

RESEARCH ARTICLE

# Serum Levels of Soluble CD26/Dipeptidyl Peptidase-IV in Type 2 Diabetes Mellitus and Its Association with Metabolic Syndrome and Therapy with Antidiabetic Agents in Malaysian Subjects

Radwan H. Ahmed<sup>1\*</sup>, Hasniza Zaman Huri<sup>2,3</sup>, Zaid Al-Hamodi<sup>4</sup>, Sameer D. Salem<sup>4</sup>, Sekaran Muniandy<sup>1\*</sup>

**1** Department of Molecular Medicine, Faculty of Medicine, University of Malaya, Kuala Lumpur, Malaysia, **2** Department of Pharmacy, Faculty of Medicine, University of Malaya, Kuala Lumpur, Malaysia, **3** Clinical Investigation Centre, University Malaya Medical Centre, Kuala Lumpur, Malaysia, **4** Department of Biochemistry and Molecular Biology, Faculty of Medicine, Sana'a University, Sana'a, Yemen

\* [radwanalhamody@yahoo.com](mailto:radwanalhamody@yahoo.com) (RHA); [sekaran@um.edu.my](mailto:sekaran@um.edu.my) (SM)



**OPEN ACCESS**

**Citation:** Ahmed RH, Huri HZ, Al-Hamodi Z, Salem SD, Muniandy S (2015) Serum Levels of Soluble CD26/Dipeptidyl Peptidase-IV in Type 2 Diabetes Mellitus and Its Association with Metabolic Syndrome and Therapy with Antidiabetic Agents in Malaysian Subjects. PLoS ONE 10(10): e0140618. doi:10.1371/journal.pone.0140618

**Editor:** Paul Proost, University of Leuven, Rega Institute, BELGIUM

**Received:** July 11, 2015

**Accepted:** September 27, 2015

**Published:** October 16, 2015

**Copyright:** © 2015 Ahmed et al. This is an open access article distributed under the terms of the [Creative Commons Attribution License](https://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

**Data Availability Statement:** All relevant data are within the paper.

**Funding:** This study was supported by research grants HIR/420001-E000046 and RP024A-14HTM and PPP Grant PG035-2014A from the University of Malaya.

**Competing Interests:** The authors have declared that no competing interests exist.

## Abstract

### Background

A soluble form of CD26/dipeptidyl peptidase-IV (sCD26/DPP-IV) induces DPP-IV enzymatic activity that degrades incretin. We investigated fasting serum levels of sCD26/DPP-IV and active glucagon-like peptide-1 (GLP-1) in Malaysian patients with type 2 diabetes mellitus (T2DM) with and without metabolic syndrome (MetS), as well as the associations between sCD26/DPP-IV levels, MetS, and antidiabetic therapy.

### Methods

We assessed sCD26/DPP-IV levels, active GLP-1 levels, body mass index (BMI), glucose, insulin, A1c, glucose homeostasis indices, and lipid profiles in 549 Malaysian subjects (including 257 T2DM patients with MetS, 57 T2DM patients without MetS, 71 non-diabetics with MetS, and 164 control subjects without diabetes or metabolic syndrome).

### Results

Fasting serum levels of sCD26/DPP-IV were significantly higher in T2DM patients with and without MetS than in normal subjects. Likewise, sCD26/DPP-IV levels were significantly higher in patients with T2DM and MetS than in non-diabetic patients with MetS. However, active GLP-1 levels were significantly lower in T2DM patients both with and without MetS than in normal subjects. In T2DM subjects, sCD26/DPP-IV levels were associated with significantly higher A1c levels, but were significantly lower in patients using monotherapy with metformin. In addition, no significant differences in sCD26/DPP-IV levels were found between diabetic subjects with and without MetS. Furthermore, sCD26/DPP-IV levels were

negatively correlated with active GLP-1 levels in T2DM patients both with and without MetS. In normal subjects, sCD26/DPP-IV levels were associated with increased BMI, cholesterol, and LDL-cholesterol (LDL-c) levels.

## Conclusion

Serum sCD26/DPP-IV levels increased in T2DM subjects with and without MetS. Active GLP-1 levels decreased in T2DM patients both with and without MetS. In addition, sCD26/DPP-IV levels were associated with A1c levels and negatively correlated with active GLP-1 levels. Moreover, metformin monotherapy was associated with reduced sCD26/DPP-IV levels. In normal subjects, sCD26/DPP-IV levels were associated with increased BMI, cholesterol, and LDL-c.

## Introduction

Diabetes mellitus is a heterogeneous group of disorders [1] that affected 387 million people worldwide in 2014 and this number is expected to rise to 592 million, or 10% of adults, by 2035 [2]. In 2014, the prevalence of diabetes mellitus among Malaysia's adult population was reported to be 16.6% [2].

Dipeptidyl peptidase-IV (DPP-IV) is a serine protease found on the apical surface of different cells that cleaves X-proline dipeptides from the N-terminus of many polypeptides, including chemokines, peptide hormones, and neuropeptides [3]. In addition to its membrane form, DPP-IV is also found in a soluble form known as cell surface antigen CD26 (sCD26/DPP-IV). This soluble form is found in a variety of biological fluids, and it originates when the transmembrane protein is shed [4]. Some studies have suggested that the soluble form of DPP-IV originates from adipocytes and immune cells [3, 5].

Glucose-dependent insulintropic polypeptides (GIP) and glucagon-like peptides (GLP-1) are major incretin hormones that are rapidly inactivated by sCD26/DPP-IV. Hence, sCD26/DPP-IV has been studied intensively for the management of T2DM [6]. The soluble form of CD26 (sCD26/DPP-IV) causes DPP-IV enzymatic activity in the extracellular domain [7, 8].

Dysfunction of pancreatic  $\beta$ -cells, insulin resistance, and chronic low-grade inflammation are the main abnormalities associated with T2DM [9]. Adipose tissue macrophages are involved in the development of insulin resistance and chronic inflammation [10]. Previous studies have demonstrated that in the visceral adipose tissue of diet-induced diabetic mice, there was an increase in the numbers of CD8+ T-cells and CD11c+ M1 macrophages, and their infiltration into visceral adipose tissue was prevented by sCD26/DPP-IV inhibition, indicating that sCD26/DPP-IV might be implicated in the inflammation of adipose tissue [11]. In addition, it was found that sCD26/DPP-IV plays a role in regulating glycemia [12, 13]. Recently, it was reported that serum DPP-IV activity in type 1 diabetics (T1DM) was associated with insulin resistance [14]. In addition, an increase of DPP-IV activity in normal subjects is a predictor for MetS and insulin resistance and could be considered a novel biomarker for insulin resistance and MetS [15].

