.142	0.713
	Light	2	389.100	126.762	8.52×10^{-9}	2	320.800	29.000	2.54×10^{-5}
	DIC x light	2	5.200	1.705	0.223	2	0.100	0.006	0.994
Dark respiration	DIC	1	18.924	2.033	0.179	1	0.002	0.003	0.957
	Light	2	23.395	2.513	0.123	2	2.584	4.697	0.031
	DIC x light	2	0.163	0.018	0.983	2	0.545	0.991	0.400
P_{max}	DIC	1	60.320	27.800	1.97×10^{-4}	1	4.320	0.514	0.487
	Light	2	190.160	87.630	6.92×10^{-8}	2	126.060	14.992	5.45×10^{-4}
	DIC x light	2	8.500	3.920	0.049	2	2.550	0.304	0.744
E_k	DIC	1	8181	39.080	4.25×10^{-5}	1	150	0.293	0.598
	Light	2	3475	16.597	3.50×10^{-4}	2	3668	7.149	0.009
	DIC x light	2	1132	5.409	0.021	2	2547	4.964	0.027
E_c	DIC	1	21.500	0.204	0.660	1	1.315	0.789	0.392
	Light	2	534.500	5.053	0.026	2	8.073	4.843	0.029
	DIC x light	2	29.200	0.276	0.763	2	8.977	5.385	0.021
α	DIC	1	1.53×10^{-2}	5.513	0.037	1	1.14×10^{-2}	2.413	0.146
	Light	2	2.24×10^{-4}	0.081	0.923	2	2.22×10^{-3}	0.469	0.637
	DIC x light	2	1.91×10^{-3}	0.686	0.522	2	0.024	5.049	0.026

All parameters were analysed with DIC and light treatments as fixed factors. N = 3. Significant p-values are in bold. Prior analysis, square-root transformation had been applied to RGR for *Cymodocea serrulata*, and to E_c , α and respiration for *Halodule uninervis*. RGR- relative growth rate, P_{max} - maximum photosynthetic rate, E_k - saturating irradiance, E_c - compensation irradiance, α - photosynthetic efficiency.

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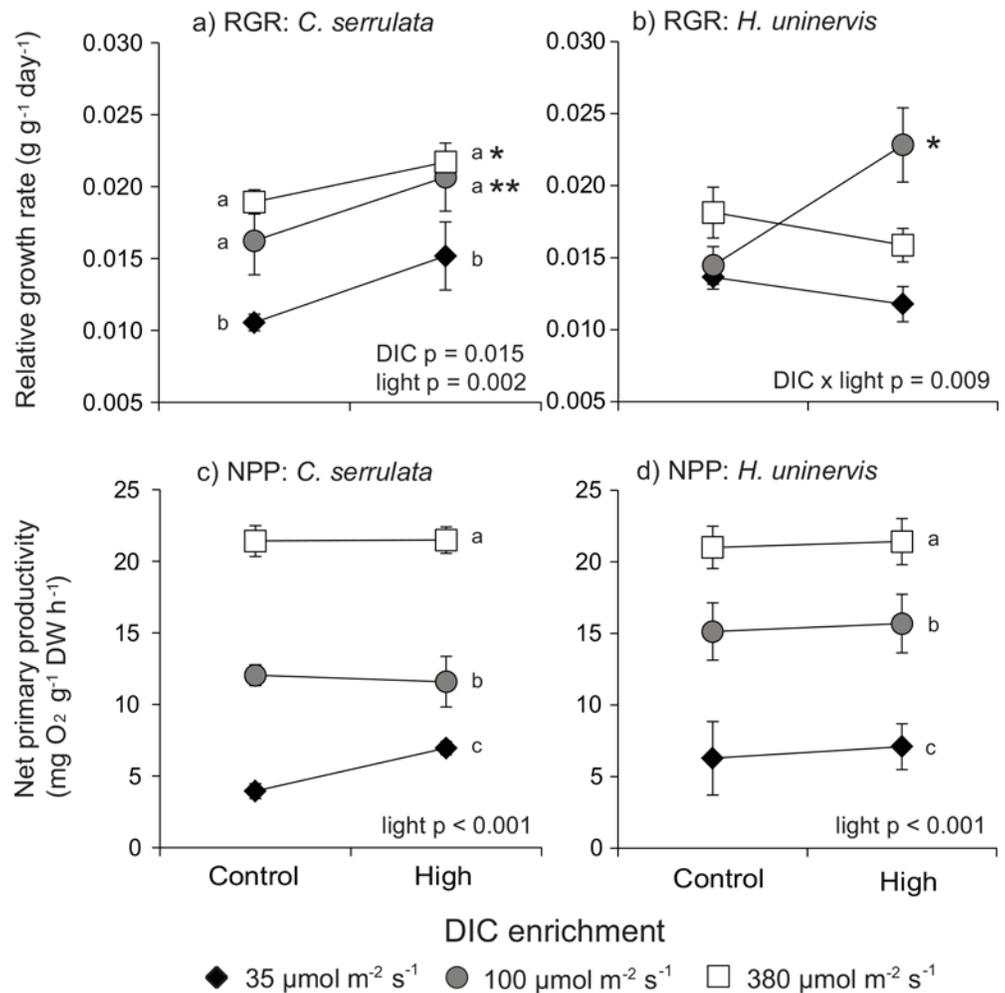


Fig 1. Comparison of relative growth rates and net primary productivity of *C. serrulata* and *H. uninervis* after two weeks exposure to light and DIC. (a-b) RGR- relative growth rate, (c-d) NPP-net primary productivity. Letters indicate significant differences between light treatments based on Tukey's HSD test; asterisks indicate significant differences between DIC treatments based on ANOVA results where there is no interaction between light and DIC, or based on Tukey's HSD test if an interaction was detected (* $p < 0.05$; ** $p < 0.01$). Control DIC = 2063 μM; high DIC = 2215 μM. Means (± S.E.) are given (n = 3).

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indicated that P_{max} significantly increased with DIC in seagrasses growing at 35 μmol m⁻² s⁻¹ (60% increase) and 100 μmol m⁻² s⁻¹ (36% increase) (Tukey HSD: 35 μmol m⁻² s⁻¹: $p = 0.014$; 100 μmol m⁻² s⁻¹: $p = 0.011$). There was no significant increase in P_{max} at 380 μmol m⁻² s⁻¹ ($p = 0.969$). For *H. uninervis*, P_{max} increased with light treatments but not with DIC enrichment (ANOVA: light $p < 0.001$; DIC $p = 0.487$) (Table 2; Fig 2B). P_{max} was significantly higher at 380 μmol m⁻² s⁻¹ than at 35 (71% increase) and 100 μmol m⁻² s⁻¹ (35% increase) (Tukey HSD: 35 μmol m⁻² s⁻¹: $p < 0.001$; 100 μmol m⁻² s⁻¹: $p = 0.014$).

