

Prevalence and Molecular Characterization of Cryptosporidium in Goats across Four Provincial Level Areas in China



Rongsheng Mi^{1,2}, Xiaojuan Wang^{1,2}, Yan Huang^{1,2}, Peng Zhou^{1,2}, Yuxuan Liu^{1,2}, Yongjun Chen^{1,2}, Jun Chen³, Wei Zhu⁴, Zhaoguo Chen^{1,2}*

1 Key Laboratory of Animal Parasitology of Ministry of Agriculture, Animal-borne Food Safety Research Center of Chinese Academy of Agricultural Sciences, Shanghai Veterinary Research Institute, Chinese Academy of Agricultural Sciences, Shanghai, China, 2 Jiangsu Co-innovation Center for Prevention and Control of Important Animal Infectious Diseases and Zoonoses, Yangzhou, China, 3 Lvxiang Town Agricultural Technology Extension Station of Jinshan District, Shanghai, China, 4 Tengzhou Animal Husbandry and Veterinary Technology Service Center, Tengzhou, China

Abstract

This study assessed the prevalence, species and subtypes of *Cryptosporidium* in goats from Guangdong Province, Hubei Province, Shandong Province, and Shanghai City of China. Six hundred and four fecal samples were collected from twelve goat farms, and the overall infection rate was 11.4% (69/604). Goats infected with *Cryptosporidium* were found in eleven farms across four provincial areas, and the infection rate ranged from 2.9% (1/35) to 25.0% (9/36). Three *Cryptosporidium* species were identified. *Cryptosporidium xiaoi* (45/69, 65.2%) was the dominant species, followed by *C. parvum* (14/69, 20.3%) and *C. ubiquitum* (10/69, 14.5%). The infection rate of *Cryptosporidium* spp. was varied with host age and goat kids were more susceptible to be infected than adult goats. Subtyping *C. parvum* and *C. ubiquitum* positive samples revealed *C. parvum* subtype IldA19G1 and *C. ubiquitum* subtype Xlla were the most common subtypes. Other *C. parvum* subtypes were detected as well, such as IlaA14G2R1, IlaA15G1R1, IlaA15G2R1 and IlaA17G2R1. All of these subtypes have also been detected in humans, suggesting goats may be a potential source of zoonotic cryptosporidiosis. This was the first report of *C. parvum* subtypes IlaA14G2R1, IlaA15G1R1 and IlaA17G2R1 infecting in goats and the first molecular identification of *C. parvum* and its subtypes in Chinese goats.

Citation: Mi R, Wang X, Huang Y, Zhou P, Liu Y, et al. (2014) Prevalence and Molecular Characterization of *Cryptosporidium* in Goats across Four Provincial Level Areas in China. PLoS ONE 9(10): e111164. doi:10.1371/journal.pone.0111164

Editor: Yung-Fu Chang, Cornell University, United States of America

Received July 17, 2014; Accepted September 24, 2014; Published October 24, 2014

Copyright: © 2014 Mi et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Data Availability: The authors confirm that all data underlying the findings are fully available without restriction. All relevant data are within the paper.

Funding: This study was supported in part by National S & T Major Program of the People's Republic of China (Grant No. 2012ZX10004220), Opening Foundation of State Key Laboratory of Veterinary Etiological Biology (Grant No. SKLVEB2013KFKT017), Shanghai Municipal Agriculture Commission (Grant No. 2005-3-4), Minhang District Human Resources and Social Security and Basic Foundation for Scientific Research of State-level Public Welfare Institutes of China (Grant No. 2013JB13). The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing Interests: The authors have declared that no competing interests exist.

* Email: zhaoguochen@shvri.ac.cn

Introduction

Cryptosporidium is an intestinal protozoan parasite that reportedly infects humans and animals worldwide [1]. In neonatal ruminants, cryptosporidiosis is a leading cause of diarrhea and mortality, and causes farmers significant economic loss [2]. Numerous molecular biological techniques have detected Cryptosporidium species/genotype and subtypes and will improve our understanding of cryptosporidiosis transmission in man and animals [3].

The first Cryptosporidium infection in a goat was reported in Australia [4] and has since then been reported worldwide [2]. Few investigations report the molecular characterization of Cryptosporidium species/genotypes in goats. Belgium [5], Egypt [6], France [7], India [8], Italy [9], Norway [10], Spain [11], Sri Lanka [12], the Czech Republic [13], and Zambia [14] have all found Cryptosporidium parvum as the dominant species. Other species/genotypes, such as C. hominis, C. xiaoi, C. ubiquitum, C. andersoni, Cryptosporidium rat genotype and a novel Cryptosporidium genotype were also described in goats [10,15–22].

Goats are economic resources in China. According to the National Bureau of Statistics, about 141.4 million goats were in China at the end of 2012 (http://data.stats.gov.cn/workspace/ index?m=hgnd), but few studies exist identifying the molecular epidemiology of cryptosporidiosis in goats. In one study, three species (C. ubiquitum, C. andersoni, and C. xiaoi) were found in Henan Province and one species (C. ubiquitum) was found in Chongqing City [20]. In another report, C. xiaoi and a novel Cryptosporidium genotype were found in goats in Qinghai Province [17]. Given the sparse data on prevalence and molecular characterization of Cryptosporidium spp. in goats in China, the present study aimed to supplement the genetic characterization of Cryptosporidium spp. in partial provinces of China, to investigate the subtypes of C. parvum and C. ubiquitum, and to evaluate the potential threat of Cryptosporidium spp. in goats to human health.

