

RESEARCH ARTICLE

Are *Salmonella*-Induced Gastroenteritis Neglected in Developing Countries? Feedback from Microbiological Investigations in N'Djamena Hospitals, Chad

Djim-adjim Tabo^{1,2,4}, Sophie A. Granier^{3*}, Colette D. Diguimbaye⁴, Muriel Marault³, Anne Brisabois³, Baizina Mama⁴, Yves Millemann^{2,3}

1 Faculté des Sciences Exactes et Appliquées (FSEA), Université de N'Djamena, N'Djamena, Chad, **2** Université Paris-Est, ENVA, Unité MASQ (Microbiologie des Aliments, Sécurité et Qualité), Maisons Alfort, France, **3** Université Paris-Est, ANSES, Laboratoire de Sécurité des Aliments, Mission Antibiorésistance, Maisons-Alfort, France, **4** Laboratoire de Recherches Vétérinaires et Zootechniques de Farcha (LRVZ/ F), N'Djamena, Chad

* Sophie.GRANIER@anses.fr



OPEN ACCESS

Citation: Tabo D-a, Granier SA, Diguimbaye CD, Marault M, Brisabois A, Mama B, et al. (2015) Are *Salmonella*-Induced Gastroenteritis Neglected in Developing Countries? Feedback from Microbiological Investigations in N'Djamena Hospitals, Chad. PLoS ONE 10(8): e0136153. doi:10.1371/journal.pone.0136153

Editor: Patrick Butaye, Ross University School of Veterinary Medicine, SAINT KITTS AND NEVIS

Received: July 30, 2014

Accepted: August 4, 2015

Published: August 27, 2015

Copyright: © 2015 Tabo et al. This is an open access article distributed under the terms of the [Creative Commons Attribution License](https://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Data Availability Statement: All relevant data are within the paper. Additional laboratory records data are available upon request to the corresponding author.

Funding: This study was carried out with a grant from the French government through a program of Franco-Chadian cooperation.

Competing Interests: The authors have declared that no competing interests exist.

Abstract

Salmonella is considered to be one of the main pathogens causing human gastroenteritis worldwide. Looking for *Salmonella* in Africa in patients suffering from gastroenteritis is rather unusual, and the use of antibiotics is not subject to any regulation. This study intends for stressing the possible prominent importance of *Salmonella* in digestive diseases in Africa as well as identifying antimicrobial resistance of *Salmonella* isolates from faeces samples of human origin. All samples were collected from five N'Djamena hospitals, from patients suffering from diarrhoea. The collecting was undertaken over two periods of six months each: from August 2010 to January 2011 and from September 2011 to February 2012. *Salmonella* isolates were obtained by standard cultivation and serotyping methods. A total of 43 *Salmonella* isolates were identified, belonging to 21 different serovars. The most prevalent serovar was *Salmonella* Stanleyville (n = 7), followed by *S. Anatum* (n = 4) and *S. Kottbus* (n = 3). The other serovars were under-represented. The majority of these isolates were susceptible to all antibiotics tested (CLSI Standards), except two *S. Enteritidis* isolates that exhibited resistance to fluoroquinolones. The different serovars and antibiotic resistance profiles that were observed highlight the substantial diversity of *Salmonella* in N'Djamena, Chad. Roughly, one out of ten patients who consulted for gastroenteritis was shedding *Salmonella* spp. and none of them would have been diagnosed outside the context of this research program. This study may encourage local clinicians to explore more often salmonellosis suspicion in their daily practice.

Introduction

Salmonella is an important food-borne pathogen responsible for infectious diseases in animals and humans worldwide. Cattle and poultry have been implicated as major sources of *Salmonella* contaminated food products that are responsible for human salmonellosis [1, 2]. Several serovars also regularly cause disease in animals and humans by transmission from one individual to another. *Salmonella* is difficult to control in food and animal environments, since animals may be asymptomatic fecal shedders. These “carrier” animals likely play an important role in the spread of infection between herds and flocks and consequently serve as sources of food contamination and human infection [3, 4].