The response in E_k to DIC enrichment was dependent on light levels in *C. serrulata*, as shown by a significant light × DIC interaction (ANOVA: $p = 0.021$) (Table 2; Fig 2C). DIC enrichment resulted in a greater increase in E_k at lower light levels (Tukey HSD: 35 μmol m⁻² s⁻¹: $p = 0.051$; 100 μmol m⁻² s⁻¹: $p < 0.001$) than at high light levels (Tukey HSD: $p = 0.713$) (Fig 2). For *H. uninervis*, there was an interactive effect of light and DIC enrichment on E_k (ANOVA: $p = 0.027$) (Table 2; Fig 2D). In low light, DIC enrichment reduced E_k by 43% (Tukey HSD:

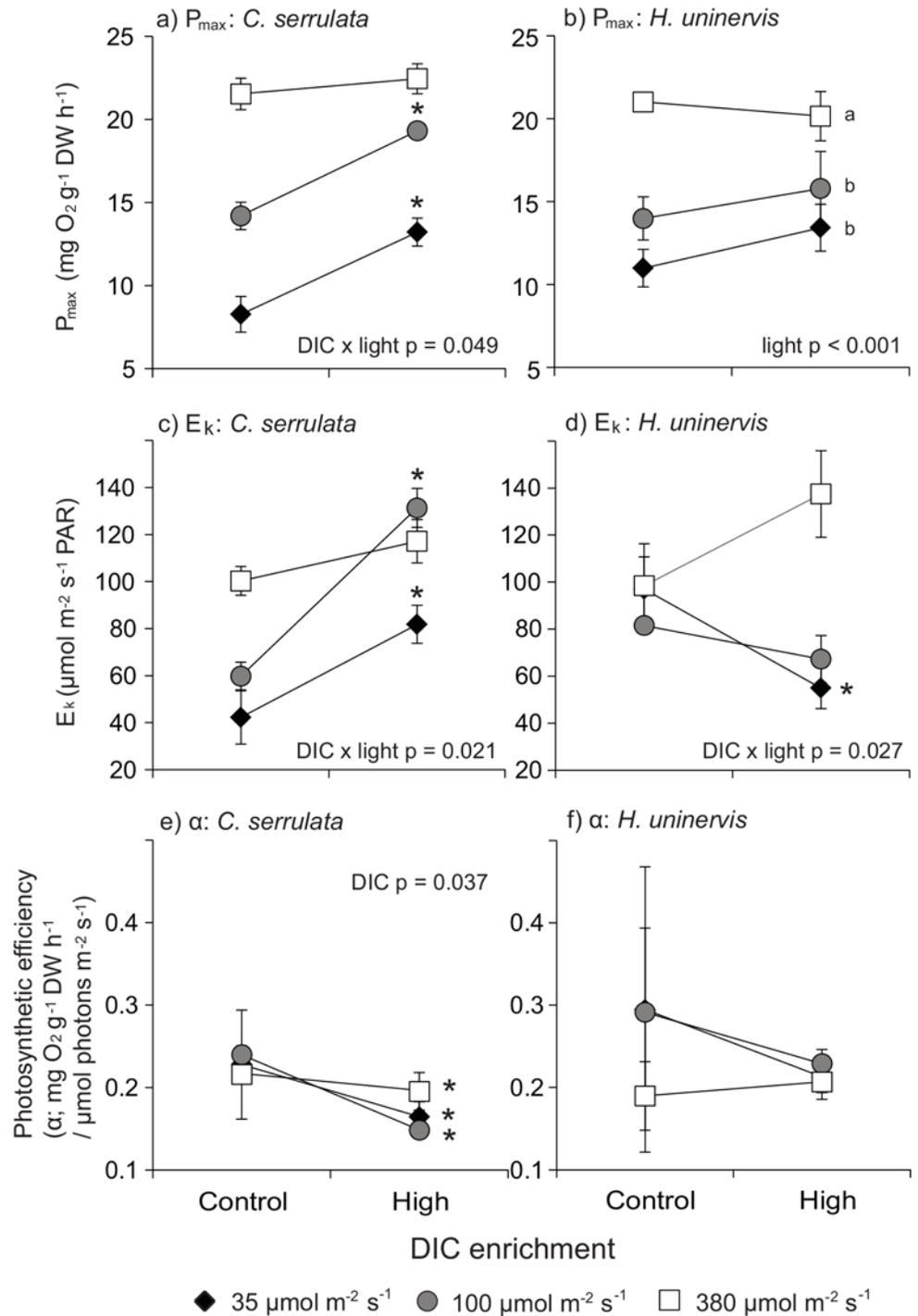


Fig 2. Comparison of photosynthetic parameters between light and DIC for *C. serrulata* and *H. uninervis* after two weeks exposure. (a-b) P_{max} —maximal photosynthetic rate, (c-d) E_k —saturating irradiances, (e-f) α —photosynthetic efficiency. Letters indicate significant differences between light treatments based on Tukey's HSD test; asterisks indicate significant differences between DIC treatments based on ANOVA results where there is no interaction between light and DIC, or based on Tukey's HSD test if an interaction was detected (* $p < 0.05$). Control DIC = 2063 μM ; high DIC = 2215 μM . Means (\pm S.E.) are given ($n = 3$).

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$p = 0.028$). However, at and above saturating light (100 and $380 \mu\text{mol m}^{-2} \text{s}^{-1}$) DIC enrichment had no significant effect on E_k .

Compensation irradiance (E_c) in *C. serrulata* varied with light (ANOVA: $p = 0.026$) but not with DIC enrichment (Table 2). At $380 \mu\text{mol m}^{-2} \text{s}^{-1}$, E_c was 102% higher than at $35 \mu\text{mol m}^{-2} \text{s}^{-1}$ (Tukey HSD: $p = 0.020$), but was not significantly different from that at $100 \mu\text{mol m}^{-2} \text{s}^{-1}$ (Tukey HSD: $p = 0.236$). In *H. uninervis*, there was no main or interactive effect of DIC enrichment and light on E_c (ANOVA: $p = 0.021$; alpha lowered to 0.01) (Table 2).

Photosynthetic efficiency (α) was significantly reduced by DIC enrichment (ANOVA: $p = 0.037$), but not between light levels (Table 2). No interaction of DIC enrichment and light was detected for α in *C. serrulata* (Table 2; Fig 2E). Overall, DIC addition reduced α by 34% for *C. serrulata*. For *H. uninervis*, there were no main or interactive effects of light and DIC enrichment on α (ANOVA: $p = 0.026$; alpha lowered to 0.01) (Table 2; Fig 2F).