Materials and Methods

Ethics Statement

The protocol of this work was approved by the Animal Care and Use of Chinese Academy of Agricultural Sciences, and authorized by the Animal Ethical Committee of Shanghai Veterinary Research Institute. All the fecal samples obtained from goat farms were permitted by the owners of farms.

Sample Collection

Fresh fecal samples were collected from 12 goat farms across four provincial areas of China from November 2007 to September 2013. Two farms from Huizhou City (22°47'N, 114°27'E) are located in Guangdong Province (South China area), four from Gongan County (30°03'N, 112°13'E) in Hubei Province (Central China area), two from Tengzhou City (35°03'N, 117°08'E) in Shandong Province (East China area), and one from Chongming County (31°36'N, 121°40'E), one from Jinshan District (30°49'N, 121°36′E) and two from Fengxian District (30°55′N, 121°22′E) in Shanghai City (East China area). The weather across four provincial areas are different. Guangdong has a tropical and sub-tropical climate (warm and humid all year round), Hubei and Shanghai belongs to the sub-tropical monsoonal climates (hot, humid summers and generally mild winters), and Shandong has a warm-temperate monsoonal climate (hot, rainy summers and cold, winters) (http://en.wikipedia.org/wiki/ Category:Geography_in_China_by_province). The flocks from all the farms were similar, and the goats were kept within fenced areas at daytime and in house with wooden, slatted floors or on the cement floor at night time. Most goats drank from water channels (source from water supply) and took native grasses. Six hundred and four samples were obtained directly from the rectums or immediately after the animals defecated using sterile gloves. The samples were sealed in bags, taken back to the laboratory and processed within a week. The goats involved in this study had not previously been examined for Cryptosporidium infection.

DNA extraction, PCR amplification and sequence analysis

Genomic DNA was extracted from 300 mg of each faecal sample. All the samples were washed twice with sterile water and further purified using FastDNA SPIN Kit for Soil and the FastPrep Instrument (MP Biomedicals, Santa Ana, CA) in accordance with the manufacturer's instructions. A two-step nested PCR of small subunit (SSU) rRNA gene was used to amplify the DNA fragment, as described by Xiao et al. [23,24]. Positive and negative control samples were used for each PCR reaction. All secondary PCR products were sequenced as previously described [25]. Sequence analysis was guided using BLAST (http://blast.ncbi.nlm.nih.gov/Blast.cgi), and Cryptospo-

ridium species/genotypes were identified preliminarily. A Neighbor-Joining (NJ) tree was constructed using/by MEGA 6 software (http://www.megasoftware.net/) to assess the genetic relationship between different *Cryptosporidium* species/genotype sequences [26]. The reference sequences of *Cryptosporidium* spp. were obtained from GenBank.

Subtype identification

For *C. parvum*, a ~400 bp fragment of the 60 kDa glycoprotein (gp60) gene was amplified by nested PCR as described by Sulaiman et al. [27]. For *C. ubiquitum*, a ~950 bp fragment of the gp60 gene was amplified by nested PCR as reported by Li et al. [28]. The second PCR products were purified using AxyPrep DNA Gel Extraction Kit (Axygen Scientific, Hangzhou, China) and sequenced as SSU rRNA gene. The *Cryptosporidium* subtypes were identified as described by Sulaiman et al. [27] and Li et al. [28].

Statistical analysis

Cryptosporidium spp. prevalence in different regions and ages was determined using IBM SPSS Statistics V21.0 for Windows (International Business Machines Corp, New York, USA). The differences were considered significant when p<0.05 by Pearson's Chi-Square test (χ^2 test) analysis.

Results

Cryptosporidium infection in different farms

The overall prevalence of *Cryptosporidium* in four provincial level districts was 11.4% (69/604) (Table 1). The highest prevalence was found in Shandong Province (18/100, 18%), a very different result than in Guangdong Province (5/91, 5.5%) ($\chi^2 = 7.03$, p < 0.05). High prevalence could also be found in Hubei Province (13/111, 11.7%) and Shanghai City (33/302, 10.9%) ($\chi^2 = 3.52$, p > 0.05). There was also no significant difference between Guangdong Province, Hubei Province, and Shanghai City ($\chi^2 = 2.67$, p > 0.05).

Eleven goat farms tested positively for *Cryptosporidium*. The highest infection rate was in Gongan-4 farm (25.0%, 9/36) of Hubei Province, while the Gongan-3 farm had no *Cryptosporidium* positive samples (Table 2). Among the eleven farms with *Cryptosporidium* infection, the infection rates ranged from 2.9% to 25.0% (Table 2).

Prevalence of Cryptosporidium species

All positive samples were sequenced and then searched using BLAST (http://blast.ncbi.nlm.nih.gov/Blast.cgi). Phylogenetic evolutionary analysis was determined using MEGA version 6 [26] (Fig. 1). Three *Cryptosporidium* species were detected,

Table 1. Prevalence and molecular characterization of Cryptosporidium spp. in different provincial areas.