Nearly 90–95% of serum DPP-IV activity is related to sCD26/DPP-IV levels [8], a finding that is supported by several epidemiological studies [16–18]. However, Cordero et al. [4] reported that DPP-IV activity and sCD26/DPP-IV concentration are not always correlated.

A number of clinical studies have been conducted to demonstrate whether DPP-IV activity is associated with either the severity or onset of diabetes or with obesity. These studies showed

controversial results, with DPP-IV activity either decreasing [19–21] or increasing [22, 23] in T2DM patients.

Despite the body of research in this area, little is known about the associations between fasting serum sCD26/DPP-IV levels, metabolic disorders, and T2DM. In addition, the associations between serum sCD26/DPP-IV levels in T2DM patients either with or without MetS are still unclear.

The aim of the current study was to evaluate fasting serum sCD26/DPP-IV levels and active GLP-1 levels in Malaysian T2DM patients with and without MetS, as well as to investigate the association of sCD26/DPP-IV level with MetS parameters and antidiabetic agents. In addition, we examined the correlation between serum sCD26/DPP-IV levels and active GLP-1 levels. Finally, this study also assessed the association between sCD26/DPP-IV levels and MetS parameters in normal subjects.

## Materials and Methods

### Subjects

This study involved diabetic and non-diabetic patients, both with and without MetS, who were diagnosed and treated at the Medical Centre, University of Malaya (UMMC). Normal subjects without either MetS or diabetes were enrolled as a control group and underwent a routine medical check-up. The study followed the principles set out in the Declaration of Helsinki and was conducted with approval by the Medical Ethics Committee of the University Malaya Medical Centre (approval number 387.15). Data were collected after obtaining written consent from each participant. Patients who had a malignancy, chronic or acute diseases of the liver, heart, or kidney, or who had received DPP-IV inhibitors were excluded from this study. The therapy regimens for diabetic patients using antidiabetic agents were obtained from patient records at UMMC.

### Anthropometric measurements

Blood pressure (BP) was measured in the morning 10 minutes after resting in a quiet room using an Omron IntelliSense Automatic Blood Pressure monitor. The average of three BP readings was recorded. The weight and height of each subject were measured in the morning after fasting for 12 hours. BMI was calculated as weight (kg) divided by height ( $m^2$ ), and waist circumference at the midpoint between the lowest rib and the frontal superior iliac spine was also measured. A 10 mL sample of fasting venous blood was taken from each participant.

### Measurements of serum sCD26/DPP-IV and active GLP-1 levels

Serum cholesterol (TC), triglyceride, HDL-c, and plasma glucose (FPG) levels were measured using an automated analyzer (Dimension® RxL Max®, Integrated Chemistry System). Fasting plasma insulin (FPI) levels were measured using the ADVIA Centaur XP Immunoassay System (Siemens Healthcare Diagnostics Inc., Deerfield, IL, USA), and the variant glycosylated hemoglobin (A1c) reorder pack (Catalogue number 270–0003) (Bio-Rad, USA) was used to measure A1c levels. Insulin resistance (HOMA-IR) and  $\beta$ -cell function (HOMA- $\beta$ ) were calculated using the Homeostasis Model Assessment (HOMA2) Calculator v2.2 (<http://www.dtu.ox.ac.uk/homacalculator/index.php>), according to procedures outlined by Matthews et al. [24]. Fasting serum levels of sCD26/DPP-IV were measured using the Human sCD26 ELISA Kit (RayBiotech, Inc., USA) according to the manufacturer's instructions. Absorbance in 96-well plates was read at 450 nm with a microplate reader (Hydroflex Elisa, Chemopharm, Austria).

Serum active GLP-1 levels, including GLP-1 (7–36) and GLP-1 (7–37), were determined by ELISA (EMD, Millipore Inc., USA), which measures active GLP-1 without cross-reacting with other inactive forms of GLP-1. The analysis was performed manually according to the manufacturer's instructions.

## Statistical analysis

Biostatistical analyses were performed using SPSS Package Version 11.5 (LEAD Technologies, Inc., USA). *P*-values < 0.05 were considered to be significant. Log<sub>10</sub> transformation was performed for the standard biochemical and demographic parameters, including serum levels of sCD26/DPP-IV and active GLP-1, because they were not normally distributed. After back transformation, means were expressed as a geometric means and standard deviation (SD). The association between fasting serum levels of sCD26/DPP-IV and active GLP-1 in T2DM and non-T2DM patients, both with and without MetS, were studied after correcting for age, race, and gender as covariates by general linear model. Correlations between serum levels of sCD26/DPP-IV and active GLP-1 were assessed by the Spearman partial correlation coefficient ( $r_s$ ).

A multiple linear regression analysis was performed in diabetic patients to investigate the associations between serum levels of sCD26/DPP-IV (as the dependent variable) and metabolic parameters, A1c levels, and metformin therapy. Hierarchical linear regression was applied to evaluate the associations between serum levels of sCD26/DPP-IV and diabetic and metabolic parameters (as the dependent variable) in normal subjects.

## Results

### Fasting serum levels of sCD26/DPP-IV and active GLP-1 in the study groups

A total of 549 subjects were enrolled in this study. Patients were divided into study groups according to diagnoses by attending physicians and endocrinologists, who performed standard biochemical tests and applied the IDF criteria model for MetS diagnosis [25]. There were 164 control subjects without either MetS or diabetes mellitus and 71 non-diabetic participants with MetS who were under treatment for hypertension and/or hyperlipidemia. The study also included 314 subjects already diagnosed with T2DM; of these, 57 did not have MetS, and 257 had MetS. The basic demographics and biochemical parameters of the normal subjects, the non-diabetic patients with MetS, the diabetic patients with MetS, and the diabetics without MetS are summarized in (Table 1)

When fasting serum levels of sCD26/DPP-IV and active GLP-1 were evaluated by general linear model, serum levels of sCD26/DPP-IV were significantly higher in T2DM subjects with MetS [1199 (245) ng/mL] than in normal subjects [1089 (281) ng/mL] ( $p = 1.2 \times 10^{-4}$ ). Likewise, serum sCD26/DPP-IV levels were significantly higher in T2DM subjects without MetS [1195 (204) ng/mL] than in normal subjects [1089 (281) ng/mL] ( $p = 0.015$ ). On the other hand, serum levels of sCD26/DPP-IV were significantly higher in T2DM subjects with MetS [1199 (245) ng/mL] than in non-diabetic MetS patients [1120 (275) ng/mL] ( $p = 0.041$ ). Additionally, no significant difference in serum sCD26/DPP-IV levels was noted between T2DM patients with or without MetS, or between control and non-diabetic MetS subjects (Table 2).

