

RESEARCH ARTICLE

Feeding Behavior of a Crab According to Cheliped Number

Diogo Nunes de Oliveira¹, Ronaldo Adriano Christofolletti², Rodrigo Egydio Barreto^{3*}

1 Universidade Estadual Paulista “Julio de Mesquita Filho”, Instituto de Biociências (IBB-UNESP), Rubião Jr. s/n, 18618–970, Botucatu, São Paulo, Brazil, **2** Universidade Federal de São Paulo, Instituto do Mar, Campus Baixada Santista (IMar/UNIFESP), Av. Alm. Saldanha da Gama, 89 - Ponta da Praia; Santos (SP)–Brazil, **3** Departamento de Fisiologia, Instituto de Biociências, Caunesp, UNESP, Rubião Jr. s/n, 18618–970, Botucatu, São Paulo, Brazil

* rebarreto@yahoo.com



Abstract

Cheliped loss through autotomy is a common reflexive response in decapod crustaceans. Cheliped loss has direct and indirect effects on feeding behavior which can affect population dynamics and the role of species in the community. In this study, we assessed the impact of autotomy (0, 1, or 2 cheliped loss) on feeding behavior in the crab *Pachygrapsus transversus*, an omnivorous and abundant species that inhabits subtropical intertidal rocky shores along the South Atlantic Ocean. Autotomy altered crab feeding patterns and foraging behavior; however, the time spent foraging on animal prey or algae was not affected. These results indicate a plasticity of feeding behavior in *P. transversus*, allowing them to maintain feeding when injured.

OPEN ACCESS

Citation: Oliveira DNd, Christofolletti RA, Barreto RE (2015) Feeding Behavior of a Crab According to Cheliped Number. PLoS ONE 10(12): e0145121. doi:10.1371/journal.pone.0145121

Editor: Erik V. Thuesen, The Evergreen State College, UNITED STATES

Received: September 1, 2015

Accepted: November 28, 2015

Published: December 18, 2015

Copyright: © 2015 Oliveira et al. This is an open access article distributed under the terms of the [Creative Commons Attribution License](https://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Data Availability Statement: All relevant data are within the paper and its Supporting Information files.

Funding: This work was supported by the Fundação de Amparo à Pesquisa do Estado de São Paulo (FAPESP) due scholarship to DNO (07/58436-8) and research funds granted to RAC (06/00559-4 and 06/60237-0). The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing Interests: The authors have declared that no competing interests exist.

Introduction

Autotomy is an efficient reflexive response which results in loss of a limb at a pre-formed breakage plane [1–4]. This mechanism is considered a useful adaptation to avoid predators and limit injuries [5–9], occurring commonly in vertebrate and invertebrate groups; therefore, it has been investigated by several researchers [8, 10–12].

Despite the immediate survivorship benefits provided by autotomy [9, 13], the loss of one or more appendages may result in long-term energetic and functional costs [6, 8, 10, 11, 14, 15]. For example, crustaceans use chelipeds for defense, capture, manipulation, and subjugation of prey [16]. Damage or loss of those appendages can cause profound effects on feeding efficiency [17, 18], growth [19, 20], reproductive success [21, 22], alter the duration of each intermolt phase [18, 23], promote defective development [11], and creates limitations on the competitiveness of animals even after limb regeneration [24].

Especially in complex environments such as rocky shores where the diversity of predators and prey is high, nonlethal damage may influence population dynamics as well as community processes. Relatively few studies have examined the effects of autotomy on decapod crustaceans and knock-on consequences to the wider community, despite these animals being an important consumer in aquatic environments [8, 25–27].

On intertidal rocky shores of the Southwest Atlantic, the grapsid crab *Pachygrapsus transversus* (Gibbes, 1850) occurs in high abundance and can control prey populations and diversity. Its feeding habit is omnivorous, although when both algae and animal prey are available, there is preference for animal prey, but not for a specific animal group [28].

Following personal observations in the natural environment where individuals of *P. transversus* have been seen foraging even without both chelipeds, this study aims to describe and evaluate the consequences of cheliped loss on feeding behavior. We assessed if crabs are able to feed themselves without chelipeds and if the foraging time and prey chosen change with the number of the chelipeds. Thus, results can contribute to our understanding of how autotomized crabs feed and survive and thus potentially the wider consequences of autotomy on prey populations.