In industrialized countries, it is primarily the economic and social burden of these infectious diseases that is important. In the U.S.A, the annual costs of salmonellosis are estimated between 400 million and 3.5 billion U.S. dollars for the whole American economy. These values take into account medical costs and loss of productivity [5, 6]. In Europe, considering that 95% of salmonellosis cases are food-borne infections, annual costs range between 560 million and 2.8 billion euros [3]. In addition to its pathogenicity, *Salmonella* isolates that are resistant to antimicrobials have become a worldwide health issue in recent years [7].

In Africa, non-typhoidal *Salmonella* (NTS) are a major cause of severe invasive disease in adults and children. Infants and young children less than 3 years of age are the most vulnerable, and clinical associations have been reported with malaria, anaemia, malnutrition, and more recently with HIV infection [8]. HIV infection with advanced immunodeficiency is the major risk factor for invasive NTS disease in African adults. The predominant NTS serovars in the African region are *Salmonella* Typhimurium and *Salmonella* Enteritidis which are increasingly resistant to many antimicrobial agents [9, 10]. *Salmonella* Enteritidis is considered to be an important human pathogen worldwide in the past decade. The consumption of eggs or derived products has been associated with a high percentage of human *S. Enteritidis* outbreaks [11, 12]. Keddy et al. [13] reported in their study 652 invasive *Salmonella* Typhimurium from patients in a provincial hospital in South Africa between 2006 and 2007. These patients were aged 15–64 years and 93% of those tested were infected with HIV, responsible for AIDS.

In Chad, *Salmonella* strains and other important bacterial pathogens are not often isolated and identified, and the resistance of these pathogens including *Salmonella* to commonly used antibiotics is rarely assessed or not at all looked for. Furthermore, the use of antibiotics for treatment of human enteric infections is not subject to any regulation. The aims of this present study were therefore, to investigate the presence and antimicrobial resistance of *Salmonella* isolates in humans and their dissemination in N’Djamena region.

Materials and Methods

Sampling

N’Djamena autonomous region has 11 functional hospitals and several private clinics whose geographic locations depend on the importance of population density in urban districts. Five hospitals agreed to participate in our study (Table 1). Permit to collect samples in Hospital 4 was obtained later during the study and that’s the reason why less samples were collected. Our study was carried out during two seasons, from August 2010 to January 2011 and from September 2011 to February 2012, and involved patients, in- or out- patients suffering from diarrhea in these five hospitals (four public and one private) in urban area of N’Djamena. Only one hospital was sampled each week, ten samples being collected on a single appointed visit on the second day of the week. Each sample was collected from laboratories of these health centers in sterile pouches (AES Chemunex, Combourg, France), placed in a cool box with ice packs and

Table 1. Prevalence of *Salmonella* strains isolated in 5 N'Djamena hospitals, Chad.

Hospitals	Number of human faeces samples (n = 420)	
	Examined	Positive (%; [95%CI])
Hosp 1	90	6 (7%, [1.5–11.8])
Hosp 2	90	4 (4%, [0.2–8.7])
Hosp 3	90	1 (1%, [0–3.3])
Hosp 4	60	11 (18%, [8.5–28.1])
Hosp 5	90	17 (19%, [10.8–27.0])
Total	420	39 (9.3%, [6.5–12.1])

doi:10.1371/journal.pone.0136153.t001

immediately transported to the Laboratory of Veterinary and Zootechnical Research (LRVZ/ N'Djamena) for analysis within the same day.

Bacterial isolation and characterization

Stool cultures were performed using food microbiology standard selective and enrichment culture techniques [14]. All *Salmonella*-like colonies were identified to the genus level by their biochemical characteristics using the Microgen ID-GNA gallery for enterobacteria (AES Chemunex, Combourg, France). *Salmonella* strains were serotyped according to the White-Kauffmann-Le Minor Scheme [15]. To reduce experiment time and cost linked to the classical serotyping method, Premi Test *Salmonella* System (PTS) Kit (Check-Points BV, Wageningen, The Netherlands) was also used for typing some *Salmonella* isolates [16]. Antimicrobial susceptibility was tested by disk diffusion according to the CLSI (Clinical and Laboratory Standards Institute) recommendations [17]. The 16 antibiotics tested were ampicillin (10 µg), amoxicillin+clavulanic acid (20, 10 µg), cephalotin (30 µg), cefotaxime (30 µg), ceftazidime (30 µg), sulphonamides (300 µg), cotrimoxazole (1.25/23.75 µg), gentamicin (10 µg), streptomycin (10 µg), kanamycin (30 µg), tetracycline (30 µg), chloramphenicol (30 µg), colistin (50 µg), nalidixic acid (30 µg), ofloxacin (5 µg), and enrofloxacin (5 µg) (Oxoid, Dardilly, France). Zone diameters were read by the automated Osiris scanner (Bio-Rad, Marne la Coquette, France) and interpreted with CLSI guidelines [18]. *Escherichia coli* ATCC 25922 was used as quality control. The set of these methods were carried out as previously described [19] at the French National Reference Laboratory for antimicrobial resistance. *qnrA*, *B*, *S* gene detection was performed by multiplex PCR as previously described [20].

