

RESEARCH ARTICLE

Quantum Chemical Study on the Antioxidation Mechanism of Piceatannol and Isorhapontigenin toward Hydroxyl and Hydroperoxyl Radicals

Yang Lu¹, AiHua Wang¹, Peng Shi¹, Hui Zhang^{2*}, ZeSheng Li³

1 College of Material Science and Engineering, Harbin University of Science and Technology, Harbin, 150080, People's Republic of China, **2** College of Chemical and Environmental Engineering, Harbin University of Science and Technology, Harbin, 150080, People's Republic of China, **3** Key Laboratory of Cluster Science of Ministry of Education & School of Chemistry, Beijing Institute of Technology, Beijing, 100081, People's Republic of China

* hust_zhanghui11@hotmail.com



OPEN ACCESS

Citation: Lu Y, Wang A, Shi P, Zhang H, Li Z (2015) Quantum Chemical Study on the Antioxidation Mechanism of Piceatannol and Isorhapontigenin toward Hydroxyl and Hydroperoxyl Radicals. PLoS ONE 10(7): e0133259. doi:10.1371/journal.pone.0133259

Editor: Dennis Salahub, University of Calgary, CANADA

Received: March 11, 2015

Accepted: June 25, 2015

Published: July 15, 2015

Copyright: © 2015 Lu et al. This is an open access article distributed under the terms of the [Creative Commons Attribution License](https://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Data Availability Statement: All relevant data are within the paper and its Supporting Information files.

Funding: This work is supported by the National Basic Research Program of China (2012CB723308), the National Natural Science Foundation of China (51337002 and 50977019), the Doctoral Foundation by the Ministry of Education of China (201112303110005), and the Science Foundation for Distinguished Young Scholar of Heilongjiang Province (JC201206). The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Abstract

A systematic study of the antioxidation mechanisms behind hydroxyl ($\bullet\text{OH}$) and hydroperoxyl ($\bullet\text{OOH}$) radical scavenging activity of piceatannol (PIC) and isorhapontigenin (ISO) was carried out using density functional theory (DFT) method. Two reaction mechanisms, abstraction (ABS) and radical adduct formation (RAF), were discussed. A total of 24 reaction pathways of scavenging $\bullet\text{OH}$ and $\bullet\text{OOH}$ with PIC and ISO were investigated in the gas phase and solution. The thermodynamic and kinetic properties of all pathways were calculated. Based on these results, we evaluated the antioxidant activity of every active site of PIC and ISO and compared the abilities of PIC and ISO to scavenge radicals. According to our results, PIC and ISO may act as effective $\bullet\text{OH}$ and $\bullet\text{OOH}$ scavengers in organism. A4-hydroxyl group is a very important active site for PIC and ISO to scavenge radicals. The introducing of $-\text{OH}$ or $-\text{OCH}_3$ group to the ortho-position of A4-hydroxyl group would increase its antioxidant activity. Meanwhile, the conformational effect was researched, the results suggest that the presence and pattern of intramolecular hydrogen bond (IHB) are considerable in determining the antioxidant activity of PIC and ISO.

Introduction

Piceatannol (4',5',3,5-tetrahydroxystilbene), a natural stilbene, can be found in various plant species, such as grape [1], peanut [2], vaccinium berries [3], euphorbia lagascae [4], etc. PIC is also an analogue of resveratrol (4',3,5-trihydroxystilbene), and can be produced from resveratrol by the action of cytochrome P450 enzyme CYP1B1 in vivo [5]. As we know resveratrol (RES) is a well known health-promoting active component with a wide variety of desirable biological activities, especially its antioxidative activity in biological systems [6,7]. The antioxidant activity of RES and other polyhydroxylated stilbenes is closely related to the location and

Competing Interests: The authors have declared that no competing interests exist.

number of hydroxyl (-OH) group which can scavenge harmful free radicals produced in vivo [8,9]. However, PIC has been pronounced to be a stronger antioxidant than RES owing to its extra-OH group [10]. Hence PIC is better than RES in increasing the lifespan of yeast cells by stimulating the activity of SIRT1 [11]. Now, PIC is getting more extensive attention because of its abirritation to age-related diseases, such as anti-inflammatory, anticarcinogenic, antiviral, antioxidative, neuroprotective and estrogenic properties [12–18].

Isorhapontigenin (3,4',5-trihydroxy-3'-methoxy-stilbene), which is isolated from *Belamcanda chinensis*, is also a derivative of stilbene, and possesses a similar chemical structure with PIC. Red wine and grapes are main sources of PIC in the human diet, and recently ISO was also identified from wine grapes [19]. Generally speaking, the compounds with similar structures would have similar biological activities. Some recent studies have proved that ISO also possesses antioxidative activity with the evidence of inhibiting oxidation of human low-density lipoprotein (LDL) and other pro-oxidant system in vitro [20,21]. Fang et al. reported that ISO could inhibit the respiratory burst of PMA-activated rat neutrophils by scavenging oxygen free radicals [22]. Liu et al. showed that ISO attenuates the proliferation of bovine aortic smooth muscle cells which are induced by oxidized LDL, by blocking the generation of reactive oxygen species (ROS) and activation of the ERKs pathway [23].

Regarding the cause of aging, one of the prevalent theories is the free radical theory. In 1956, Harman proposed the concept that free radicals play an important role in the aging process of biological systems [24]. In light of his theory, the continual generation of free radicals in the process of cellular metabolism would result in the accumulation of oxidative damage to proteins, lipids and DNA, which is the primary causal factor of the aging process. Then in 1969, McCord and Fridovich discovered the superoxide dismutase (SOD) and indicated that it can disintegrate the generation of ROS from the mitochondrial respiratory chain [25]. This provided convincing evidence about the importance of free radicals in living body. Recently, it was discovered that ROS lead to the accumulation of oxidative damage to cellular constituents. This gives rise to a new development of the oxidative stress theory of aging, which holds that the increase in ROS accompanying aging can cause the functional alterations, pathological conditions, and even death [26].

ROS are certain radicals derived from oxygen. As the byproducts of normal metabolism, they represent the most important class of radicals in biological systems. ROS can cause the oxidative decomposition of cellular membranes by lipid peroxidation, and induce the DNA damage [27]. In addition, ROS are responsible for the apoptosis of cells [28]. In order to counteract the excess ROS and then avoid oxidative damage, natural antioxidant as a kind of phytoalexin, can also protect the human body from the damage of free radicals and prevent the advancement of certain chronic diseases related to oxidative stress. PIC was found to have inhibitory effect on the toxic action to neutrophils and oxidative damage to tissues, by scavenging free radicals produced by neutrophils [29].

Besides experimental studies, some theoretical research on the antioxidant activity of PIC have been conducted. Recently, Rossi et al. solved the crystal structure of PIC, and based on the structure they pointed out that the strong antioxidant activity of this polyphenol is due to the formation of the extensive hydrogen bond by its hydroxyl groups which facilitates the transfer of the hydrogen atom. They also compared the ability of RES and PIC to scavenge free radicals, and proved that PIC is a more efficient •OH and •OOH scavenger than RES [30]. Rossi et al. performed a theoretical investigation on the structural features of PIC and the effect of water using DFT method and COSMO solvation model [30]. Mikulski et al. first completed a theoretical research of the electronic structural feature of PIC using MP2 and DFT method [31]. Perez-Gonzalez et al. investigated the free radical scavenging activity of a series of polyphenols. By comparing their activity indexes and bond dissociation energy (BDE), they found that PIC

has a smaller BDE, which suggested that it is a good antioxidant via H transfer [32]. Cordova-Gomez et al. studied the ability of RES and PIC anions to scavenge $\bullet\text{OOH}$ in water and pentyl ethanoate solution using DFT method. They indicated that PIC is a better hydroperoxyl radical scavenger than RES, regardless of the polarity of the environment [33]. Above studies provide us some knowledge on the microscopic mechanisms of the antioxidation reactions of PIC. However the theoretical research of the antioxidant reaction of ISO has not been reported. Therefore, we proceed a quantum chemical study on the activity of ISO scavenging radical for the first time. The main objective of this study is to establish a qualitative and quantitative relationship among the thermodynamic, kinetic and structural properties for PIC and ISO. By comparing the obtained barrier height of each pathway, we found out the effect of different substituent group on the ability of PIC and ISO scavenging radicals. Hope this research can provide some theoretical basis for the subsequent drug design involving the stilbenes antioxidation.

Hydroxyl and hydroperoxyl radical are the main ROS generated in vivo. Particularly to $\bullet\text{OH}$, it is the most active ROS with a very short half-life of approx 10^{-9} s in vivo [34]. In contrast, $\bullet\text{OOH}$ is the relatively slow-reacting species with half-life of the order of seconds [35], and be capable of diffusing to the remote cellular locations [36].

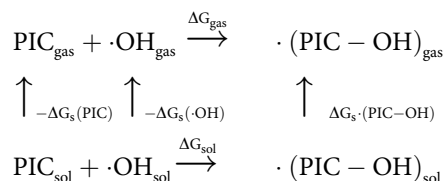
To research the antioxidant activity of PIC and ISO, we conducted a systematic study on the reaction mechanisms behind the $\bullet\text{OH}$ and $\bullet\text{OOH}$ radical scavenging activities of PIC and ISO, using DFT method. Two reaction mechanisms, ABS and RAF, were investigated. In addition, we obtained the thermodynamic and kinetic properties of all pathways in water to investigate the solvation effect on reactions.

Computational Methods

All quantum chemical calculations were performed using the GAUSSIAN 09 computational package [37]. Unrestricted calculations were used for open shell systems. Geometry optimization and frequency calculation of all stationary points (reactants, complexes, transition states and products) were carried out at the M05-2X level with the 6-311++G(d,p) basis set. All of the reactants, complexes and products have only real frequencies; while all of the transition states (TS) present a single imaginary frequency which corresponds to the expected vibrational mode. The M05-2X function is a new DFT method that was specifically developed for kinetics calculation by Truhlar group [38]. So far, this functional yields satisfactory overall performance for the calculations of thermochemistry and thermochemical kinetics in organic, biological systems involving free radical reaction [39–41].

Solvation effect was introduced using SMD Continuum Solvation Model [42] which is recommended by Gaussian Manual to compute solvation energy. Because water is the major component in organism, in this work we use water as the solvent to simulate the cellular environment. The solvation effect was assessed by the single point calculations on the optimized geometries of the gas phase, with SMD model at the M05-2X/6-311++G(d,p) level of theory.

Relative Gibbs energies in solution were computed using thermodynamic cycles and Hess law which explicitly include solvation energies. For example, the thermodynamic cycle for the addition reaction of $\bullet\text{OH}$ with PIC is as follow:



Using this strategy the Gibbs energy of reaction in solution (ΔG_{sol}) can be obtained as the sum of the Gibbs energy of reaction in the gas phase (ΔG_{gas}) and the difference in solvation energies ($\Delta\Delta G_s$):

$$\Delta G_{\text{sol}} = \Delta G_{\text{gas}} + \Delta\Delta G_s \quad (1)$$

where $\Delta\Delta G_s$ is calculated as:

$$\Delta\Delta G_s = \Delta G_s(\cdot\text{PIC} - \text{OH}) - \Delta G_s(\text{PIC}) - \Delta G_s(\cdot\text{OH}) \quad (2)$$

where ΔG_s represents the solvation energy. In all cases, the reference state is 1M. The solvent cage effect has been included with the corrections proposed by Okuno [43], which takes into account the free volume theory. These corrections are in good agreement with those independently obtained by Ardura et al. [44]. In this work the expression used to correct the Gibbs energy is:

$$\Delta G_{\text{sol}}^{\text{FV}} \cong \Delta G_{\text{sol}}^0 - RT\{\ln[n 10^{(2n-2)}] - (n-1)\} \quad (3)$$

where n represents the molecularity of the reaction. According to the Expression 3, the solvent cage effect causes a decrease of 10.63 kJ/mol in ΔG for a bimolecular reaction at 298.15 K [43].