Materials and Methods

Ethics statement

This research was authorized by the System of authorization and information on biodiversity (Sistema de Autorização e Informação em Biodiversidade—SISBIO, Brazil) from the Ministry of Environment of Brazilian government (Ministério do Meio Ambiente—MMA, Brazil), protocol number 30870–4.

Crabs

P. transversus adults, with carapace width (CW) ranging from 13 to 25 mm, were collected during the spring season on rocky shores of the north coast of São Paulo State, Brazil. Crabs were classified in accordance to the number of chelipeds (0, 1, or 2) and, in the absence of sufficient specimens without cheliped, autotomy was induced in the laboratory by cutting the cheliped according to the methods previously described [29]. The gender and level of sexual maturity were not considered and analyzed in the present study based on previous observations showing no influence of these variables on stomach contents in this species (D. N. Oliveira unpubl. data). In this case, autotomized crabs were chosen randomly and the cheliped autotomized was also randomly chosen. After autotomization, crabs were maintained in individual tanks for 48 hours before starting the experiment.

Experimental design and procedures

The experiment was carried out in two consecutive treatment blocks of 18 tanks and 12 randomized replications were performed in each treatment (0, 1 or 2 cheliped). For experimentation, crabs were acclimated individually into aquaria with a central shelter and flowing sea water for 48 hours fasting. Food types were chosen based on previous analysis of stomach contents of this species [28], which determined the most consumed prey. After this acclimatization period, 7 cm² each food type (ascidians, the bivalve *Brachidontes solisianus* (d'Orbigny, 1842), biofilm, and macroalgae) were arranged in parallel in a randomly chosen corner in each aquarium. The bivalve *B. solisianus* had a length ranging from 0.34 to 1.72 cm. Only intact organisms were used as prey to avoid any influence of chemical cues. All food items were washed in sea water and visually examined to remove any other organisms that could serve as a food source.

Crabs were monitored using digital cameras for one hour to quantify the time spent foraging. Foraging time was determined as the ratio of time spent handling a specific food to the total time of filming, a procedure based on [28]. Foraging time was considered as the time that crabs spent holding a food item and taking the chelipeds to the oral appendages (Figs 1B and 2I). For crabs without chelipeds, foraging time consisted of the period during which the crab performed