In summary, both DIC enrichment and light levels influenced the response of photosynthetic parameters in *C. serrulata*. The change in parameters such as P_{max} and E_k depended on either DIC enrichment or light levels, and also the interaction of both factors. Conversely, photosynthesis in *H. uninervis*— P_{max} and E_k —seemed to be strongly influenced by light treatment but was independent of DIC enrichment.

Effects of light levels on HCO_3^- uptake. In *C. serrulata*, addition of AZ significantly reduced net photosynthesis (paired t-test: $t = 4.261$, $df = 11$, $p < 0.001$) (Fig 3A), indicating that activity of external CA is an important mechanism aiding in HCO_3^- uptake. The reduction in net photosynthesis due to the inhibition of CA-catalysed conversion of HCO_3^- (AZ treatment) was not significantly different between light levels (paired t-test: $t = 1.851$, $df = 5$, $p = 0.123$) (Fig 3A). Addition of TRIS to *C. serrulata* also resulted in a strong reduction in net photosynthesis (paired t-test: $t = 3.962$, $df = 11$, $p = 0.001$) (Fig 3A). This suggests that presence of the H^+ gradient is essential for HCO_3^- transport into the cell. Dissipation of H^+ gradient alone (TRIS addition) resulted in a higher reduction in net photosynthesis under saturating light (>90%), compared to that under limiting light (60%) (paired t-test: $t = 10.126$, $df = 5$,

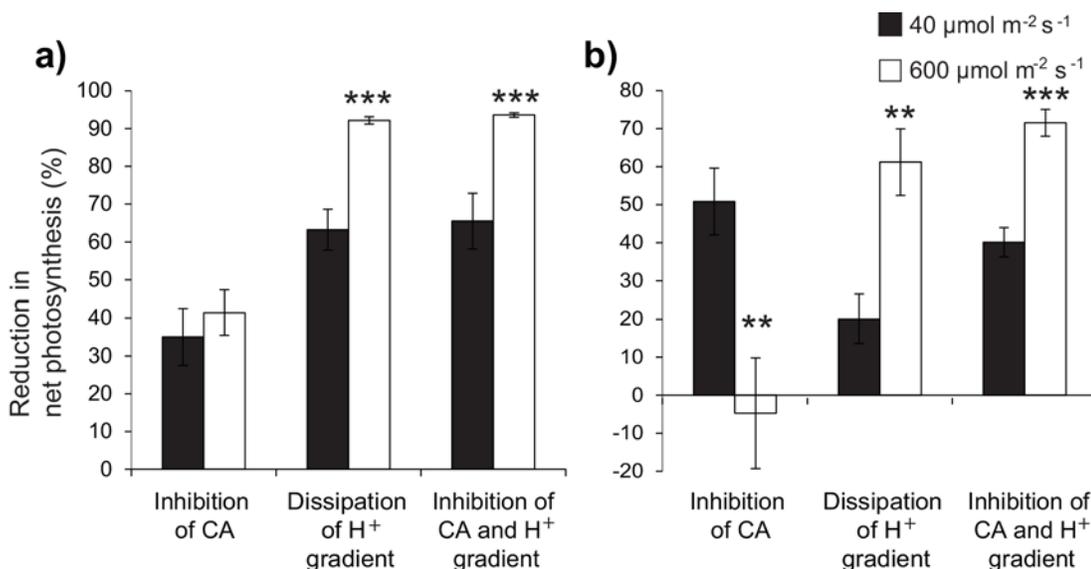


Fig 3. Reduction in net photosynthetic responses of (a) *C. serrulata* and (b) *H. uninervis*, when subjected to HCO_3^- uptake inhibitors. Net photosynthetic responses were expressed relative to control rates in normal seawater. 0.1 mM acetazolamide (inhibition of extracellular CA), 45 mM TRIS buffer at pH 8.0 (dissipation of H^+ gradient), and a combination of TRIS and acetazolamide (inhibition of CA and H^+ gradient). Asterisks indicate significant differences between light treatments for each inhibitor or inhibitor combination (** $p < 0.01$; *** $p < 0.001$). Means (\pm S.E) are given ($n = 6$).

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$p < 0.001$) (Fig 3A). When both CA and H^+ extrusion were inhibited (addition of both AZ and TRIS), net photosynthesis in *C. serrulata* was reduced significantly (paired t-test: $t = 4.095$, $df = 11$, $p < 0.001$) (Fig 3A). Net photosynthesis was reduced to a greater extent under saturating light than under limiting light (paired t-test: $t = 7.855$, $df = 5$, $p < 0.001$) (Fig 3A).

In *H. uninervis*, inhibition of extracellular CA (AZ treatment) reduced net photosynthesis by 50% in limiting light (paired t-test: $t = -4.188$, $df = 5$, $p = 0.009$) (Fig 3B), but had no effect in saturating light. This indicated that when light is limiting, CA played a more significant role in HCO_3^- uptake. The dissipation of H^+ gradient (TRIS addition) decreased net photosynthesis in *H. uninervis* (paired t-test: $t = 2.755$, $df = 11$, $p < 0.001$) (Fig 3B). Net photosynthesis decreased more under saturating light conditions (60%) than under low light (20%) (paired t-test: $t = 4.380$, $df = 5$, $p = 0.007$) (Fig 3B). Inhibition of both CA and H^+ extrusion (addition of both AZ and TRIS) lowered net photosynthesis under both light levels (paired t-test: $t = 4.079$, $df = 11$, $p < 0.001$) (Fig 3B). The extent of inhibition was greater under saturating light than in limiting light (paired t-test: $t = 8.315$, $df = 5$, $p < 0.001$).

Discussion

Light availability can affect the ability of tropical seagrass to respond to increased DIC provisioning under OA. The present study examined whether *C. serrulata* and *H. uninervis* were able to adjust growth and photosynthesis responses when exposed to an enriched DIC concentration approximating predicted end-of-century pCO_2 level (1077ppm) [37] over a range of light levels. Growth of *C. serrulata* was stimulated by both DIC enrichment and light availability while growth of *H. uninervis* was strongly influenced by light availability only. Interactive effects of DIC enrichment and light treatment were evident in P-E curve parameters in *C. serrulata*, while photosynthetic potential in *H. uninervis* was affected by the experimental light treatment. The use of CA and H^+ gradient inhibitors highlighted important differences in carbon uptake mechanisms which may explain some of the differences in responses of the species on a physiological level.

