

RESEARCH ARTICLE

Immune Activation in the Female Genital Tract: Expression Profiles of Soluble Proteins in Women at High Risk for HIV Infection

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OPEN ACCESS

Citation: Francis SC, Hou Y, Baisley K, van de Wijgert J, Watson-Jones D, Ao TT, et al. (2016) Immune Activation in the Female Genital Tract: Expression Profiles of Soluble Proteins in Women at High Risk for HIV Infection. PLoS ONE 11(1): e0143109. doi:10.1371/journal.pone.0143109

Editor: Vikrant Sahasrabudhe, National Cancer Institute, UNITED STATES

Received: March 24, 2015

Accepted: October 30, 2015

Published: January 27, 2016

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Data Availability Statement: Due to ethical restrictions, interested researchers may submit request for de-identified data to Saidi.Kapiga@lshtm.ac.uk.

Funding: Financial support for the Biomarkers Project was provided by the European and Developing Countries Clinical Trials Partnership (project code: CT_ct_05_32070_002), the European Combined Highly Active Retroviral Microbicides (CHAARM-FP7 - Grant agreement n°242135 -www.chaarm.eu), the International Partnership for Microbicides, the Medical Research Council (MRC) and Department for International Development

Abstract

Soluble cervicovaginal biomarkers of inflammation, immune activation and risk of HIV acquisition are needed to reliably assess the safety of new biomedical prevention strategies including vaccines and microbicides. However, a fuller understanding of expression profiles in women at high risk for HIV infection is crucial to the effective use of these potential biomarkers in Phase 3 trial settings. We have measured 45 soluble proteins and peptides in cervicovaginal lavage samples from 100 HIV negative women at high risk for HIV infection. Women were followed over one menstrual cycle to investigate modulation by hormonal contraception, menstrual cycle phase, recent sexual exposure and intravaginal practices. Women using injectable DMPA had increased concentration of several soluble proteins of the innate and adaptive immune system, including IL-1 α , IL-1 β , IL-2, MIP-1 β , IP-10, IL-8, TGF- β , HBD4, IgA, IgG1, and IgG2. Women using combined oral contraceptives had a similar signature. There were differences in concentrations among samples from post-ovulation compared to pre-ovulation, notably increased immunoglobulins. Increased prostate-specific antigen, indicative of recent sexual exposure, was correlated with increased IL-6, MCP-1, and SLPI, and decreased GM-CSF and HBD3. The identified signature profiles may prove critical in evaluating the potential safety and impact on risk of HIV acquisition of different biomedical intervention strategies.

(DFID) with S.C. Francis (G1002369), R.J. Hayes and K. Baisley (G0700837) receiving MRC & DFID support. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing Interests: The authors have declared that no competing interests exist.

Introduction

The HIV pandemic continues to expand, with an average of 2.5 million new infections per year [1]. The majority of these new infections are in sub-Saharan Africa, where the epidemic is driven by heterosexual transmission and women make up 60% of the epidemic [1]. Safe, effective, female-controlled HIV prevention methods are urgently needed. Although there have been recent successes with oral pre-exposure prophylaxis, a topical vaginal microbicide and a parenteral vaccine [2–7], these products have been only partially protective, and the search continues for more robust methods. Investigations following unsuccessful products in Phase 3 clinical trials, especially for products associated with increased rates of HIV infection such as nonoxynol-9 [8], cellulose sulphate [9], and recombinant Adenovirus-5 HIV vaccines [10], have highlighted the need to better understand immune activation in the female genital tract for early safety assessment.

Immune activation in the female genital tract can be caused by infection, irritation or epithelial trauma, and results in increased or decreased expression of soluble immune proteins [11], and has been shown to result in attraction of cells expressing HIV co-receptors to the cervicovaginal mucosa thereby increasing susceptibility to HIV infection [8]. Evidence from several trials of ineffective or harmful microbicides has shown that some candidate products can increase concentration of inflammatory immune proteins [8]. Increasingly, clinical studies are measuring soluble immune biomarkers to screen for product-induced mucosal toxicity/irritation in pre-clinical and clinical trials [12–17]. The most common soluble proteins evaluated in trials have been interleukin (IL)-1 α , IL-1 β , IL-1-receptor antagonist, IL-6, IL-8, tumour necrosis factor (TNF)- α and secretory leukocyte peptidase inhibitor (SLPI) [13–17]. Soluble immune biomarkers may also be useful for vaccine development; not only providing safety information for mucosal vaccines, but also for parenteral vaccines such as Adenovirus 5 that may increase immune activation at mucosal sites [18,19].

A number of biomedical and behavioural factors can influence expression of immune proteins in the female genital tract [12], and more research is needed to understand this background variation for future clinical trials. Two studies have investigated baseline variation in low risk populations appropriate for Phase I clinical trials [20,21]; however, only one study has investigated baseline variation among women at high risk for HIV infection in sub-Saharan Africa [22]. There is evidence that soluble protein concentrations vary by menstrual cycle phase [23,24], hormonal contraception use [20,25], seminal plasma exposure [26], the composition of the vaginal microbiota [22,27], and the presence of infections, including sexually transmitted infections (STIs) [28–30]. In sub-Saharan Africa, the effect of highly prevalent vaginal practices, such as intravaginal cleansing, on immune proteins has only been investigated in one study [22,31]. Lastly, many of the studies have focused on pro-inflammatory cytokines and chemokines and, to a lesser extent, growth factors and antimicrobial proteins or peptides. Investigating a wide array of soluble proteins in the female genital tract, including immunoglobulins, may be useful for improving our understanding of their interactions.

To address these gaps, we measured the levels of 45 different soluble immune and antimicrobial proteins and peptides in cervicovaginal lavages (CVL) from 100 women participating in an intensive longitudinal sub-study of a larger microbicide feasibility study in North-West Tanzania. Soluble analytes evaluated included pro-inflammatory cytokines, anti-inflammatory cytokines, growth factors, chemokines, antimicrobial proteins and immunoglobulins. We present data from visits without known STIs, and report on the concentrations of these analytes and the association with menstrual cycle, hormonal contraception, seminal plasma exposure, reported intravaginal practices and clinical findings.

Materials and Methods

Ethics Statement

All study procedures were approved by the ethics committees of the London School of Hygiene and Tropical Medicine and the Medical Research Coordinating Committee of the Tanzanian National Institute for Medical Research. All participants received detailed information about the study to ensure that they understood why the study was being carried out and what the study involved. Informed consent was obtained by signature if literate, or thumb-printed and witnessed (if illiterate) prior to their participation in the study.

Study participants

This study was nested within a 12 month microbicide feasibility study of 970 HIV-negative women aged 18–44 years working in bars, hotels, and other food and recreational facilities in the three towns of Geita, Shinyanga and Kahama in northwestern Tanzania from 2008–2010. These towns are located near large-scale gold or diamond mines in which there are large populations of migratory male workers. This study showed a high HIV prevalence (18%) and incidence (3.7 cases per 100 woman-years) [32]. Similar populations have been targeted for recent HIV prevention trials in Tanzania due to high prevalence and incidence of HIV [33,34]. The purpose of the microbicide feasibility study was to assess feasibility, retention and appropriateness of this population for future trials, and screening, enrolment and follow-up procedures for this cohort have been described elsewhere [32]. We enrolled 100 women into a sub-study between August and October 2009. Women were sampled purposefully by reported intravaginal practices at cohort enrolment in order to ensure inclusion of women using less common intravaginal practices (e.g. insertion with detergents). Women who were HIV positive, pregnant, currently breastfeeding, or had previous known cervical or uterine abnormalities or surgery (“Have you ever been told that you have a problem with your cervix or uterus?” and “Have you ever had an operation on your cervix or uterus?”) were excluded from the study. Women who were menstruating during the enrolment visits were asked to return after the completion of menses.

Study design

Participants enrolled in the sub-study were followed up three times a week for 4 weeks (12 visits total). At enrolment, interviews were carried out to obtain information about sexual behaviour, vaginal practices, current contraception, and STI symptoms. On the first and last visit (visits 1 & 12), a clinical and colposcopic examination were performed; cervical and vaginal swabs were collected to test for vaginal pH and reproductive tract infections (*Chlamydia trachomatis*, *Neisseria gonorrhoeae*, *Trichomonas vaginalis*, bacterial vaginosis, and yeast); and a CVL was obtained for the detection of *Herpes simplex virus, types 1 and 2* (HSV), prostate-specific antigen (PSA) to measure seminal plasma exposure in the last 48 hours [35], soluble immune proteins, haemoglobin and white blood cells (WBCs). During visits 2 to 11, a shortened interview was conducted to obtain updated sexual behaviours and intravaginal practices. A brief clinical examination was performed to obtain vaginal swabs for testing vaginal pH, bacterial vaginosis and yeast, and a CVL for the detection of HSV, PSA, soluble immune proteins, haemoglobin and WBCs. Urine was collected at every visit to test for pregnancy and menstrual cycle phase. If a woman was menstruating, no genital samples were obtained on that visit. No blood samples were collected; however laboratory data from the main cohort on HSV antibody status, HIV status and syphilis results were available for the statistical analysis and methods have been reported in a previous study [32]. Colposcopy was carried out by trained clinicians according to the CONRAD/WHO revised manual for the Standardization of Colposcopy for the Evaluation of Vaginal Products [36].

Sample processing and laboratory testing

CVL sample processing. CVL samples were obtained by flushing 5ml of sterile normal saline with a sterile pipette over the cervix and the lateral vaginal walls. After 60 seconds, the fluid was aspirated from the posterior fornix using the same pipette and collected in a 15 ml conical polypropylene tube that was then stored temporarily in a cool box with ice (2–8°C) before processing. Visual appearance was noted and documented, and 10µl of samples was used to assess the sample for haemoglobin using Hemastix® reagent indicator strips (Bayer Diagnostics, Tarrytown, NY, USA), comparing the indicator colouring with colour categories representing approximate quantities of erythrocytes (ery) per uL as specified by the manufacturer: none, low (25 ery/µL), moderate (80 ery/µL), high (200 ery/µL). CVLs were centrifuged onsite at 3500 rpm for 10 minutes within two hours of collection. Protease inhibitor (Cocktail Set I, Calbiochem, Merck Millipore, Darmstadt, Germany) was added to two 1 ml aliquots for immune protein testing, and two 1ml aliquots were stored for batch testing for PSA and HSV viral quantification. CVL supernatant aliquots were stored at -20°C for 3 days and then at -80°C for long-term storage before shipping or batch testing.