In contrast with the noted increase in sCD26/DPP-IV levels, the fasting serum levels of active GLP-1 were significantly lower in T2DM subjects with MetS [3.74 (2.29) pmol/L] than in normal subjects [4.26 (3.89) pmol/L] ( $p = 0.020$ ). Likewise, serum active GLP-1 levels were significantly lower in T2DM subjects without MetS [3.53 (2.24) pmol/L] than in normal subjects [4.26 (3.89) pmol/L] ( $p = 0.028$ ) (Table 2).

**Table 1. Demographic and standard biochemical parameters among normal control, non-diabetes metabolic syndrome, type 2 diabetes mellitus subjects with and without metabolic syndrome.**

Parameters	Non-diabetes n = 235			Type 2 diabetes n = 314			
	Normal Control n = 164 (29.9%)	MetS n = 71 (12.9%)	P-Value	Without MetS n = 57 (10.4%)	P-Value	With MetS n = 257 (46.8%)	P-Value
Gender % (Male/Female)	39.0/61.0	59.2/40.8		56.1/43.9		44.4/55.6	
Race (Malay %)	52.4	43.6		36.8		51.4	
Race (Chinese %)	31.7	28.2		26.3		14.8	
Race (Indian %)	15.9	28.2		36.9		33.8	
Age (years)	48.3(12.30)	50.9(9.12)	<sup>a</sup> 0.443	50.0(8.91)	<sup>a, b</sup> 0.999	50.7(7.59)	<sup>a</sup> 0.123, <sup>b, c</sup> 0.999
Weight (kg)	60.5(13.80)	74.0 (13.80)	<sup>a</sup> <b>1.3×10<sup>-10</sup></b>	62.6(13.49)	<sup>a</sup> 0.999, <sup>b</sup> <b>4.3×10<sup>-5</sup></b>	72.1(15.14)	<sup>a</sup> <b>1.3×10<sup>-15</sup></b> , <sup>b</sup> 0.999, <sup>c</sup> <b>2.5×10<sup>-5</sup></b>
Height (m)	1.61(0.09)	1.64(0.09)	<sup>a</sup> 0.426	1.60(0.10)	<sup>a</sup> 0.999, <sup>b</sup> 0.273	1.58(0.10)	<sup>a</sup> <b>0.005</b> , <sup>b</sup> <b>7.6×10<sup>-5</sup></b> , <sup>c</sup> 0.660
Body Mass Index (kg/m <sup>2</sup> )	23.3(4.27)	27.6(4.68)	<sup>a</sup> <b>3.2×10<sup>-10</sup></b>	24.3(4.68)	<sup>a</sup> 0.765, <sup>b</sup> <b>3.8×10<sup>-4</sup></b>	28.9(5.25)	<sup>a</sup> <b>3.8×10<sup>-29</sup></b> , <sup>b</sup> 0.344, <sup>c</sup> <b>4.5×10<sup>-10</sup></b>
Waist Circumference (cm)	81.5(12.3)	94.9 (10.72)	<sup>a</sup> <b>1.01×10<sup>-14</sup></b>	86.0(12.30)	<sup>a</sup> 0.047, <sup>b</sup> <b>1.5×10<sup>-4</sup></b>	96.6(11.48)	<sup>a</sup> <b>2.3×10<sup>-33</sup></b> , <sup>b</sup> 0.999, <sup>c</sup> <b>1.1×10<sup>-8</sup></b>
Systolic Blood Pressure (mmHg)	130(19.50)	143(15.14)	<sup>a</sup> <b>1.3×10<sup>-5</sup></b>	120(16.60)	<sup>a</sup> <b>0.001</b> , <sup>b</sup> <b>4.02×10<sup>-11</sup></b>	138(18.62)	<sup>a</sup> <b>1.7×10<sup>-4</sup></b> , <sup>b</sup> 0.315, <sup>c</sup> <b>2.1×10<sup>-10</sup></b>
Diastolic Blood Pressure (mmHg)	81(9.77)	86(8.91)	<sup>a</sup> <b>3.5×10<sup>-4</sup></b>	76(8.71)	<sup>a</sup> <b>0.012</b> , <sup>b</sup> <b>3.4×10<sup>-8</sup></b>	83(10.23)	<sup>a</sup> 0.052, <sup>b</sup> 0.118, <sup>c</sup> <b>3.4×10<sup>-6</sup></b>
Fasting Plasma Glucose (mmol/L)	5.01(0.45)	5.55(0.68)	<sup>a</sup> 0.083	8.09(4.27)	<sup>a</sup> <b>1.8×10<sup>-23</sup></b> , <sup>b</sup> <b>8.8×10<sup>-12</sup></b>	7.79(3.55)	<sup>a</sup> <b>1.4×10<sup>-42</sup></b> , <sup>b</sup> <b>3.5×10<sup>-16</sup></b> , <sup>c</sup> 0.999
Glycosylated A1c (%)	5.58(0.44)	6.00(0.50)	<sup>a</sup> 0.171	7.91(2.75)	<sup>a</sup> <b>2.7×10<sup>-23</sup></b> , <sup>b</sup> <b>4.2×10<sup>-12</sup></b>	8.18(2.04)	<sup>a</sup> <b>1.9×10<sup>-48</sup></b> , <sup>b</sup> <b>1.2×10<sup>-22</sup></b> , <sup>c</sup> 0.999
Insulin (pmol/L)	48.8 (38.0)	83.1 (51.29)	<sup>a</sup> <b>1.8×10<sup>-8</sup></b>	60.8(57.86)	<sup>a</sup> 0.133, <sup>b</sup> <b>0.029</b>	114(85.11)	<sup>a</sup> <b>5.4×10<sup>-36</sup></b> , <sup>b</sup> <b>0.001</b> , <sup>c</sup> <b>7.7×10<sup>-11</sup></b>
HOMA-β (%)	98(40.74)	116(47.86)	<sup>a</sup> 0.518	48(47.54)	<sup>a</sup> <b>1.99×10<sup>-10</sup></b> , <sup>b</sup> <b>1.01×10<sup>-11</sup></b>	81(78.72)	<sup>a</sup> <b>0.023</b> , <sup>b</sup> <b>4.7×10<sup>-4</sup></b> , <sup>c</sup> <b>2.4×10<sup>-6</sup></b>
HOMA-IR	1.04(0.79)	1.81(1.10)	<sup>a</sup> <b>1.09×10<sup>-8</sup></b>	1.60(1.58)	<sup>a</sup> <b>9.4×10<sup>-5</sup></b> , <sup>b</sup> 0.999	2.88(2.57)	<sup>a</sup> <b>1.5×10<sup>-46</sup></b> , <sup>b</sup> <b>5.6×10<sup>-7</sup></b> , <sup>c</sup> <b>3.3×10<sup>-9</sup></b>
Total-Cholesterol (mmol/L)	5.16(1.05)	5.04(1.00)	<sup>a</sup> 0.999	4.61(1.15)	<sup>a</sup> <b>0.004</b> , <sup>b</sup> 0.111	4.75(1.15)	<sup>a</sup> <b>0.001</b> , <sup>b</sup> 0.250, <sup>c</sup> 0.999
High-Density Lipoprotein Cholesterol (mmol/L)	1.50(0.36)	1.18(0.26)	<sup>a</sup> <b>1.2×10<sup>-12</sup></b>	1.36(0.26)	<sup>a</sup> <b>0.025</b> , <sup>b</sup> <b>0.003</b>	1.11(0.27)	<sup>a</sup> <b>3.4×10<sup>-34</sup></b> , <sup>b</sup> 0.335, <sup>c</sup> <b>1.5×10<sup>-8</sup></b>
Low-Density Lipoprotein Cholesterol (mmol/L)	4.19(1.00)	3.37(0.89)	<sup>a</sup> 0.300	2.73(1.12)	<sup>a</sup> <b>0.002</b> , <sup>b</sup> 0.777	2.75(0.98)	<sup>a</sup> <b>3.4×10<sup>-07</sup></b> , <sup>b</sup> 0.324, <sup>c</sup> 0.999
Triglycerides (mmol/L)	1.04(0.58)	1.72(0.71)	<sup>a</sup> <b>7.8×10<sup>-12</sup></b>	0.96(0.34)	<sup>a</sup> 0.999, <sup>b</sup> <b>2.5×10<sup>-10</sup></b>	1.75(1.45)	<sup>a</sup> <b>2.1×10<sup>-23</sup></b> , <sup>b</sup> 0.999, <sup>c</sup> <b>1.9×10<sup>-15</sup></b>