			Species		
Province/City	No. Sample	No. Positive (%)	C. xiaoi (%)	C. parvum (%)	C. ubiquitum (%)
Guangdong	91	5(5.5)	3	0	2
Hubei	111	13(11.7)	10	3	0
Shandong	100	18(18.0)	18	0	0
Shanghai	302	33(10.9)	14	11	8
Total	604	69(11.4)	45(65.2)	14(20.3)	10(14.5)

doi:10.1371/journal.pone.0111164.t001

 Table 2.
 Prevalence and molecular characterization of Cryptosporidium spp. in different farms.

				Species			Gp60 Subtypes	
Province/City	Farm	No. Sample	No. Positive (%)	C. xiaoi (%)	C. parvum (%)	C. ubiquitum (%)	C. parvum	C. ubiquitum
Guangdong	Huizhou-1	35	1(2.9)	-	0	0		
	Huizhou-2	56	4(7.1)	2	0	2		XIIa(2)
Hubei	Gongan-1	19	2(10.5)	0	2	0	llaA14G2R1(1) Unknown(1)	
	Gongan-2	33	2(6.1)	2	0	0		
	Gongan-3	23	0	0	0	0		
	Gongan-4	36	9(25.0)	80	-	0	llaA15G1R1(1)	
Shandong	Tengzhou-1	50	10(20.0)	10	0	0		
	Tengzhou-2	50	8(16.0)	8	0	0		
Shanghai	Chongming	155	18(11.6)	8	8	2	IIdA19G1(8)	XIIa(2)
	Fengxian-1	55	7(12.7)	-	0	9		XIIa(6)
	Fengxian-2	22	3(13.6)	8	0	0		
	Jinshan	70	5(7.1)	5	m	0	llaA17G2R1(1) llaA15G2R1(1) Unknown(1)	
Total		604	69(11.4)	45(65.2)	14(20.3)	10(14.5)		

doi:10.1371/journal.pone.0111164.t002

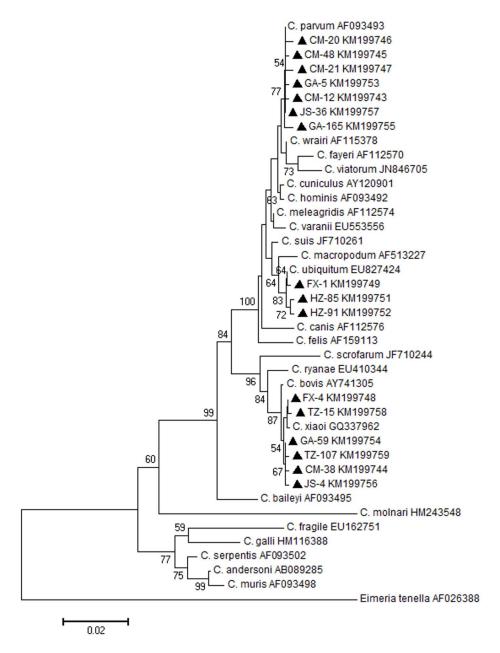


Figure 1. Phylogenetic tree of *Cryptosporidium* spp. constructed based on partial SSU rRNA nucleotide sequence. The phylogenetic tree was constructed using the neighbor-joining algorithm of the phylogeny program of MEGA 6.0. Bootstrap method via 1000 pseudo replicates was used to assess the reliability of the tree, and bootstrap value more than 50% were shown. ▲: Partial sequences obtained in the present study; CM: The sample from Chongming County; FX: The sample from Fengxian District; GA: The sample from Gongan County; HZ: The sample from Huizhou City; JS: The sample from Jinshan District; TZ: The sample from Tengzhou City. doi:10.1371/journal.pone.0111164.g001

including C. xiaoi (45/69, 65.2%), C. parrum (14/69, 20.3%) and C. ubiquitum (10/69, 14.5%) (Table 2). They were found in 10 (Gongan-2, Gongan-4, Huizhou-1, Huizhou-2, Tengzhou-1, Tengzhou-2, Chongming, Fengxian-1, Fengxian-2 and Jinshan), 4 (Gongan-1, Gongan-4, Chongming and Jinshan), and 3 (Huizhou-2, Chongming and Fengxian-1) goat farms, respectively (Table 2). C. xiaoi was the most prevalent species in Gongan-4 (22.2%), while in Huizhou-2 and Chongming, C. xiaoi with C. ubiquitum and C. xiaoi with C. parvum were equally prevalent (Table 2). C. ubiquitum was the most prevalent species in Fengxian-1, and C. parvum was most prevalent in Gongan-1. Infections mixing two species were found in four farms (Gongan-4,

Huizhou-2, Fengxian-1 and Jinshan), and infection from all three species was found in Chongming. The unique partial SSU rRNA gene sequences of *Cryptosporidium* spp. were submitted to the GenBank database under accession numbers KM199743 to KM199759.

Prevalence of *Cryptosporidium* species in different goat age ranges

The infection rates of goats in four different age groups were investigated in the present study. The highest infection rate of *Cryptosporidium* spp. was observed in pre-weaned kids (11/55, 20.0%), followed by post weaned kids (34/227, 15%), yearling

Table 3. Prevalence and molecular characterization of *Cryptosporidium* spp. in different host age groups.