Growth and net productivity response

The experimental light treatments represented a range of ecologically relevant light conditions that occur in the natural environment. Both *C. serrulata* and *H. uninervis* were grown in light-saturated conditions under both moderate ($100 \mu mol m^{-2} s^{-1}$) and high ($380 \mu mol m^{-2} s^{-1}$) light treatments, where light levels were generally at or above E_k . The only exception to this was for *C. serrulata* at $100 \mu mol m^{-2} s^{-1}$ under DIC enrichment, in which E_k was $120 \mu mol m^{-2} s^{-1}$. In contrast, the lowest light treatment ($35 \mu mol m^{-2} s^{-1}$) was below E_k for both species under all DIC levels, and so photosynthesis was light-limited. All light treatments were above E_c , and so both seagrass species were in net carbon surplus.

For *C. serrulata*, the effect of light and DIC enrichment on growth appeared to be additive, meaning that increasing both DIC and light increased their growth rates. Thus, the highest growth rate was observed at the high light level under DIC enrichment. Further, short term exposure to DIC enrichment meant that plants growing at 35 and $100 \mu mol m^{-2} s^{-1}$ were able to grow as fast as plants without DIC enrichment at 100 and $380 \mu mol m^{-2} s^{-1}$ respectively, such that DIC enrichment somewhat compensated for lower light availability [19]. While a previous *ex-situ* study showed that growth rates of *C. serrulata* did not respond to CO_2 enrichment [21], our current study showed an increase in growth rates for this species with DIC enrichment, albeit at a very modest 13%. Higher increases in shoot density (194–350%) and above-ground biomass (32–987%) of *C. serrulata* were observed in natural CO_2 vent sites with considerably greater DIC enrichment, compared to adjacent non- CO_2 enriched sites [51]. Net primary production of *C.*

serrulata in this study appeared to be strongly limited by light, not DIC concentration. However, previous studies had indicated an increase in net primary productivity with CO₂ enrichment in this species [21, 51]. Here, the results suggest that light availability plays an upstream role, relative to DIC, in the hierarchical control on seagrass photosynthesis.

Net production and growth in *H. uninervis* did not appear to be DIC-limited, as it was less sensitive to an increase in DIC than *C. serrulata* under all light treatments. However, previous work has shown that the same population of *H. uninervis* can respond to DIC addition by increasing net productivity and growth under similar treatment conditions [21]. Seasonal variation in carbon demand for growth and metabolism might have contributed to the observed differences in response to DIC enrichment between studies [52, 53].

Both DIC enrichment and light availability had been known to stimulate productivity and growth in seagrasses [17, 19, 22, 54]. However, while *C. serrulata* increased growth rates with DIC enrichment and light availability, *H. uninervis* did not demonstrate a growth response to DIC enrichment. The growth response of *H. uninervis* here was limited by light availability, consistent with its net productivity response. Growth responses to DIC enrichment can also be influenced by nutrient availability [55] and water temperature [56]. Sediment pore water nutrients were not measured in the present study and hence it was not possible to assess if overall nutrient availability was limiting seagrass productivity. Knowledge of the interactive effects of environmental factors (light, temperature, nutrients) with DIC enrichment is needed to predict future seagrass productivity responses in the field.

Photosynthetic potential

Photosynthetic response of *C. serrulata* to DIC enrichment depended on treatment light levels. Photosynthetic capacity (P_{\max}) was higher in *C. serrulata* exposed to higher DIC levels for two weeks. *C. serrulata* at similar CO₂ and light conditions to those used here—DIC enrichment and at 400 $\mu\text{mol m}^{-2} \text{s}^{-1}$ —did increase P_{\max} by ~20% [21]. Increases in P_{\max} in response to CO₂ enrichment have also been observed in *Z. marina* [19] and *Z. noltii* [18]. A larger increase in P_{\max} was observed in plants from the limiting (60% increase) compared to the saturating light treatments (0% increase). DIC enrichment can enhance maximum photosynthetic capacity by providing more substrate for fixation and simultaneously lowering photorespiration rates [57]. Light increases maximum photosynthetic capacity by boosting the production of reducing intermediates (e.g. NADPH and ATP) for the carbon reduction cycle [58]. In theory, since both factors have independent mode of actions, their combined effect should be synergistic [59, 60]. The combined sub-additive effect on P_{\max} observed in results suggested that the extent of DIC limitation was greater under low light than high light.

Saturating irradiance (E_k) in *C. serrulata* increased with DIC enrichment, with a greater rise in E_k observed at lower light levels. Higher saturating light requirements could be driven by the higher photosynthetic capacity due to greater DIC availability, as similarly observed in *Thalassia hemprichii* [17]. The lowering of photosynthetic efficiency with DIC enrichment was unexpected, as increased CO₂ availability would mean less resources (i.e. ATP) were needed to procure HCO₃⁻ for photosynthesis [39]. Under CO₂ enrichment, *Zostera marina* tripled the rate of light-saturated photosynthesis (i.e. P_{\max}) to reduce the daily photoperiod required for a positive carbon balance [19]. Overall, the studies conducted so far suggest that while DIC enrichment increases the intensity of saturating irradiance required to reach maximum photosynthetic rates, it also reduces the daily period of saturating irradiance required to achieve a net carbon surplus.

Photosynthesis in *H. uninervis*, on the other hand, was limited by light, and not by DIC availability. Photosynthetic capacity (P_{\max}) and E_k increased with increasing light levels.

Interestingly, increased DIC concentration lowered E_k for *H. uninervis* shoots growing under low light (35 and 100 $\mu\text{mol m}^{-2} \text{s}^{-1}$). This implies that DIC enrichment could, to a certain extent, compensate for low light levels by reducing light requirements in this species. However, the increase in productivity with light availability was higher than the increase with DIC enrichment [61], with maximum photosynthetic rates remaining the highest under high light treatment. Similarly in *Z. marina*, CO_2 enrichment only increased shoot production and below-ground biomass under light-replete but not light-deplete treatments [16].

Effects of light on DIC utilization

Normally the supply of CO_2 , the preferred DIC species for seagrasses [23, 24], is limited by low free CO_2 concentration and diffusion rates, and slow conversion from HCO_3^- to CO_2 [62]. While most seagrasses can utilise HCO_3^- as a DIC source [26, 27] and the concentration of total DIC is non-limiting, the high energetic cost of HCO_3^- uptake makes it a less preferred substrate under low light levels [28, 63]. Light fuels the generation of ATP for both carbon fixation and HCO_3^- uptake [48, 58]. This may explain the apparent paradox that *C. serrulata* in our experiment was more DIC-limited at lower light levels than at higher levels. This finding was consistent with the postulation that HCO_3^- utilization is limited at lower light levels [34]. In [34], both deep water (i.e. low light) *Halophila ovalis* and *C. serrulata* showed a greater increase in relative electron transport rates (100% and 66% respectively) compared to their intertidal (i.e. high light) counterparts (30% and 20% respectively) when subjected to an 180% increase in DIC concentration. Low light availability can lower HCO_3^- utilization, and OA conditions could boost DIC supply by providing more dissolved CO_2 [22, 64].