The results presented represent geometric means (SD)

<sup>a</sup>vs normal control group

<sup>b</sup>vs non-diabetes MetS group

<sup>c</sup>vs type 2 diabetes mellitus without metabolic syndrome evaluated using ANOVA.

doi:10.1371/journal.pone.0140618.t001

Additionally, sCD26/DPP-IV levels were negatively correlated with active GLP-1 levels in both T2DM patients with MetS ( $r_s = -0.324$ ;  $p < 0.001$ ) and T2DM patients without MetS ( $r_s = -0.299$ ;  $p < 0.001$ ) after adjusting for age, gender, and race (Table 3).

**Table 2. Comparison of fasting serum levels of sCD26/DPP-IV and active GLP-1 between normal, non-diabetic metabolic syndrome, type 2 diabetes mellitus subjects with and without metabolic syndrome and total type 2 diabetes mellitus.**

Parameters	Non-diabetes n = 235			Type 2 diabetes n = 314			Total type 2 diabetes n = 314		
	Normal Control n = 164	MetS n = 71	P-Value	Without MetS n = 57	P-Value	With MetS n = 257	P-Value	Total type 2 diabetes n = 314	
<b>sCD26/DPP-IV (ng/mL)</b>	1089 (281)	1120 (275)	<sup>a</sup> 0.427	1195 (204)	<sup>a</sup> <b>0.015</b> , <sup>b</sup> 0.139	1199 (245)	<sup>a</sup> <b>1.2×10<sup>-4</sup></b> , <sup>b</sup> <b>0.041</b> , <sup>c</sup> 0.933	1198 (239)	<sup>a</sup> <b>5.7×10<sup>-5</sup></b> , <sup>b</sup> <b>0.038</b>
<b>Active GLP-1 (pmol/L)</b>	4.26 (3.89)	3.94 (3.71)	<sup>a</sup> 0.340	3.53 (2.24)	<sup>a</sup> <b>0.028</b> , <sup>b</sup> 0.270	3.74 (2.29)	<sup>a</sup> <b>0.020</b> , <sup>b</sup> 0.487, <sup>c</sup> 0.479	3.71 (2.28)	<sup>a</sup> <b>0.010</b> , <sup>b</sup> 0.397

The results presented represent geometric means (SD), adjusted for age, race and gender.

<sup>a</sup>vs control group

<sup>b</sup>vs non-diabetic metabolic syndrome group

<sup>c</sup>vs type 2 diabetes mellitus without metabolic syndrome which evaluated using univariate (General Linear Model).

Bold values are significant. MetS: metabolic syndrome.

doi:10.1371/journal.pone.0140618.t002

### Associations between sCD26/DPP-IV levels, MetS, and A1c levels among T2DM patients

The multiple linear regression analysis (adjusted for age, gender, race, and duration of diabetes) showed that serum levels of sCD26/DPP-IV in diabetic patients were associated with increased A1c levels (B = 19.96, *p* = 0.009), but not associated with insulin resistance (B = 2.44, *p* = 0.525) (Table 4).

### Associations between sCD26/DPP-IV levels, MetS, and T2DM parameters among controls

The associations between serum levels of sCD26/DPP-IV, MetS, and T2DM parameters in the control group were investigated by hierarchical linear regression after adjusting for age, race, and gender. This analysis showed that serum levels of sCD26/DPP-IV were associated with increased BMI (B = 0.003, *p* = 0.023), cholesterol (B = 0.001, *p* = 0.001), and LDL-c (B = 0.001, *p* = 0.001) (Table 5).