			Species		
Age	No. Sample	No. Positive (%)	C. xiaoi (%)	C. parvum (%)	C. ubiquitum (%)
Pre-weaned kids	55	11(20.0)	6	5	0
Post weaned kids	227	34(15.0)	22	4	8
Yearling goats	306	23(7.5)	17	5	1
Adult goats	16	1(6.3)	0	0	1
Total	604	69(11.4)	45(65.2)	14(20.3)	10(14.5)

doi:10.1371/journal.pone.0111164.t003

goats (23/306, 7.5%) and adult goats (1/16, 6.3%) (Table 3), with significant differences between each age groups ($\chi^2 = 11.9$, p < 0.05).

In addition to adult goats, *C. xiaoi* and *C. parvum* were found in three age groups. *C. ubiquitum* also found in three age groups except for pre-weaned kids. *C. xiaoi* (22/45, 48.9%) and *C. ubiquitum* (8/10, 80%) were mainly prevalence in post weaned kids, while *C. parvum* had a similar infection rates in three age groups (Table 3).

Prevalence of Cryptosporidium subtypes

Twelve (85.7%) *C. parvum* positive samples were subtyped successfully based on the gp60 gene. Two subtypes, IIa and IId, were found. Four *C. parvum* IIa subtypes were found in 4 samples, including an IIaA14G2R1 from Gongan-1 farm and an IIaA15G1R1 from Gongan-4 farm of Hubei Province, and an IIaA15G2R1 and an IIaA17G2R1 from Jinshan farm of Shanghai City (Table 2). One IId subtype (IIdA19G1) was found in 8 samples from Chongming farm of Shanghai City (Table 2). All *C. ubiquitum* positive samples (10/10) were subtyped successfully, and all of them belonged to subtype XIIa (Table 2). The various sequences we acquired in this study were put in the GenBank database under accession numbers KM199736 to KM199742.

Discussion

Cryptosporidium infection in goats has been reported all over the world in the past ten years, including in Belgium [5], Brazil [29], Cyprus [30], Egypt [6], England and Wales [31], France [15,18], Greece [21], India [8], Italy [9], Malawi [32], Norway [10], Papua New Guinea [22], Spain [16,33,34], the Republic of Korea [19], Turkey [35], and Zambia [14]. However, most of these studies used microscopic examinations, immunofluorescence tests, or modified Ziehl-Neelsen staining techniques. Few reports used molecular methods, and the features of Cryptosporidium species/genotypes were still unclear.

In this study, the prevalence and molecular characterization of *Cryptosporidium* in goats across four provincial level areas in China were detected with nested PCR technique. Eleven goat farms had *Cryptosporidium* infection, and the infection rates ranged from 2.9% (1/35) to 25.0% (9/36). Only one farm (Gongan-3) from Hubei Province was not found *Cryptosporidium* infection. We found different provinces had different infection rates, and the highest prevalence was detected in Shandong Province (18/100, 18.0%), followed by Hubei Province (13/111, 11.7%), Shanghai City (33/302, 10.9%), and Guangdong Province (5/91, 5.5%). The total prevalence of *Cryptosporidium* spp. in goats from 12 goat farms was 11.4% (69/604), which was higher than the 4.4% (10/228) infection rate in Papua New

Guinea used the same detection method [22], and lower than the 17.9% (980/5468) infection rate in goats summarized by Wang et al [20]. In China, partial provinces reported *Cryptosporidium* infection in goats, but the reported infection rates varied greatly. For instance, the infection rate was 35.7% (15/42) by immunofluorescence test (IFT) and 4.8% (2/42) by PCR approach in Wulan County of Qinghai Province [17]. The infection rates were 27.5% (14/51) by IFT and 15.7% (8/51) by PCR in Qinghai Province [36]. A recent survey reported the percentage of *Cryptosporidium* infection was 2.75% (28/1017) in Henan Province and 6.45% (16/248) in Chongqing City by microscopical detection [20]. The difference in infection rates between our survey and previous surveys may be affected by host age, examination methods, feeding levels and the raising density of animals.

All positive samples were sequenced successfully, and three species were identified in this study. *C. xiaoi* was the most common species (45/69, 65.2%), followed by *C. parvum* (14/69, 20.3%) and *C. ubiquitum* (10/69, 14.5%). The results were similar to a recent study in Greece, in which same *Cryptosporidium* species were observed in goat kids [21]. The *Cryptosporidium* species presented differently depending on geographical location. *C. xiaoi* by itself was found in Shandong Province, *C. xiaoi* and *C. parvum* mixed were found in Hubei Province, *C. xiaoi* and *C. ubiquitum* mixed were found in Guangdong Province. All three species were found in Shanghai City. These results agree with previous surveys in China, which detected *C. andersoni*, *C. ubiquitum* and *C. xiaoi* in Henan Province, *C. ubiquitum* in Chongqing City [20], and *C. xiaoi* and a new genotype in Qinghai Province [17], but we did not find *C. andersoni* in goats for this survey.