Application of CA and H^+ gradient inhibitors showed that, in general, both CA and H^+ gradients are important mechanisms to allow utilisation of HCO_3^- as a carbon source for the two species investigated. This was observed previously in several other seagrass species [22, 28, 38]. For *C. serrulata*, the inhibition of carbonic anhydrase (CA) and H^+ extrusion both reduced net photosynthesis. The strong dependence on H^+ extrusion for HCO_3^- utilisation in *C. serrulata* meant that adequate light levels were needed before HCO_3^- can be used efficiently for photosynthesis. Therefore, at limiting light levels this species probably depends more on CO_2 diffusion. CA-mediated HCO_3^- dehydration in acidic zones was thought to be a more efficient means of HCO_3^- utilization than CA conversion alone, as the CO_2 concentration at equilibrium is higher within acidic zones than at normal seawater pH, thus driving the inward diffusion of CO_2 [28]. Despite this, [26] and [38] demonstrated that extracellular CA-catalysed HCO_3^- conversion to CO_2 (without H^+ extrusion sites) was enough to support photosynthetic demand in *C. serrulata*. Differences between their findings and ours could be due to long term acclimation of conspecifics to different source meadow environments, or that there were genotypic variation between seagrass populations [65, 66].

Halodule uninervis appeared to vary HCO_3^- uptake mechanisms, enabling it to use HCO_3^- over a wider light range for photosynthesis. Under low light conditions, CA-mediated conversion of HCO_3^- contributed substantially to the carbon supply for photosynthesis [28]; when light became saturating, HCO_3^- uptake was supplemented by H^+ co-transport with HCO_3^- . Extracellular CA-mediation of HCO_3^- uptake is less likely to depend on light availability, when compared to the H^+ extrusion-driven co-transport of HCO_3^- [26, 29]. This flexibility between HCO_3^- utilization pathways suggested *H. uninervis* was able to mediate, to a certain extent, DIC limitation under low light conditions. Similarly, efficient use of HCO_3^- utilization mechanisms was thought to account for the lack of photosynthetic responses to DIC enrichment in marine macroalgae [67].

Differential sensitivity to photosynthetic carbon between seagrass species could be due to species variation in DIC utilization mechanisms [38, 64] and extent of carbon-limitation [9, 64]. Species such *C. serrulata* (this study) and *Thalassia testudinum* [64] would benefit more from increased CO₂ supply under OA conditions since they were limited in their ability to utilise the dominant HCO₃⁻ in seawater. Less DIC limited species, like *H. uninervis* (this study), *Halodule wrightii* and *Syringodium filiforme* might show a smaller response to DIC enrichment [64].

Light and OA as drivers of seagrass productivity

Under future scenarios of OA, marine macrophytes like seagrass could benefit, from both increased DIC concentration and a proportional rise in CO₂ [15, 25]. While short-term (e.g. this study) and long-term [68] studies have documented varying degrees of DIC limitation in seagrasses, physiological processes that could regulate responses to increased DIC over intermediate time-scales remain unexplored for seagrasses. Epiphytic algae may also benefit from higher CO₂ [35], and in turn compete with seagrasses for the same resources. Their effects would vary with their composition, biomass and the turnover rates of seagrass leaves [69]. Furthermore, many seagrass habitats are primarily light-limited [8]. The range of light levels used in this experiment, representing the recent light exposure history (~ five years) of the seagrasses at their source meadow, was already much reduced compared to pre-European settlement due to a four-times increase in sediment runoff [70]. Reduced light availability, due to increased sedimentation and epiphytic algal growth, can negate positive OA effects on seagrass growth [35]. Our study showed that the rise in light availability elicited a greater increase in seagrass productivity than DIC enrichment. Compared to OA, variation in water clarity occurs over shorter frequencies and with greater intensities, thus playing a more imminent role in controlling seagrass productivity. Hence, to ensure continued productivity in seagrass meadows in the future, changes in water quality and OA has to be studied and managed in unison.

Supporting Information

S1 Fig. Distribution of daily light sums (photosynthetically active radiation levels) at Cockle Bay, Magnetic Island, Great Barrier Reef during the dry and wet seasons. Distribution of PAR was measured at seagrass canopy height using planar 2π PAR loggers from September 2009 to July 2014.
(EPS)

S2 Fig. Light spectra of Aqua Illumination LED Sol at different settings. At 100% blue channel only (blue), 100% white channel only (orange) and 100% both blue and white channels (black). Light spectra were measured using Jazz spectrometer on SpectraSuite software (OceanOptics, USA).
(JPG)

S1 Table. Average incubation time, and approximate time range for DIC limitation to occur in 200 mL of ambient seawater during trials, for each light step of P-E curve. Leaf material used during trials was 5–12 mg in dry weight. Light steps in bold indicate a replacement of fresh filtered seawater prior to incubation during actual measurements.
(DOCX)

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Author Contributions

Conceived and designed the experiments: YO SU CC. Performed the experiments: YO. Analyzed the data: YO SU CC. Contributed reagents/materials/analysis tools: SU CC. Wrote the paper: YO SU CC.