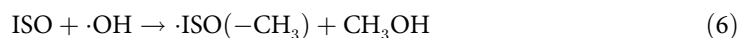
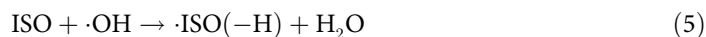
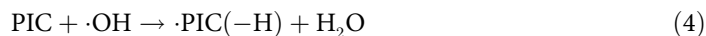
Results and Discussion

It has been proved by experiments that PIC and ISO have remarkable activity of scavenging radicals. To clarify the mechanisms behind their scavenging activities, we researched the reactions of PIC and ISO with two classic ROS ($\cdot\text{OH}$ and $\cdot\text{OOH}$) produced in organism.

The optimized structures of PIC and ISO with atom labels are shown in Fig 1. By our calculation with the method of M05-2X/6-311++G(d,p), the dihedral angle between two benzene rings of PIC is 179.95°, which is similar with the value of 179.50° calculated by MP2/6-311G(d,p) method [31]. The theoretical results are in agreement well with the corresponding experimental value of 179.23° [30]. For each kind of free radical, we considered four abstraction reaction pathways (from the site A4, A5, B3 and B5 in Fig 1) by ABS mechanism as well as two addition reaction pathways (adding to the site α and β) by RAF mechanism. At the same time, the solvent effect was also taken into account.

Hydroxyl Radical ($\cdot\text{OH}$)

$\cdot\text{OH}$ abstracts the H atom from the hydroxyl group of PIC or ISO, followed by forming a water molecule and a corresponding radical; while at the A5 site of ISO, $\cdot\text{OH}$ abstracts CH_3 from the methoxyl group. In general, the -OH group of different site possesses different activity of scavenging radicals. So we defined eight ABS pathways: A4, A5, B3, B5 for PIC and A'4, A'5, B'3, B'5 for ISO. The ABS reactions can be expressed as follow:



The main products after the H atom has been abstracted are semiquinone radicals which are stable as that the unpaired electron can delocalize through both aromatic rings. The optimized structures of all product complexes (PC) are shown in Fig 2.

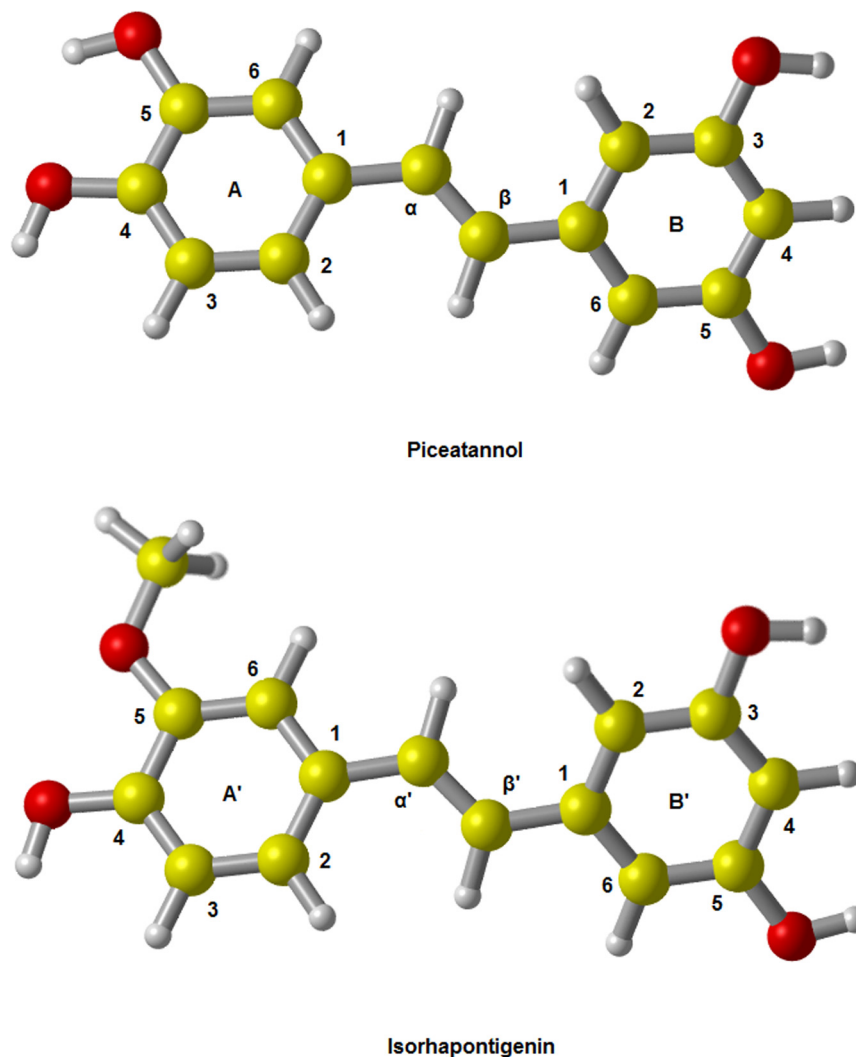


Fig 1. The optimized geometries of PIC and ISO in the gas phase. The results show that both of PIC and ISO have an approximately planar structure. The dihedral angle between two benzene rings of PIC is about 179.95° , which is very consistent with the experimental result of 179.23° ; the corresponding dihedral angle of ISO is -179.90° .

doi:10.1371/journal.pone.0133259.g001

For completely separated reactants and products, the reaction Gibbs energies (ΔG_{gas}) and reaction enthalpies (ΔH_{gas}) of ABS reactions are presented in Tables 1 and 2 respectively. It can be seen, for both of PIC and ISO, all ABS pathways are exothermic and thermodynamically favored. Among them, the most exergonic pathway is from the same position, A4 and A'4. Therefore the semiquinone radicals produced from A4 and A'4 pathway are expected to be the major products of ABS reactions. This is because the resonance effect between two aromatic rings can donate the electron pairs which can be transmitted via the stilbene bridge, and this type of conjugation in electron transfer process has been reported previously [45].

We identified all TS of ABS reactions in the gas phase at the M05-2X/6-311++G(d,p) level, their fully optimized geometries are shown in Fig 3. For the TS structures of A4, A5, B3 and B5, the lengths of O-H bonds that need to break are increased by 4.3%, 7.1%, 6.6% and 6.6% respectively comparing to their equilibrium bond lengths in PIC reactant; the lengths of H-O bonds that are ready to form are stretched by 59.6%, 47.9%, 47.4% and 47.4% respectively

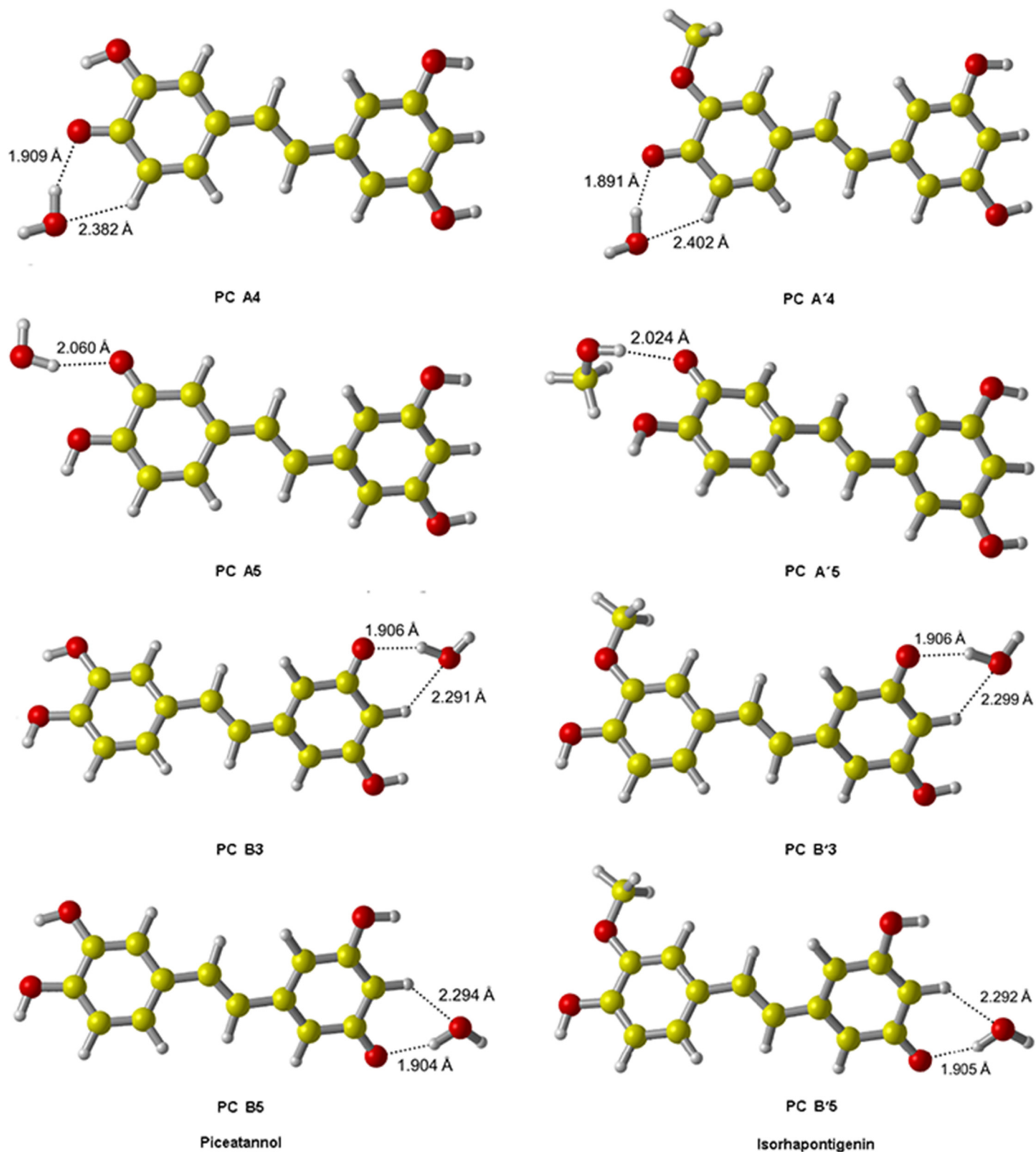


Fig 2. The optimized geometries of the product complexes of PIC and ISO from ABS reactions initiated by $\bullet\text{OH}$. In these structures, the water molecule forms a hydrogen bond with the PIC/ISO semiquinone radical.

doi:10.1371/journal.pone.0133259.g002

Table 1. The reaction Gibbs energies for reactions of PIC and ISO with •OH in the gas phase and water (in kJ/mol).

PIC+•OH			ISO+•OH		
site	ΔG_{gas}	ΔG_{sol}	site	ΔG_{gas}	ΔG_{sol}
A4	-181.55	-196.19	A'4	-162.75	-190.32
A5	-121.18	-164.78	A'5	-133.75	-161.70
B3	-40.23	-42.61	B'3	-39.94	-42.32
B5	-38.17	-40.01	B'5	-40.86	-43.29
α	-105.84	-104.00	α'	-108.12	-108.04
β	-105.03	-108.12	β'	-108.93	-111.07

doi:10.1371/journal.pone.0133259.t001

comparing to their equilibrium bond lengths in isolated H₂O product. Thus, all the breaking bonds are much less elongated than all the forming bonds, indicating that all TS of PIC in ABS pathways are reactant-like. Similarly, all TS from ABS reactions of ISO are reactant-like. These are consistent with the Hammond's postulate [46] for exothermic reaction.