C. xiaoi (previously identified as Cryptosporidium bovis-like genotype) was first reported in a goat in Qinghai Province of China [17] and subsequently identified in countries like France, Norway, Papua New Guinea, and Spain [10,15,16,22]. Recently, C. xiaoi was also found in Henan Province of China [20]. In this study, C. xiaoi as the dominant species was found in all the detected provincial areas and most of the detected goat farms. A previous French study found that C. xiaoi was the most common species in pre-weaned kids [15], and a recent study also reported a high prevalence of C. xiaoi infection in goat kids in Greece [21]. In China, C. xiaoi was also detected in sheep [37,38] and a yearling yak [39]. A recent study reported that C. xiaoi was identified in two HIV/AIDS patients in Ethiopia [40], suggesting that it pose a potential threat to human health.

Most previous surveys found that *C. parvum* was the dominant species in goats [5,8,9,11]. In this study, *C. parvum* was the second most prevalent species in goats. This agrees with a recent report from France in which *C. xiaoi* had a higher prevalence rate than *C. parvum* [15], and in Papua New Guinea where *C. hominis* was

more prevalent than *C. parvum* [22]. To our knowledge, this was the first molecular identification of *C. parvum* infection in goats in China

C. ubiquitum (formerly known as Cryptosporidium cervine genotype) as a zoonotic species has been reported in humans worldwide and in many animals, including ruminants, primates, and rodents, and has also been found in water [1,27,41,42]. Few studies report it in goats. A recent survey in China have reported that C. ubiquitum was the predominant species in Chongqing City [20]. By contrast, we found C. ubiquitum was the least frequently detected species in goats. C. ubiquitum was found in only three goat farms (Huizhou-2, Chongming, and Fengxian-1). Interestingly, in Fengxian-1 farm of Shanghai City, C. ubiquitum was the dominant species.

Our survey, and previous surveys, revealed that young animals were more susceptible to infection than adult animals [29,43]. Four age groups in goats were identified in this study, and the infection rate was decreased significantly with the increase of host age. All three species were found in post weaned kids and yearling goats, and C. xiaoi and C. parvum were found in pre-weaned kids, while C. ubiquitum as a unique species was identified in an adult goat. C. xiaoi was the major species, followed by C. parvum and C. ubiquitum, and these results were different than in previous studies. Most previous reports found C. parvum as the dominant species in goat kids [5.8,10,11,14]. Other studies in France and Spain listed C. xiaoi as the most common species in goat kids [15,16], and C. ubiquitum as the unique species was found in periparturient goats in one flock in western France [18]. However, a recent study found that C. parvum and C. xiaoi existed in adult goats in Papua New Guinea [22], and C. ubiquitum and C. xiaoi were found in goats of all ages in China [20]. In this study, only one positive sample was detected in adult goats, it may be attributed to the small number of fecal samples. In future research, larger numbers of fresh samples of adult goats in these areas will be collected and detected to understand the relationship between the host age and Cryptosporidium species infection.

To date, only a few studies analyzed C. parvum subtypes in goats, and two subtype families (IIa and IId) were reported. The Ha family has been found in Belgium, Norway, and Papua New Guinea [5,10,22], and the IId family was only found in Europe (Belgium, Greece, Italy and Spain) [5,9,11,21]. Similar to previous studies, the IIa and IId families were also found in China in this survey, and IId subtype family (8/12, 66.7%) was the dominant subtype in goats. This agrees with previous surveys that said goats were more susceptible to infection C. parvum IId subtypes in Belgium (8/11, 72.7%), Greece (2/2, 100%) and Spain (17/17, 100%) [5,11,21]. The distribution of C. parvum subtypes had geographic differences. Ha subtypes were found in the Gongan-1 (IIaA14G2R1) and Gongan-4 (IIaA15G1R1) goat farms of Hubei Province and in Jinshan farm (IIaA15G2R1 and IIaA17G2R1) of Shanghai City. The subtype IId was identified only in Chongming farm (IIdA19G1) of Shanghai City. The subtype IIdA19G1 had been detected in goats in previous studies from Spain [11] and in HIV-positive patients in China [44] and Portugal [45]. IIdA19G1 also had been found in calves in Henan Province [46], but had never been found in goats in China. Subtype IIaA15G2R1 was the

References

- Fayer R (2010) Taxonomy and species delimitation in Cryptosporidium. Exp. Parasitol 124: 90–97.
- Santín M (2013) Clinical and subclinical infections with Cryptosporidium in animals. N Z Vet I 61: 1–10.
- Xiao L (2010) Molecular epidemiology of cryptosporidiosis: an update. Exp Parasitol 124: 80–89.

most common subtype in calves and humans [3,47]. It had been found in goats in Belgium [5] and Papua New Guinea [22]. In China, this subtype was also detected in lambs and yaks [38,48] but not goats. Subtype IIaA14G2R1 was mainly found in calves in Europe, including in Belgium [49], England [50], Germany [51], and the Netherlands [52]. It was also found in humans in Ethiopia [40]. In China, this subtype had been found in yaks [48]. Other subtypes, IIaA15G1R1 and IIaA17G2R1, have been reported in humans in many previous studies. For example, subtype IIaA15G1R1 had been found in humans in Australia [53], Egypt [54], Kuwait [26], and Slovenia [55]. Subtype IIaA17G2R1 had been reported in Australia [53,56,57], Canada [58], Ethiopia [40], the United Kingdom [59], and the United States [60]. The three Ha subtypes detected in this study, including HaA14G2R1, IIaA15G1R1 and IIaA17G2R1, are the first reported in goats. Taken together, all the C. parvum subtypes have been detected previously in humans, suggesting that goats may be involved in zoonotic transmission of cryptosporidiosis.