References

1. Costanza R, d'Arge R, de Groot R, Farber S, Grasso M, Hannon B, et al. The value of the world's ecosystem services and natural capital. *Nature*. 1997; 387:253–60.
2. Hemminga MA, Duarte CM. *Seagrass ecology*. 1st ed. Cambridge: Cambridge University Press; 2000.
3. Duarte CM, Chiscano CL. Seagrass biomass and production: a reassessment. *Aqua Bot*. 1999; 65:159–74. doi: [10.1016/S0304-3770\(99\)00038-8](https://doi.org/10.1016/S0304-3770(99)00038-8)
4. Dennison WC. Effects of Light in Seagrass Photosynthesis, Growth and Depth Distribution. *Aqua Bot*. 1987; 27:15–26. doi: [10.1016/0304-3770\(87\)90083-0](https://doi.org/10.1016/0304-3770(87)90083-0)
5. Schaffelke B, Mellors J, Duke NC. Water quality in the Great Barrier Reef region: responses of mangrove, seagrass and macroalgal communities. *Mar Poll Bull*. 2005; 51(1–4):279–96. doi: [10.1016/j.marpolbul.2004.10.025](https://doi.org/10.1016/j.marpolbul.2004.10.025)
6. Orth RJ, Carruthers TJB, Dennison WC, Duarte CM, Fourqurean JW, Heck KL Jr., et al. A global crisis for seagrass ecosystems. *BioScience*. 2006; 56(12):987–96. doi: [10.1641/0006-3568\(2006\)56\[987:AGCFSE\]2.0.CO;2](https://doi.org/10.1641/0006-3568(2006)56[987:AGCFSE]2.0.CO;2)
7. Waycott M, Duarte CM, Carruthers TJB, Orth RJ, Dennison WC, Olyamik S, et al. Accelerating loss of seagrasses across the globe threatens coastal ecosystems. *Proc Nat Acad Sci*. 2009; 106(30):12377–81. doi: [10.1073/pnas.0905620106](https://doi.org/10.1073/pnas.0905620106) PMID: [19587236](https://pubmed.ncbi.nlm.nih.gov/19587236/)
8. Ralph PJ, Durako MJ, Enriquez S, Collier CJ, Doblin MA. Impact of light limitation on seagrasses. *J Exp Mar Biol Ecol*. 2007; 350:176–93. doi: [10.1016/j.jembe.2007.06.017](https://doi.org/10.1016/j.jembe.2007.06.017)
9. Beer S, Koch E. Photosynthesis of marine macroalgae and seagrasses in globally changing CO₂ environments. *Mar Ecol Prog Ser*. 1996; 141:199–204.
10. Invers O, Zimmerman RC, Alberte RS, Perez M, Romero J. Inorganic carbon sources for seagrass photosynthesis: an experimental evaluation of bicarbonate use in species inhabiting temperate waters. *J Exp Mar Biol Ecol*. 2001; 365:203–17. doi: [10.1016/S0022-0981\(01\)00332-X](https://doi.org/10.1016/S0022-0981(01)00332-X)
11. Gattuso JP, Hansson L. Ocean acidification: background and history. In: Gattuso JP, Hansson L, editors. *Ocean Acidification*. Oxford, New York: Oxford University Press; 2011. p. 1–16.
12. Raven JA, Caldeira K, Elderfield H, Hoegh-Guldberg O, Liss P, Riebesell U, et al. *Ocean acidification due to increasing atmospheric carbon dioxide*. The Royal Society. The Ciyvedon Press Ltd, Cardiff, UK: 2005.
13. Koch M, Bowes G, Ross C, Zhang XH. Climate change and ocean acidification effects on seagrasses and marine macroalgae. *Glob Change Biol*. 2013; 19:103–32. doi: [10.1111/j.1365-2486.2012.02791.x](https://doi.org/10.1111/j.1365-2486.2012.02791.x)
14. Short FT, Neckles HA. The effects of global climate change on seagrasses. *Aqua Bot*. 1999; 63:169–96. doi: [10.1016/S0304-3770\(98\)00117-X](https://doi.org/10.1016/S0304-3770(98)00117-X)
15. Fabricius KE, Langdon C, Uthicke S, Humphrey C, Noonan S, Death G, et al. Losers and winners in coral reefs acclimatized to elevated carbon dioxide concentrations. *Nat Clim Change*. 2011; 1:165–9.
16. Palacios SL, Zimmerman RC. Response of eelgrass *Zostera marina* to CO₂ enrichment: possible impacts of climate change and potential for remediation of coastal habitats. *Mar Ecol Prog Ser*. 2007; 344:1–13.
17. Jiang ZJ, Huang X, Zhang J. Effects of CO₂ enrichment on photosynthesis, growth and biochemical composition of seagrass *Thalassia hemprichii* (Ehrenb.) Aschers. *J Integr Plant Biol*. 2010; 52(10):904–13. doi: [10.1111/j.1744-7909.2010.00991.x](https://doi.org/10.1111/j.1744-7909.2010.00991.x) PMID: [20883442](https://pubmed.ncbi.nlm.nih.gov/20883442/)
18. Alexandre A, Silva J, Buapet P, Bjork M, Santos R. Effects of CO₂ enrichment on photosynthesis, growth, and nitrogen metabolism of the seagrass *Zostera noltii*. *Ecol Evol*. 2012; 2(10):2620–30. doi: [10.1002/ece3.333](https://doi.org/10.1002/ece3.333) PMID: [23145346](https://pubmed.ncbi.nlm.nih.gov/23145346/)
19. Zimmerman RC, Kohrs DG, Stellar DL, Alberte RS. Impacts of CO₂ enrichment on productivity and light requirements of eelgrass. *Plant Physiol*. 1997; 115:599–607. doi: [10.1104/pp.115.2.599](https://doi.org/10.1104/pp.115.2.599) PMID: [12223828](https://pubmed.ncbi.nlm.nih.gov/12223828/)