We developed a method to enumerate the WBCs in the cell pellet modified from a manual blood count: the cell pellet was spread across a glass slide, fixed with methanol and air-dried. Leishman's stain was applied to the slide for 10 minutes. With a light microscope at 100 times magnification, WBCs were counted and recorded on a differential cell counter until 100 WBCs were counted. Erythrocytes and epithelial cells were enumerated on a separate counter, and were not included in the 100-cell differential count.

Measurement of vaginal pH and reproductive tract infections. At the first and last visit, an endocervical swab was tested for detection of *N. gonorrhoeae* and *C. trachomatis* by AmpliCor PCR kits (Roche Diagnostics, Branchburg, USA). All positive tests for *N. gonorrhoeae* were confirmed using specific primers to the 16S DNA coding region in PCR in-house assays [37]. A vaginal swab was obtained to inoculate a culture (TV InPouch, Biomed Diagnostics, San Jose, USA), which was read for the presence of motile trichomonads by light microscopy at 72 hours after incubation at 36–37°C.

At each visit, CVL supernatants were tested for HSV shedding using Artus HSV-1/2 PCR kits (Qiagen, Hilden, Germany). A vaginal swab was Gram-stained and examined and Nugent scored for diagnosis of bacterial vaginosis [38]. A third swab was rolled onto a pH indicator strip (ThermoFisher, Waltham, MA, USA); the indicator colouring was compared with colour zones representing the following pH values: 3.6; 4.1; 4.4; 4.7; 5.0; 5.3; 5.6 and 6.1. The same swab was used for wet mount microscopy for the detection of yeast hyphae or buds.

Measurement of PSA. We measured PSA at every visit using a quantitative PSA ELISA Kit (Calbiotech, Inc, Spring Valley, CA, USA) for the detection of PSA in human serum for cancer detection. A random selection of CVL supernatant aliquots were shipped to the Institute of Tropical Medicine in Antwerp, Belgium for evaluation against the SERATEC PSA Semi-quant (Göttingen, Germany). The SERATEC test is a semi-quantitative chromatographic immunoassay also originally developed for human serum for cancer detection, and subsequently validated for the detection of PSA in vaginal fluid for professional forensic purposes with a sensitivity and specificity of 100% [39]. The Calbiotech test was found to be 70% sensitive and 100% specific for PSA in the cervical vaginal lavages against SERATEC when combining low positive (<4ng/ml) and negative values.

Measurement of immune proteins and peptides. Twenty-three soluble immune protein were quantified by in house multiplex bead immunoassay as previously described (Panels a and b of Figure A in S1 Fig) [40,41]. Following the same protocols, two more panels were added to measure human beta defensins(HBD)3, apolipoprotein(APO)A1, squamous cell

carcinoma antigen(SCCA)-1, polymeric immunoglobulin receptor (PIGR), IL-10, IL-17, IL-18 and transforming growth factor(TGF)- α (Panel c of Figure A in [S1 Fig](#)) and SLPI, elafin, HBD2, HBD3, α -defensin/human neutrophil peptide (HNP) 1–3, involucrin and S100 calcium binding protein A8 (S100a8) (Panel e of Figure A in [S1 Fig](#)). Additionally, six immunoglobulin (Ig) isotypes (IgG1, IgG2, IgG3, IgG4, IgA, IgM) were measured using Milliplex map kit (Merck Millipore, Billerica, USA; Cat No: HGAM-301) following the manufacturer's protocol (Panel d of Figure A in [S1 Fig](#)). Total protein was measured using Quick Start™ Bradford Protein Assay (Bio-Rad Laboratories, Hercules, USA).

Measurement of menstrual cycle phase. Urine aliquots were stored at -80°C for testing for Pregnanediol-3-Glucuronide (PDG, EIA, Immunometrics UK Ltd, London, United Kingdom), the principal metabolite of progesterone, and for testing of creatinine (CRT, R&D Systems, Inc., Minneapolis, MN, USA). Each PDG concentration was indexed by the CRT concentration of the same sample to adjust for urine dilution. We used a modified Kassam method to identify ovulatory cycles during the study; we used the minimum PDG/CRT value for each woman during the study as the denominator, and the per-visit PDG/CRT value as the numerator [42]. A ratio threshold of greater than 4.0 signalled that ovulation had taken place [42,43]. This method has been used for twice weekly and every other day sampling, and found to be 100% sensitive and 77% specific with 6% misclassification [43]. In those women who reported no hormonal contraceptive use, who showed evidence of ovulation, and who had one observed menstruation, we then examined the PDG curves to identify the rise in PDG. We defined “Day 1” of the menstrual cycle as the first visit with observed menstruation. All visits after Day 1 were defined as pre-ovulation until a rise in PDG was seen, and all subsequent visits were defined as post-ovulation until menstruation. If there was no obvious PDG rise, visits were not categorized for analysis.

Statistical analysis

Data were analyzed using Stata, version 12 (StataCorp, College Station, USA). We restricted the data set to “healthy visits” defined as visits without a positive STI test or reproductive tract infection. Visits were excluded from this analysis if results were positive for HSV shedding, vaginal yeast or bacterial vaginosis. In addition, all follow-up visits for women who tested positive at the first or last visit for *C. trachomatis*, *N. gonorrhoeae*, and *T. vaginalis* were excluded from this analysis.

CVL samples with analyte concentrations below the lower limit of quantification (LLOQ) of the assay were assigned a concentration of half the LLOQ. Those above the upper limit of quantification (ULOQ) were assigned values twice the ULOQ. Spearman's rank correlation coefficient was calculated for each of the analytes within a Luminex panel to look for evidence of cross-reactivity (Figure A in [S1 Fig](#)).

The proportion of samples with concentration above the LLOQ, and the median and range of concentrations above the LLOQ, were calculated for each analyte. Since most analytes showed skewed distribution, concentrations were log₁₀ transformed. To characterize the variation of the analytes over time between and within women, we used mixed-effects linear regression to estimate the ICC of each log-transformed value; variance components were estimated using residual maximum likelihood. The ICC was calculated as $\sigma_B^2 / (\sigma_B^2 + \sigma_W^2)$, where σ_B^2 is the between-women variance and σ_W^2 is the within-woman variance. An ICC of 0 implies that observations from the same woman are no more similar to each other than they are to observations from different women. An ICC of 1 implies that all observations from the same woman are identical, so that the variation is due to between-woman differences. We also explored the effects of the dilution factor of the CVL by ICC (Table A in [S1 Table](#) and [S1 Text](#)).

We examined the association of analytes concentrations with the following exposures: menstrual cycle stage, hormonal contraception (reported use of combined oral contraceptive [COC] or injectable depot medroxyprogesterone acetate [DMPA]), seminal plasma, reported vaginal practices, cervical ectopy, colposcopic findings, and vaginal pH. Since detailed clinical examinations were only done at Visits 1 and 12, ectopy and colposcopy results were not available at other time points. To explore the association with the exposures of interest, we used mixed-effect linear regression for analytes with concentration >85% above LLOQ, or logistic regression with random effects for analytes with concentration \leq 85% above LLOQ.

For the analysis, PSA was categorised into three levels: no PSA detected; low positive (<4ng/ml); and high positive (\geq 4ng/ml). Neutrophil counts were categorised into the following levels per 100 WBCs: no cells, 1–10 cells, 11–50 cells, >50 cells, and lymphocytes were analysed as presence/ absence. Vaginal pH results were categorised into the following levels: 3.6–4.1 (normal pH); 4.4–4.7 (high normal pH); and 5.0 and above (abnormal pH).

For each of the analyses of intravaginal cleansing with soap, intravaginal cleansing with cloth, and intravaginal insertion (i.e. the insertion of pulverized herbs or detergents), we compared samples from visits in which women reported the specific intravaginal practice to visits in which no cleansing or cleansing with fingers and water only was reported. We excluded visits in which women reported insertion of prescribed medications (e.g. treatment for candidiasis), leaving all but two women inserting detergents.

In the multivariable analysis, we considered age, reported sex in the past 3 days, and the presence of haemoglobin in the CVL as *a priori* confounders based on a conceptual model (Figure B in [S1 Fig](#)). For menstrual cycle phase, hormonal contraceptive use, intravaginal practices, clinical cervical ectopy, colposcopic abnormalities, and vaginal pH we controlled for the effects of age, reported sexual intercourse in the past three days and presence of haemoglobin; for PSA we controlled for age and presence of haemoglobin.

Results

Demographics, behavioural characteristics and clinical findings of the participants

One hundred participants attended 1,108 (92%) of 1,200 possible visits over follow-up. Of the 1,108 visits, 956 (86%) were non-menstruating, and of these, 370 (39%) met the definition of healthy visits; 67 women contributed at least one visit to this analysis. The participant demographic, behavioural, contraception and menstrual cycle data, and clinical and laboratory findings by visit are presented in [Table 1](#). The mean age was 26 years old (range 18–44 years). Approximately one-quarter (28%) reported current use of hormonal contraception (i.e. COC or DMPA) at enrolment, 81% reported sexual intercourse during the study period, and 100% reported practicing vaginal cleansing during the study period. For cleansing, 81% used soap, 21% used a cloth (versus fingers alone), and 39% reported insertion of a substance into the vagina (e.g. herbs) at least once during the study.

There was evidence of ectopy at 28 of 71 (39%) possible healthy visits; the majority of cases of ectopy involved less than 20% of the cervix face. Colposcopy findings were detected at 21% (16/71) of visits: 15 cases were cervical findings (13 petechiae, 1 erythema and 1 ecchymosis); and one was on the anterior fornix (petechiae). There were no findings of oedema, grossly white findings, peeling, ulcer, abrasion or laceration.

In over half of the visits (58%), the vaginal pH was between 4.7 and 5.0, with only 14% of visits having a pH >5.0. Overall, 49% tested positive for PSA. Two-thirds of the samples (67%) tested positive for haemoglobin; 20% of samples with detectable haemoglobin were at visits flanking an observed menstrual period. However, of the 65 samples with high haemoglobin

Table 1. Demographics, behaviour and clinical characteristics.