### Associations between sCD26/DPP-IV levels and metformin

Diabetic subjects were divided into 4 subgroups: those receiving monotherapy with metformin (n = 34); those receiving combination therapy without metformin [instead receiving

**Table 3. Correlation between of fasting serum levels of sCD26/DPP-IV and active GLP-1 among normal, non-diabetic metabolic syndrome, type 2 diabetes mellitus subjects with and without metabolic syndrome.**

Group	r <sub>s</sub>	P-value
<b>Active GLP-1 (pmol/L)</b>		
<b>Normal Control n = 164</b>	-0.198	<b>0.019</b>
<b>Non-diabetes MetS n = 71</b>	-0.139	0.303
<b>Type 2 diabetes without MetS n = 57</b>	-0.299	<b>&lt; 0.001</b>
<b>Type 2 diabetes with MetS n = 257</b>	-0.324	<b>&lt; 0.001</b>

The results are presented as r<sub>s</sub> and (P-value) assessed by Spearman partial correlation adjusted for, age, race and gender. Bold values are significant.

doi:10.1371/journal.pone.0140618.t003

**Table 4. Association of fasting serum levels of sCD26/DPP-IV with MetS and A1c levels among type 2 diabetes patients.**

Parameters	B	P-value
Body Mass Index (Kg/m <sup>2</sup> )	4.64	0.100
HOMA-β (%)	-0.068	0.693
Insulin resistance	2.44	0.525
Glycosylated A1c (%)	19.96	0.009
Triglycerides (mmol/L)	0.492	0.964
HDL-cholesterol (mmol/L)	27.60	0.610

The results are presented as unstandardized coefficients; B and (P-value) assessed using multiple linear regression adjusted for, age, race, gender, and duration of diabetes. Bold values are significant. B: coefficient for the relationship between the dependent variable “DPP-IV level” and the independent variable “diabetic and metabolic biomarker”. The positive sign of the coefficient implies a direct relationship, and the negative sign implies an inverse relationship.

doi:10.1371/journal.pone.0140618.t004

sulfonylurea (SU), and thiazolidinedione (TZD) with or without insulin] (n = 28); those receiving combination therapy with metformin (metformin, SU, and TZD with or without insulin) (n = 241); and non-treated diabetes (n = 8).

The general linear model indicated that patients treated with monotherapy (metformin) had significantly lower serum levels of sCD26/DPP-IV [1048 (275) ng/mL] than patients receiving combination therapy without metformin [1355 (166) ng/mL] ( $p = 2.9 \times 10^{-5}$ ). Likewise, patients treated with combination therapy that included metformin had significantly lower serum levels of sCD26/DPP-IV [1205 (229) ng/mL] than patients receiving combination therapy without metformin [1335 (166) ng/mL] ( $p = 0.023$ ) (Table 6).

This association was confirmed by multiple linear regression analysis, and the results remained significant only for patients treated with monotherapy that included metformin (B = -201.6,  $p = 0.041$ ), after adjusting for age, race, gender, BMI, A1c, and duration of diabetes (Table 7).

**Table 5. Association of fasting serum levels of sCD26/DPP-IV with diabetic and metabolic parameters among normal subjects.**

Parameters	B/r <sup>2</sup>	P-value
Body Mass Index (kg/m <sup>2</sup> )	<b>0.003/0.105</b>	<b>0.023</b>
Waist circumference (cm)	0.006/0.284	0.055
Triglyceride (mmol/L)	0.0003/0.116	0.134
Cholesterol (mmol/L)	0.001/0.107	<b>0.001</b>
LDL-cholesterol (mmol/L)	0.001/0.094	<b>0.001</b>
HDL-cholesterol (mmol/L)	-2.461-5/0.333	0.796
Fasting Blood Sugar (mmol/L)	0.0001/0.077	0.420
Glycosylated A1c (%)	0.0001/0.026	0.531
HOMA-β (%)	0.017/0.053	0.125
Insulin resistance	0.0003/0.070	0.073

The results are presented as unstandardized coefficients; B, r<sup>2</sup> and (P-value) assessed using hierarchical linear regression adjusted for, age, race, and gender. Bold values are significant. B: coefficient for the relationship between the dependent variable “metabolic syndrome and T2DM parameters” and the independent variable “DPP-IV level.” The positive sign of the coefficient implies a direct relationship, and the negative sign implies an inverse relationship.

doi:10.1371/journal.pone.0140618.t005

**Table 6. Comparison of fasting serum levels of sCD26/DPP-IV between combination therapy without metformin, combination therapy with metformin, monotherapy with metformin and non- treated among subjects with type 2 diabetes.**

Parameter	* SU +TZD w/wo insulin(n = 28)	Metformin + SU + TZD w/wo insulin (n = 241)	P-Value	Monotherapy with metformin (n = 34)	P-Value	Non-treated (n = 8)	P-Value
sCD26/DPP-IV (ng/mL)	1355 (166.0)	1205 (229.1)	<b><sup>a</sup>0.023</b>	1048 (275.4)	<sup>a</sup> <b>2.9×10<sup>-5</sup></b> , <sup>b</sup> <b>0.001</b>	1123 (302.0)	<sup>a</sup> 0.056, <sup>b</sup> 0.385, <sup>c</sup> 0.432

The results presented represent geometric means (SD), adjusted for age, gender, and race

<sup>a</sup>vs combination therapy without metformin (SU + TZD with or without insulin)

<sup>b</sup>vs combination therapy with metformin (metformin + SU + TZD with or without insulin)

<sup>c</sup>vs monotherapy with Metformin which evaluated by univariate (General Linear Model).

Bold values are significant.

\*Sulfonylurea (SU), Thiazolidinedione (TZD) with or without insulin.

doi:10.1371/journal.pone.0140618.t006

## Discussion

In this study, the observed fasting serum levels of sCD26/DPP-IV in T2DM patients were higher than that in normal subjects; these results are in agreement with findings by Lee et al., who excluded patients treated with metformin and/or thiazolidinedione therapy [17]. These results are in contrast to findings by Meneilly et al. and Korosi et al. [26, 27], who reported decreased sCD26/DPP-IV levels in diabetic patients.

There are several studies that support increased DPP-IV activity in T2DM patients [23, 28]; however, the cause for the increase in DPP-IV activity in diabetic patients remains unclear. Pala et al. [29] indicated that human glomerular endothelial cells that are exposed to high concentrations of glucose promote the biosynthesis of DPP-IV in vitro. In addition, another study by Pala et al. [30] reported that DPP-IV activation was not induced in control subjects, T2DM patients, or patients with impaired glucose tolerance according to oral glucose loading. Similarly, research by Ryskjaer et al. [31] indicated that DPP-IV activity was increased in T2DM patients. However, observed DPP-IV activity was not altered after meal ingestion and subsequent acute changes in plasma glucose. Recently, Aso et al. demonstrated that sCD26/DPP-IV levels in healthy subjects exhibited an acute increase after oral glucose loading, and this abrupt increase may be associated with the presence of nonalcoholic fatty liver and/or insulin resistance [32]. According to these findings, it is believed that DPP-IV biosynthesis is associated with long-term exposure to high levels of glucose.