The barrier heights including zero-point energy corrections for ABS pathways are reported in Table 3. It can be found that most pathways in Table 3 have negative barrier heights, these reactions are barrierless reactions. The reactant complexes (IMA4, IMA'4, IMB3, IMB'3, IMB5, IMB'5, IM α , IM α' , IM β and IM β') are formed in the entrance of pathway A4, A'4, B3, B'3, B5, B'5, α , α' , β and β' . The energies of IM are lower than that of reactants, the relative energies (the reactant energies are set to zero) were plotted in Figs 4 and 5.

We can also find, in the gas phase, the pathway A4 has the lowest barrier height in all ABS reactions of PIC with •OH. Combining with the thermodynamic analysis, we conclude that the pathway A4 is more thermodynamically and kinetically favorable than other ABS pathways, and it is the major •OH scavenging pathway of PIC by ABS mechanism. This finding could be further confirmed by our calculation of BDE. The homolytic BDE is an important factor in determination of the effectiveness of an antioxidant: the weaker this bond, the higher its antioxidant activity. The values of O-H BDE for sites of PIC are 310 kJ/mol (A4), 363 kJ/mol (A5), 444 kJ/mol (B3) and 445 kJ/mol (B5). The O-H BDE of A4 site is the smallest, suggesting that its antioxidant activity is the strongest. This is coincident with the calculation result of the previous research [32]. For ISO, the pathway A'4 has the lowest barrier height in all ABS reactions in the gas phase. So the pathway A'4 is the leading channel for ISO to scavenge •OH by ABS mechanism.

In addition, addition reactions may occur as follows:

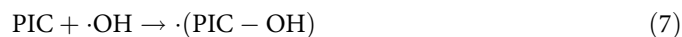


Table 2. The reaction enthalpies at 298K for reactions of PIC and ISO with •OH in the gas phase and water (in kJ/mol).

PIC+•OH			ISO+•OH		
site	ΔH_{gas}	ΔH_{sol}	site	ΔH_{gas}	ΔH_{sol}
A4	-171.31	-176.54	A'4	-156.76	-174.16
A5	-118.10	-150.86	A'5	-122.39	-139.56
B3	-36.63	-129.55	B'3	-36.43	-132.32
B5	-36.47	-130.70	B'5	-36.92	-131.56
α	-150.60	-141.15	α'	-151.34	-146.56
β	-150.45	-147.02	β'	-151.96	-147.29

doi:10.1371/journal.pone.0133259.t002

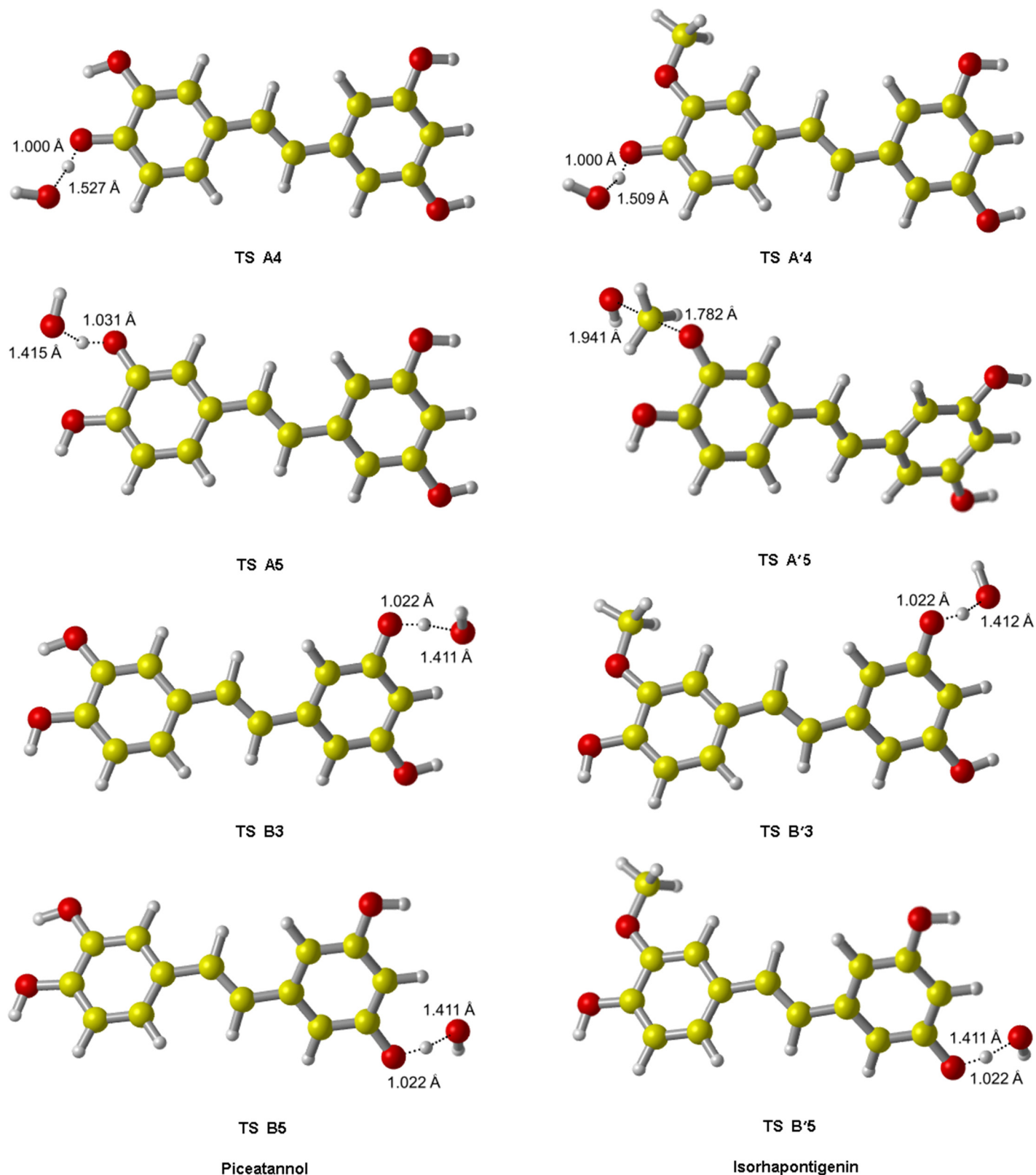


Fig 3. The transition state geometries of PIC and ISO from ABS reactions initiated by •OH. The elongations of the breaking bonds are smaller than those of the forming bonds, indicating these TS are all reactant-like, i.e. these reactions are all exothermic in light of the Hammond's postulate. These agree with our calculated results in [Table 2](#).

doi:10.1371/journal.pone.0133259.g003

Table 3. The barrier heights of TS with zero-point energy corrections for reactions of PIC and ISO with •OH in the gas phase and water (in kJ/mol).

PIC+•OH			ISO+•OH		
site	ΔE_{gas}	ΔE_{sol}	site	ΔE_{gas}	ΔE_{sol}
A4	-15.54	-58.86	A'4	-12.50	-56.45
A5	5.08	-30.58	A'5	132.02	98.28
B3	-5.33	-16.52	B'3	-7.68	-25.83
B5	-5.72	-16.44	B'5	-6.59	-25.25
α	-13.94	-15.64	α'	-15.83	-13.38
β	-17.94	-16.30	β'	-16.86	-13.52

doi:10.1371/journal.pone.0133259.t003

where •OH is added to either carbon atom of >C = C< moiety (at site α/α' and β/β' , corresponding to the pathway α and β for PIC, α' and β' for ISO), and an OH-adduct radical is produced. The reaction Gibbs energies (ΔG_{gas}) and reaction enthalpies (ΔH_{gas}) of RAF pathways were also calculated and presented in Tables 1 and 2 respectively. The results indicate that RAF pathways of PIC and ISO are all exothermic and thermodynamically favored.

The optimized geometries of TS and PC for addition reactions are shown in Fig 6, and the barrier heights are listed in Table 3. By comparing the value of ΔE_{gas} , we estimated that the site β/β' is always more active than α/α' for PIC and ISO.

Considering the application of antioxidant scavenger in vivo environment, the reaction Gibbs energies (ΔG_{sol}) and reaction enthalpies (ΔH_{sol}) of all pathways in water were calculated using SMD model. The results are included in Tables 1 and 2 respectively. As same as in the gas phase, all pathways of PIC and ISO with •OH are exothermic and thermodynamically

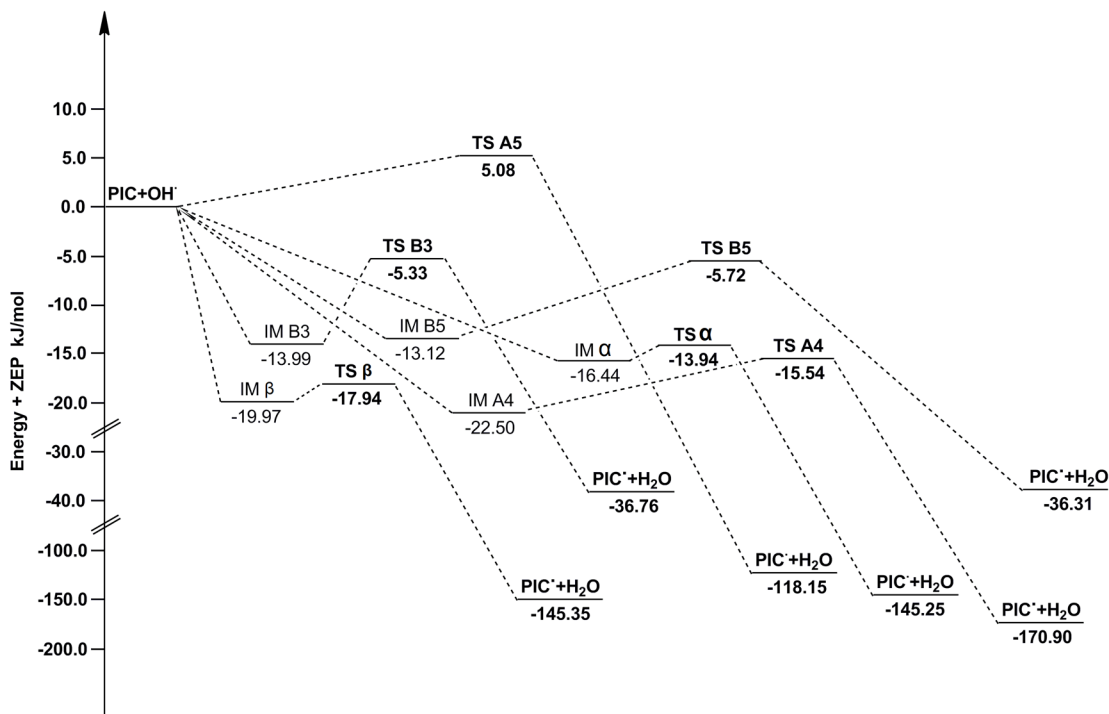


Fig 4. The potential energy surfaces of the reactions of PIC with •OH in the gas phase. The relative energies (in kJ/mol) were calculated at the M05-2X/6-311++G(d,p) + ZPE level. To facilitate comparison, the energy of the reactants are set to zero.