Ethics statement

This study was approved by the Regional Health Delegation of N'Djamena, Public Health Ministry, Chad, and by the Review Board of the N'Djamena University. All adult subjects provided informed consent, and a parent or guardian of any child participant provided informed consent on their behalf. Informed consent was oral, as the majority of the people are illiterate in Chad. Informed consent was collected by the technicians in charge of samples, and recorded in laboratory notebooks. However, one must emphasize all the data were analyzed anonymously.

Results

A total of 420 samples of human faeces collected from 5 N'Djamena hospitals were analysed for the presence of *Salmonella* and of these, 39 (9.3% %) samples were positive for *Salmonella* (Table 1). Out of the total of 39 positive samples, 43 *Salmonella* isolates were identified, belonging to 21 different serovars. The most frequent serovar was *Salmonella* Stanleyville (n = 7),

Table 2. Distribution and antimicrobial susceptibility profiles of *Salmonella* serovars isolated from 5 N'Djamena hospitals, Chad.

Hospitals Isolates per hospital	Serovars	Isolates sources by Gender/age of patients	Number of patients per serovar	Antimicrobial Resistance profiles
Hosp5 19 isolates of 9 serovars	S. Anatum	F/17ys, M/48ys, F/50ys, F/20ys ^a	4	Pan-susceptible
	S. Braenderup	M/20ys, F/5ys,	2	Pan-susceptible
	S. Stanleyville	M/34ys ^b , M/13ys, F/18ys	3	Pan-susceptible
	S. Altona	M/6ys, M/16ys	2	Pan-susceptible
	S. Farcha	M/38ys, F/20ys ^a	2	Pan-susceptible
	S. Hato	M/32ys, M/52ys	2	Pan-susceptible
	S. Havana	F/12ys	1	Pan-susceptible
	S. Urbana	M/59ys, M/22ys	2	Pan-susceptible
	S. Hull	M/34ys ^b	1	Pan-susceptible
Hosp4 12 isolates of 6 serovars	S. Stanleyville	M/32ys, F/66ys, F/32ys, F/15ys	4	Pan-susceptible
	S. Colindale	F/23ys, F/16ys	2	Pan-susceptible
	S. Idikan	F/14ys, M/1ys	2	Pan-susceptible
	S. Enteritidis	M/23ys, M/15ys	2	NAL, OFX, CIP
	S. Teshie	M/30ys ^c	1	Pan-susceptible
	S. Vom	M/30ys ^c	1	Pan-susceptible
Hosp1 6 isolates of 4 serovars	S. Herston	F/2ys, F/50ys	2	Pan-susceptible
	S. Ona	M/24ys, F/22ys	2	Pan-susceptible
	S. Kottbus	F/12ys	1	Pan-susceptible
	S. 8:i:-	F/31ys	1	Pan-susceptible
Hosp2 5 isolates of 3 serovars	S. Kottbus	M/54ys, F/14ys ^d	2	Pan-susceptible
	S. Amsterdam	M/45ys, F/6ys	2	Pan-susceptible
	S. 6, 7:a:-	F/14ys ^d	1	Pan-susceptible
Hosp3 1 isolates of 1 serovar	S. Muenchen	F/2ys	1	Pan-susceptible
Total 43 isolates of 21 serovars	21 serovars	39 patients	43 isolates	2 profiles

ys: years; M: male; F: female;