20. Invers O, Tomas F, Perez M, Romero J. Potential effect of increased global CO₂ availability on the depth distribution of the seagrass *Posidonia oceanica* (L.) Delile: A tentative assessment using a carbon balance model. *B Mar Sci.* 2002; 71(3):1191–8.
21. Ow YX, Collier CJ, Uthicke S. Responses of three tropical seagrass species to CO₂ enrichment. *Mar Biol.* 2015; 162:1005–17. doi: [10.1007/s00227-015-2644-6](https://doi.org/10.1007/s00227-015-2644-6)
22. Burnell OW, Connell SD, Irving AD, Watling JR, Russell BD. Contemporary reliance on bicarbonate acquisition predicts increased growth of seagrass *Amphibolis antarctica* in a high-CO₂ world. *Conserv Physiol.* 2014; 2. doi: [10.1093/conphys/cou052](https://doi.org/10.1093/conphys/cou052)
23. Durako MJ. Photosynthetic utilization of CO_{2(aq)} and HCO₃⁻ in *Thalassia testudinum* (Hydrocharitaceae). *Mar Biol.* 1993; 115:373–80. doi: [10.1007/BF00349834](https://doi.org/10.1007/BF00349834)
24. Beer S, Waisel Y. Some photosynthetic carbon fixation properties of seagrasses. *Aqua Bot.* 1979; 7:129–38. doi: [10.1016/0304-3770\(79\)90017-2](https://doi.org/10.1016/0304-3770(79)90017-2)
25. Hall-Spencer JM, Rodolfa-Metalpa R, Martin S, Ransome E, Fine M, Turner SM, et al. Volcanic carbon dioxide vents show ecosystem effects of ocean acidification. *Nature.* 2008; 454:96–9. doi: [10.1038/nature07051](https://doi.org/10.1038/nature07051) PMID: [18536730](https://pubmed.ncbi.nlm.nih.gov/18536730/)
26. Bjork M, Weil A, Semesi S, Beer S. Photosynthetic utilisation of inorganic carbon by seagrasses from Zanzibar, East Africa. *Mar Biol.* 1997; 129:363–6. doi: [10.1007/s002270050176](https://doi.org/10.1007/s002270050176)
27. Hellblom F, Beer S, Bjork M, Axelsson L. A buffer sensitive inorganic carbon utilisation system in *Zostera marina*. *Aqua Bot.* 2001; 69:55–62. doi: [10.1016/S0304-3770\(00\)00132-7](https://doi.org/10.1016/S0304-3770(00)00132-7)
28. Beer S, Bjork M, Hellblom F, Axelsson L. Inorganic carbon utilisation in marine angiosperms (seagrasses). *Funct Plant Biol.* 2002; 29:349–54. doi: [10.1071/PP01185](https://doi.org/10.1071/PP01185)
29. Hellblom F, Axelsson L. External HCO₃⁻ dehydration maintained by acid zones in the plasma membrane is an important component of the photosynthetic carbon uptake in *Ruppia cirrhosa*. *Photosynth Res.* 2003; 77(173–181). doi: [10.1023/A:1025809415048](https://doi.org/10.1023/A:1025809415048)
30. Beer S, Rehnberg J. The acquisition of inorganic carbon by the seagrass *Zostera marina*. *Aqua Bot.* 1997; 56:277–83. doi: [10.1016/S0304-3770\(96\)01109-6](https://doi.org/10.1016/S0304-3770(96)01109-6)
31. Masini RJ, Manning CR. The photosynthetic responses to irradiance and temperature of four meadow-forming seagrasses. *Aquat Bot.* 1997; 58:21–36. doi: [10.1016/S0304-3770\(97\)00008-9](https://doi.org/10.1016/S0304-3770(97)00008-9)
32. Schubert N, Colombo-Pallota MF, Enriquez S. Leaf and canopy scale characterization of the photoprotective response to high-light stress of the seagrass *Thalassia testudinum*. *Limnol Oceanogr.* 2015; 60:286–302. doi: [10.1002/lno.10024](https://doi.org/10.1002/lno.10024)
33. Enriquez S, Pantoja-Reyes NI. Form-function analysis of the effect of canopy morphology on leaf self-shading in the seagrass *Thalassia testudinum*. *Oecologia.* 2005; 145:235–43. doi: [10.1007/s00442-005-0111-7](https://doi.org/10.1007/s00442-005-0111-7) PMID: [15942763](https://pubmed.ncbi.nlm.nih.gov/15942763/)
34. Schwarz AM, Bjork M, Buluda T, Mtolera M, Beer S. Photosynthetic utilisation of carbon and light by two tropical seagrass species as measured in situ. *Mar Biol.* 2000; 137:755–61.
35. Burnell OW, Russell BD, Irving AD, Connell SD. Seagrass response to CO₂ contingent on epiphytic algae: indirect effects can overwhelm direct effects. *Oecologia.* 2014; 176:871–82. doi: [10.1007/s00442-014-3054-z](https://doi.org/10.1007/s00442-014-3054-z) PMID: [25193313](https://pubmed.ncbi.nlm.nih.gov/25193313/)
36. Liu SL, Wang WL, Dy DT, Fu CC. The effect of ulvoid macroalgae on the inorganic carbon utilization by an intertidal seagrass *Thalassia hemprichii*. *Bot Bull Acad Sin.* 2005; 46:197–203.
37. Collins M, Knutti R, Arblaster J, Dufresne JL, Fichet T, Friedlingstein P, et al. Long-term Climate Change: Projections, Commitments and Irreversibility. In: Stocker TF, Qin D, Plattner GK, Tignor M, Allen SK, Boschung J, et al., editors. *Climate Change 2013: The Physical Science Basis Contribution of Working Group I to the Fifth Assessment Report of the Intergovernmental Panel on Climate Change.* Cambridge, United Kingdom and New York, NY, USA: Cambridge University Press; 2013.
38. Uku J, Beer S, Bjork M. Buffer sensitivity of photosynthetic carbon utilisation in eight tropical seagrasses. *Mar Biol.* 2005; 147:1085–90. doi: [10.1007/s00227-005-0019-0](https://doi.org/10.1007/s00227-005-0019-0)
39. Kubler JE, Raven JA. The interaction between inorganic carbon acquisition and light supply in *Palmaria palmata* (Rhodophyte). *J Phycol.* 1995; 31:369–75. doi: [10.1111/j.0022-3646.1995.00369.x](https://doi.org/10.1111/j.0022-3646.1995.00369.x)
40. Spalding MH, Ogren WL. Photosynthesis is required for induction of the CO₂-concentrating system in *Chlamydomonas reinhardtii*. *FEBS Lett.* 1982; 145:41–4.
41. McKenzie LJ, Collier C, Waycott M. Reef Rescue Marine Monitoring Program: Inshore seagrass, annual report for the sampling period 1st June 2012–31st May 2013. Book ER. Cairns: James Cook University, 2015.
42. Vogel N, Uthicke S. Calcification and photobiology in symbiont-bearing benthic foraminifera and responses to a high CO₂ environment. *J Exp Mar Biol Ecol.* 2012; 424–425:15–24. doi: [10.1016/j.jembe.2012.05.008](https://doi.org/10.1016/j.jembe.2012.05.008)