doi:10.1371/journal.pone.0133259.g004

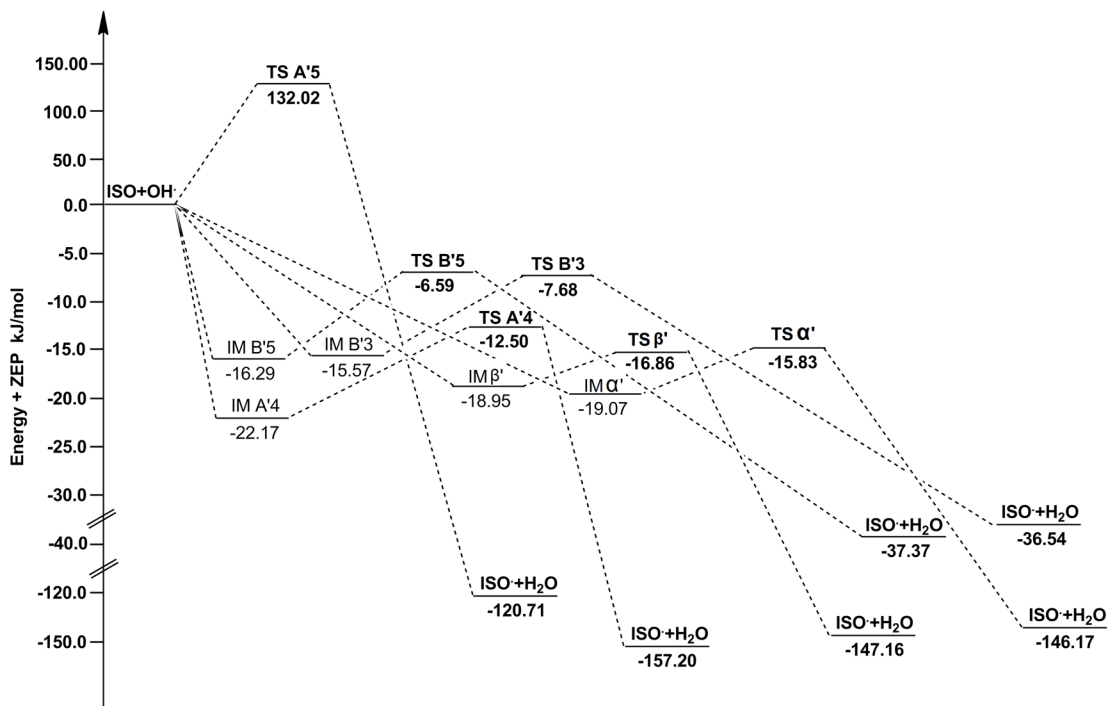


Fig 5. The potential energy surfaces of the reactions of ISO with •OH in the gas phase. The relative energies (in kJ/mol) were calculated at the M05-2X/6-311++G(d,p) + ZPE level. To facilitate comparison, the energy of the reactants are set to zero.

doi:10.1371/journal.pone.0133259.g005

feasible in the water phase. In addition, we found that for ABS pathway, ΔG_{sol} is always smaller than ΔG_{gas} , which suggests that the ABS reactions in water are more exergonic and easier than those in the gas phase. In water, the most exergonic pathways are still A4 and A'4 corresponding to the reaction systems of PIC and ISO. Thus, the thermodynamic properties of ABS reactions in water is same as in the gas phase.

The barrier heights for reactions of PIC and ISO with •OH in water were obtained and presented in Table 3. As shown in this table, the barrier heights of ABS reactions in water are significantly decreased in comparison with reactions in the gas phase. It proves that water solvent promotes the scavenging activity of PIC and ISO toward •OH. In water, the pathway A4 and A'4 respectively have the lowest barrier heights of ABS reactions just like in the gas phase. Therefore we inferred that, in the gas phase and water, A4/A'4-OH group is always the most active site of PIC/ISO by ABS mechanism, while β/β' site is always more active for RAF mechanisms.

In conclusion, A4-OH group is the dominant active site for the antioxidant reaction of PIC and ISO toward •OH. The common feature of A4-OH groups in PIC and ISO is that there is a substituent group on their ortho-position, where is a hydroxyl group in PIC and a methoxyl group in ISO. So the existence of the adjacent substituent may have a promoting effect on the antioxidant activity of A4-OH group. In addition, by comparing the best active site of PIC with that of ISO, we found that the barrier height of A4 is smaller than that of A'4 wherever in the gas phase or water, hence the activity of A4-OH group of PIC is stronger than that of ISO. This implies that, for the activity of -OH group scavenging •OH, the contribution of the adjacent-OH group is bigger than that of the adjacent-OCH₃ group. It is probably because two adjacent hydroxyl groups of PIC can form a IHB which make the ABS product more stable, so A4-OH group becomes a better H atom donor [47].

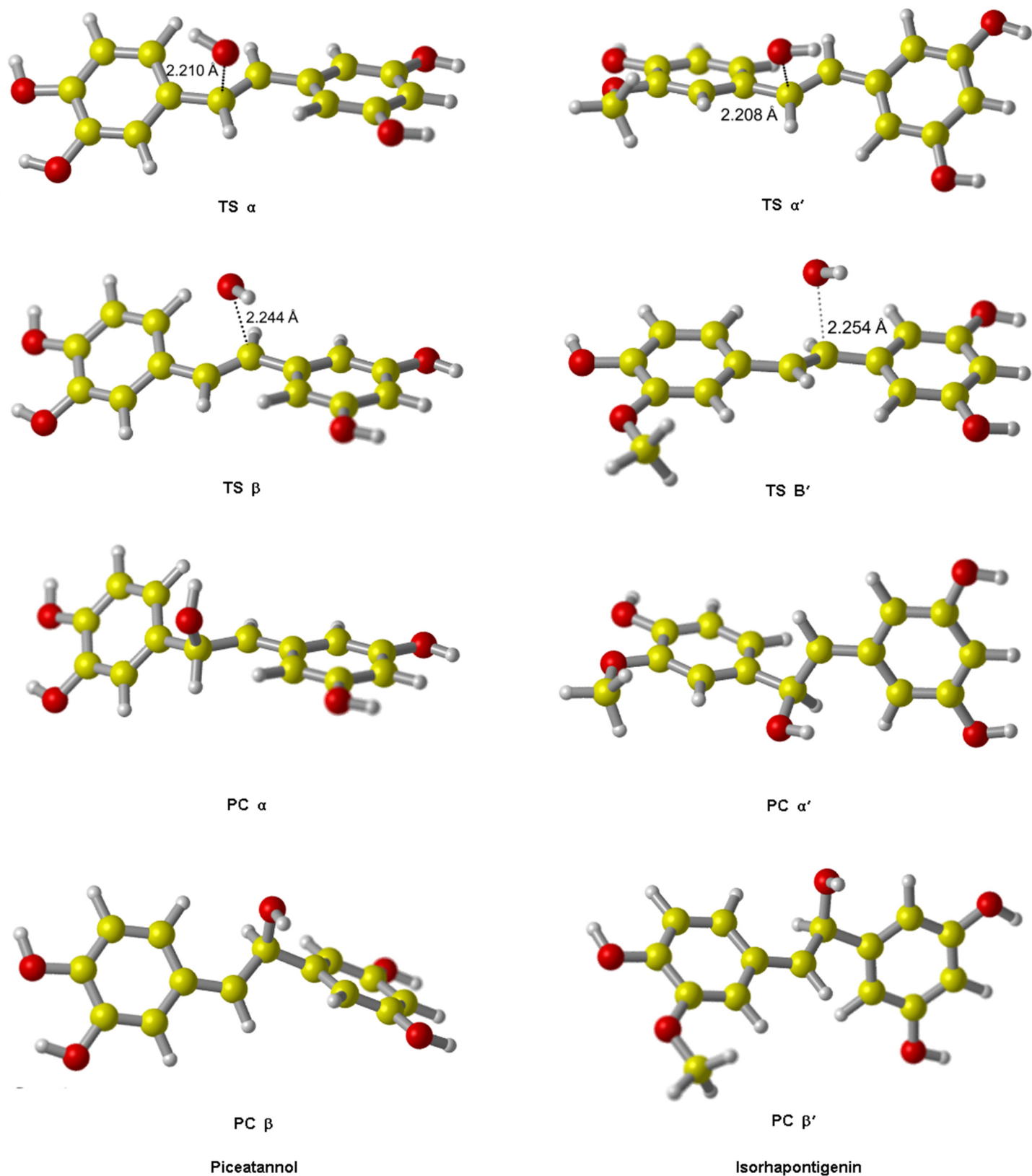


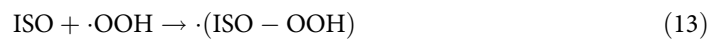
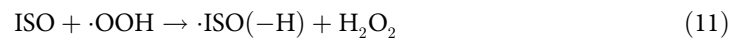
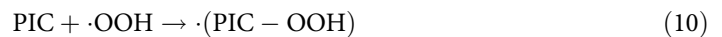
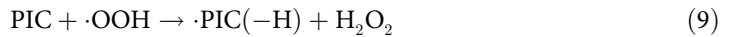
Fig 6. The transition states and product complexes of PIC and ISO from RAF reaction initiated by $\bullet\text{OH}$. All TS have only one imaginary frequency, and in their vibrational mode, $\bullet\text{OH}$ moves in a direction which is perpendicular to that of the carbon atom. The H-atom attached to that carbon atom folds back slightly to accommodate the incoming of $\bullet\text{OH}$.

doi:10.1371/journal.pone.0133259.g006

To sum up, PIC and ISO are capable of scavenging •OH by two mechanisms and the most active site is A4-OH group for ABS mechanism and β carbon atom for RAF mechanism.

Hydroperoxyl Radical (•OOH)

In order to investigate the scavenging activity of PIC and ISO toward •OOH, similar to •OH, eight ABS sites (A4p /A'4p, A5p/A'5p, B3p/B'3p and B5p/B'5p) and four RAF sites (αp/α'p and βp/β'p) were studied, and the corresponding PIC/ISO reaction pathways were denoted as A4p/A'4p, A5p/A'5p, B3p/B'3p, B5p/B'5p, αp/α'p and βp/β'p respectively, according to the following reactions:



For the reactions initiated by •OOH, the reaction Gibbs energies and reaction enthalpies are reported in Tables 4 and 5 respectively (including in the gas phase and water).

Among the pathways of PIC in the gas phase, only A4p is thermodynamically feasible. So the semiquinone radical A4p is the main product of ABS reactions in the gas phase. In water, the ΔG_{sol} of pathway A4p and A5p are both negative as shown in Table 4, i.e. the ABS reactions in ring A are thermodynamically feasible. In contrast, the pathways in ring B are not thermodynamically viable. These are in accord with the previous study [33]. In addition, all RAF pathways are endergonic both in the gas phase and water, so none of addition reaction can happen spontaneously. It is possible to conclude that PIC cannot scavenge •OOH by RAF mechanism.

For ISO, pathway A'4p and A'5p are exergonic and exothermic both in the gas phase and water. As same as PIC, only the reactions in ring A of ISO can happen spontaneously, the ring B has no contribution to scavenge •OOH. In addition, the RAF pathway β'p is thermodynamically feasible. So ISO can scavenge •OOH radical by both of ABS and RAF mechanisms.

All TS of ABS reactions were identified and shown in Fig 7. In each of these structures, •OOH is approximately perpendicular to the molecular plane of PIC or ISO, and orientated toward the H atom of the-OH group. All TS and PC of RAF reactions initiated by •OOH are shown in Fig 8.