Demographic and behavioural characteristics (N = 67)	N(%)
Age (<i>Mean, range</i>)	26 (18–44)
Reported vaginal intercourse during the study	54 (81%)
Reported intravaginal cleansing during the study	
Any cleansing	67 (100%)
Use of soap ¹	54 (81%)
Use of cloth	14 (21%)
Reported intravaginal insertion during the study	
Any insertion	26 (39%)
Medication	6 (9%)
Detergent ¹	18 (27%)
Herbs	1 (2%)
Tobacco	1 (2%)
Contraception at enrolment (N = 67)	
No contraception	19 (28%)
Condom use only	23 (34%)
Depot medroxyprogesterone acetate (DMPA)	13 (19%)
Combined oral contraceptive (COC)	6 (9%)
Sterilization	3 (5%)
IUD	1 (2%)
Other	2 (3%)
Menstrual cycle	
The number of cycles that could be assigned as ovulatory ² (n = 48)	46 (96%)
The number of menstruations during the study per participant ³ (n = 46)	
No menstruation observed	11 (24%)
One menstruation	33 (72%)
Two menstruations	2 (4%)
Visits that could be assigned a menstrual cycle phase ⁴ (n = 102 visits)	
Pre-ovulation	56 (55%)
Post-ovulation	46 (45%)
Clinical and laboratory findings by visit (N = 370)	
Cervical ectopy at V1 & V12 (n = 71 visits)	
Absent	43 (61%)
<20%	27 (38%)
≥20%	1 (1%)
Colposcopic findings ⁵ at V1 & V12 (n = 71 visits)	
Absent	55 (78%)
Vaginal	0 (0%)
Fornices	1 (0.01%)
Cervical	15 (21%)
Vaginal pH measured at each visit ⁶ (V1-V12; n = 361 visits)	
3.6–4.1	66 (19%)
4.4–4.7	145 (40%)
5.0 +	150 (72%)
Prostate specific antigen (PSA) measured at each visit (V1-V12; n = 370 visits)	
Negative	189 (51%)
Low positive (<4ng/ml)	118 (32%)
High positive (≥4ng/ml)	63 (17%)

(Continued)

Table 1. (Continued)

Demographic and behavioural characteristics (N = 67)	N(%)
Haemoglobin presence in CVL measured at each visit ⁷ (V1-V12; n = 369 visits)	
Negative	123 (33%)
25 Ery/uL (+)	121 (33%)
80 Ery/uL (++)	60 (16%)
200 Ery/uL (+++)	65 (18%)

1. Soaps were defined as soaps meant for cleansing the body, while detergents were for cleaning clothes, dishes, etc.
2. We used a modified Kassam method to identify ovulatory cycles during the study; we used the minimum pregnanediol 3-glucuronide/ creatinine (PDG/CRT) value for each women during the study as the denominator, and the per-visit PDG/CRT value as the numerator. A ratio threshold of greater than 4.0 signalled that ovulation had taken place [51,52].
3. In those women who reported no hormonal contraceptive use and showed evidence of ovulation.
4. In those women who reported no hormonal contraceptive use, showed evidence of ovulation, and with one observed menstruation, pre and post ovulation was defined by the rise in pregnanediol 3-glucuronide.
5. The colposcopic findings on the fornix were petechiae (1 case), and on the cervix were petechiae (13 cases) erythema (1 case) and ecchymosis (1 case). There were no findings of oedema, grossly white findings, peeling, ulcer, abrasion or laceration.
6. Vaginal pH was measured using ThermoFisher pH test strips (range 3.6–6.1) with a colour chart.
7. Haemoglobin was measured using Hemastix® test strips with a colour chart.

doi:10.1371/journal.pone.0143109.t001

(≥200 ery/uL), only 46% were at the visit before and after an observed menstrual period. Thus, menstrual blood may have accounted for some, but not all of the haemoglobin found in the samples. WBCs were found in 51% of the CVL cell pellets, and these were mostly neutrophils with a lower proportion of lymphocytes.

In the 48 women who did not report hormonal contraception use, 46 (96%) showed signs of having ovulatory cycles; however, of these, 11 (24%) did not have an observed menstruation during the study period, and two had two separate menstruations. In the 33 women who showed evidence of ovulation and had one observed menstruation, 102 of 169 (63%) visits could be assigned as either pre-ovulatory or post-ovulatory.

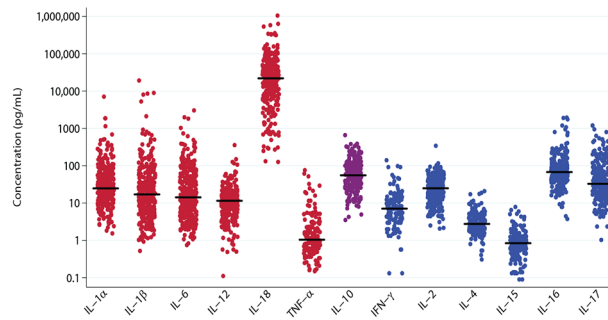
Expression patterns for soluble immune proteins in cervicovaginal lavages (CVLs)

Distributions of the analyte concentrations are visualized in Fig 1. Most analytes were detectable by the assays that were used, with only 13 out of 38 analytes having concentrations below LLOQ in ≥15% of visits at which they were measured (Table 2). For the comparison of biomedical and behavioural factors, these analytes were analysed as binary variables (i.e. presence/absence). Details of the median, range, mean and standard deviation, as well as percentages detected for each analyte from healthy visits are given in Table 2.

Inter- and intra-woman variation in analyte concentration differed from one analyte to another (Table 2). For all analytes, there was strong evidence of a correlation between repeated samples within the same woman over time ($p < 0.001$). For most analytes, the ICC was close to 0.50 (median = 0.45, IQR = 0.38–0.51).

Menstrual cycle phase signature. Compared to samples from visits during pre-ovulation, samples from visits during post-ovulation had increases in IL-18, IL-10, IL-17, and TGF-β, and a decrease in monocyte chemoattractant protein 1 (MCP-1), stromal cell-derived factor 1 (SDF-1β),

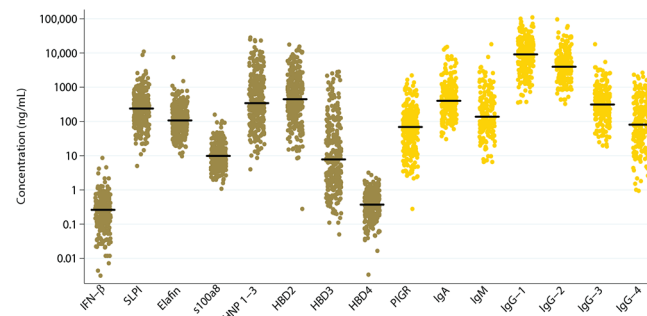
(a) Inflammatory cytokines, anti-inflammatory cytokines, and adaptive cytokines



(b) Chemokines and growth factors



(c) Antimicrobial proteins and immunoglobulins



(d) Controls

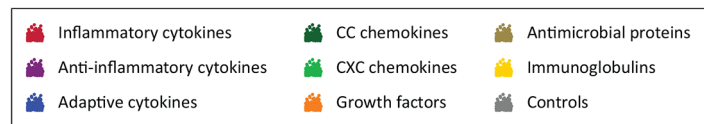
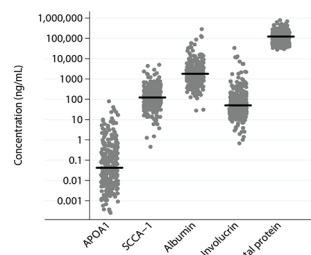


Fig 1. Distribution of analyte concentrations in cervicovaginal lavage samples for 370 healthy visits. Each data point represents a single sample and the line through data points represents the median concentration. APOA1 and albumin are serum controls and SCCA-1 and involucrin are vaginal epithelial controls.

doi:10.1371/journal.pone.0143109.g001

Table 2. The proportion of samples with analytes detected, median, mean and intra-class correlation coefficients (ICC) from 370 CVL samples.