^{a, b, c, d}: patient followed by the same superscript letter = same patient; Pan-susceptible means susceptible to all antimicrobials tested; NAL: nalidixic acid, OFX: ofloxacin, CIP: ciprofloxacin

doi:10.1371/journal.pone.0136153.t002

followed by *S. Anatum* (n = 4) and *S. Kottbus* (n = 3). The other serovars were under-represented. Descriptions of each *Salmonella* isolate serovar, source and resistance profile are shown in Table 2. Among the 43 *Salmonella* isolates tested for resistance to 16 different antimicrobials, only 2 (less than 5%) *S. Enteritidis* isolates were found resistant to nalidixic acid, ofloxacin and ciprofloxacin (Table 2). The resistance mechanism involved in this low level of resistance to fluoroquinolone was investigated by multiplex PCR. Both isolates were carrying a *qnrB* gene. The 2 fluoroquinolone resistant *S. Enteritidis* were isolated in the same hospital at the same date from 2 different patients and no link has been established between those 2 patients.

Discussion

We identified several serovars of *Salmonella* at the end of our epidemiological investigations in the N'Djamena hospitals. These human *Salmonella* strains were isolated from a single type of

stool samples in the following clinical situations: gastroenteritis, diarrhea and dysentery. In this study, 9.3% of 420 faeces samples collected from 5 hospitals in the capital city and examined by stool cultures were positive for *Salmonella* (Table 1). This prevalence rate observed highlights an alarming clinical picture knowing that all *Salmonella* serovars can, theoretically, cause a systemic infection in humans having a weak immune status [21]. This study provides a first recent description of the human *Salmonella* serovars present in the N'Djamena area.

In most African countries, non-typhoidal *Salmonella* is cited among the three most common pathogenic bacteria, responsible for bloodstream infections in adults and children. The cases of death related to diseases due to invasive non typhoidal *Salmonella* among hospitalized patients range from 4.4% to 27% in children and 22% to 47% in adults [22, 9].

From one African country to another, these prevalence data may vary depending on socio-economic and demographic situations. The study conducted by Hill et al. [23] revealed that non-typhoidal *Salmonella* represented 8.6% of bacteraemia cases, especially linked to young children in Gambian hospitals. In Senegal, relying on the data obtained on patients with HIV by Dakar University Hospital from 1996 to 2005, Seydi et al. [24] reported 62 cases (8.8%) of bacteraemia due to non-typhoidal *Salmonella*. In Malawi, 64% of newborns admitted to a pediatric hospital died after meningitis caused by non-typhoidal *Salmonella* [9]. Kariuki et al. [25] reported through their study, a total of 336 non-typhi *Salmonella* isolated from blood cultures from 1994 to 2005 in children aged 0–13 years admitted to a rural pediatric hospital in Kenya. This must be stressed that these studies focused on one clone, namely Typhimurium ST313. Nevertheless, co-factors like HIV will favour invasive *Salmonella* infection whatever the serovar.

Indeed, far from being exhaustive, these data show the importance of human salmonellosis in different African countries. It must be emphasized that the prevalence rate (9.3%) revealed by this study does not take into account specifically the age group or sex, and the immune status of respondents. It rather highlights a global situation of non-typhoidal *Salmonella* gastroenteritis in patients of all ages and both genders who consulted the five hospitals of N'Djamena involved in our study (Table 2). Before this study in these five N'Djamena hospitals, the medical diagnosis of gastroenteritis and other human food-borne infections was never oriented by clinicians, to a suspicion of possible salmonellosis. Furthermore, to discuss *Salmonella* prevalence in humans, it is necessary to look back to the 1960s and '70s to gather information on *Salmonella* serovars in the N'Djamena area [26, 27]. Results from this study provide therefore, relevant information to clinicians about diagnosis and medical care of human gastroenteritis and other severe infections that can be caused by *Salmonella* serovars: one out of ten patients consulting on gastroenteritis purposes are infected by *Salmonella*.

In the present study, the analysis of stool samples from 5 hospitals in N'Djamena, revealed many isolates of *Salmonella*, which were particularly heterogeneous. All strains were isolated from patients with episodes of digestive pain and diarrhea. We identified a total of 43 *Salmonella* isolates belonging to 21 different serovars. The diversity of serovars observed in humans in this study reflects the singularity of this result, unlike results published by Galanis et al. [3] and EFSA [4] concerning industrialized countries and many African countries where the distribution of *Salmonella* serovars seems quite homogeneous. However, one of the studies conducted in N'Djamena in the 60s and 70s already pointed out such a variety of isolated serovars, leading to the description of 10 previously unidentified serovars [26].