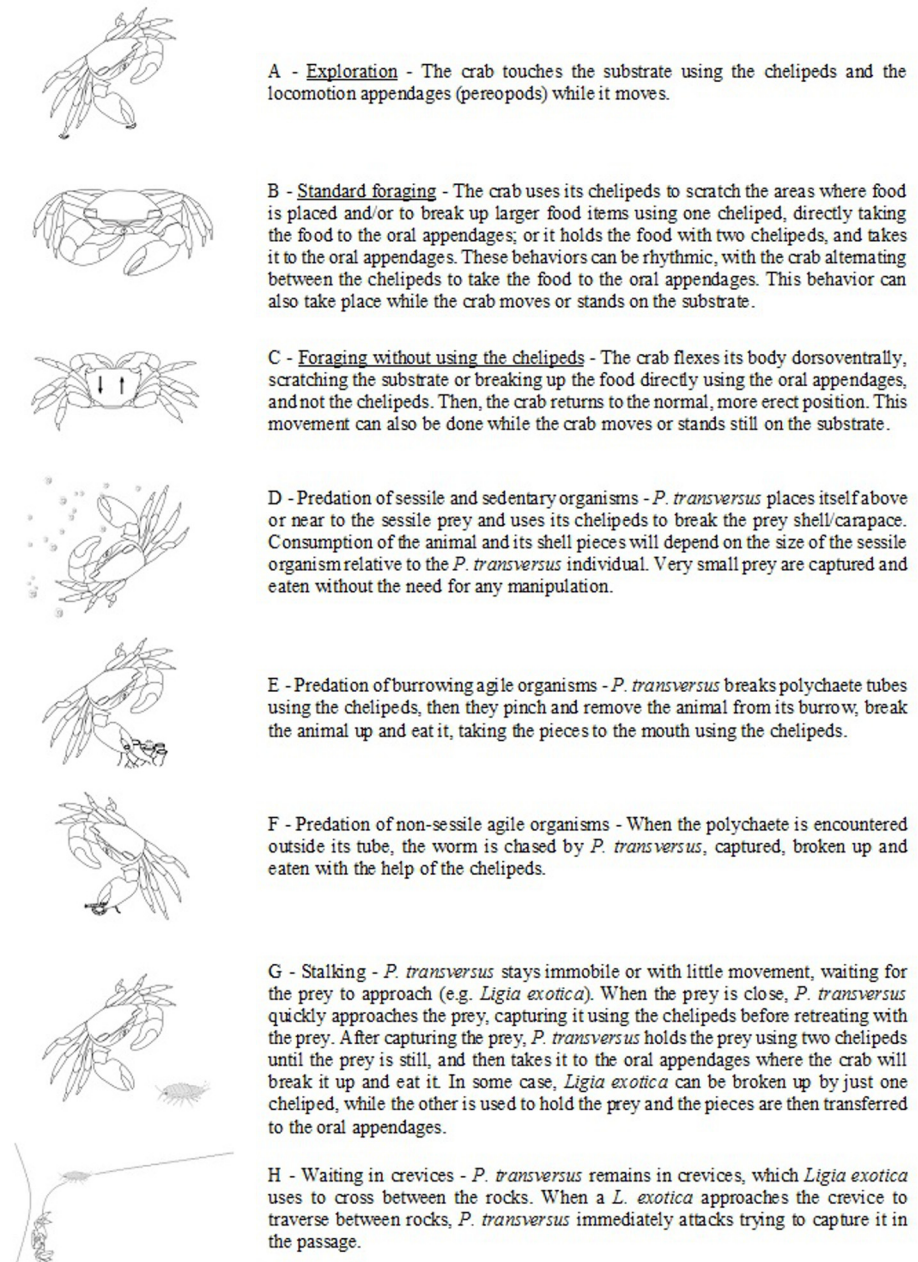


Fig 1. Ethogram of the feeding behavior of *P. transversus* observed in the natural environment and in the laboratory.

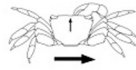
doi:10.1371/journal.pone.0145121.g001

the movement to flex the body dorsoventrally by putting oral appendages in contact with food on substract ([Fig 2K–2L](#)).

Ethograms of feeding behavior as a function of the number of chelipeds were described for *P. transversus* foraging on sessile prey based on two laboratory filming periods of 36 and 99 hours. Furthermore, 21 hours of film, pictures, and in situ observations of predation behavior of *P. transversus* on mobile prey such as polychaetes and the isopod *Ligia exotica* (Roux, 1828) were added.



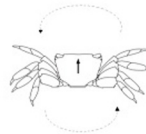
I - Foraging using only one cheliped - Similar to the intact crab (Fig 1, A), the autotomized crab uses its single cheliped to scratch the substrate in areas where food is placed, or to break food up into smaller pieces, directly taking them to the oral appendages. However, due to the absence of one cheliped, the first locomotory appendage is used to help with food manipulation, and also used to clean the oral structures. Only having one cheliped, movements to take food to the oral appendages are rhythmic and faster than in the uninjured crabs.



J - Foraging using oral appendages - Unlike the pattern observed in the uninjured crabs (Fig 1, C), autotomized *P. transversus* dorsoventrally flexes the body, but, with the body flexed, the crab moves around directly scratching the substrate using the oral appendages, and then returns to the normal, erect position.



K - Foraging without the chelipeds - Crabs without chelipeds dorsoventrally flex their bodies, scratching the substrate with their oral appendages and then return to the normal, more erect position. Unlike the pattern observed in the uninjured crabs or crabs with one cheliped (Fig 1, C and Fig 2, J), this behavior is rhythmic, while the crab stands or moves. The use of the anterior locomotory appendages was also observed to manipulate and hold the food, while the crab dorsoventrally flexes its body to consume food. This behavior pattern was observed whilst the crab was foraging on bivalves and the biofilm.



L - Foraging without the chelipeds (variation) - The crab dorsoventrally flexes its body and, with the body still flexed, it spins 360° around its own axis scratching the substrate using its oral appendages. This behavior was observed whilst crabs were foraging on bivalves or the biofilm, but only in individuals who had suffered autotomy of both chelipeds, both in the natural environment and laboratory.

Fig 2. Ethogram of the feeding behavior of autotomized *P. transversus* observed in the natural environment and in the laboratory.

doi:10.1371/journal.pone.0145121.g002

Statistical analysis

Data on foraging activity rate (dependent variable) were arcsine transformed. Following transformation, data were normally distributed and homoscedastic according to the Kolmogorov-Smirnov test and Levene's test, respectively. Data were analyzed using a two-way mixed ANOVA (one-between-subjects categorical predictor and one-within-subjects repeated measures ANOVA) followed by a Newman-Keuls post-hoc test, with number of claws as independent factor and food types as repeated measures. Statistical differences were considered significant when $P < 0.05$.