**Table 7. Association of fasting serum levels of sCD26/DPP-IV with antidiabetes medications groups among type 2 diabetes patients.**

Parameters	B	P-value
*SU+TZD w/wo insulin	24.0	0.760
metformin +SU+ TZD w/wo insulin	-131.6	0.120
Monotherapy with Metformin	-201.6	<b>0.041</b>
Non-treated	-97.4	0.085

The results are presented as unstandardized coefficients; B and (P-value) assessed using multiple linear regression adjusted for, age, race, gender, BMI, HbA1c, and duration of diabetes. Bold values are significant. B: coefficient for the relationship between the dependent variable “DPP-IV level” and the independent variable “diabetic drugs”. The positive sign of the coefficient implies a direct relationship, and the negative sign implies an inverse relationship.

\*Sulfonylurea (SU), Thiazolidinedione (TZD) with or without insulin.

doi:10.1371/journal.pone.0140618.t007



Interestingly, in line with the observed increased sCD26/DPP-IV levels and decreased active GLP-1 levels in T2DM subjects, sCD26/DPP-IV levels showed a negative correlation with active GLP-1 levels in T2DM patients both with and without MetS. In Chinese subjects with T2DM, it was observed that GLP-1 levels were lower than in the normal glucose tolerance subjects [33]. A similar finding in Caucasian subjects indicated that GLP-1 levels decreased significantly in T2DM patients than in the control subjects [34]. Even in patients with T1DM, GLP-1 levels were lower than in healthy subjects [35]. Overall, in subjects with T2DM, the significant reduction in the incretin effect has been attributed primarily to decreased circulating levels of GLP-1, which may be secondary to either increased degradation by DPP-IV or its decreased secretion by the gut [36]. Pala et al. suggested that the decrease in GLP-1 levels in the early stage of the disease is the most dominant because of impairment in its secretion. However, increased sCD26/DPP-IV activity has a major role after the disease has been in place for longer durations [30].

This study also demonstrated that higher serum levels of sCD26/DPP-IV in T2DM patients were associated with increased A1c, which is in agreement with findings by Lee et al. [17]. On the other hand, it has been reported that treatment with DPP-IV inhibitors may improve A1c levels in T2DM patients [37]. In addition, previous studies [31,38] have demonstrated that DPP-IV activity showed significant correlations with serum A1c levels in diabetic patients. However, these results differ from recently published research by Fadini et al. [23].

We found that serum levels of sCD26/DPP-IV in diabetic patients were not associated with insulin resistance (HOMA-IR). Such findings were in contrast to the findings of Lee et al. [17], which could be due to differences in treatment profiles, as these may affect insulin resistance. Associations between sCD26/DPP-IV levels and MetS parameters were assessed in the control group, since non-diabetic subjects with MetS were under treatment. Research by Lamers et al. [5] demonstrated that adipose tissue inflammation and enlargement of adipocytes enhances the release of soluble DPP-IV from fat cells into the circulation. Furthermore, other studies [5, 23] have reported that circulating DPP-IV correlated with different markers for MetS, including plasma TG, BMI, and waist circumference. Recently, Yang et al. demonstrated that increased DPP-IV activity in healthy Chinese could independently predict MetS, insulin resistance [15], and the risk of developing hypertension [39]. Our current study showed that increased serum levels of sCD26/DPP-IV were associated with increased BMI, total cholesterol, and LDL-c. Our research provides evidence that sCD26/DPP-IV may be useful as a biomarker for increased risk of obesity or metabolic syndrome.

Our findings are also consistent with a previous study that suggested that metformin reduced serum sCD26/DPP-IV levels [17]. We found lower sCD26/DPP-IV levels in diabetic patients on monotherapy (e.g., metformin). Numerous studies [23, 40, 41] have demonstrated that sCD26/DPP-IV activity in metformin users was lower. Some studies have suggested that metformin lowered DPP-IV activity by repressing the release of its soluble isoforms from cells [41, 42]. On the other hand, metformin was also postulated to lower plasma DPP-IV activity indirectly through upregulation of GLP-1 receptors' expression in pancreatic  $\beta$ -cells and increasing plasma GLP-1 levels [43].

## Conclusion

Our results demonstrated that fasting serum levels of sCD26/DPP-IV were increased in T2DM patients both with MetS and without MetS. In contrast, active GLP-1 levels were decreased in T2DM patients both with and without MetS. In addition, sCD26/DPP-IV levels were associated with A1c and were negatively correlated with active GLP-1 levels. However, sCD26/DPP-IV levels were decreased in patients on monotherapy that included metformin.

In control subjects, sCD26/DPP-IV levels were found to be associated with increased BMI, cholesterol, and LDL-c cholesterol. Further studies are necessary to explore the reasons for increased sCD26/DPP-IV levels in T2DM patients and to ascertain whether sCD26/DPP-IV level is an early marker for MetS and/or T2DM.

## Acknowledgments

We are grateful to all the participants of this research and all medical and nursing staff at UMMC for their dedication in this study.

## Author Contributions

Conceived and designed the experiments: RHA SM ZA. Performed the experiments: RHA. Analyzed the data: RHA SDS ZA. Contributed reagents/materials/analysis tools: RHA HZH ZA SDS. Wrote the paper: RHA HZH SM.