The different serovars identified are characterized by their specific distribution linked to the geographical location of each hospital. Only serovars Kottbus and Stanleyville were found in two different hospitals located in two urban areas of N'Djamena quite far from each other. Most of these serovars were already recovered by Le Minor et al. [26] and Guard et al. [27] in stool samples collected, at least forty years ago, at the main hospital of Fort-Lamy region, renamed meanwhile N'Djamena.

In Africa, despite monitoring systems, non-typhoidal *Salmonella* serovars remain the leading causes of infant bacteraemia and are responsible for high rates of morbidity and mortality, particularly among young children and elderly or immunocompromised people, when treatment with appropriate antibiotics is not available [25, 28, 29]. Based on the work of Vandenberg et al. [10], out of 59% of invasive non-typhi *Salmonella* isolated from blood cultures of children in a rural district hospital of the Democratic Republic of Congo, where the malaria disease is endemic, the most represented serovars were *S. Typhimurium* (61%) and *S. Enteritidis* (22%). The mortality rate associated with these serovars was of 23% and most of the isolates were resistant to multiple antimicrobial (92%).

Out of the total of 43 *Salmonella* serovars isolated from humans in this study, only two *S. Enteritidis* were resistant to nalidixic acid, ofloxacin and ciprofloxacin. Even if no link has been established at the hospital between those 2 patients, Hospital 4 gathers patients from 6 wards of N'Djamena, shopping exclusively on the same single market and offering a unique big high school; we cannot assess that those 2 men have nothing in common. The other *Salmonella* isolates were pan-susceptible to the 16 antimicrobials tested. The emergence of quinolone resistance in the most common *Salmonella* serovar worldwide is a serious public health concern. Resistance to nalidixic acid has been associated with reduced efficacy of fluoroquinolones such as ciprofloxacin. Several studies conducted in many countries worldwide have demonstrated that *Salmonella* Enteritidis encountered in animal and human infections have showed various resistances to antimicrobial molecules, particularly to quinolones, including fluoroquinolones [30]. This finding is of concern because quinolone and fluoroquinolone molecules are first-line drugs for ambulatory treatment of human invasive salmonellosis. All these data highlight the real burden of non-typhoidal *Salmonella* infections in African countries and the associated risk factors. It therefore appears necessary to improve the current knowledge on food and environmental risk factors linked to the mechanisms of *Salmonella* contaminations and infections. The results shown by this study in hospitals of N'Djamena region are very significant and complete this severe epidemiological situation of *Salmonella* infections in the African context. Also, we point out that the comparison with avian *Salmonella* isolated at the same period using some molecular methods to assess the possible contribution of avian *Salmonella* isolates to human salmonellosis is under way.

This study provides data concerning prevalence rate (9.3%) and resistance to antimicrobials of *Salmonella* isolates among human in N'Djamena, Chad. In the course of this study, we revealed the great diversity among the *Salmonella* serovars isolated from human samples in the different health centers investigated and have shown a relatively low proportion of these isolates resistant to antimicrobial agents in the N'Djamena area. Currently, the real burden of human diseases due to *Salmonella* serovars in Chad remains unknown. A comprehensive epidemiological investigation of invasive non-typhoidal *Salmonella* and diarrhea caused by non-typhoidal *Salmonella* is needed. A clearer understanding of the incidence, complications, and case-fatality rates related to non-typhoidal *Salmonella* serovars at the population level would also be important in deciding how to improve health care management in N'Djamena, Chad.

Acknowledgments

This study was carried out with a grant from the French government through a program of Franco-Chadian cooperation. We wish to acknowledge the MASQ-ENVA staff, the CEB unit staff of ANSES, Maisons-Alfort and the LRVZ staff, N'Djamena-Chad for their entire collaboration. We also do not want to forget to express our gratitude to the WHO collaborating center for *Salmonella* at the Pasteur Institute.