Results

Feeding behavior

The first activity of *P. transversus* during foraging is exploration of the environment touching the substrate with chelipeds and/or pereopods to find the food source. Observations show that autotomy does not have an influence on this behaviour (Fig 1A and 1D). Crabs with 1 or 2 chelipeds used them as their principal feeding tool, using the cheliped to tear the food and then to eat it (Figs 1B and 2I).

Furthermore, crabs without chelipeds used the oral appendages as the principal feeding tool. Crabs without chelipeds flex their body dorsoventrally, lowering the body down to the substrate, scratching and breaking up the food by using the oral appendages (Fig 2K–2L). This behavior was observed for both uninjured and autotomized crabs, but for uninjured crabs, it was less frequent and lasted only for a short period of time before the body was returned to the erect position (Figs 1C and 2J). Predatory behavior on mobile organisms (e.g., the isopod *L. exotica* and polychaetes) were only observed in uninjured *P. transversus* specimens *in situ* (Fig 1E–1H).

Foraging activity

Autotomy neither influenced the foraging activity ($F_{(2,33)} = 1.26$; $P = 0.297$), nor the interaction with food types ($F_{(6,99)} = 0.98$; $P = 0.443$). However, crabs foraged significantly more on the

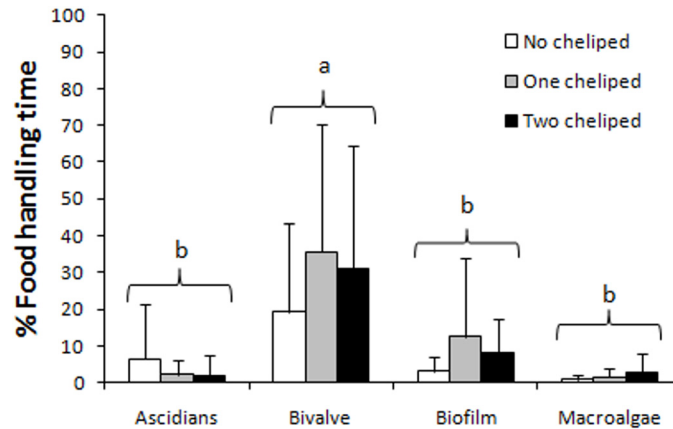


Fig 3. Foraging time (% mean \pm SD, see S1 Table for details) in relation to food type and cheliped number in *P. transversus*. Different letters represent significant differences among food types (two-way repeated measures ANOVA; $p < 0.05$).

doi:10.1371/journal.pone.0145121.g003

bivalve *B. solisianus* compared to other food types ($F_{(3,99)} = 17.36$; $P < 0.00001$; Fig 3) regardless of the number of chelipeds.

Discussion

Autotomy had no influence on foraging time, because the crab *P. transversus* showed clear behavioral plasticity in their feeding pattern according to the number of chelipeds, which allow crabs to handle different food items. Change in feeding behavior is an adaptive mechanism which enables crabs to continue feeding on the same type of prey.

The number of chelipeds present influenced the activity patterns of *P. transversus* and reduced their behavioral repertoire. The lower the number of chelipeds, the smaller the behavioural repertoire became. All autotomized crabs used locomotor appendages to capture and manipulate food during foraging. This behavior has also been observed for *Menippe mercenaria* [30], *Callinectes sapidus* [17, 31], *Hemigrapsus sanguineus* [32], and for the congener *Pachygrapsus crassipes* [33, 34]. Thus, we suggest that the use of locomotor appendages is a compensatory mechanism to overcome the absence of chelipeds in crustaceans, allowing them to maintain nutrient acquisition for essential biological processes, such as molt and regeneration of a new limb. However, this issue was not the aim of the present study and must be assessed in future studies.

Feeding behavior of *P. transversus* showed that autotomy influences the way the crabs feed but does not influence food choice. Bivalves were previously identified as the preferred prey of this crab [28] corroborating with the present study where crabs spent significantly more time handling bivalve prey instead of algae, biofilm, or ascidians, regardless of the number of chelipeds. A predator does not need a second cheliped to hold a prey, to open or crush the shell, explaining why there is no change in food preference with loss of chelipeds [35].