## References

1. Bell GI, Polonsky KS. Diabetes mellitus and genetically programmed defects in  $\beta$ -cell function. *Nature*. 2001; 414(6865):788–91. PMID: [11742410](#)
2. International Diabetes Federation. *Diabetes Atlas*, 6th edn. International Diabetes Federation, 2013. 2014.
3. Matteucci E, Giampietro O. Dipeptidyl Peptidase-4 (CD26): Knowing the Function before Inhibiting the Enzyme. *Current Medicinal Chemistry*. 2009; 16(23):2943–51. PMID: [19689275](#)
4. Cordero OJ, Salgado FJ, Nogueira M. On the origin of serum CD26 and its altered concentration in cancer patients. *Cancer immunology, immunotherapy*. 2009; 58(11):1723–47. doi: [10.1007/s00262-009-0728-1](#) PMID: [19557413](#)
5. Lamers D, Famulla S, Wronkowitz N, Hartwig S, Lehr S, Ouwens DM, et al. Dipeptidyl peptidase 4 is a novel adipokine potentially linking obesity to the metabolic syndrome. *Diabetes*. 2011; 60(7):1917–25. doi: [10.2337/db10-1707](#) PMID: [21593202](#)
6. Ussher JR, Drucker DJ. Cardiovascular biology of the incretin system. *Endocrine reviews*. 2012; 33(2):187–215. doi: [10.1210/er.2011-1052](#) PMID: [22323472](#)
7. Morimoto C, Schlossman SF. The structure and function of CD26 in the T-cell immune response. *Immunological reviews*. 1998; 161(1):55–70.
8. Durinx C, Lambeir A-M, Bosmans E, Falmagne J-B, Berghmans R, Haemers A, et al. Molecular characterization of dipeptidyl peptidase activity in serum: Soluble CD26/dipeptidyl peptidase IV is responsible for the release of X-Pro dipeptides. *FEBS European Journal of Biochemistry*. 2000; 267(17):5608–13.
9. Matsuzawa Y. The role of fat topology in the risk of disease. *International journal of obesity*. 2008; 32: S83–S92. doi: [10.1038/ijo.2008.243](#) PMID: [19136997](#)
10. Heilbronn LK, Campbell LV. Adipose tissue macrophages, low grade inflammation and insulin resistance in human obesity. *Current pharmaceutical design*. 2008; 14(12):1225–30. PMID: [18473870](#)
11. Shirakawa J, Fujii H, Ohnuma K, Sato K, Ito Y, Kaji M, et al. Diet-Induced Adipose Tissue Inflammation and Liver Steatosis Are Prevented by DPP-4 Inhibition in Diabetic Mice. *Diabetes*. 2011; 60(4):1246–57. doi: [10.2337/db10-1338](#) PMID: [21330637](#)
12. Barnett A. DPP-4 inhibitors and their potential role in the management of type 2 diabetes. *IJCP International Journal of Clinical Practice*. 2006; 60(11):1454–70. PMID: [17073841](#)
13. Scheen AJ. DPP-4 inhibitors in the management of type 2 diabetes: a critical review of head-to-head trials. *Diabetes & metabolism*. 2012; 38(2):89–101. PMID: [26442284](#)
14. Blaslov K, Bulum T, Duvnjak L. Circulating dipeptidyl peptidase-4 activity is associated with insulin resistance in type 1 diabetic patients. *Journal of diabetes and its complications*. 2015; 29(3):390–4. doi: [10.1016/j.jdiacomp.2014.12.019](#) PMID: [25641023](#)
15. Yang F, Zheng T, Gao Y, Baskota A, Chen T, Ran X, et al. Increased plasma DPP4 activity is an independent predictor of the onset of metabolic syndrome in Chinese over 4 years: result from the China National Diabetes and Metabolic Disorders Study. *PLoS one*. 2014; 9(3). PMID: [24687099](#)

16. Kobayashi H, Hosono O, Mimori T, Kawasaki H, Dang NH, Tanaka H, et al. Reduction of serum soluble CD26/dipeptidyl peptidase IV enzyme activity and its correlation with disease activity in systemic lupus erythematosus. *The Journal of rheumatology*. 2002; 29(9):1858–66. PMID: [12233879](#)
17. Lee SA, Kim YR, Yang EJ, Kwon E- J, Kim SH, Kang SH, et al. CD26/DPP4 levels in peripheral blood and T cells in patients with type 2 diabetes mellitus. *The Journal of Clinical Endocrinology & Metabolism*. 2013; 98(6):2553–61.
18. Busso N, Wagtmann N, Herling C, Chobaz-Péclat V, Bischof-Delaloye A, So A, et al. Circulating CD26 is negatively associated with inflammation in human and experimental arthritis. *The American journal of pathology*. 2005; 166(2):433–42. PMID: [15681827](#)
19. McKillop AM, Duffy NA, Lindsay JR, O'Harte FP, Bell PM, Flatt PR. Decreased dipeptidyl peptidase-IV activity and glucagon-like peptide-1 (7–36) amide degradation in type 2 diabetic subjects. *Diabetes research and clinical practice*. 2008; 79(1):79–85. PMID: [17904681](#)
20. Feron D, Begu-Le Corroller A, Piot J-M, Frelicot C, Vialettes B, Fruitier-Arnaudin I. Significant lower VVH7-like immunoreactivity serum level in diabetic patients: evidence for independence from metabolic control and three key enzymes in hemorphin metabolism, cathepsin D, ACE and DPP-IV. *Peptides*. 2009; 30(2):256–61. doi: [10.1016/j.peptides.2008.11.004](#) PMID: [19061927](#)
21. Firneisz G, Varga T, Lengyel G, Fehér J, Ghyczy D, Wichmann B, et al. Serum dipeptidyl peptidase-4 activity in insulin resistant patients with non-alcoholic fatty liver disease: a novel liver disease bio-marker. *PLoS One*. 2010; 5(8):e12226. doi: [10.1371/journal.pone.0012226](#) PMID: [20805868](#)
22. Ryskjaer J. Plasma dipeptidyl peptidase-IV activity in patients with type-2 diabetes mellitus correlates positively with HbA1c levels, but is not acutely affected by food intake. *European Journal of Endocrinology*. 2006; 155(3):485–93. PMID: [16914604](#)
23. Fadini GP, Albiero M, Menegazzo L, de Kreutzenberg SV, Avogaro A. The increased dipeptidyl peptidase-4 activity is not counteracted by optimized glucose control in type 2 diabetes, but is lower in metformin-treated patients. *DOM Diabetes, Obesity and Metabolism*. 2012; 14(6):518–22. doi: [10.1111/j.1463-1326.2011.01550.x](#) PMID: [22171692](#)
24. Matthews DR, Hosker JP, Rudenski AS, Naylor BA, Treacher DF, Turner RC. Homeostasis model assessment: insulin resistance and b-cell function from fasting plasma glucose and insulin concentrations in man. *Diabetologia: Clinical and Experimental Diabetes and Metabolism*. 1985; 28(7):412–9.
25. Alberti K, Eckel RH, Grundy SM, Zimmet PZ, Cleeman J, Donato K, et al. A joint interim statement of the International Diabetes Federation Task Force on Epidemiology and Prevention; National Heart, Lung, and Blood Institute; American Heart Association; World Heart Federation; International Atherosclerosis Society; and International Association for the Study of Obesity. *Circulation*. 2009; 120(16):1640–5. doi: [10.1161/CIRCULATIONAHA.109.192644](#) PMID: [19805654](#)
26. Korosi J, McIntosh CH, Pederson RA, Demuth HU, Habener JF, Gingerich R, et al. Effect of aging and diabetes on the enteroinsular axis. *The journals of gerontology Series A, Biological sciences and medical sciences*. 2001; 56(9):575–9.
27. Meneilly GS, Demuth HU, McIntosh CHS, Pederson RA. Effect of ageing and diabetes on glucose-dependent insulinotropic polypeptide and dipeptidyl peptidase IV responses to oral glucose. *DME Diabetic Medicine*. 2000; 17(5):346–50.
28. Mannucci E, Pala L, Ciani S, Bardini G, Pezzatini A, Sposato I, et al. Hyperglycaemia increases dipeptidyl peptidase IV activity in diabetes mellitus. *Diabetologia*. 2005; 48(6):1168–72. PMID: [15864535](#)
29. Pala L, Mannucci E, Pezzatini A, Ciani S, Sardi J, Raimondi L, et al. Dipeptidyl peptidase-IV expression and activity in human glomerular endothelial cells. *Biochemical and Biophysical Research Communications*. 2003; 310(1):28–31. PMID: [14511643](#)
30. Pala L, Ciani S, Dicembrini I, Bardini G, Cresci B, Pezzatini A, et al. Relationship between GLP-1 levels and dipeptidyl peptidase-4 activity in different glucose tolerance conditions. *Diabetic Medicine*. 2010; 27(6):691–5. doi: [10.1111/j.1464-5491.2010.03010.x](#) PMID: [20546289](#)
31. Ryskjær J, Deacon CF, Carr RD, Krarup T, Madsbad S, Holst J, et al. Plasma dipeptidyl peptidase-IV activity in patients with type-2 diabetes mellitus correlates positively with HbA1c levels, but is not acutely affected by food intake. *European journal of endocrinology*. 2006; 155(3):485–93. PMID: [16914604](#)
32. Aso Y, Terasawa T, Kato K, Jojima T, Suzuki K, Iijima T, et al. The serum level of soluble CD26/dipeptidyl peptidase 4 increases in response to acute hyperglycemia after an oral glucose load in healthy subjects: association with high-molecular weight adiponectin and hepatic enzymes. *Translational Research*. 2013; 162(5):309–16. doi: [10.1016/j.trsl.2013.07.011](#) PMID: [23994650](#)
33. Zhang F, Tang X, Cao H, Lü Q, Li N, Liu Y, et al. Impaired secretion of total glucagon-like peptide-1 in people with impaired fasting glucose combined impaired glucose tolerance. *International journal of medical sciences*. 2012; 9(7):574. PMID: [22991496](#)
34. Legakis IN, Tziaras C, Phenekos C. Decreased glucagon-like peptide 1 fasting levels in type 2 diabetes. *Diabetes Care*. 2003; 26(1):252–. PMID: [12502699](#)