Recent research about *C. ubiquitum* subtyping that targeted the gp60 gene was reported by Li et al. [28], and 6 subtype families (XIIa-XIIf) were identified. In this study, all the *C. ubiquitum* positive samples were identified as XIIa, in accordance with a recent survey that asserted XIIa was the unique subtype in goats in Henan Province and Chongqing City [20]. This subtype was also found in a goat from Algeria [28]. A recent study found *C. ubiquitum* subtype XIIa not only existed in domestic and wild ruminants, but also commonly seen in humans [28], which suggests this subtype may be a potential source of *C. ubiquitum* infection between animals and humans.

In summary, the prevalence and molecular characterizations of *Cryptosporidium* species and subtypes in goats from 12 farms across four different provincial level regions of China indicate that *Cryptosporidium* spp. are common infections in goats in China. The overall infection rate was 11.4% and the highest infection rate was found in pre-weaned kids. *C. xiaoi*, five subtypes of *C. parvum*, and one subtype of *C. ubiquitum* were found in goats. To our knowledge, this was the first report of *C. parvum* subtypes IIaA14G2R1, IIaA15G1R1 and IIaA17G2R1 infecting in goats and the first molecular identification of *C. parvum* and its subtypes in Chinese goats. The zoonotic *C. parvum* subtype IIdA19G1 and *C. ubiquitum* subtype XIIa were the dominant subtypes in the present study, suggesting that monitoring goat populations for the presence of *Cryptosporidium* is important to public health.

Acknowledgments

We thank Ke Lu at Shanghai Veterinary Research Institute and Yanfu Xia at Animal Disease Control and Prevention Center of Huiyang District for their help in sample collecting.

Author Contributions

Conceived and designed the experiments: ZC. Performed the experiments: RM XW YL. Analyzed the data: RM XW PZ YH ZC. Contributed reagents/materials/analysis tools: YC JC WZ. Contributed to the writing of the manuscript: RM ZC. Approved the final version of the manuscript: YH PZ XW YL.

- Mason RW, Hartley WJ, Tilt L (1981) Intestinal cryptosporidiosis in a kid goat. Aust Vet J 57: 386–388.
- Geurden T, Thomas P, Casaert S, Vercruysse J, Claerebout E (2008) Prevalence and molecular characterisation of *Cryptosporidium* and *Giardia* in lambs and goat kids in Belgium. Vet Parasitol 155: 142–145.

- Shoukry NM, Dawoud HA, Haridy FM (2009) Studies on zoonotic cryptosporidiosis parvum in Ismailia Governorate, Egypt. J Egypt Soc Parasitol 30: 470-488
- Ngouanesavanh T, Guyot K, Certad G, Le Fichoux Y, Chartier C, et al. (2006) Cryptosporidium population genetics: evidence of clonality in isolates from France and Haiti. J Eukaryot Microbiol 53 Suppl 1: S33–S36.
- Maurya PS, Rakesh RL, Pradeep B, Kumar S, Kundu K, et al. (2013) Prevalence and risk factors associated with *Cryptosporidium* spp. infection in young domestic livestock in India. Trop Anim Health Prod 45: 941–946.
- Drumo R, Widmer G, Morrison LJ, Tait A, Grelloni V, et al. (2012) Evidence of host-associated populations of *Cryptosporidium parvum* in Italy. Appl Environ Microbiol 78: 3523–3529.
- Lange H, Johansen OH, Vold L, Robertson LJ, Anthonisen IL, et al. (2013) Second outbreak of infection with a rare Cryptosporidium parvum genotype in schoolchildren associated with contact with lambs/goat kids at a holiday farm in Norway. Epidemiol Infect DOI: http://dx.doi.org/10.1017/ S0950268813003002.
- Quílez J, Torres E, Chalmers RM, Hadfield SJ, Del CE, et al. (2008) Cryptosporidium genotypes and subtypes in lambs and goat kids in Spain. Appl Environ Microbiol 74: 6026–6031.
- Noordeen F, Horadagoda NU, Faizal AC, Rajapakse RP, Razak MA, et al. (2002) Infectivity of *Cryptosporidium parvum* isolated from asymptomatic adult goats to mice and goat kids. Vet Parasitol 103: 217–225.
- goats to mice and goat kids. Vet Parasitol 103: 217–225.