In other crab species, autotomy also showed no effect on time spent foraging but feeding rate decreased in *C. pagurus* [29], *Cancer magister* [34], *Hemigrapsus sanguineus* [32] and *Carcinus maenas* [35]. In the present study, we did not quantify feeding rate, but we observed shell fragments and bivalves without some parts in all treatments, indicating that mussels had been ingested independently of the number of chelipeds.

Some behaviors are considered patterns and are well known for decapods. For example, the behavior of manipulating food with the locomotor appendages was observed here in all three

treatments (0, 1, and 2 chelipeds), although mainly in autotomized crabs. Such behavior has also been reported for other species [17, 30–33]. In this study we specifically observed the behavior in which the crabs flexed dorsoventrally their body and rotated 360° around their own axis, in order to manipulate the food with the oral appendices. This behavior was observed for crabs that had autotomized both chelipeds and allowed feeding to occur.

Our study demonstrated that autotomy has the capability of modulating behavioral changes in *P. transversus*, but autotomy is not a limitation large enough to modify the food preferences of this species. Also, a new behavior was observed in crabs with both chelipeds autotomized that allowed crabs to retain feeding capacity. Further studies on the use of locomotor appendages as a compensatory mechanism to overcome the absence of chelipeds in crustaceans are warranted.

Supporting Information

S1 Table. Foraging time (% , mean \pm SD) in relation to food type and cheliped number in *P. transversus*.

(DOC)

Acknowledgments

Our acknowledgements to João Bosco for confection of the pictures in Tables 1 and 2, for the anonymous reviewers for the suggestions and for Prof Stuart Jenkins, Prof Gray Williams and Daiane Oliveira for the comments on the last version of this manuscript.

Author Contributions

Conceived and designed the experiments: RAC. Performed the experiments: DNO. Analyzed the data: RAC REB. Contributed reagents/materials/analysis tools: RAC REB. Wrote the paper: RAC DNO REB.

References

1. Woodland WNF. Some observations on caudal autotomy and regeneration in the gecko (*Hemidactylus flaviviridis*, Rüppel), with notes on the tails of sphenodon and Pygopus. *Quart J Microsc Sci*. 1920; 65: 63–100.
2. Wood FD, Wood HE. Autotomy in decapod Crustacea. *J Exp Zool*. 1932; 62: 1–55.
3. McVean A. The nervous control of autotomy in *Carcinus maenas*. *J Exp Biol*. 1974; 60: 423–436.
4. Knope ML, Larson RJ. Autotomy in porcelain crabs is an effective escape mechanism from rockfish predation. *Mar Ecol*. 2014; 35: 471–477.
5. McVean A. Autotomy. In: Bliss DE, editor. *The Biology of Crustacea*. Academic Press; 1982. pp: 107–132.
6. Arnold EN. Evolutionary aspects of tail shedding in lizards and their relatives. *J Nat Hist*. 1984; 18: 127–169.
7. Endler JA. Defence against predators. In: Feder ME, Lauder GV, editors. *Predator-prey relationships: perspectives and approaches from the study of lower vertebrates*. University of Chicago Press. 1986. pp: 109–134.
8. Juanes F, Smith LD. The ecological consequences of limb damage and loss in decapod crustaceans: a review and prospectus. *J Exp Mar Biol Ecol*. 1995; 193: 197–223.
9. Wasson K, Lyon BE, Knope M. Hair-trigger autotomy in porcelain crabs is a highly effective escape strategy. *Behav Ecol*. 2002; 13: 481–486.
10. Maginnis TL. The costs of autotomy and regeneration in animals: a review and framework for future research. *Behav Ecol*. 2006; 17: 857–872.
11. Fleming PA, Muller D, Bateman PW. Leave it all behind: a taxonomic perspective of autotomy in invertebrates. *Biol Rev*. 2007; 82: 481–510. PMID: [17624964](#)

