

Functional Fcγ Receptor Polymorphisms Are Associated with Human Allergy

Jianming Wu^{1,2*}, Rui Lin¹, Jinhai Huang¹, Weihua Guan³, William S. Oetting⁴, P. Sriramarao^{1,2}, Malcolm N. Blumenthal²

1 Department of Veterinary and Biomedical Sciences, University of Minnesota, St. Paul, Minnesota, United States of America, **2** Department of Medicine, University of Minnesota, Minneapolis, Minnesota, United States of America, **3** Department of Biostatistics, University of Minnesota, Minneapolis, Minnesota, United States of America, **4** Department of Experimental and Clinical Pharmacology, University of Minnesota, Minneapolis, Minnesota, United States of America

Abstract

Objective: IgG Fc receptors (FcγRs) play important roles in immune responses. It is not clear whether FcγR receptors play a role in human asthma and allergy. The aim of current study was to investigate whether functional single nucleotide polymorphisms (SNPs) of FcγR genes (*FCGR*) are associated with human asthma and allergy.

Methods: Functional SNPs of *FCGR2A* (FcγRIIA-131His>Arg, rs1801274), *FCGR2B* (FcγRIIB-187Ile>Thr, rs1050501), *FCGR2C* (FcγRIIC-13Gln>Stop, rs10917661), *FCGR3A* (FcγRIIIA-158Val>Phe, rs396991), and *FCGR3B* variants (FcγRIIB NA1 and NA2) were genotyped in an asthma family cohort including 370 atopy positive, 239 atopy negative, and 169 asthma positive subjects. The genotype and phenotype data (asthma, bronchial hyper-responsiveness, and atopy) of subjects were analyzed using family-based association tests (FBAT) and logistic regression adjusted for age and sex.

Result: The FcγRIIA-131His>Arg SNP is significantly associated with atopy in a family-based association test ($P=0.00287$) and in a logistic regression analysis ($P=0.0269$, OR 0.732, 95% CI: 0.555–0.965). The FcγRIIA-131His (or rs1801274-A) allele capable of binding human IgG2 has a protective role against atopy. In addition, the rare FcγRIIB-187Thr (or rs1050501-C) allele defective for the receptor-mediated inhibitory signals is a risk factor for atopy ($P=0.0031$, OR 1.758, 95% CI: 1.209–2.556) and IgE production ($P<0.001$). However, variants of activating FcγRIIIA (rs396991), and FcγRIIB (NA1 and NA2), and FcγRIIC (rs10917661) are not associated with asthma, BHR, and atopy ($P>0.05$).

Conclusions: FcγRIIA and FcγRIIB functional polymorphisms may have a role in the pathogenesis of allergy.

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* E-mail: jmwu@umn.edu

Introduction

Asthma is a complex syndrome characterized by airflow obstruction, bronchial hyper-responsiveness (BHR), and airway inflammation. Both genetic and environmental factors contribute to the development of asthma. Evidence for a genetic component in asthma includes familial clustering and higher concordance rates in monozygotic twins than in dizygotic twins [1,2]. Approximately 48–79% of asthma risk is attributable to genetic factors [1,2]. According to the American Academy of Allergy, Asthma and Immunology, half of the 20 million Americans with asthma have allergic asthma. Thus, allergic reactions to foreign antigens are considered as the most common causes for asthma. To date, no genes have been definitely shown to influence asthma/allergy development. It is well-known that IgE and its cognate receptor (FcεRI) are important mediators in allergic reactions [3]. However, the role of human IgG Fc receptors (FcγRs) in asthma and allergy remains unknown.

A recent meta-analysis of human genome-wide association study (GWAS) revealed a significant asthma susceptibility locus on chromosome 1q23, where FcγR (*FCGR*) genes are located [4]. Human FcγRs are glycoproteins that bind the Fc region of immunoglobulin G (IgG). FcγRs mediate a variety of immune functions such as antigen presentation, immune complex clearance, phagocytosis of pathogens, degranulations, ADCC, and cytokine production [5]. In humans, five genes (*FCGR2A*, *FCGR2B*, *FCGR2C*, *FCGR3A*, and *FCGR3B*) in the 1q23 chromosome region code for five classical low affinity Fcγ receptors (FcγRIIA, FcγRIIB, FcγRIIC, FcγRIIIA, and FcγRIIIB). Coordination between the activating FcγRs (FcγRIIA, FcγRIIC, FcγRIIIA, and FcγRIIIB) and the inhibitory FcγR (FcγRIIB) is crucial in balancing immune responses and determining the outcomes of local and systemic inflammations [6]. FcγRs have important roles in the pathogenesis of a variety of human inflammatory diseases [7]. Not surprisingly, functional polymorphisms of FcγR have robust effects on susceptibility and severity of inflammatory diseases as demonstrated in genetic association

studies by our group and others [8,9,10,11]. However, comprehensive genetic analysis of human *FCGR* genes in asthma/allergy patients has yet been performed. It remains unknown whether human Fc γ Rs play a role in the development of allergy.