 13. Hajdušek O, Ditrich O, Šlapeta J (2004) Molecular identification of *Cryptosporidium* spp. in animal and human hosts from the Czech Republic. Vet Parasitol 122: 183–192.
- Goma FY, Geurden T, Siwila J, Phiri IGK, Gabriel S, et al. (2007) The prevalence and molecular characterisation of *Cryptosporidium* spp. in small ruminants in Zambia. Small Ruminant Res 72: 77–80.
- Rieux A, Paraud C, Pors I, Chartier C (2013) Molecular characterization of Cryptosporidium spp. in pre-weaned kids in a dairy goat farm in western France. Vet Parasitol 192: 268–272.
- Diaz P, Quilez J, Robinson G, Chalmers RM, Diez-Baños P, et al. (2010) Identification of Cryptosporidium xiaoi in diarrhoeic goat kids (Capra hircus) in Spain. Vet Parasitol. 172: 132–134.
- Karanis P, Plutzer J, Halim NA, Igori K, Nagasawa H, et al. (2007) Molecular characterization of *Cryptosporidium* from animal sources in Qinghai province of China. Parasitol Res 101: 1575–1580.
- Paraud C, Pors I, Rieux A, Brunet S (2014) High excretion of Cryptosporidium ubiquitum by peri-parturient goats in one flock in western France. Vet Parasitol 202: 301–304.
- Park JH, Guk SM, Han ET, Shin EH, Kim JL, et al. (2006) Genotype analysis of Cryptosporidium spp. prevalent in a rural village in Hwasun-gun, Republic of Korea. Korean J Parasitol 44: 27–33.
- Wang R, Li G, Cui B, Huang J, Cui Z, et al. (2014) Prevalence, molecular characterization and zoonotic potential of *Cryptosporidium* spp. in goats in Henan and Chongqing, China. Exp Parasitol 142: 11–16.
- Tzanidakis N, Sotiraki S, Claerebout E, Ehsan A, Voutzourakis N, et al. (2014)
 Occurrence and molecular characterization of Giardia duodenalis and Cryptosporidium spp. in sheep and goats reared under dairy husbandry systems in Greece. Parasite 21: 45.
- Koinari M, Lymbery AJ, Ryan UM (2014) Cryptosporidium species in sheep and goats from Papua New Guinea. Exp Parasitol 141: 134–137.
- Xiao L, Morgan UM, Limor J, Escalante A, Arrowood M, et al. (1999) Genetic diversity within Cryptosporidium parvum and related Cryptosporidium species. Appl Environ Microbiol 65: 3386–3391.
- Xiao L, Bern C, Limor J, Sulaiman I, Roberts J, et al. (2001) Identification of 5 types of *Cryptosporidium* parasites in children in Lima, Peru. J Infect Dis 183: 492–497.
- 25. Chen Z, Mi R, Yu H, Shi Y, Huang Y, et al. (2011) Prevalence of *cryptosporidium* spp. in pigs in Shanghai, China. Vet Parasitol 181: 113–119.
- Tamura K, Stecher G, Peterson D, Filipski A, Kumar S (2013) MEGA6: Molecular Evolutionary Genetics Analysis version 6.0. Mol Biol Evol 30: 2725–2729.
- Sulaiman IM, Hira PR, Zhou L, Al-Ali FM, Al-Shelahi FA, et al. (2005) Unique endemicity of cryptosporidiosis in children in Kuwait. J Clin Microbiol 43: 2805–2809.
- Li N, Xiao L, Alderisio K, Elwin K, Cebelinski E, et al. (2014) Subtyping Cryptosporidium ubiquitum, a zoonotic pathogen emerging in humans. Emerg Infect Dis 20: 217–224.
- Bomfim TC, Huber F, Gomes RS, Alves LL (2005) Natural infection by *Giardia* sp. and *Cryptosporidium* sp. in dairy goats, associated with possible risk factors of the studied properties. Vet Parasitol 134: 9–13.
- Giadinis ND, Symeoudakis S, Papadopoulos E, Lafi SQ, Karatzias H (2012)
 Comparison of two techniques for diagnosis of cryptosporidiosis in diarrhoeic goat kids and lambs in Cyprus. Trop Anim Health Prod 44: 1561–1565.
- Smith RP, Chalmers RM, Mueller-Doblies D, Clifton-Hadley FA, Elwin K, et al. (2010) Investigation of farms linked to human patients with cryptosporidiosis in England and Wales. Prev Vet Med 94: 9–17.
- Banda Z, Nichols RA, Grimason AM, Smith HV (2009) Cryptosporidium infection in non-human hosts in Malawi. Onderstepoort J Vet Res 76: 363–375.
- 33. Castro-Hermida JA, Almeida A, González-Warleta M, Correia Da Costa JM, Rumbo-Lorenzo C, et al. (2007) Occurrence of *Cryptosporidium parvum* and