12. Bateman PW, Fleming PA. To cut a long tail short: a review of lizard caudal autotomy studies carried out over the last 20 years. *J Zool.* 2009; 277: 1–14.
13. Formanowicz DR Jr. The antipredator efficacy of spider leg autotomy. *Anim Behav.* 1990; 40: 400–401.
14. Naya DE, Veloso C, Muñoz JLP, Bozinovic F. Some vaguely explored (but not trivial) costs of tail autotomy in lizards. *Compar Biochem Physiol.* 2007; 146: 189–193.
15. Allen BJ, Levinton JS. Costs of bearing a sexually selected ornamental weapon in a fiddler crab. *Func Ecol.* 2007; 21: 154–161.
16. Lawton P. Predatory interaction between the brachyuran crab *Cancer pagurus* and decapod crustacean prey. *Mar Ecol Prog Ser.* 1989; 52: 169–179.
17. Smith LD, Hines AH. The effect of cheliped loss on blue crab *Callinectes sapidus* Rathbun foraging rate on soft-shelled clams *Mya arenaria* L. *J Exp Mar Biol Ecol.* 1991; 151: 245–256.
18. Brock RE, Smith LD. Recovery of claw size and function following autotomy in *Cancer productus* (Decapoda: Brachyura). *Biol Bull.* 1998; 194: 53–62.
19. Bennett DB. The effect of limb loss and regeneration on the growth of the edible crab, *Cancer pagurus* (L). *J Exp Mar Biol Ecol.* 1973; 13: 45–53.
20. Smith LD. Patterns of the limb loss in the blue crab, *Callinectes sapidus* Rathbun, and the effects of autotomy on growth. *Bull Mar Sci.* 1990; 46: 23–36.
21. Smith LD. The impact of limb autotomy on mate competition in blue crabs *Callinectes sapidus* Rathbun. *Oecologia.* 1992; 89: 494–501.
22. Reaney LT, Milner RNC, Detto T, Backwell PRY. The effects of claw regeneration on territory ownership and mating success in the fiddler crab *Uca mjoebergi*. *Anim Behav.* 2008; 75: 1473–1478.
23. Skinner DM, Graham DE. Molting in land crabs: stimulation by leg removal. *Science.* 1970; 169: 383–385. PMID: [5450373](#)
24. Maginnis TL, Isikbay M, Degerstedt SG, Luethke TJ, Ortman M. The effects of cheliped autotomy and regeneration on aggression in purple shore crabs (*Hemigrapsus nudus*). *Mar Fresh Behav Physiol.* 2015; 48: 241–251.
25. Harris RN. Nonlethal injury to organisms as a mechanism of population regulation. *Amer Natur.* 1989; 134: 835–847.
26. Coen LD. Herbivory by crabs and the control of algal epibionts on Caribbean host corals. *Oecologia.* 1988; 75: 198–203.
27. Hines AH, Haddon AM, Wiechert LA. Guild structure and foraging impact of blue crabs and epibenthic fish in a subestuary of Chesapeake Bay. *Mar Ecol Prog Ser.* 1990; 67: 105–126.
28. Christofolletti RA, Murakami VA, Oliveira DN, Barreto RE, Flores AAV. Foraging by the omnivorous crab *Pachygrapsus transversus* affects the structure of assemblages on sub-tropical rocky shores. *Mar Ecol Prog Ser.* 2010; 420: 125–134.
29. Patterson L, Dick JTA, Elwood RW. Claw removal and feeding ability in the edible crab, *Cancer pagurus*: Implications for fishery practice. *Appl An Behav Sci.* 2009; 116: 302–305.
30. Savage T, Sullivan JR. Growth and regeneration of the stone crab, *Menippe mercenaria*. *Fla Mar Res Publ.* 1978; 32: 23.
31. Smith LD, Hines AH. Autotomy in blue crab (*Callinectes sapidus* Rathbun) populations: geographic, temporal, and ontogenetic variation. *Biol Bull.* 1991; 180: 416–431.
32. Davis JLD, Dobroski NA, Carlton JT, Prevas J, Parks S, Hong D et al. Autotomy in the Asian shore crab (*Hemigrapsus sanguineus*) in a non-native area of its range. *J Crust Biol.* 2005; 25: 655–660.
33. Hiatt RW. The biology of the lined shore crab *Pachygrapsus crassipes* Randall. *Pac Sci.* 1948; 2: 135–213.
34. Juanes F, Hartwick EB. Prey size selection in Dungeness crabs: the effect of claw damage. *Ecology.* 1990; 71: 744–758.
35. Flynn PST, Mellish CL, Pickering TR, Quijón PA. Effects of claw autotomy on green crab (*Carcinus maenas*) feeding rate. *J Sea Res.* 2015; 103: 113–119.