Analyte	Percent with concentrations above LLOQ ¹	Median (range) CVL concentration	Mean (standard deviation) CVL concentration	σ ; ICC; σ_w of log-transformed data ² (N = 302) ^{3,4}
Inflammatory cytokines				
		pg/ml	pg/ml	
IL-1 α /IL-1F1	100%	24.7 (1.5–7093.6)	83.4 (401.1)	0.35; 0.51; 0.35
IL-1 β /IL-1F2	93%	17.0 (0.5–19069.8)	204.5 (1330.4)	0.58; 0.52; 0.55
IL-6	95%	14.2 (0.7–3049.1)	68.0 (234.2)	0.50; 0.48; 0.53
IL-12 ⁵	77%	11.5 (0.1–356.1)	16.2 (26.7)	0.46; 0.55; 0.42
IL-18 (n = 301)	92%	21960.0 (125.0–1055770.0)	49244.1 (100235.8)	0.54; 0.46; 0.58
TNF- α ⁵	41%	1.0 (0.2–75.2)	3.9 (9.8)	0.36; 0.52; 0.35
Anti-inflammatory cytokines				
		pg/ml	pg/ml	
IL-10 (n = 301)	86%	55.3 (3.5–660.5)	78.1 (78.3)	0.34; 0.43; 0.39
Adaptive cytokines				
		pg/ml	pg/ml	
IFN- γ ⁵	36%	7.1 (0.1–140.0)	12.7 (19.3)	0.31; 0.42; 0.37
IL-2	93%	24.8 (2.1–342.0)	30.5 (25.8)	0.30; 0.55; 0.27
IL-4 ⁵	49%	2.8 (0.3–20.9)	3.6 (3.1)	0.25; 0.40; 0.31
IL-15 ⁵	44%	0.8 (0.1–7.8)	1.1 (1.2)	0.33; 0.57; 0.29
IL-16 ⁵	69%	67.5 (3.7–1931.3)	128.2 (238.4)	0.36; 0.50; 0.36
IL-17 ⁵ (N = 301)	82%	32.7 (1.0–1203.1)	87.9 (162.9)	0.45; 0.49; 0.46
CC Chemokines				
		pg/ml	pg/ml	
MIP-1 α ⁵ /CCL3	60%	7.8 (0.6–49.3)	10.1 (7.8)	0.27; 0.41; 0.33
MIP-1 β /CCL4	89%	23.1 (0.5–760.6)	46.1 (83.5)	0.30; 0.41; 0.37
MCP-1/CCL2	88%	14.8 (1.8–6034.7)	75.4 (368.0)	0.38; 0.31; 0.57
MCP-2 ⁵ /CCL8	63%	8.4 (0.2–74.0)	9.8 (8.5)	0.33; 0.44; 0.38
RANTES/ CCL5	86%	4.0 (0.2–555.2)	17.5 (56.8)	0.52; 0.54; 0.48
CXC Chemokines				
		pg/ml	pg/ml	
IP-10/ CXCL10	99%	287.5 (7.4–4686.0)	574.8 (768.2)	0.42; 0.52; 0.41
SDF-1 β / CXCL12 ⁵	41%	71.7 (17.4–2071.5)	163.0 (295.9)	0.30; 0.51; 0.30
MIG/CXCL9 ⁵	75%	5354.9 (174.2–115344.4)	9778.1 (14319.5)	0.58; 0.61; 0.46
IL-8	100%	491.1 (4.6–82557.8)	1317.0 (4957.9)	0.37; 0.46; 0.40
Growth Factors				
		pg/ml	pg/ml	
GM-CSF	95%	4.4 (0.2–84.9)	5.9 (6.2)	0.31; 0.49; 0.32
G-CSF	92%	30.4 (2.2–225.6)	261.1 (476.0)	0.50; 0.55; 0.45
TGF- α	89%	40.1 (0.4–322.9)	36.8 (28.2)	0.33; 0.57; 0.28
TGF- β	99%	104.3 (0.8–4507.0)	54.5 (47.5)	0.34; 0.37; 0.45
Antimicrobial proteins				
		ng/ml	ng/ml	
IFN- β ⁵	64%	0.3 (0.0–8.6)	0.4 (0.8)	0.42; 0.42; 0.50
SLPI	100%	238.5 (5.0–10892.2)	441.8 (869.3)	0.28; 0.33; 0.40
Elafin	100%	107.0 (9.6–7548.3)	185.7 (459.0)	0.22; 0.41; 0.26
s100a8 (n = 302)	100%	9.9 (1.1–159.7)	13.9 (15.8)	0.21; 0.37; 0.27
HNP 1–3	100%	342.9 (4.0–28582.2)	1635.1 (3703.9)	0.45; 0.38; 0.57
HBD2 (n = 302)	95%	446.2 (0.3–17689.2)	1374.6 (2435.7)	0.49; 0.38; 0.62
HBD3 (n = 301)	97%	7.8 (0.1–2807.2)	132.8 (410.9)	0.57; 0.32; 0.83
HBD4 (n = 302)	86%	0.4 (0.0–3.2)	0.5 (0.5)	0.32; 0.49; 0.33
Immunoglobulins				
		ng/ml	ng/ml	
pIgR (n = 301)	99%	68.9 (0.3–2198.1)	144.2 (250.5)	0.41; 0.42; 0.49
IgA (n = 230)	96%	401.1 (30.3–14977.4)	1016.9 (2024.7)	0.37; 0.36; 0.50
IgM ⁵ (n = 230)	75%	137.6 (6.6–17905.0)	562.0 (1594.1)	0.51; 0.44; 0.57

(Continued)

Table 2. (Continued)

Analyte	Percent with concentrations above LLOQ ¹	Median (range) CVL concentration	Mean (standard deviation) CVL concentration	σ ; ICC; σ_w of log-transformed data ² (N = 302) ^{3,4}
IgG1 (n = 230)	99%	9123.7 (355.3–110238.7)	14815.8 (17047.5)	0.23; 0.14; 0.57
IgG2 (n = 230)	90%	3963.0 (325.8–96848.8)	7478.8 (10841.0)	0.41; 0.48; 0.43
IgG3 (n = 230)	97%	312.3 (18.6–18261.1)	666.5 (1466.2)	0.33; 0.20; 0.67
IgG4 (n = 230)	90%	80.7 (0.9–2662.5)	250.7 (422.5)	0.71; 0.68; 0.48
Controls		ng/ml	ng/ml	
APOA1 (n = 302)	100%	0.04 (0.0–78.6)	1.3 (6.0)	0.65; 0.36; 0.86
SCCA-1 (n = 301)	97%	122.7 (0.5–5072.3)	254.2 (503.8)	0.36; 0.31; 0.55
Albumin (n = 302)	100%	1802.4 (27.7–284126.9)	5761.3 (20514.8)	0.38; 0.46; 0.42
Involucrin (n = 302)	100%	50.1 (0.7–33668.8)	431.9 (2351.2)	0.47; 0.46; 0.51
Total protein	100%	123290.0 (27517.2–786809.2)	148872.1 (104461.5)	0.19; 0.42; 0.22

1. Samples with analyte concentrations below the lower limit of quantification (LLOQ) of the assay were assigned a concentration of half the LLOQ.
2. σ = variance between women; ICC = intra-class correlation coefficient; σ_w = within-woman variance.
3. Restricted to 302 samples in which involucrin was measured.
4. For all of the analytes, there is strong evidence of clustering within women—i.e. strong evidence that ICC >0; the p-value is <0.001 calculated by ANOVA, using an F test of between-women variation/within-women variation.
5. Biomarkers with <85% observations below the limit of quantification

doi:10.1371/journal.pone.0143109.t002

IgA, IgG2 and IgM (n = 102; Fig 2; Table B in S1 Table). There were no detectable difference in expression of antimicrobial proteins with menstrual cycle phase.

Hormonal contraception signatures. Compared to women who reported no hormonal contraceptive use, women reporting DMPA use had an increase in concentration of several soluble proteins of the innate and adaptive immune system, including IL-1 α , IL-1 β , IL-6, TNF- α , IL-2, IL-4, IL-16, interferon (IFN)- γ , macrophage inflammatory protein (MIP)-1 α , MIP-1 β , MCP-2, IP-10, SDF- β , monokine induced by gamma interferon (MIG), IL-8, TGF- β , IFN- β , HBD4, IgA, IgG1, and IgG2 (n = 327; Fig 3A and 3B; Table B in S1 Table). Women reporting COC use had a similar signature with increases in IL-1 β , IL-2, IL-6, IL-8, MIP-1 β , MCP-2, G-CSF, HNP 1–3, HBD4, and SCCA-1 (n = 305; Fig 3C and 3D; Table B in S1 Table).

Signature for the detection of prostate-specific antigen. There was modest evidence of a positive linear correlation between PSA levels and IL-6, MCP-1, SLPI, APOA1, and evidence of a negative linear correlation with s100a8, GM-CSF, and HBD3 (n = 370; Fig 4; Table B in S1 Table).

Intravaginal practice signatures. For the analysis of cleansing with soap, the analyte signature showed no evidence of a difference between women who used soap versus no cleansing or cleansing with water alone (Table B in S1 Table). For the analysis of cleansing with cloth, there was evidence of an increase in the innate and adaptive immune system proteins, including IL-1 β , IL-2, IL-8, MIP-1 β , SDF-1 β , RANTES, IP-10, HNP 1–3, HBD4, IFN- β , and immunoglobulins (n = 155; Fig 5A and 5B; Table B in S1 Table). For the analysis of insertion, with exception of IL- β , TNF- α and IFN- γ , most of the other analytes trend towards a decrease for visits with reported insertion (n = 145; Fig 5C and 5D; Table B in S1 Table).

Analyte signatures from clinical findings. For the analysis of cervical ectopy, we compared samples from women who had any ectopy at the first and last visit to samples from

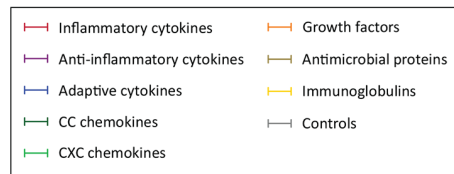
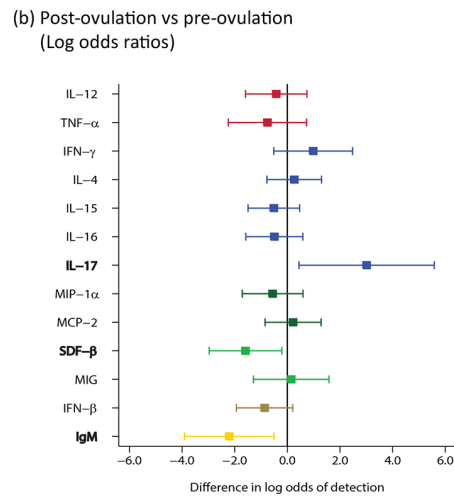
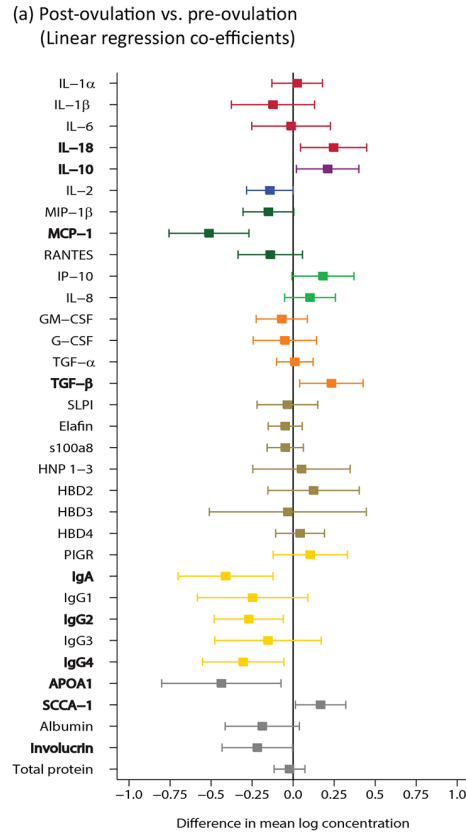
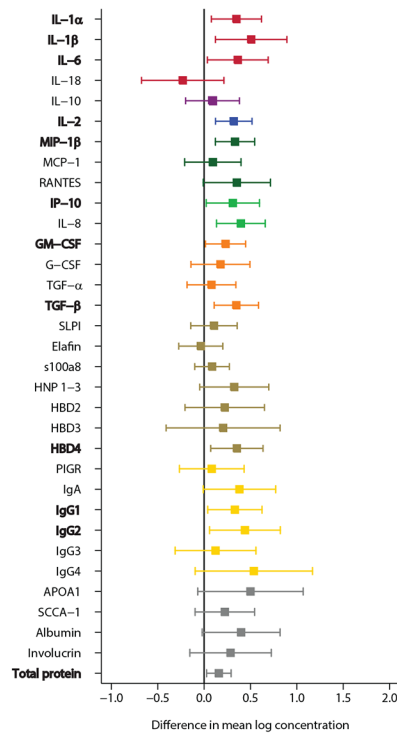