43. Robbins LL, Hansen ME, Kleypas JA, Meylan SC. CO₂ calc—a User-friendly Seawater Carbon Calculator for Windows, Mac OS X, and iOS (iPhone) Open-file Report 2010. Reston, USA: U.S. Geological Survey, 2010.
44. Ryle VD, Mueller HR, Gentien P, Science AloM. Automated analysis of nutrients in tropical sea waters. Townsville, Australia: Australian Institute of Marine Science; 1981. 24 p.
45. Short FT, Duarte CM. Methods for the measurement of seagrass growth and production. In: Short FT, Coles RC, editors. Global Seagrass Research Methods. Amsterdam, The Netherlands: Elsevier Science B.V.; 2001. p. 155–82.
46. Collier CJ, Uthicke S, Waycott M. Thermal tolerance of two seagrass species at contrasting light levels: Implications for future distribution in the Great Barrier Reef. *Limnol Oceanogr.* 2011; 56(6):2200–10. doi: [10.4319/lo.2011.56.6.2200](https://doi.org/10.4319/lo.2011.56.6.2200)
47. Jassby AD, Platt T. Mathematical formulation of the relationship between photosynthesis and light for phytoplankton. *Limnol Oceanogr.* 1976; 21:540–7. doi: [10.4319/lo.1976.21.4.0540](https://doi.org/10.4319/lo.1976.21.4.0540)
48. Axelsson L, Mercado J, Figueroa F. Utilization of HCO₃⁻ at high pH by the brown macroalga *Laminaria saccharina*. *Eur J Phycol.* 2000; 35:53–9. doi: [10.1080/09670260010001735621](https://doi.org/10.1080/09670260010001735621)
49. Underwood AJ. Experiments in Ecology: Their logical design and interpretation using analysis of variance. Cambridge, United Kingdom: Cambridge University Press; 1997. 504 p.
50. R Development Core Team. R: A Language and Environment for Statistical Computing. Vienna, Austria: R Foundation for Statistical Computing; 2014.
51. Russell BD, Connell SD, Uthicke S, Muehlehner N, Fabricius K, Hall-Spencer J. Future seagrass beds: Can increased productivity lead to increased carbon storage? *Mar Poll Bull.* 2013; 73:463–9. doi: [10.1016/j.marpolbul.2013.01.031](https://doi.org/10.1016/j.marpolbul.2013.01.031)
52. Lee K, Park SR, Kim J. Production dynamics of the eelgrass, *Zostera marina* in two bay systems on the south coast of the Korean peninsula. *Mar Biol.* 2005; 147:1091–108. doi: [10.1007/s00227-005-0011-8](https://doi.org/10.1007/s00227-005-0011-8)
53. Kaldy JE, Dunton KH. Above- and below-ground production, biomass and reproductive ecology of *Thalassia testudinum* (turtle grass) in a subtropical coastal lagoon. *Mar Ecol Prog Ser.* 2000; 193:271–83.
54. Collier CJ, Waycott M, Ospina AG. Responses of four Indo-West Pacific seagrass species to shading. *Mar Poll Bull.* 2012; 65:342–54. doi: [10.1016/j.marpolbul.2011.06.017](https://doi.org/10.1016/j.marpolbul.2011.06.017)
55. Stitt M, Krapp A. The interaction between elevated carbon dioxide and nitrogen nutrition: the physiological and molecular background. *Plant Cell Environ.* 1999; 22:583–621. doi: [10.1046/j.1365-3040.1999.00386.x](https://doi.org/10.1046/j.1365-3040.1999.00386.x)
56. Touchette BW, Burkholder JM. Overview of the physiological ecology of carbon metabolism in seagrasses. *J Exp Mar Biol Ecol.* 2000; 250:169–205. PMID: [10969168](https://pubmed.ncbi.nlm.nih.gov/10969168/)
57. Long SP, Ainsworth EA, Rogers A, Ort DR. Rising atmospheric carbon dioxide: Plants FACE the future. *Annu Rev Plant Biol.* 2004; 55:591–628. PMID: [15377233](https://pubmed.ncbi.nlm.nih.gov/15377233/)
58. Hall DO, Rao KK. Photosynthesis, 6th ed. Great Britain: Cambridge University Press; 1994. 198 p.
59. van Dam JW, Negri AP, Mueller JF, Altenburger R, Uthicke S. Additive pressures of elevated sea surface temperatures and herbicides on symbiont-bearing foraminifera. *PLoS one.* 2012; 7(3):e33900. doi: [10.1371/journal.pone.0033900](https://doi.org/10.1371/journal.pone.0033900) PMID: [22439012](https://pubmed.ncbi.nlm.nih.gov/22439012/)
60. Bliss CI. The toxicity of poisons applied jointly. *Ann Appl Biol.* 1939; 26:585–615.
61. Zou DH, Gao KS. Effects of elevated CO₂ on the red seaweed *Gracilaria lemaneiformis* (Gigartinales, Rhodophyta) grown at different irradiance levels. *Phycologia.* 2009; 48:510–7. doi: [10.2216/08-99.1](https://doi.org/10.2216/08-99.1)
62. Stumm W, Morgan JJ. Aquatic Chemistry: Chemical Equilibria and Rates in Natural Waters. Hoboken: Wiley; 2013.
63. Hepburn CD, Pritchard DW, Cornwall CE, McLeod RJ, Beardalls J, Raven JA, et al. Diversity of carbon use strategies in a kelp forest community: implications for a high CO₂ ocean. *Glob Change Biol.* 2011; 17:2488–97. doi: [10.1111/j.1365-2486.2011.02411.x](https://doi.org/10.1111/j.1365-2486.2011.02411.x)
64. Campbell JE, Fourqurean JW. Mechanisms of bicarbonate use influence the photosynthetic carbon dioxide sensitivity of tropical seagrasses. *Limnol Oceanogr.* 2013; 58:839–48. doi: [10.4319/lo.2013.58.3.0839](https://doi.org/10.4319/lo.2013.58.3.0839)
65. Salo T, Reusch TBH, Bostrom C. Genotype-specific responses to light stress in eelgrass *Zostera marina*, a marine foundation plant. *Mar Ecol Prog Ser.* 2015; 519:129–40.
66. Reusch TBH, Veron AS, Preuss C, Weiner J, Wissler L, Beck A, et al. Comparative analysis of expressed sequence tag (EST) libraries in the seagrass *Zostera marina* subjected to temperature stress. *Mar Biotechnol.* 2008; 10:297–309. doi: [10.1007/s10126-007-9065-6](https://doi.org/10.1007/s10126-007-9065-6) PMID: [18239962](https://pubmed.ncbi.nlm.nih.gov/18239962/)
67. Israel A, Hophy M. Growth, photosynthetic properties and Rubisco activities and amounts of marine macroalgae grown under current and elevated seawater CO₂ concentrations. *Glob Change Biol.* 2002; 8:831–40. doi: [10.1046/j.1365-2486.2002.00518.x](https://doi.org/10.1046/j.1365-2486.2002.00518.x)

68. Takahashi M, Noonan SHC, Fabricius KE, Collier CJ. The effects of long-term in situ CO₂ enrichment on tropical seagrass communities at volcanic vents. *ICES Journal of Marine Science: Journal du Conseil*. 2015. doi: [10.1093/icesjms/fsv157](https://doi.org/10.1093/icesjms/fsv157)
69. Campbell JE, Fourqurean JW. Ocean acidification outweighs nutrient effects in structuring seagrass epiphyte communities. *Journal of Ecology*. 2014; 102(3):730–7. doi: [10.1111/1365-2745.12233](https://doi.org/10.1111/1365-2745.12233)
70. Kroon FJ, Kuhnert PM, Henderson BL, Wilkinson SN, Kinsey-Henderson A, Abbott B, et al. River loads of suspended solids, nitrogen, phosphorus and herbicides delivered to the Great Barrier Reef lagoon. *Mar Poll Bull*. 2012; 65:167–81. doi: [10.1016/j.marpolbul.2011.10.018](https://doi.org/10.1016/j.marpolbul.2011.10.018)