Table 4. The reaction Gibbs energies for reactions of PIC and ISO with •OOH in the gas phase and water (in kJ/mol).

site	PIC+•OOH		site	ISO+•OOH	
	ΔG _{gas}	ΔG _{sol}		ΔG _{gas}	ΔG _{sol}
A4p	-44.59	-55.72	A'4p	-25.78	-49.84
A5p	15.78	-24.30	A'5p	-27.29	-52.02
B3p	96.74	97.87	B'3p	97.03	98.16
B5p	98.80	100.47	B'5p	96.10	97.19
αp	5.20	7.80	α'p	0.99	4.26
βp	1.11	2.66	β'p	-2.61	-4.78

doi:10.1371/journal.pone.0133259.t004

Table 5. The reaction enthalpies at 298K for reactions of PIC and ISO with •OOH in the gas phase and water (in kJ/mol).

site	PIC+•OOH		site	ISO+•OOH	
	ΔH_{gas}	ΔH_{sol}		ΔH_{gas}	ΔH_{sol}
A4p	-38.19	-41.34	A'4p	-23.65	-38.95
A5p	15.02	-15.66	A'5p	-22.01	-38.94
B3p	96.49	5.66	B'3p	96.68	3.89
B5p	96.65	4.51	B'5p	96.20	3.56
α p	-47.09	-44.97	α' p	-48.25	-44.62
β p	-52.81	-43.06	β' p	-51.23	-49.26

doi:10.1371/journal.pone.0133259.t005

The barrier heights of all reactions in the gas phase and water are reported in Table 6. Potential energy diagrams are plotted in Figs 9 and 10. As shown in Table 6, for ABS reactions of PIC in the gas phase, the pathway A4p has the lowest barrier height. While in water, A5p becomes the pathway with the lowest barrier height, where the barrier height decreases from 53.69 kJ/mol (in the gas phase) to 38.14 kJ/mol. Therefore, A5p-OH group is also an important active site for PIC to scavenge •OOH in vivo. The high activities of site A4p and A5p are due to the fact that there are two ortho-OH groups in the benzene ring A, and their respective semiquinone radical products yielded by abstraction reactions are more stable due to the formation of IHB interactions [48].

All above indicate that the scavenging activity of PIC toward •OOH is ruled by the hydroxyl groups in ring A.

For ABS reactions of ISO, the pathway A'4p has the lowest barrier height wherever in the gas phase or water. Along with its thermodynamic superiority, A'4p is the absolute leading channel for ISO to scavenge •OOH by ABS mechanism. The higher activity of A'4p is as the presence of the adjacent-OCH₃ group, because its character of electron-donating group (-OCH₃) promotes the H atom transfer of A'4 group. Moreover, unlike PIC, the barrier height of A'5p of ISO are very big. It is probably because the steric hindrance effect brought by-OCH₃ group is bigger than that of-OH group, which results in the decrease of its activity. For the RAF reactions initiated by •OOH, the site β' p is always more active than α' p, as same as the RAF mechanism of ISO with •OH.

In the gas phase, A4p and A'4p are the best active sites for PIC and ISO respectively, and the pathway A4p has a smaller ΔE than A'4p, indicating that the ability of PIC eliminating •OOH is superior to ISO. This is consistent with the result of eliminating •OH. But in water, ISO (A'4p) possess a smallest barrier height among all PIC and ISO pathways, so it is a better scavenger toward •OOH in vivo.

On the basis of the thermodynamic and kinetic results obtained from the reactions of scavenging •OH and •OOH, we also found that both of PIC and ISO have a higher ability to scavenge •OH than •OOH. This is reasonable, since the half-time of •OOH is several orders longer than that of •OH, and the activity of •OH is higher than •OOH.

Conformational Analysis

Conformational analysis is an important tool to investigate the antioxidant capacity of polyphenols, since the antioxidant behavior of-OH group is strongly influenced by the geometry, such as IHB. In order to clarify the probable effect of IHB to antioxidant activity of the studied compounds, we optimized the conformers of PIC and ISO which structures exist different IHB. The geometries of the conformers (PIC-I and ISO-I) with labels are shown in Fig 11. The

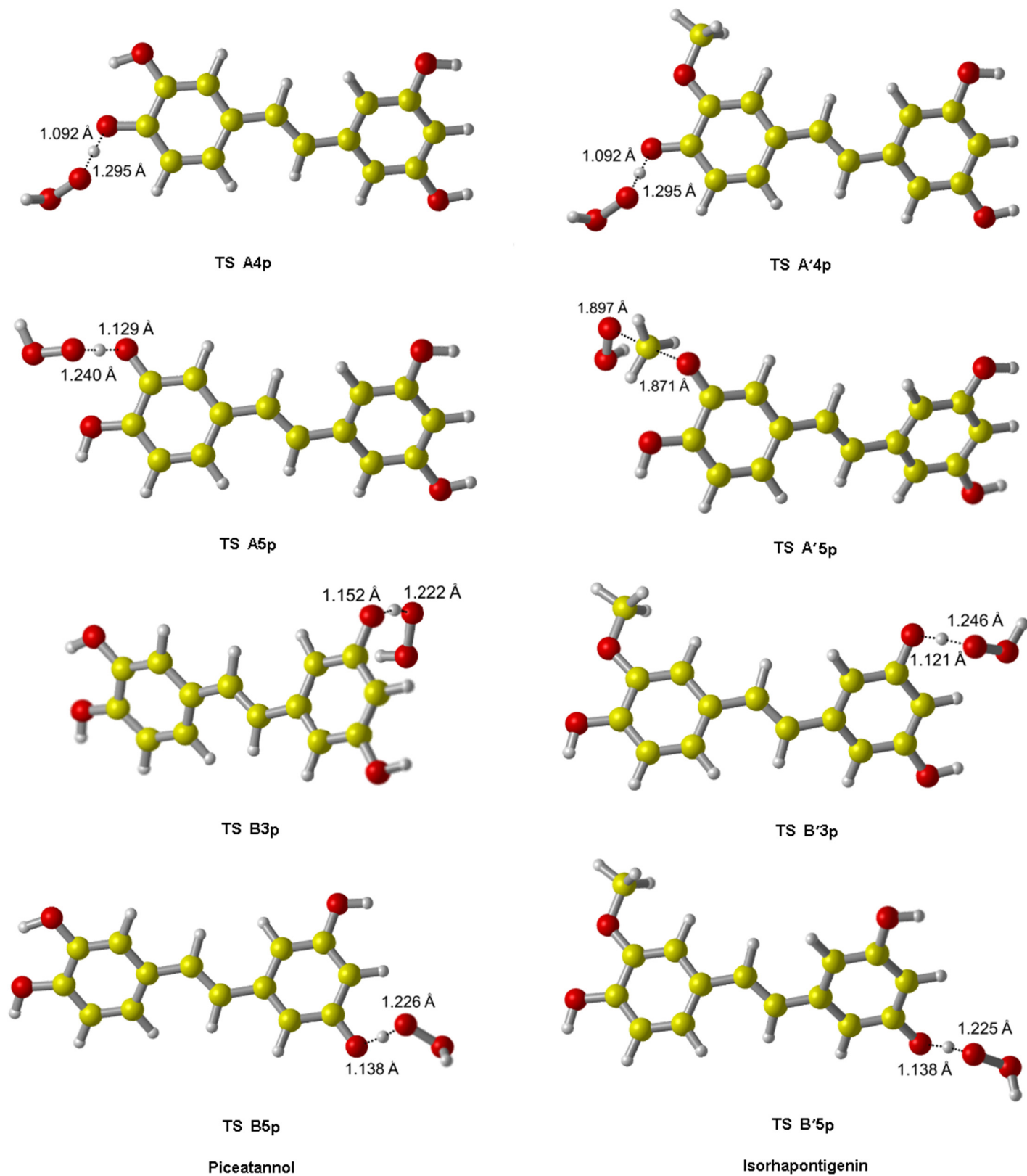


Fig 7. The transition state geometries of PIC and ISO from ABS reaction initiated by •OOH. For the same active site of PIC and ISO, the corresponding TS structures are very similar.

doi:10.1371/journal.pone.0133259.g007

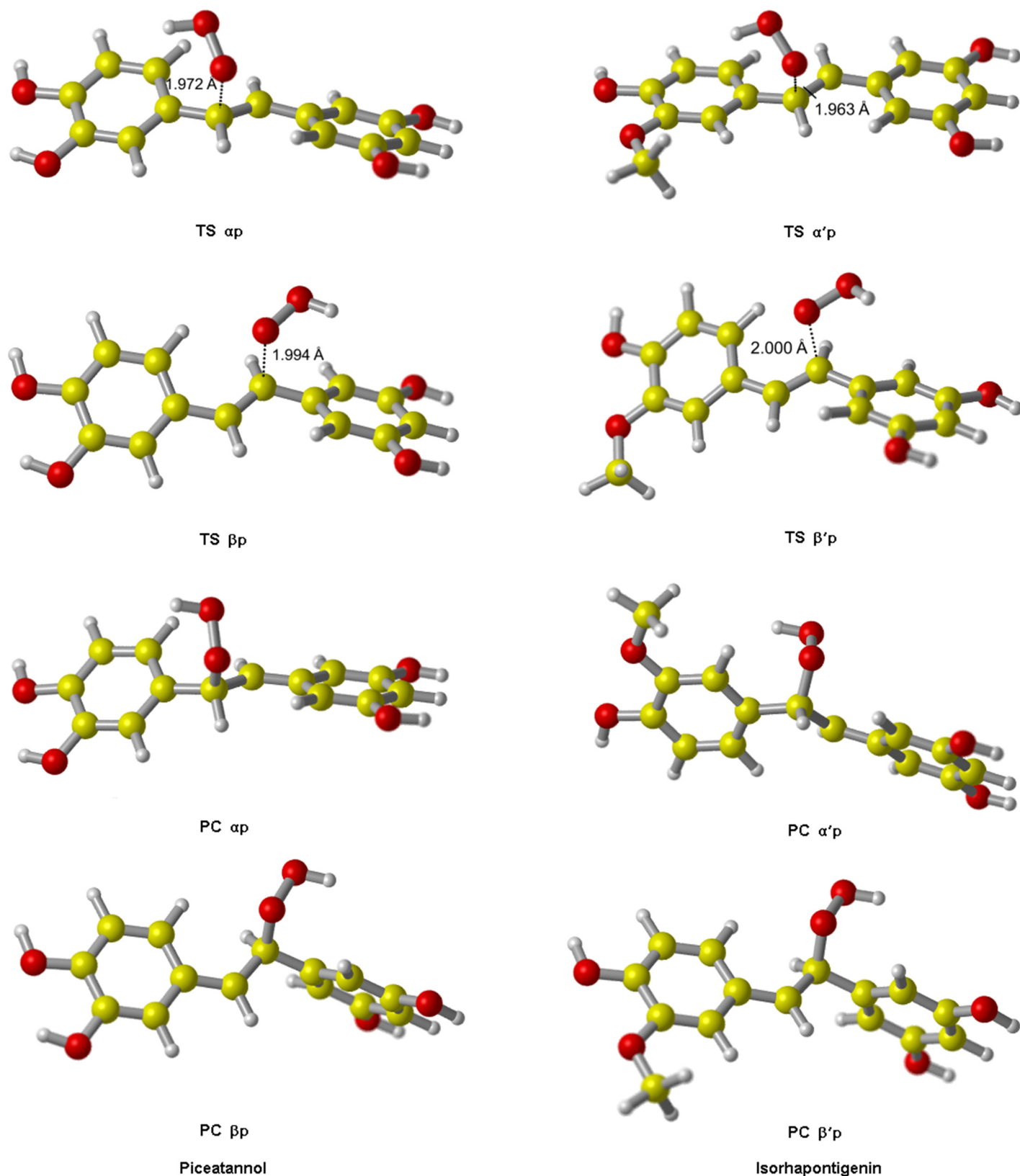


Fig 8. The transition states and product complexes of PIC and ISO from RAF reaction initiated by \bullet OOH. All TS have only one imaginary frequency, and in their vibrational mode, \bullet OOH moves in a direction which is perpendicular to that of the carbon atom. The H atom attached to that carbon atom folds back slightly to accommodate the incoming of \bullet OOH.

doi:10.1371/journal.pone.0133259.g008

Table 6. The barrier heights of TS with zero-point energy corrections for reactions of PIC and ISO with •OOH in the gas phase and water (in kJ/mol).