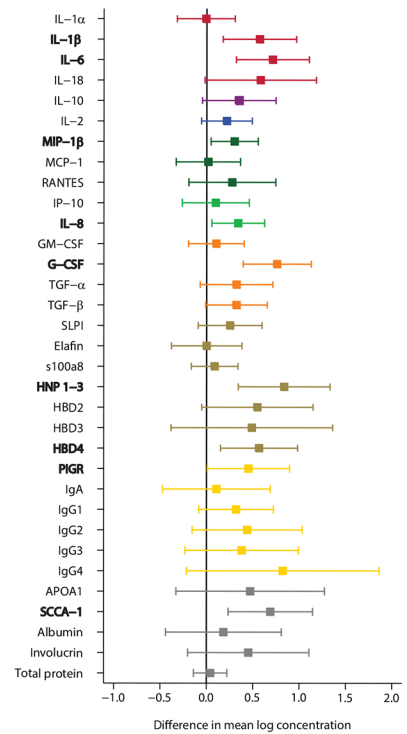
Fig 2. Menstrual cycle phase (n = 102). A comparison of analyte concentration between samples from visits occurring post-ovulation to pre-ovulation (reference). Menstrual cycle stage was assessed by measurement of urine pregnanediol 3-glucuronide. Bolded analytes represent associations with a p-value ≤ 0.05 . (A) Analytes with $\geq 85\%$ detection using linear regression showing coefficients (boxes) and confidence intervals (lines) (B) Analytes with $< 85\%$ detection using logistic regression showing odds ratios (boxes) and confidence intervals (lines).

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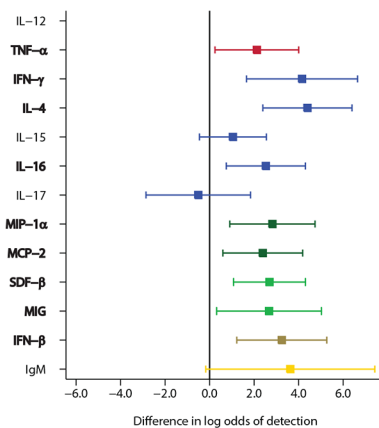
(a) DMPA vs no hormonal contraception (Linear regression coefficients)¹



(c) COCs vs no hormonal contraception (Linear regression coefficients)¹



(b) DMPA vs no hormonal contraception (Log odds ratio)



(d) COCs vs no hormonal contraception (Log odds ratio)

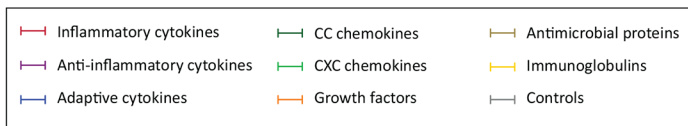
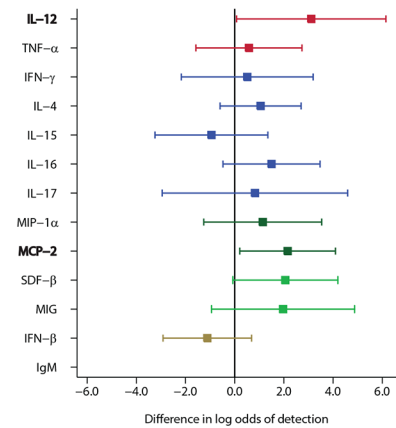


Fig 3. Reported hormonal contraceptive use (DMPA, n = 327; and COC, n = 305). Bolded analytes represent associations with a p-value ≤ 0.05 . (A) A comparison of analyte concentration $\geq 85\%$ between samples DMPA and women who reported no hormonal contraception use (reference) using linear regression.

(B) A comparison of analyte concentration <85% between samples from women who reported use of DMPA and women who reported no hormonal contraception use (reference) using logistic regression. (C) A comparison of analyte concentration $\geq 85\%$ between samples from women who reported use of COC and women who reported no hormonal contraception use (reference) using linear regression. (D) A comparison of analyte concentration <85% between samples from women who reported use of COC and women who reported no hormonal contraception use (reference) using logistic regression. Footnotes: 1. The x-axis range is from -1 to +2 which is wider than for all other figures; 2. For the association with DMPA and IL-12, odds ratios could not be estimated as all DMPA visits had detectable IL-12 levels; 3. For the association with COC use and IgM, odds ratios could not be estimated as all COC visits had detectable IgM levels.

doi:10.1371/journal.pone.0143109.g003

women who had no clinical ectopy. Samples from visits with clinical ectopy showed some evidence of an increase in IL-6 and G-CSF ($n = 67$; [Fig 6](#); Table B in [S1 Table](#)).

For the analysis of colposcopy, we compared samples from women who had any colposcopic finding (88% [14/16] were petechiae) at the first and last visit to samples from women who had no colposcopic findings. Samples from visits with colposcopic findings showed a decrease in IL-2, RANTES, and GM-CSF compared to visits without colposcopic findings ($n = 67$; [Fig 7](#); Table B in [S1 Table](#)).

There was some evidence for a negative linear correlation between pH and the concentration of inflammatory cytokine IL-18; adaptive cytokine IL-2; growth factors GM-CSF and TGF- β ; and HBD3 ($n = 361$; [Fig 8](#); Table B in [S1 Table](#)). There was evidence of a positive linear correlation of pH with IgA, IgG4 and APOA1.

Signatures from WBCs identified in the CVL cell pellet. We carried out analyses for neutrophils and lymphocytes only because there were too few other WBCs reported. There was evidence for strong positive linear correlation between neutrophil count and the concentration of analytes in the innate and adaptive immune system, including IL-1 α , IL-1 β , IL-6, MIP-1 β , IP-10, IL-8, G-CSF, TGF- α , HNP 1–3, HBD2, HBD4, immunoglobulins, APOA1, albumin, and total protein ($n = 361$; [Fig 9](#); Table B in [S1 Table](#)). For the analysis of lymphocytes, we compared samples that had any lymphocytes to those that had none. There was strong evidence of a positive association between samples that had detectable lymphocytes and IL-1 β , IL-6, RANTES, G-CSF, IgG2, and APOA1, albumin and total protein, and a negative association between samples with detectable lymphocytes and IgG1 ($n = 361$; [Fig 9](#); Table B in [S1 Table](#)).

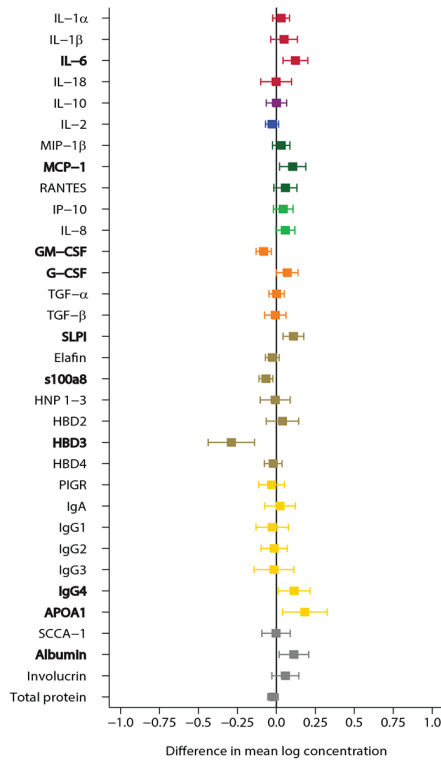
Signatures from samples with haemoglobin. There was evidence for strong positive linear correlation between haemoglobin category and concentration of IL-1 β , IL-6, IL-10, MIP-1 β , MCP-1, RANTES, IP-10, IL-8, G-CSF, TGF- α , immunoglobulins, APOA1, albumin and total protein ($n = 369$; [Fig 10](#); Table B in [S1 Table](#)).

Adjustments for age, recent sex and samples with haemoglobin. We controlled for the effects of age, reported sexual intercourse in the past three days and presence of haemoglobin. All signatures showed little change after adjustment (Figs C-K in [S1 Fig](#)). Adjusted coefficients can be found in Table C in [S1 Table](#).

Discussion

We measured 45 different soluble immune proteins and peptides in CVLs from women taking part in a microbicide feasibility study, and who would be appropriate for enrolment into Phase 3 clinical trials for HIV prevention candidate products. During healthy visits, we found differential expression of soluble immune proteins by hormonal contraception use, menstrual cycle stage, recent sexual exposure, clinical findings, and some types of traditional intravaginal practices. Increasingly, inflammatory cytokines and other soluble immune proteins are being used to assess the potential safety of candidate microbicides, vaccines or other HIV prevention

(a) Linear trend for change in log concentrations of PSA (Linear regression coefficients)



(b) Change in odds of analytes detection with one unit increase in PSA (Log odds ratio)