Author Contributions

Conceived and designed the experiments: DT SAG CDD AB YM. Performed the experiments: DT SAG MM BM. Analyzed the data: DT SAG YM. Wrote the paper: DT SAG CDD MM AB BM YM.

References

1. Voetsch AC, Van Gilder TJ, Angulo FJ, Farleu MM, Shallow S, Marcus R, et al. (2004) FoodNet estimate of the burden of illness caused by non-typhoidal *Salmonella* infections in the United States. *CID* 38: 127–134.
2. Delmas G, Jourdan-Da Silva N, Pihier N, Weill FX, Vaillant V, De Valk H (2010) Les toxi-infections alimentaires collectives en France entre 2006 et 2008. *Bull Epidemiol Hebd* 31–32: 44–48.
3. Galanis E, Lo Fo Wong DMA, Patrick ME, Binsztein N, Cieslik A, Chalermchaikit T et al. (2006) Web-based surveillance and global *Salmonella* distribution, 2000–2002. *Emerg Infect Dis* 12(3): 381–388. PMID: [16704773](#)
4. EFSA (2011) EU summary report on trends and sources of zoonoses and zoonotic agents and food-borne outbreaks 2009. *EFSA J* 9(3): 2090.
5. Frenzen PD, Riggs TL, Buzby JC, Breuer T, Roberts T, Voetsch D (1999) *Salmonella* cost estimated update using foodnet data. *Food Rev* 22: 10–15.
6. Sarwari A, Magder L, Levine P, McNamara A, Knower S, Armstrong GL et al. G (2001) Serotypes distribution of *Salmonella* isolates from food animals after slaughter differs from that of isolates found in humans. *J Infect Dis* 183: 1295–1299. PMID: [11262216](#)
7. Butaye P, Michael GB, Schwarz S, Baret TJ, Brisabois A, White DG (2006) The clonal spread of multi-drug resistant non-typhoidal *Salmonella* serotypes. *Microb and Infect* 8: 1891–1897.
8. Graham SM, Molyneux EM, Walsh AL, Cheesbrough JS, Molyneux ME, Hart CA (2000) Non-typhoidal *Salmonella* infections of children in tropical Africa. *Pediatr Infect Dis J* 19: 1189–1196. PMID: [11144383](#)
9. Gordon MA, Graham SM, Walsh AL, Wilson L, Phiri A, Molyneux E et al. (2008) Epidemics of invasive *Salmonella enterica* serovars Enteritidis and *Salmonella enterica* serovar Typhimurium infection associated with multidrug resistance among adults and children in Malawi. *Clin Infect Dis* 46: 963–969. doi: [10.1086/529146](#) PMID: [18444810](#)
10. Vandenberg O, Nyarukweba DZ, Ndeba PM, Hendriksen RS, Barzilay EJ, Schirvel C et al. (2010) Microbiologic and clinical features of *Salmonella* species isolated from bacteremic children in eastern Democratic Republic of Congo. *Pediatr Infect Dis J* 29: 504–510. doi: [10.1097/INF.0b013e3181cd615a](#) PMID: [20104200](#)
11. Mølbak K, Gerner-Smidt P, Wegener HC (2002) Increasing Quinolone Resistance in *Salmonella enterica* serotype Enteritidis. *Emerg Infect Dis* 8(5): 514–515. PMID: [11996688](#)
12. Suresh T, Hatha AAM, Sreenivasan D, Sangeetha N, Lashmanaperumalsamy P (2006) Prevalence and antimicrobial resistance of *Salmonella* Enteritidis and other salmonellas in the eggs and egg-storing trays from retail markets of Coimbatore, South India. *Food Microbiol* 23: 294–299 PMID: [16943017](#)
13. Keddy KH, Dwarika S, Crowther P, Perovic O, Wadula J, Hoosen A et al. (2009) Genotypic and demographic characterization of invasive isolates of *Salmonella* Typhimurium in HIV co-infected patients in South Africa. *J Infect Dev Ctries* 3: 585–592. PMID: [19801800](#)
14. Norme AFNOR NF ISO 6579: méthode horizontale pour la recherche de *Salmonella* (décembre 2002).
15. Guibourdenche M, Roggentin P, Mikoleit M, Fields PI, Bockemuhl J, Grimont PA, et al. (2010) Supplement 2003–2007 (No. 47) to the White-Kauffmann-Le Minor scheme. *Res Microbiol* 161: 26–29. doi: [10.1016/j.resmic.2009.10.002](#) PMID: [19840847](#)
16. Wattiau P, Van Hesse M, Schlicker C, Vander Veken H, Imberechts H (2008) Comparison of classical serotyping and Premi-Test assay for routine identification of common *Salmonella enterica* serovars. *J clin Microbiol* 46: 4037–4040. doi: [10.1128/JCM.01405-08](#) PMID: [18842945](#)
17. CLSI (2009) Performance standards for antimicrobial disk susceptibility tests, approved standard-tenth edition. M2-A10, Wayne, PA. USA. Clinical and Laboratory Standard Institute.
18. CLSI (2011) Performance standards for antimicrobial susceptibility testing, twenty-first Informational Supplement. M100-S21. Wayne, PA. USA: Clinical and Laboratory Standard Institute.
19. Tabo D, Diguimbaye CD, Granier SA, Moury F, Brisabois A, Elgroud R, et al. (2013) Prevalence and antimicrobial resistance of non-typhoidal *Salmonella* serotypes isolated from laying hens and broiler chicken farms in N'Djamena, Chad. *Vet Microbiol* 166: 293–298. doi: [10.1016/j.vetmic.2013.05.010](#) PMID: [23810700](#)