PIC+•OOH			ISO+•OOH		
site	ΔE_{gas}	ΔE_{sol}	site	ΔE_{gas}	ΔE_{sol}
A4p	22.31	50.53	A'4p	28.27	15.70
A5p	53.69	30.36	A'5p	181.33	166.99
B3p	30.81	48.87	B'3p	43.72	56.29
B5p	45.13	74.63	B'5p	43.96	50.36
α p	30.69	35.32	α' p	29.71	33.33
β p	27.92	35.15	β' p	26.26	33.26

doi:10.1371/journal.pone.0133259.t006

structure features of PIC-I and ISO-I are A4-I hydroxy group form a IHB with the O atom of the adjacent A5-I hydroxy group, while the structures of ring B are not changed. So we select two active sites (A4-I and A5-I) in ring A as the target and make comparing between the same sites of PIC and ISO.

By the same method of M05-2X/6-311++G(d,p), we calculated the thermodynamic data of PIC-I and ISO-I scavenging •OH radicals (in Table 7). This table shows that the ABS reactions of PIC-I and ISO-I with •OH are all exothermic and thermodynamically favored.

As we have mentioned above, BDE is an important indicator of evaluating the activity of -OH groups in polyphenols, the relative low O-H BDE will facilitate their H-transfer ability. So we computed the O-H BDE of PIC/ISO along with their conformers, and listed in Table 8.

It has been reported that IHB could stabilize the parent molecule and the radical formed after H atom has been abstracted, but the effect to the latter is more strong. Therefore the BDE

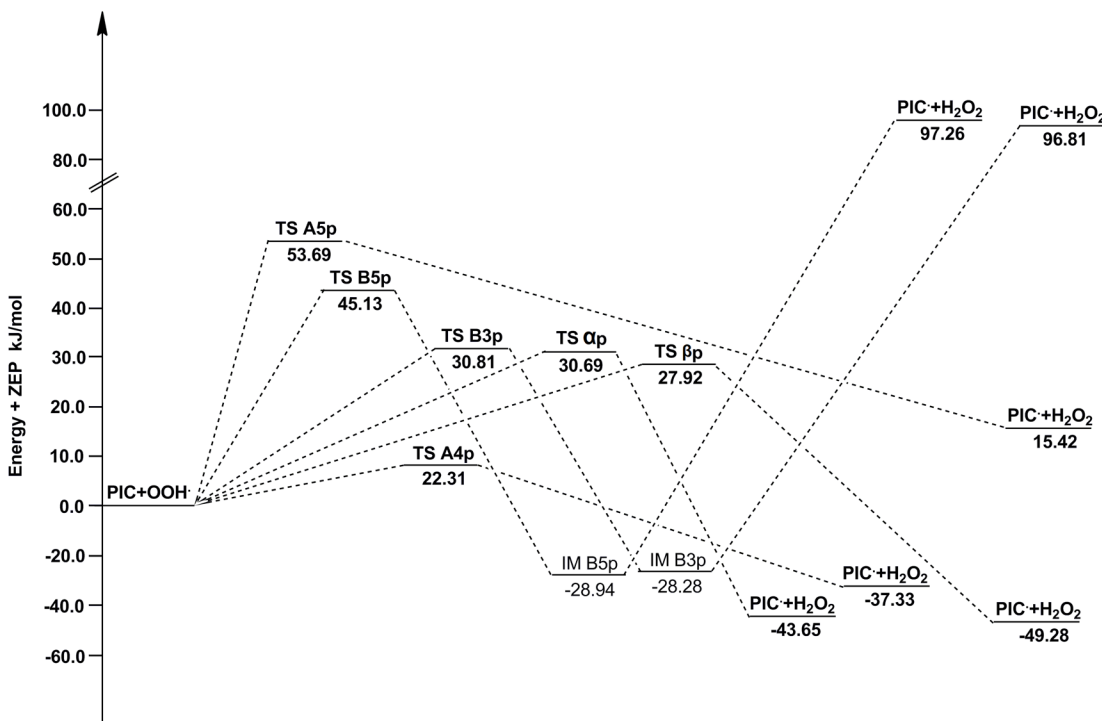


Fig 9. The potential energy surfaces of the reactions of PIC with •OOH in the gas phase. The relative energies (in kJ/mol) were calculated at the M05-2X/6-311++G(d,p) + ZPE level. To facilitate comparisons, the energy of the reactants are set to zero.

doi:10.1371/journal.pone.0133259.g009

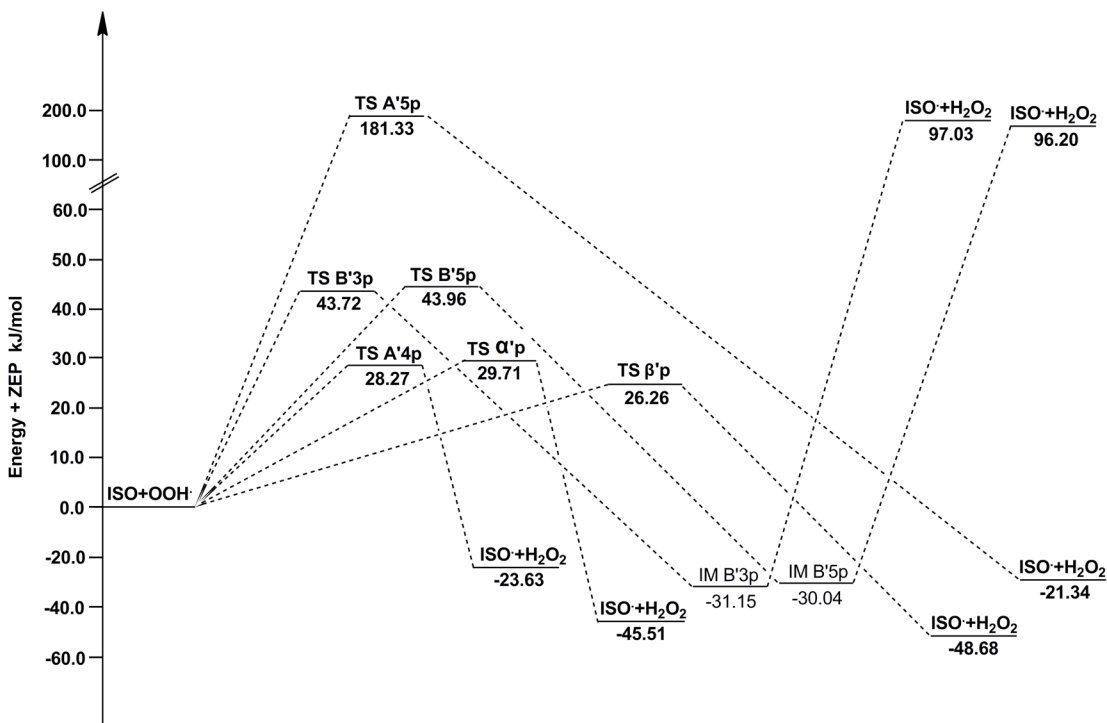


Fig 10. The potential energy surfaces of the reactions of ISO with •OOH in the gas phase. The relative energies (in kJ/mol) were calculated at the M05-2X/6-311++G(d,p) + ZPE level. To facilitate comparisons, the energy of the reactants are set to zero.

doi:10.1371/journal.pone.0133259.g010

of-OH group which participates the formation of IHB will be increased and the BDE of free-OH group will be decreased [49]. For the A4-I hydroxy group of PIC-I, it involves a OH...O hydrogen bond with the O atom of the adjacent hydroxy group; in contrast, the A4-OH group of PIC is free. So from Table 8, the O-H BDE of A4-I is higher than that of A4, this means the activity of A4-I is decreased. What is more, the BDE of A5-I hydroxy group which is free in PIC-I is lower than that of A5 hydroxy group which involved in IHB in PIC, so the activity of A5-I is increased. To sum up, the O-H BDE of PIC shows remarkable dependence on the pattern of IHB.

Then we identified all TS of PIC-I and ISO-I scavenging •OH in the gas phase, their fully optimized geometries are shown in Fig 12. We also obtained the barrier heights of all pathways, along with the corresponding barrier heights of PIC and ISO, listed in Table 8. According to our previous discuss, the most active ABS site of PIC scavenging •OH is A4. But from this Table, it was interestingly found that the most active site of the PIC-I has changed: the barrier height of pathway A5-I is lower than that of A4-I, A5-I becomes the easiest site abstracted by •OH. This can be explained that, for PIC-I, the H atom of A4-I hydroxy group which involved in IHB is more stable, therefore it needs more energy to be abstracted. And this is agreement with the change of its BDE. While for the free-OH group of A5-I, its product radical is stabilized by the IHB where adjacent A4-I OH interacts with its O atom. The presence of IHB decreased the O-H BDE of A5-I, hence it becomes the most active-OH group of PIC-I with •OH both in the gas phase and water phase.

By comparison the Figs 1 and 11, we can see that there is no IHB in the structure of ISO. While for ISO-I, its H atom of A4-I group forms a IHB with the O atom of the adjacent methoxy group (A5-I). Table 8 shows that the difference of O-H BDE of A4 and A4-I is minor. But the barrier height of pathway A4-I is especially higher than that of A4 in ISO. This proved that