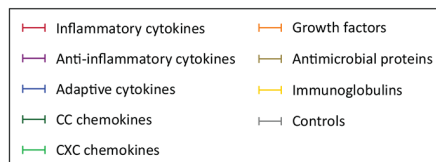
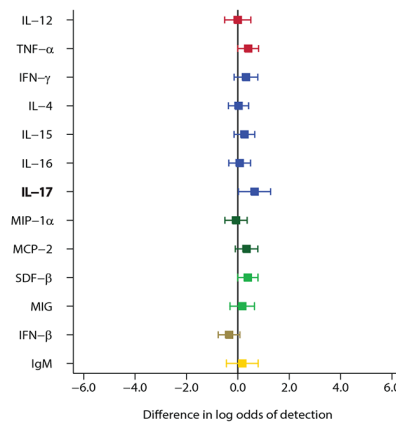
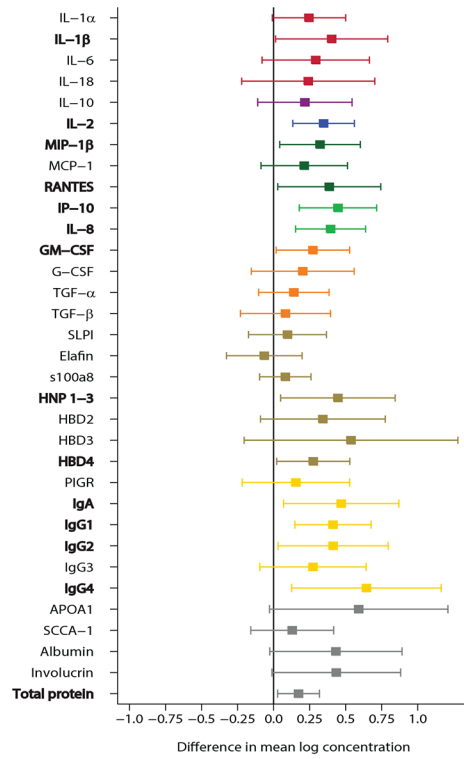


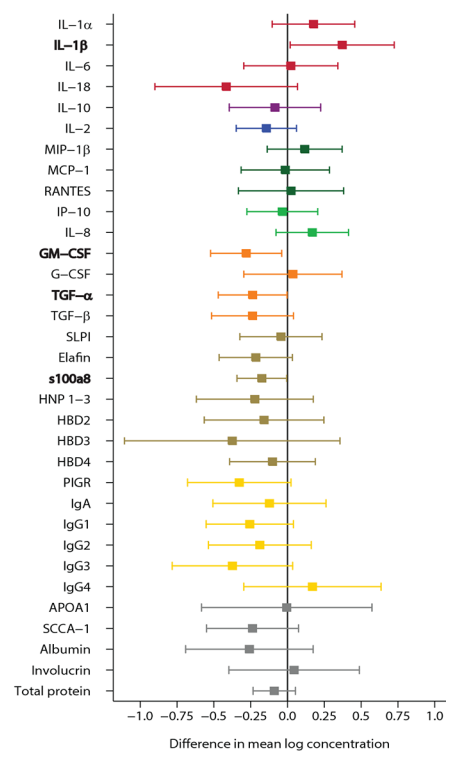
Fig 4. Prostate-specific antigen (PSA, n = 370). PSA categories: None, low positive (<4 ng/mL), high positive (≥ 4 ng/mL). Bolded analytes represent associations with a p-value ≤ 0.05 . (A) Analytes with $\geq 85\%$ detection using linear regression, linear trend for change in log concentration. (B) Analytes with $< 85\%$ detection using logistic regression, change in odds of analyte detection (if $< 85\%$ LLOQ) with one unit increase in exposure category.

doi:10.1371/journal.pone.0143109.g004

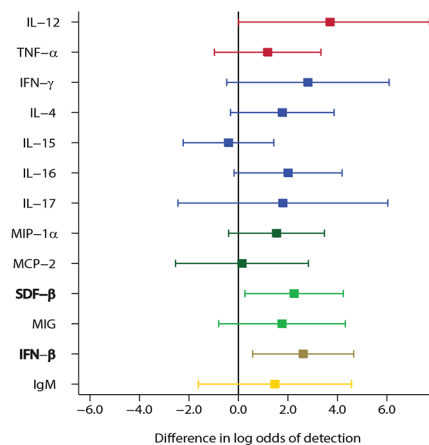
(a) Cleansing with clothes vs. no IVP or cleansing with water and fingers alone (Linear regression coefficients)



(c) Insertion vs. no IVP or cleansing with water and fingers alone (Linear regression coefficients)



(b) Cleansing with clothes vs. no IVP or cleansing with water and fingers alone (Logs odds ratios)



(b) Insertion vs. no IVP or cleansing with water and fingers alone (Logs odds ratios)

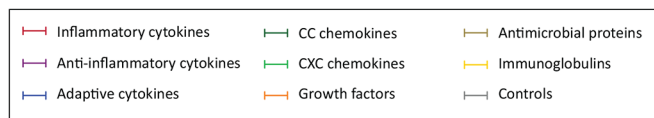
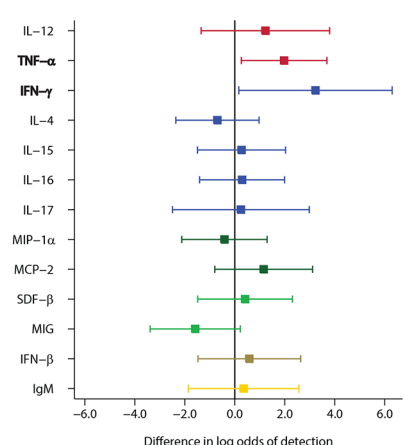


Fig 5. Traditional intravaginal practices (intravaginal cleansing with cloth, n = 155; and intravaginal insertion, n = 145). Bolded analytes represent associations with a p-value ≤ 0.05 . (A) A comparison of analyte concentration $\geq 85\%$ between samples from women who reported intravaginal cleansing with cloth

and women who reported no intravaginal cleansing use or intravaginal cleansing with water alone (reference) using linear regression. (B) A comparison of analyte concentration $<85\%$ between samples from women who reported intravaginal cleansing with cloth and women who reported no intravaginal cleansing use or intravaginal cleansing with water alone (reference) using logistic regression. (C) A comparison of analyte concentration $\geq 85\%$ between samples from women who reported intravaginal insertion and women who reported no intravaginal insertion (reference) using linear regression. (D) A comparison of analyte concentration $<85\%$ between samples from women who reported intravaginal insertion and women who reported no intravaginal insertion (reference) using logistic regression.

doi:10.1371/journal.pone.0143109.g005

products [12,13]. These behavioural and biological factors should be taken into account in the design, analysis and interpretation of clinical trials using these analytes.

DMPA and COCs are the most popular forms of contraception in East and southern Africa [44]. A recent meta-analysis has suggested that DMPA, but not COCs, increase risk for HIV acquisition [45]. Our study showed an increase in innate and adaptive, inflammatory and anti-inflammatory analytes, signalling an overall heightened inflammatory state among women using DMPA and a similar signature for women using COCs. This is consistent with two recent studies showing increases in cytokines and chemokines among women using DMPA [46,47] and COCs [46]. Both studies suggest that the reported increase in RANTES among DMPA users may be a key factor underlying epidemiological associations between DMPA use and increased HIV acquisition; while COCs users showed no evidence of an increase in RANTES [46]. In our study, we see a trend towards an increase in RANTES among DMPA users; however, confidence intervals are wide. More studies are needed to investigate safety of effective contraception among women at high risk for HIV infection.

In contrast to hormonal contraception signatures, menstrual cycle phase signatures had relatively less differential expression. Consistent with our findings, both human and macaque studies have shown that IgA and IgM levels in the cervical mucus are decreased during the post-ovulation phase [48,49]. Published literature report differential expression of other soluble immune proteins by menstrual cycle phase [50], but the evidence is less consistent in *in vivo* studies. For example, concentrations of IL-1 β and IL-6 have been found to be lower post-ovulation in one study of HIV-negative, pre-menopausal women in the US [24], but in a study among women living with HIV infection, levels of IL-1 β and IL-6 were similar during pre-ovulation and post-ovulation phases [23]. In a study of healthy Belgian women, levels of IL-1 β and IL-6 were also similar during pre-ovulation and post-ovulation phases, but there were increases in IL-1 α and HBD1-3 post-ovulation. Additionally, in a small study that obtained vaginal biopsies from women pre-ovulation and post-ovulation, there were no differences in immune cell populations [51].

We measured PSA as a biomarker of recent seminal plasma exposure and found some evidence of differential expression with increasing PSA. A study of healthy Belgian women also reported a strong correlation between PSA and IL-6 [21]. Seminal plasma has been reported to stimulate pro-inflammatory cytokines and chemokines such as IL-8, IL-6, MCP-1 and GM-CSF in immortalized cervical cells, as well as stimulate infiltration of the cervix by macrophages, dendritic cells, and T lymphocytes [26]. It is thought that inflammation may facilitate preparation of the female reproductive tissues for pregnancy through clearance of pathogens, sperm selection, and induction of immune tolerance toward the semi-allogeneic embryo [52]. Additionally, epithelial microabrasions from sexual intercourse may cause immune activation; a small study reported that microabrasions were detected in 60% of healthy women after consensual intercourse [53].

Cervical ectopy typically occurs during times of increased oestrogen levels, including adolescence, pregnancy and among women taking oestrogen-containing hormonal contraceptives.

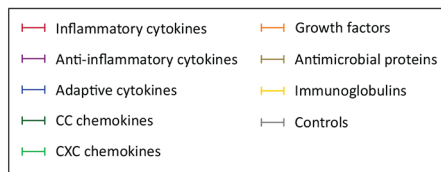
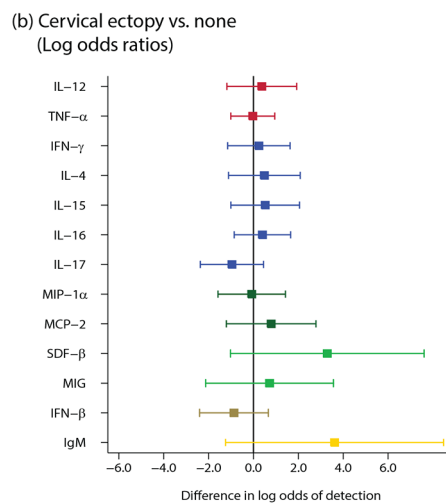
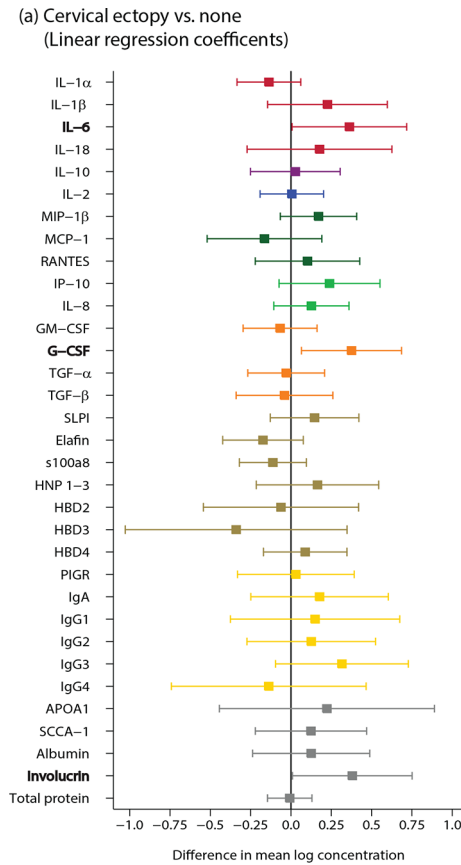