20. Cattoir V, Poirel L, Rotimi V, Soussy CJ, Nordmann P. Multiplex PCR for detection of plasmid-mediated quinolone resistance *qnr* genes in ESBL-producing enterobacterial isolates. *J Antimicrob Chemother.* 2007; 60(2):394–7. PMID: [17561500](#)
21. Bäümle A, Tsolis R, Heffron F (2000) Virulence mechanisms of *Salmonella* and their genetic basis. In: Wray C., Wray A. (Eds.), *Salmonella in Domestic Animals*. CABI Publishing, Oxon 57–72.
22. Brent AJ, Oundo JO, Mwangi I, Ochola L, Lowe B, Berkley JA (2006) *Salmonella* bacteremia in Kenyan children. *Pediatr Infect Dis J* 25: 230–6. PMID: [16511385](#)
23. Hill PC, Onyema CO, Ikumapayi UN, Secka O, Ameyaw S, Simmonds N et al. (2007) Bacteraemia in patients admitted to an urban hospital in West Africa. *BMC Infect Dis* 26: 7:2.
24. Seydi M, Soumare M, Sow AI, Diop SA, Sow I, Dieng AB et al. (2008) Non-typhoidal *Salmonella* bacteremia cases in AIDS patients in a Dakar University Hospital (Senegal). *Med Mal Infect* 38: 25–28. PMID: [18093773](#)
25. Kariuki S, Revathi G, Kiiru J, Lowe B, Berkley JA, Hart CA (2006) Decreasing prevalence of antimicrobial resistance in NTS isolated from children with bacteraemia in a rural district hospital, Kenya. *Intern J Antimicrob Agents* 28:166–171.
26. Le Minor L, Chamoiseau G, Barbé E, Charie-Marsaines C, Egron L (1969) Dix nouveaux serotypes de *Salmonella* isolés au Tchad. *Ann Instit Past*, 116(6): 775–780.
27. Guard O, Delpy P, Sirol J (1973) Les infections à *Salmonella* au Tchad. A propos de 152 cas observés à l'Hôpital de Fort Lamy en 1970. *Rev Elev Méd Vét Pays Trop*: 33–57.
28. Morpeth SC, Ramadhani HO, Crump JA (2009) Invasive non-Typhi *Salmonella* disease in Africa. *Clin Infect Dis* 49: 606–611. doi: [10.1086/603553](#) PMID: [19591599](#)
29. Feasey NA, Dougan G, Kigsley RA, Heyderman RS, Gordon MA (2012) Invasive non typhoidal *Salmonella* disease: an emerging and neglected tropical disease in Africa. *Lancet* 372: 2489–2499.
30. Aarestrup FM, Wiuff C, Molbak K, Threlfall EJ (2003) Is it time to change fluoroquinolone breakpoints for *Salmonella* spp.? *Antimicrob Agents Chemother* 47: 827–829. PMID: [12543704](#)