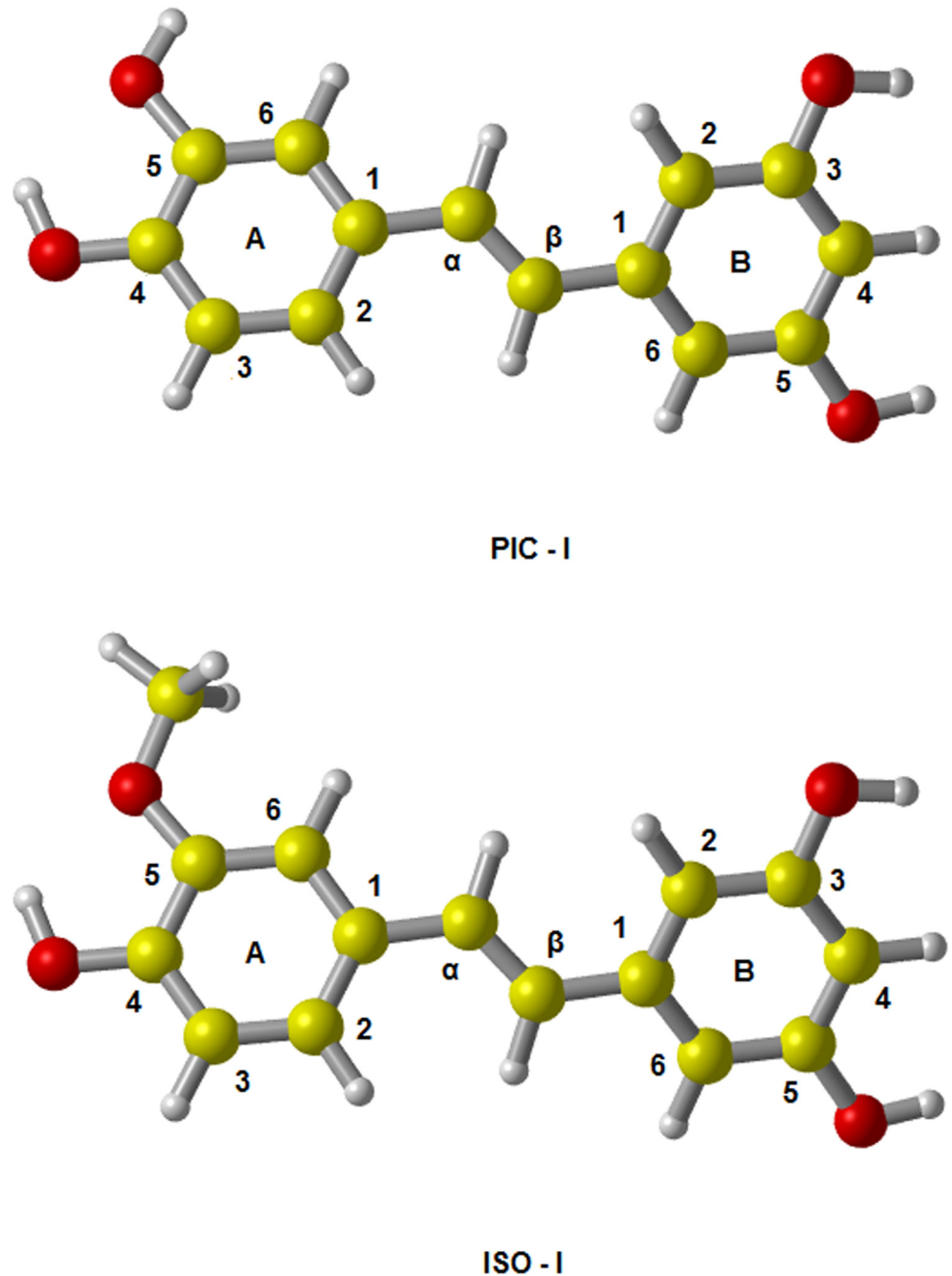


Fig 11. The optimized geometries of PIC-I and ISO-I in the gas phase. The dihedral angle between two benzene rings of PIC-I is 179.87° , and the dihedral angle between two benzene rings of ISO-I is -179.99° .

doi:10.1371/journal.pone.0133259.g011

the presence of IHB could decrease the activity of the -OH group involved in the H-bond formation.

In conclusion, IHB has an important effect on the activity of the -OH group of PIC and ISO. The existence and pattern of IHB can potentially lead to completely different theoretically estimated H-transfer activity. Therefore, the conformational effect is an important factor that should not be neglected in investigating the antioxidant activity of PIC and ISO analogues.

Table 7. The reaction Gibbs energies and reaction energies at 298K for reactions of PIC-I and ISO-I with •OH in the gas phase and water (in kJ/mol).

site	PIC-I + •OH				ISO-I + •OH			
	ΔG_{gas}	ΔG_{sol}	ΔH_{gas}	ΔH_{sol}	ΔG_{gas}	ΔG_{sol}	ΔH_{gas}	ΔH_{sol}
A4-I	-137.56	-182.75	-133.28	-167.15	-140.80	-185.73	-135.11	-170.12
A5-I	-160.40	-173.91	-157.12	-160.23	-149.80	-164.24	-140.10	-145.10

doi:10.1371/journal.pone.0133259.t007

Table 8. The bond dissociation enthalpies of O-H bonds and the barrier heights of TS with zero-point energy corrections for reactions of PIC-I/PIC and ISO-I/ISO with •OH in the gas phase and water (in kJ/mol).

site	PIC-I/PIC + •OH			ISO-I/ISO + •OH		
	BDE	ΔE_{gas}	ΔE_{sol}	BDE	ΔE_{gas}	ΔE_{sol}
A4-I	347.41	2.39	-39.16	324.00	0.83	-42.91
A5-I	324.42	-12.87	-50.51	-	-	-
A4	310.30	-15.54	-58.86	324.00	-12.50	-56.45
A5	363.05	5.08	-30.58	-	-	-

doi:10.1371/journal.pone.0133259.t008

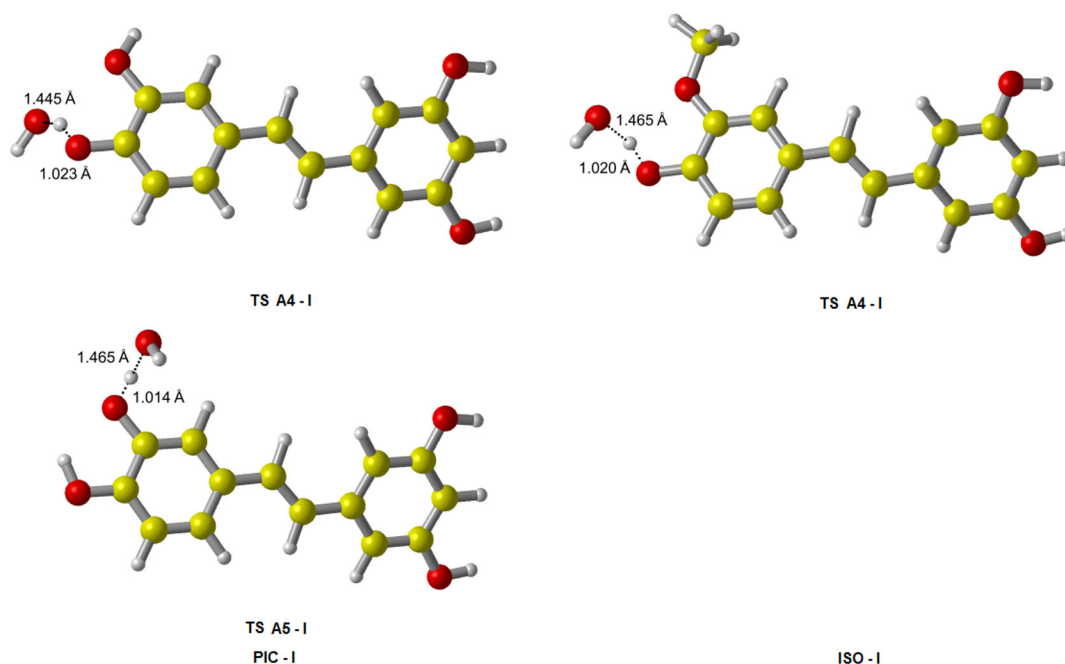


Fig 12. The transition state geometries of PIC-I and ISO-I from ABS reactions initiated by •OH. The elongations of the breaking bonds are smaller than those of the forming bonds, indicating that these TS are all reactant-like, i.e. these reactions are all exothermic in light of the Hammond's postulate. These agree with our calculated results in Table 7.

doi:10.1371/journal.pone.0133259.g012

Conclusions

In this work, the antioxidation mechanisms behind •OH and •OOH scavenging activities of PIC and ISO were studied by using a quantum mechanical method at the M05-2X/6-311++G (d,p) level of theory. The reaction Gibbs energy, reaction enthalpy and reaction potential barrier height were computed in the gas phase and water. Two mechanisms, ABS and RAF, were investigated.

To scavenge $\bullet\text{OH}$, both of ABS and RAF mechanisms are thermodynamically and kinetically feasible for PIC and ISO. For ABS mechanism, A4-hydroxyl group is the most active site of PIC and ISO both in the gas phase and water. The structure difference of A4/A'4 with other hydroxyl groups of PIC/ISO is that there is a substituent group on its ortho-position. So we conclude that introducing adjacent-OH and-OCH₃ group would increase the radical scavenging activity of PIC and ISO. PIC is more effective than ISO in scavenging $\bullet\text{OH}$, because its ortho-dihydroxyl moiety allows to share the H atom by forming the IHB, so the corresponding product is more stable. For addition mechanism, the sites of β are always more active than α for both of PIC and ISO.

To scavenge $\bullet\text{OOH}$, we find that ABS is the only thermodynamically feasible mechanism for PIC; while for ISO, both of ABS and RAF mechanisms are viable. In the gas phase, A4-hydroxyl group is still the most active site for PIC and ISO to scavenge $\bullet\text{OOH}$ by ABS mechanism, and the activity of PIC is more prominent than ISO. But in water, due to the solvent effect, A5-hydroxyl group becomes the most active site of PIC, at the same time PIC loses its superiority of scavenging $\bullet\text{OOH}$ in contrast to ISO. So the antioxidation activity of PIC is mainly controlled by the benzene ring A, and ISO is more effective than PIC in eliminating $\bullet\text{OOH}$ in organisms.

The changes of conformation have great influence on the antioxidant activity of PIC and ISO. The activity of the-OH group which participates in the formation of IHB will be decreased, while the activity of free-OH group will be increased.

Although both of PIC and ISO have a lower ability to scavenge $\bullet\text{OOH}$ than $\bullet\text{OH}$, considering $\bullet\text{OOH}$ may also have a significant contribution to the oxidation in biological media, we conclude that PIC and ISO are very good antioxidants. To sum up, our study offers a deep understanding on the relationship of antioxidation mechanism with structure property of PIC and ISO.

Supporting Information

S1 Fig. The optimized geometries of the reactant complexes for the reactions of PIC with $\bullet\text{OH}$ in the gas phase.

(TIF)

S2 Fig. The optimized geometries of the reactant complexes for the reactions of ISO with $\bullet\text{OH}$ in the gas phase.

(TIF)

S3 Fig. The optimized geometries of the product complexes of PIC and ISO from ABS reactions initiated by $\bullet\text{OOH}$ in the gas phase.

(TIF)

Acknowledgments

We thank Dr.s Niu Huang and Hua Lan (National Institute of Biological Science, Beijing 102206, China) for their fruitful discussions and checking the language. We also thanks the grid computing server provided by the Chinese Academy of Sciences.

Author Contributions

Conceived and designed the experiments: YL HZ ZL. Performed the experiments: YL AW PS. Analyzed the data: YL AW PS. Contributed reagents/materials/analysis tools: HZ. Wrote the paper: YL. Literature research: YL AW PS. Manuscript revision/review: HZ ZL.