Fig 6. Clinical cervical ectopy (n = 67). Bolded analytes represent associations with a p-value ≤ 0.05 . A comparison of analyte concentration between samples among women with cervical ectopy and women without ectopy (reference). (A) Analytes with >85% detection using linear regression; (B) Analytes with <85% detection using logistic regression.

doi:10.1371/journal.pone.0143109.g006

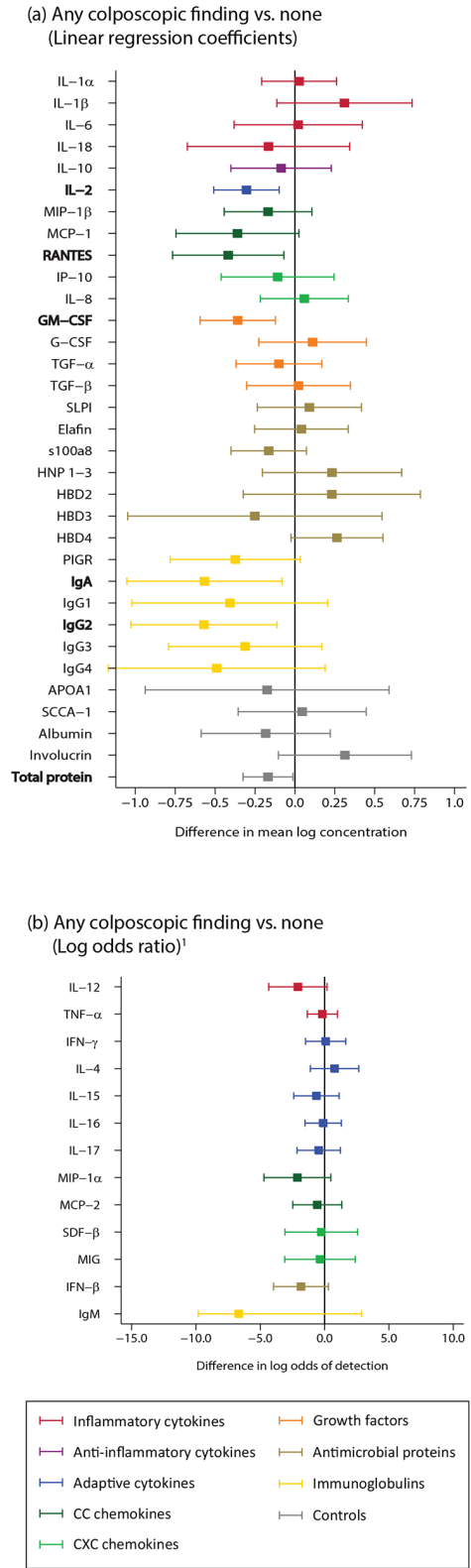
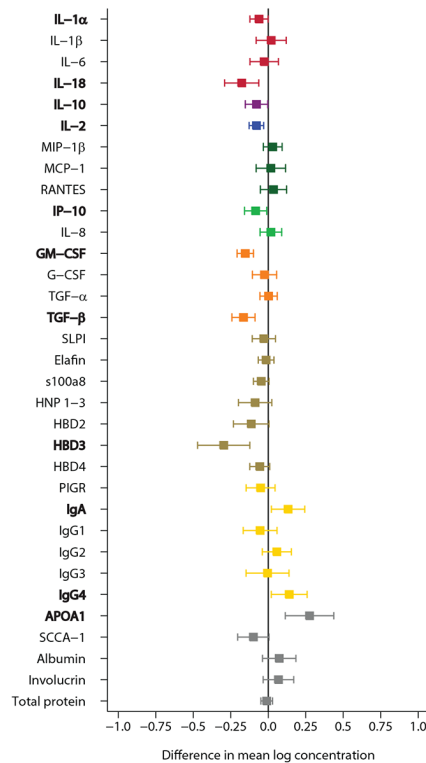


Fig 7. Colposcopy examination (n = 67). Bolded analytes represent associations with a p-value ≤ 0.05 . A comparison of analyte concentration between samples among women with colposcopic findings and women without colposcopic findings (reference). (A) Analytes with $>85\%$ detection using linear regression; (B) Analytes with $<85\%$ detection using logistic regression. Footnote: 1. The x-axis range is from -15.0 to +10.0 which is wider than for all other figures.

doi:10.1371/journal.pone.0143109.g007

(a) Linear trend for change in log concentration of vaginal pH (Linear regression coefficients)



(b) Change in odds of analyte detection with one unit increase in vaginal pH (Log odds ratio)

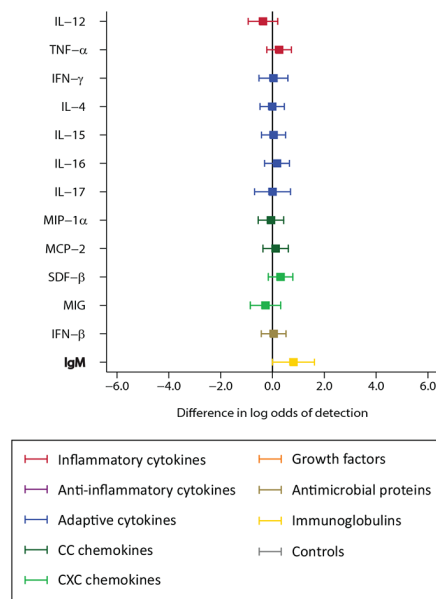
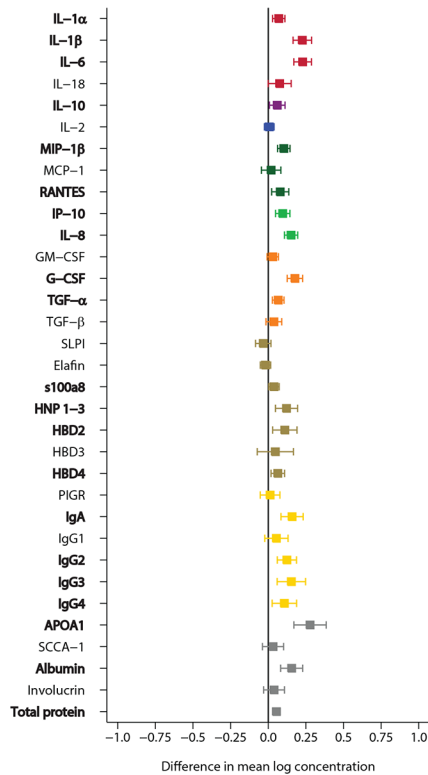


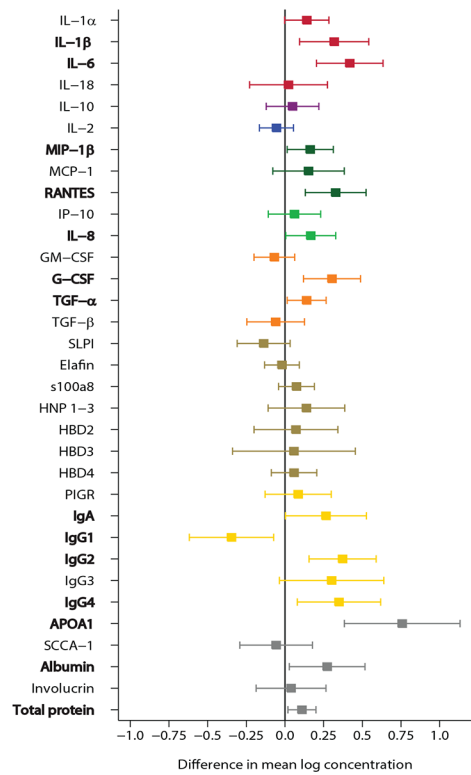
Fig 8. Vaginal pH (n = 361). Bolded analytes represent associations with a p-value ≤ 0.05 . Vaginal pH was measured with test strips during the clinical examination: 3.6–4.1 (normal pH); 4.4–4.7 (high normal pH); and 5.0 and above (abnormal pH). (A) For analytes with $>85\%$ detection using linear regression, linear trend for change in log concentration. (B) Analytes with $<85\%$ detection using logistic regression, change in odds of analyte detection with one unit increase in exposure category.

doi:10.1371/journal.pone.0143109.g008

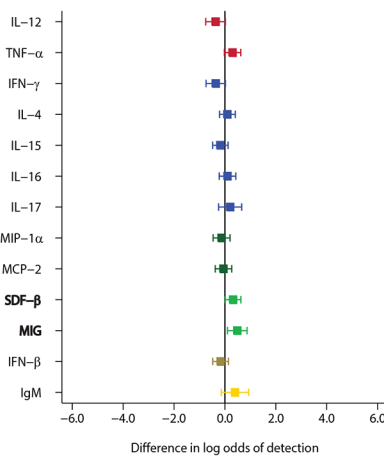
(a) Linear trend for change in log concentration of neutrophils (Linear regression coefficients)



(c) Any lymphocytes vs. none (Linear regression coefficients)



(b) Change in odds of analyte detection with one unit increase in neutrophils (Log odds ratios)



(b) Any lymphocytes vs. none (Log odds ratios)

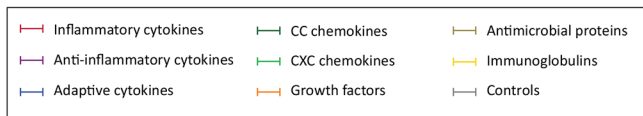
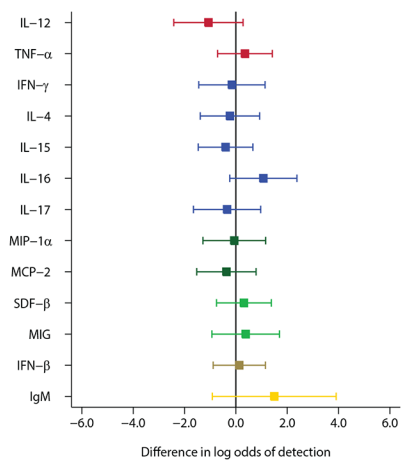


Fig 9. White blood cells (WBCs; neutrophils, n = 361; lymphocytes, n = 361). WBCs from the CVL cell pellet were identified and enumerated. For the statistical analysis, lymphocytes were either present or absent, and neutrophil were categorized as follows: no cells, 1–10 cells, 11–50 cells, >50 cells. Bolded analytes

represent associations with a p-value ≤ 0.05 . (A) For analytes with >85% detection using linear regression, linear trend for change in log concentration in neutrophils. (B) Analytes with <85% detection using logistic regression, change in odds of analyte detection with one unit increase in neutrophil category. (C) A comparison of analyte concentration >85% between samples with presence of lymphocytes and samples with absent lymphocytes (reference) using linear regression. (D) A comparison of analyte concentration >85% between samples with presence of lymphocytes and samples with absent lymphocytes (reference) using logistic regression.