35. Blaslov K, Bulum T, Zibar K, Duvnjak L. Relationship between metabolic syndrome and meal-induced glucagon like peptide-1 response in type 1 diabetic patients. *Journal of diabetes*. 2015; 7(3):340–6. doi: [10.1111/1753-0407.12194](https://doi.org/10.1111/1753-0407.12194) PMID: [25042812](https://pubmed.ncbi.nlm.nih.gov/25042812/)
36. Freeman JS. Role of the incretin pathway in the pathogenesis of type 2 diabetes mellitus. *Cleveland Clinic journal of medicine*. 2009; 76(Suppl 5):S12–S9. doi: [10.3949/ccjm.76.s5.03](https://doi.org/10.3949/ccjm.76.s5.03) PMID: [19952298](https://pubmed.ncbi.nlm.nih.gov/19952298/)
37. Kusunoki M, Sato D, Nakamura T, Oshida Y, Tsutsui H, Natsume Y, et al. The Beneficial Effects of the DPP-4 Inhibitor Alogliptin on Hemoglobin A1c and Serum Lipids in Japanese Patients with Type 2 Diabetes. *Drug research*. 2015. PMID: [26418415](https://pubmed.ncbi.nlm.nih.gov/26418415/)
38. Mannucci E, Pala L, Ciani S, Bardini G, Pezzatini A, Sposato I, et al. Hyperglycaemia increases dipeptidyl peptidase IV activity in diabetes mellitus. *Diabetologia: Clinical and Experimental Diabetes and Metabolism*. 2005; 48(6):1168–72.
39. Zheng T, Chen T, Liu Y, Gao Y, Tian H. Increased plasma DPP4 activity predicts new-onset hypertension in Chinese over a 4-year period: possible associations with inflammation and oxidative stress. *Journal of human hypertension*. 2014. PMID: [25518897](https://pubmed.ncbi.nlm.nih.gov/25518897/)
40. Lindsay JR, Duffy NA, McKillop AM, Ardill J, O'Harte FPM, Flatt PR, et al. Inhibition of dipeptidyl peptidase IV activity by oral metformin in Type 2 diabetes. *DME Diabetic Medicine*. 2005; 22(5):654–7.
41. Green BD, Irwin N, Duffy NA, Gault VA, O'Harte FPM, Flatt PR. Inhibition of dipeptidyl peptidase-IV activity by metformin enhances the antidiabetic effects of glucagon-like peptide-1. *European Journal of Pharmacology*. 2006; 547(1–3):192–9. PMID: [16945366](https://pubmed.ncbi.nlm.nih.gov/16945366/)
42. Lenhard JM, Croom DK, Minnick DT. Reduced serum dipeptidyl peptidase-IV after metformin and pioglitazone treatments. *Biochemical and Biophysical Research Communications*. 2004; 324(1):92–7. PMID: [15464987](https://pubmed.ncbi.nlm.nih.gov/15464987/)
43. Liu Y, Hong T. Combination therapy of dipeptidyl peptidase-4 inhibitors and metformin in type 2 diabetes: rationale and evidence. *DOM Diabetes, Obesity and Metabolism*. 2014; 16(2):111–7. doi: [10.1111/dom.12128](https://doi.org/10.1111/dom.12128) PMID: [23668534](https://pubmed.ncbi.nlm.nih.gov/23668534/)