References

1. Bavaresco L, Fregoni M, Trevisan M, Mattivi F, Vrhovsek U, Falchetti R (2002) The occurrence of the stilbene piceatannol in grapes. *Vitis* 41: 133–136.
2. Ku KL, Chang PS, Cheng YC, Lien CY (2005) Production of stilbenoids from the callus of arachis hypogaea: A novel source of the anticancer compound piceatannol. *Journal of Agricultural and Food Chemistry* 5: 33877–3881.
3. Rimando AM, Kalt W, Magee JB, Dewey J, Ballington JR (2004) Resveratrol, pterostilbene, and piceatannol in vaccinium berries. *Journal of Agricultural and Food Chemistry* 52: 4713–4719. PMID: [15264904](#)
4. Ferrigni NR, McLaughlin JL, Powell RG, Smith CR (1984) Use of potato disc and brine shrimp bioassays to detect activity and isolation of piceatannol as the antileukemic principle from the seeds of euphorbia lagascae. *Journal of Natural Products* 47: 347–352. PMID: [6736971](#)
5. Potter GA, Patterson LH, Wanogho E, Perry PJ, Butler PC, Ijaz T, et al. (2002) The cancer preventative agent resveratrol is converted to the anticancer agent piceatannol by the cytochrome P450 enzyme CYP1B1. *British Journal of Cancer* 86: 774–778. PMID: [11875742](#)
6. Cai YJ, Fang JG, Ma LP, Yang L, Liu ZL (2003) Inhibition of free radical-induced peroxidation of rat liver microsomes by resveratrol and its analogues. *Biochimica et Biophysica Acta* 1637: 31–38. PMID: [12527404](#)
7. Surh YJ, Hurh JY, Lee E, Kong G, Lee S (1999) Resveratrol, an antioxidant present in red wine, induces apoptosis in human promyelocytic leukemia (HL-60) cells. *Cancer Letters* 140: 1–10. PMID: [10403535](#)
8. Wang MF, Li JG, Rangarajan M, Shao Y, Lavoie EJ, Huang TC, et al. (1998) Antioxidative phenolic compounds from sage (*Salvia officinalis*). *Journal of Agricultural and Food Chemistry* 46: 4869–4873.
9. Fang JG, Lu M, Chen ZH, Zhu HH, Li Y, Yang L, et al. (2002) Antioxidant effects of resveratrol and its analogues against the free-radical-induced peroxidation of linoleic acid in micelles. *Chemistry-A European Journal* 8: 4191–4198.
10. Lee SK, Mbwanbo ZH, Chung H, Luyengi L, Gamez EJC, Mehta RG, et al. (1998) Evaluation of the antioxidant potential of natural products. *Combinatorial Chemistry and High Throughput Screening* 1: 35–46. PMID: [10499128](#)
11. Howitz KT, Bitterman KJ, Cohen HY, Lamming DW, Lavu S, Wood JG, et al. (2003) Small molecule activators of sirtuins extend *Saccharomyces cerevisiae* lifespan. *Nature* 425: 191–196. PMID: [12939617](#)
12. Kimura Y, Okuda H, Arichi S (1985) Effects of stilbenes on arachidonate metabolism in leukocytes. *Biochimica et Biophysica Acta* 834: 275–278. PMID: [3922423](#)
13. Jang M, Cai L, Udeani GO, Slowing KV, Thomas CF, Beecher CWW, et al. (1997) Cancer chemopreventive activity of resveratrol, a natural product derived from grapes. *Science* 275: 218–220. PMID: [8985016](#)
14. Docherty JJ, Fu MMH, Stiffler BS, Limperos RJ, Pokabla CM, Delucia AL (1999) Resveratrol inhibition of herpes simplex virus replication. *Antiviral Research* 43: 135–145. PMID: [10551373](#)
15. Stivala LA, Savio M, Carafoli F, Perucca P, Bianchi L, Maga G, et al. (2001) Specific structural determinants are responsible for the antioxidant activity and the cell cycle effects of resveratrol. *Journal of Biological Chemistry* 276: 22586–22594. PMID: [11316812](#)
16. King RE, Bomser JA, Min DB (2006) Bioactivity of resveratrol. *Comprehensive Reviews in Food Science and Food Safety* 5: 65–69.
17. Djoko B, Chiou RYY, Shee JJ, Liu YW (2007) Characterization of immunological activities of peanut stilbenoids, arachidin-1, piceatannol, and resveratrol on lipopolysaccharide-induced inflammation of RAW 264.7 macrophages. *Journal of Agricultural and Food Chemistry* 55: 2376–2383. PMID: [17316017](#)
18. Bastianetto S, Dumont Y, Han Y, Quirion R (2009) Comparative neuroprotective properties of stilbene and catechin analogs: action via a plasma membrane receptor site? *CNS Neuroscience & Therapeutics* 15: 76–83.
19. Fernandez-Marin MI, Guerrero RF, Garcia-Parrilla MC, Puertas B, Richard T, Rodriguez-Werner MA, et al. (2012) Isorhapontigenin: a novel bioactive stilbene from wine grapes. *Food Chemistry* 135: 1353–1359. doi: [10.1016/j.foodchem.2012.05.086](#) PMID: [22953865](#)
20. Stojanovic S, Sprinz H, Brede O (2001) Efficiency and mechanism of the antioxidant action of trans-resveratrol and its analogues in the radical liposome oxidation. *Archives of Biochemistry and Biophysics* 391: 79–89. PMID: [11414688](#)
21. Wang QL, Lin M, Liu GT (2001) Antioxidative activity of natural isorhapontigenin. *The Japanese Journal of Pharmacology* 87: 61–66.

22. Fang YN, Liu GT (2002) Effect of isorhapontigenin on respiratory burst of rat neutrophils. *Phytomedicine* 9: 734–738. PMID: [12587695](#)
23. Liu YL, Liu GT (2004) Isorhapontigenin and resveratrol suppress oxLDL-induced proliferation and activation of ERK1/2 mitogen-activated protein kinases of bovine aortic smooth muscle cells *Biochemical Pharmacology* 67: 777–785. PMID: [14757178](#)
24. Harman D (1956) Aging: A theory based on free-radical and radiation chemistry. *Journals of Gerontology* 11: 298–300. PMID: [13332224](#)
25. McCord JM, Fridovich I (1969) Superoxide dismutase an enzymic function for erythrocyte hemocuprein. *Journal of Biological Chemistry* 244: 6049–6055. PMID: [5389100](#)
26. Hagen TM (2003) Oxidative stress, redox imbalance, and the aging process. *Antioxidants & Redox Signaling* 5: 503–506.
27. Esterbauer H, Eckl P, Ortner A (1990) Possible mutagens derived from lipids and lipid precursors. *Mutation Research* 238: 223–233. PMID: [2342513](#)
28. Simon HU, Haj-Yehia A, Levi-Schaffer F (2000) Role of reactive oxygen species (ROS) in the apoptosis induction. *Apoptosis* 5: 415–418. PMID: [11256882](#)
29. Jancinova V, Perecko T, Nosal R, Svitekova K, Drabivoka K (2013) The natural stilbenoid piceatannol decreases activity and accelerates apoptosis of human neutrophils: involvement of protein kinase C. *Oxidative Medicine and Cellular Longevity* 136539: 1–8.
30. Rossi M, Caruso F, Opazo C, Salciccioli J (2008) Crystal and molecular structure of piceatannol; scavenging features of resveratrol and piceatannol on hydroxyl and peroxy radicals and docking with trans-thyretin. *Journal of Agricultural and Food Chemistry* 56: 10557–10566. doi: [10.1021/jf801923j](#) PMID: [18959413](#)
31. Mikulski D, Molski M (2012) Quantum-mechanical computations on the electronic structure of trans-resveratrol and trans-piceatannol: a theoretical study of the stacking interactions in trans-resveratrol dimers. *Journal of Molecular Modeling* 18: 3255–3266. doi: [10.1007/s00894-011-1342-7](#) PMID: [22249749](#)
32. Pérez-González A, Rebollar-Zepeda AM, León-Carmona JR, Galano A (2012) Reactivity indexes and O-H bond dissociation energies of a large series of polyphenols: Implications for their free radical scavenging activity. *Journal of the Mexican Chemical Society* 56: 241–249.
33. Cordova-Gomez M, Galano A, Alvarez-Idaboy JR (2013) Piceatannol, a better peroxy radical scavenger than resveratrol. *RSC Advances* 3: 20209–20218.
34. Pastor N, Weinstein H, Jamison E, Brenowitz M (2000) A detailed interpretation of OH radical footprints in a TBP DNA complex reveals the role of dynamics in the mechanism of sequencespecific binding. *Journal of Molecular Biology* 304: 55–68. PMID: [11071810](#)
35. Pryor WA (1986) Oxy-radicals and related species: their formation, lifetimes and reactions. *Annual Review of Physiology* 48: 657–667. PMID: [3010829](#)
36. Marnett LJ (1987) Peroxyl free radicals: potential mediators of tumor initiation and Promotion. *Carcinogenesis* 8: 1365–1373. PMID: [3477337](#)
37. Gaussian 09, Revision A. 02: Frisch MJ, Trucks GW, Schlegel HB, Scuseria GE, Robb MA, Cheeseman JR, Scalmani G, Barone V, Mennucci B, Petersson GA, Nakatsuji H, Caricato M, Li X, Hratchian HP, Izmaylov AF, Bloino J, Zheng G, Sonnenberg JL, Hada M, Ehara M, Toyota K, Fukuda R, Hasegawa J, Ishida M, Nakajima T, Honda Y, Kitao O, Nakai H, Vreven T, Montgomery JA. Jr, Peralta JE, Ogliaro F, Bearpark M, Heyd JJ, Brothers E, Kudin KN, Staroverov VN, Kobayashi R, Normand J, Raghavachari K, Rendell A, Burant JC, Iyengar SS, Tomasi J, Cossi M, Rega N, Millam JM, Klene M, Knox JE, Cross JB, Bakken V, Adamo C, Jaramillo J, Gomperts R, Stratmann RE, Yazyev O, Austin AJ, Cammi R, Pomelli C, Ochterski JW, Martin RL, Morokuma K, Zakrzewski VG, Voth GA, Salvador P, Dannenberg J.J., Dapprich S, Daniels A.D, Farkas O, Foresman JB, Ortiz JV, Cioslowski J, Fox DJ Gaussian, Inc., Wallingford CT, 2009.
38. Zhao Y, Schultz NE, Truhlar DG (2006) Design of density functionals by combining the method of constraint satisfaction with parameterization for thermochemistry, thermochemical kinetics, and noncovalent interactions. *Journal of Chemical Theory Computation* 2: 364–382.
39. Zavala-Oseguera C, Alvarez-Idaboy JR, Merino G, Galano A (2009) OH radical gas phase reactions with aliphatic ethers: a variational transition State Theory Study. *The Journal of Physical Chemistry A* 113: 13913–13920. doi: [10.1021/jp906144d](#) PMID: [19908880](#)
40. Galano A, Macías-Ruvalcaba NA, Campos ONM, Pedraza-Chaverri J (2010) Mechanism of the OH radical scavenging activity of nordihydroguaiaretic acid: a combined theoretical and experimental study. *The Journal of Physical Chemistry B* 114: 6625–6635. doi: [10.1021/jp912001c](#) PMID: [20415502](#)

41. Iuga C, Alvarez-Idaboy JR, Vivier-Bunge A (2011) ROS initiated oxidation of dopamine under oxidative stress conditions in aqueous and lipidic environments. *The Journal of Physical Chemistry B* 115: 12234–12246. doi: [10.1021/jp206347u](https://doi.org/10.1021/jp206347u) PMID: [21919526](https://pubmed.ncbi.nlm.nih.gov/21919526/)
42. Marenich AV, Cramer CJ, Truhlar (2009) DG Universal solvation model based on solute electron density and on a continuum model of the solvent defined by the bulk dielectric constant and atomic surface tensions. *Journal of Physical Chemistry B* 113: 6378–6396.
43. Okuno Y (1997) Theoretical investigation of the mechanism of the baeyer-villiger reaction in nonpolar solvents. *Chemistry-A European Journal*. 3: 212–218.
44. Ardura D, Lopez R, Sordo TL (2005) Relative gibbs energies in solution through continuum models: effect of the loss of translational degrees of freedom in bimolecular reactions on gibbs energy barriers. *Journal of Physical Chemistry B* 109: 23618–23623.
45. Ricks AB, Solomon GC, Colvin MT, Scott AM, Chen K, Ratner MA, et al. (2010) Controlling electron transfer in donor-bridge-acceptor molecules using cross-conjugated bridges. *Journal of the American Chemical Society* 132: 15427–15434. doi: [10.1021/ja107420a](https://doi.org/10.1021/ja107420a) PMID: [20942407](https://pubmed.ncbi.nlm.nih.gov/20942407/)
46. Hammond GS (1955) A correlation of reaction rates. *Journal of the American Chemical Society* 77: 334–338.
47. Shang YJ, Jin XL, Shang XL, Tang JJ, Liu GY, Dai F, et al. (2010) Antioxidant capacity of curcumin-directed analogues: structure-activity relationship and influence of microenvironment. *Food Chemistry* 119:1435–1442.
48. Leopoldini N, Russo N, Toscano (2011) The molecular basis of working mechanism of natural polyphenolic antioxidants. *Food Chemistry* 125: 288–306.
49. Zhang HY, Sun YM, Wang XL (2003) Substituent effects on O-H bond dissociation enthalpies and ionization potentials of catechols: a DFT study and its implications in rational design of phenolic antioxidants and elucidation of structure—activity relationships for flavonoid antioxidants. *Chemistry—A European Journal* 9: 502–508.