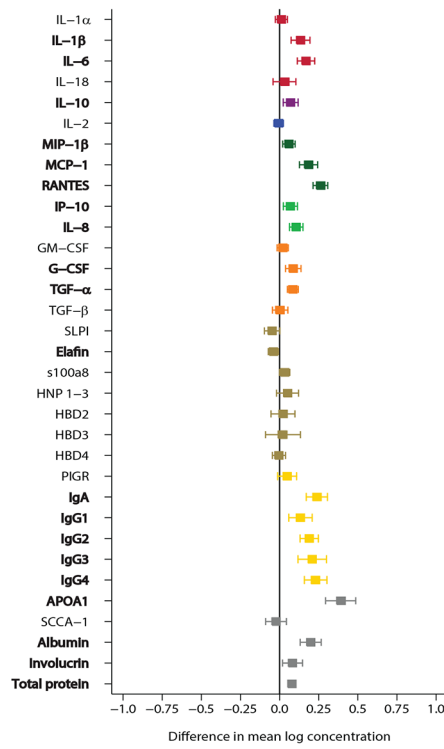
doi:10.1371/journal.pone.0143109.g009

Consistent with our results, Kyongo and colleagues also found a strong correlation between cervical ectopy and IL-6 and G-CSF, but also IL-1 β , IL-8, MIP-1 β among Belgian and sub-Saharan African women [21,22]. Additionally, Hwang and colleagues found that CVL samples among women with ectopy had high levels of IL-1 β , IL-6 and IL-8, but also IL-1 α , MIP-1 α , RANTES, TNF- α , IL-10, IL-12 and IFN- γ [54]. There is evidence from published studies that ectopy is associated with COCs, but not with DMPA [55]. In our study, the signatures for COCs and ectopy are similar, and it may be that most of the differential expression seen among women with ectopy could be explained by increased levels of hormones (i.e. oestrogens).

Vaginal practices are very common in sub-Saharan Africa [56], and have been implicated in HIV acquisition and associated changes in vaginal microbiota [57]. In this population, intravaginal cleansing is highly prevalent, with many women using soap, while fewer women reported using cloth for cleansing or inserting substances [58]. Kyongo and colleagues report an increase in IL-6 and MIP-1 among women reporting using substances other than water for IVP in South Africa [22]. Our study showed no differential expression of analytes for intravaginal cleansing with soap; however, most analytes were *decreased* during visits with reported insertion use, and detergent was the most common substance inserted in our study. In previous *in vitro* studies, Nonoxynol-9 (N9), a surfactant (i.e. a detergent), has been shown to cause IL-1 release and activation of NF- κ B in cervical and vaginal epithelial cells [8]. Our results suggest that soaps and detergents may not have a similar effect on the vaginal milieu as N9, though results should be interpreted with caution. In contrast, cloth use showed increased expression of the innate and adaptive, inflammatory and anti-inflammatory analytes, signalling an overall heightened inflammatory state among women using cloth. The mechanism behind this immune activation may be microabrasions caused by cloth use or introduction of bacteria. These results are consistent with epidemiological findings from a large individual person data analysis that showed an association between cloth use and HIV acquisition (pooled adjusted hazard ratio of 1.47, 95% confidence interval 1.18–1.83) indicating that cloth use may increase susceptibility to HIV acquisition [57]. More research is needed to understand the inflammatory effects of well-defined, prevalent IVP.

This study has a number of important strengths. We measured 45 different analytes, and reported concentrations found in CVLs in realistic trial conditions. We presented unadjusted raw data to show the differential expression of these analytes associated with biomedical and behavioural factors that are likely to be present during a clinical trial (e.g. hormonal contraception use, menstrual cycle stage, seminal plasma exposure). We have presented the data with figures showing unadjusted point estimates with confidence intervals in order to show signatures by factors. However, caution is needed when interpreting any one analyte estimate as this study has multiple comparisons using repeated significance testing which will lead to Type 1 errors or false positives. Therefore, it is more useful to look at the overall signatures, the patterns in types of analytes (e.g. inflammatory cytokines, immunoglobulins), and the strength of associations.

(a) Linear trend for change in log concentrations of Hemastix category (Linear regression coefficients)



(b) Change in odds of analytes detectin with one unit increase in Hemastix (Log odds ratio)

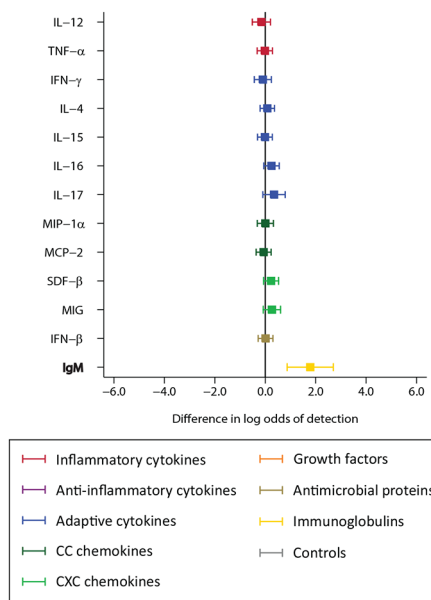


Fig 10. Haemoglobin (n = 369). Haemoglobin was measured by Hemostix test strips in the CVLs, categories were none (25 erythrocytes [ery]/ μ L), moderate (80 ery/ μ L), high (200 ery/ μ L). Bolded analytes represent associations with a p-value ≤ 0.05 . (A) For analytes with >85% detection using linear regression, linear trend

for change in log concentration. (B) Analytes with <85% detection using logistic regression, change in odds of analyte detection with one unit increase in exposure category.

doi:10.1371/journal.pone.0143109.g010

This study also has some limitations. Several biomedical and behavioural factors (e.g. menstrual phase, COCs, DMPA, rarer types of vaginal practices) had lower statistical power to detect associations and wide confidence intervals; therefore, caution is warranted in interpreting these results. However, even with lower power, signatures show marked immune activation for hormonal contraception and vaginal practices with cloth, and relatively less with menstrual cycle, intravaginal insertion, ectopy and colposcopic findings. Secondly, our methods for assigning menstrual phase did not identify the exact visit during ovulation, and samples were not collected during menstruation; therefore differential expression of analytes occurring during ovulation or menses only would have been missed. Lastly, we defined healthy visits by the absence of STIs measured in this study, and there may be unmeasured effects of other STIs (e.g. Human papillomavirus). Additionally, there may be unmeasured effects of persistent immune activation after clearance of HSV shedding [59].

Conclusions

Soluble immune proteins may be useful biomarkers to assess immune activation to predict harm or indicate the risk for HIV-1 acquisition in clinical trials for vaginal microbicides, mucosal vaccines and other interventions. Biomedical and behavioural factors such as hormonal changes, by menstrual cycle stage or hormonal contraception use, recent sex, vaginal practices, and blood contamination may affect concentration of these proteins. Although, randomization should ensure that the timing of visits and presence of these factors are similar between trial arms, investigators should be aware of background variability and collect reliable data on these factors. Additionally, understanding the modulation of total antibody expression over a menstrual cycle should inform the assessment of specific antibody responses following a trial of a candidate HIV vaccine, and provide evidence for the standardization of obtaining samples during the menstrual cycle phase.

Supporting Information

S1 Fig. Figure A, Spearman rank correlations by analyte panel. Figure B, Conceptual model for exploring confounding. Figure C, Adjusted menstrual cycle phase signature. Figure D, Adjusted reported hormonal contraceptive signatures. Figure E, Adjusted prostate-specific antigen signature. Figure F, Adjusted traditional intravaginal practices signatures. Figure G, Adjusted clinical cervical ectopy signature. Figure H, Adjusted colposcopy findings signature. Figure I, Adjusted vaginal pH signature. Figure J, Adjusted white blood cells signature. Figure K, Adjusted haemoglobin signature.

(DOCX)

S1 Table. Table A, The intra-class correlation coefficients and standard deviations for raw data, protein normalized and involucrin normalized analyte data. Table B, Unadjusted coefficients for associations of analytes with biomedical and behavioural factors. Table C, Adjusted coefficients for associations of analytes with biomedical and behavioural factors.

(DOCX)

S1 Text. Normalisation to account for dilution factor in the cervicovaginal lavage.

(DOCX)

Acknowledgments

We thank Joseph Chilongani, the field teams and participants in Geita, Shinyanga and Kahama, without whom this project would not have been possible. We are grateful to National Institute of Medical Research (NIMR) in Mwanza led by John Changelucha. Special thanks to Charles Mmbando and Claire von Mollendorf for on-site training and capacity building in colposcopy. We also thank Tania Crucitti at the Institute of Tropical Medicine in Antwerp, Belgium for external quality control of *T. vaginalis* and PSA.

Author Contributions

Conceived and designed the experiments: SCF RJH RS DWJ JvdW SK GC. Performed the experiments: YH SCF TTA SK KM AA. Analyzed the data: KB SCF. Wrote the paper: JvdW YH CH RS KB RJH SCF.

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