- $\it Giardia~duodenalis$ in healthy adult domestic ruminants. Parasitol Res 101: 1443–1448.
- Sanz Ceballos L, Illescas Gómez P, Sanz Sampelayo MR, Gil Extremera F, Rodríguez Osorio M (2009) Prevalence of *Cryptosporidium* infection in goats maintained under semi-extensive feeding conditions in the southeast of Spain. Parasite 16: 315–318.
- Çiçek M, Körkoca H, Gül A (2008) Investigation of Cryptosporidium sp. in workers of the Van municipality slaughterhouse and in slaughtered animals. Turkiye Parazitol Derg 32: 8–11.
- Ma L, Sotiriadou I, Čai Q, Karanis G, Wang G, et al. (2014) Detection of Cryptosporidium and Giardia in agricultural and water environments in the Qinghai area of China by IFT and PCR. Parasitol Res.
- Wang Y, Feng Y, Cui B, Jian F, Ning C, et al. (2010) Cervine genotype is the major *Cryptosporidium* genotype in sheep in China. Parasitol Res 106: 341–347.
- Ye J, Xiao L, Wang Y, Wang L, Amer S, et al. (2013) Periparturient transmission of *Cryptosporidium xiaoi* from ewes to lambs. Vet Parasitol 197: 627–633.
- Ma J, Cai J, Ma J, Feng Y, Xiao L (2014) Occurrence and molecular characterization of *Cryptosporidium* spp. in yaks (*Bos grunniens*) in China. Vet Parasitol 202: 113–118.
- Adamu H, Petros B, Zhang G, Kassa H, Amer S, et al. (2014) Distribution and Clinical Manifestations of *Cryptosporidium* Species and Subtypes in HIV/AIDS Patients in Ethiopia. PLoS Negl Trop Dis 8: e2831.
- Feng Y, Alderisio KA, Yang W, Blancero LA, Kuhne WG, et al. (2007) Cryptosporidium genotypes in wildlife from a New York watershed. Appl Environ Microbiol 73: 6475–6483.
- Galván AL, Magnet A, Izquierdo F, Fernández Vadillo C, Peralta RH, et al. (2014) A year-long study of *Cryptosporidium* species and subtypes in recreational, drinking and wastewater from the central area of Spain. Sci Total Environ 468– 469: 368–375.
- Noordeen F, Faizal AC, Rajapakse RP, Horadagoda NU, Arulkanthan A (2001) Excretion of *Cryptosporidium* oocysts by goats in relation to age and season in the dry zone of Sri Lanka. Vet Parasitol 99: 79–85.
- Wang L, Zhang H, Zhao X, Zhang L, Zhang G, et al. (2013) Zoonotic Cryptosporidium species and Enterocytozoon bieneusi genotypes in HIV-positive patients on antiretroviral therapy. J Clin Microbiol 51: 557–563.
- Alves M, Xiao L, Antunes F, Matos O (2006) Distribution of Cryptosporidium subtypes in humans and domestic and wild ruminants in Portugal. Parasitol Res 99: 287–292.
- Wang R, Wang H, Sun Y, Zhang L, Jian F, et al. (2011) Characteristics of Cryptosporidium transmission in preweaned dairy cattle in Henan, China. J Clin Microbiol 49: 1077–1082.
- Feng Y, Torres E, Li N, Wang L, Bowman D, et al. (2013) Population genetic characterisation of dominant *Cryptosporidium parvum* subtype IIaA15G2R1. Int J Parasitol 43: 1141–1147.
- Mi R, Wang X, Li C, Huang Y, Zhou P, et al. (2013) Prevalence and genetic characterization of *Cryptosporidium* in yaks in Qinghai Province of China. PLoS One 8: e74985.
- Geurden T, Berkvens D, Martens C, Casaert S, Vercruysse J, et al. (2007) Molecular epidemiology with subtype analysis of *Cryptosporidium* in calves in Belgium. Parasitology 134: 1981–1987.
- Brook EJ, Anthony HC, French NP, Christley RM (2009) Molecular epidemiology of *Cryptosporidium* subtypes in cattle in England. Vet J 179: 378–382.
- Broglia A, Reckinger S, Cacció SM, Nöckler K (2008) Distribution of Cryptosporidium parvum subtypes in calves in Germany. Vet Parasitol 154: 8– 13.
- Wielinga PR, de Vries A, van der Goot TH, Mank T, Mars MH, et al. (2008) Molecular epidemiology of *Cryptosporidium* in humans and cattle in The Netherlands. Int J Parasitol 38: 809–817.
- Waldron LS, Dimeski B, Beggs PJ, Ferrari BC, Power ML (2011) Molecular epidemiology, spatiotemporal analysis, and ecology of sporadic human cryptosporidiosis in Australia. Appl Environ Microbiol 77: 7757–7765.
- 54. Helmy YA, Krücken J, Nöckler K, von Samson-Himmelstjerna G, Zessin KH (2013) Molecular epidemiology of *Cryptosporidium* in livestock animals and humans in the Ismailia province of Egypt. Vet Parasitol 193: 15–24.
- Soba B, Logar J (2008) Genetic classification of Cryptosporidium isolates from humans and calves in Slovenia. Parasitology 135: 1263–1270.
- Waldron LS, Ferrari BC, Power ML (2009) Glycoprotein 60 diversity in C. hominis and C. parvum causing human cryptosporidiosis in NSW, Australia. Exp Parasitol 122: 124–127.
- Ng J, MacKenzie B, Ryan U (2010) Longitudinal multi-locus molecular characterisation of sporadic Australian human clinical cases of cryptosporidiosis from 2005 to 2008. Exp Parasitol 125: 348–356.
- Trotz-Williams LA, Martin DS, Gatei W, Cama V, Peregrine AS, et al. (2006) Genotype and subtype analyses of *Cryptosporidium* isolates from dairy calves and humans in Ontario. Parasitol Res 99: 346–352.
- Chalmers RM, Smith RP, Hadfield SJ, Elwin K, Giles M (2011) Zoonotic linkage and variation in *Cryptosporidium parvum* from patients in the United Kingdom. Parasitol Res 108: 1321–1325.
- Feltus DC, Giddings CW, Schneck BL, Monson T, Warshauer D, et al. (2006) Evidence supporting zoonotic transmission of *Cryptosporidium* spp. in Wisconsin. J Clin Microbiol 44: 4303–4308.