Patients and Methods

Study Subjects

Genomic DNA was isolated from anti-coagulated peripheral blood of human subjects from 27 multigenerational families with multiple asthmatic members, which were originally recruited as part of the Collaborative Study on the Genetics of Asthma (CSGA) [12]. For the CSGA, asthma families were ascertained through two asthmatic siblings. Additional relatives in the families were then recruited either by extending the families through asthmatic relatives or by including no more than one unaffected relative to permit a lineage to incorporate other relatives with asthma. The inclusion criteria for each family consisted of each of the two asthmatic siblings having met the following criteria for the proband: (1) being at least 6 years of age; (2) having either bronchial hyper-responsiveness (BHR), defined as a fall from baseline FEV1 greater than 20% in one second after inhalation of 25 mg/ml or less of methacholine, or reversibility, defined as a 15% or greater increase from baseline FEV1 after inhaled bronchodilator (albuterol) for those with reduced baseline FEV1; (3) having the presence of two or more of the symptoms of coughing, wheezing and shortness of breath; (4) having less than three pack/years of cigarette smoking; and (5) having a physician's diagnosis of asthma with no conflicting pulmonary disease. All family members went through a standardized protocol consisting of an interviewer administered questionnaire, pulmonary function studies including a methacholine challenge and/or reversibility studies, blood drawing for serum IgE levels at a single time not during an acute exacerbation and skin prick testing using standardized allergens [12]. Additional details of the study design can be found in an earlier publication [12]. The 27 multigenerational Caucasian families were recruited in Minnesota as previously described [13]. These families had 169 asthmatic members, 347 who were not asthmatic and 129 for whom the diagnosis was unavailable [13]. Pulmonary function data were available on 619 individuals. The study (Title: Genetics of Asthma. Study Number: 920M05150) was approved by The Institutional Review Board of Human Study at the University Of Minnesota. The informed written consent was obtained from all participants recruited in this study. The written consents containing participants' signatures were kept in locked file cabinets for record. The traits of asthma, BHR, atopy, and IgE levels were analyzed in the current genetic study.

Genotyping of *FCGR* SNPs

FCGR family member genes were generated through duplication and divergence during evolution [14]. SNPs in five *FCGR* genes are not suitable for direct TaqMan assays due to near 100% sequence identity surrounding the functional SNPs between homologous genes. Consequently, we used a modified *FCGR* SNP TaqMan assay in which *FCGR* gene-specific PCR fragments were used as templates instead of genomic DNA for TaqMan assays. The genomic DNA fragments containing functional SNPs of *FCGR2A* and *FCGR3A* were amplified using the gene specific primers as described previously [9]. For the *FCGR2B* SNP, a genomic DNA fragment containing Fc γ RIIB-187Ile>Thr was amplified using the gene specific primers as described [8,15]. To genotype Fc γ RIIC-13Gln>STP, a long *FCGR2C* genomic fragment (6227 bps) containing the SNP was amplified using Platinum

Taq DNA Polymerase High Fidelity (Invitrogen) with a sense primer (5'-CTG CAT ATG TTG TCC CCC TGT GTT GCT AAA T-3') annealing to the *FCGR2C* intron 2 and an antisense primer (5'-AAC ATG AGA GAG AAA AAG AGA GGC AGG GAG GGA GCT TA-3') annealing to the *FCGR2C* intron 6. The TaqMan assays for *FCGR2A* SNP (Fc γ RIIA-131His>Arg), *FCGR2B* SNP (Fc γ RIIB-187Ile>Thr), *FCGR2C* SNP (Fc γ RIIC-13Gln>STP), and *FCGR3A* SNP (Fc γ RIIA-158Val>Phe) were designed using the Software Primer Express v3.0 (Applied Biosystems Inc.). TaqMan genotyping assays were carried out according to the standard protocol on an ABI 7500 Real-Time PCR System using Genotyping Master Mix (Applied Biosystems). The primers and probes used in *FCGR* TaqMan genotyping assays are listed in Table 1. Genotyping of the respective SNPs of *FCGR2A*, *FCGR2B*, *FCGR2C*, and *FCGR3A* was carried out with four independent TaqMan allele discrimination assays that were developed and validated in the lab. The specificity and accuracy of individual TaqMan assays were validated by the perfect match (100%) with at least 300 genotyped human subjects published previously [8,9,15]. For *FCGR3B* allele determination, a primer pair that specifically amplifies the *FCGR3B* fragment containing *FCGR3B* coding SNPs (cSNPs) was used. The 1.6 kb *FCGR3B* PCR fragment was treated with ExoSAP-IT PCR Product Cleanup reagent (Affymetrix) before being sequenced on an ABI 3730xl DNA Analyzer with BigDye Terminator kit (Applied Biosystems) with the sequencing primer (5'-TCC TCA CCC CAC ATT ATC TTG-3'). The *FCGR3B* alleles and genotypes were determined based on the published reference [16,17].

Statistical Analysis

The IgE levels were log-transformed to correct for skewed distribution. Family-based association tests (FBAT) [18] were used to examine whether individual *FCGR* SNPs are associated with phenotypes of human subjects in the asthma family cohort. Alternatively, we used conditional logistic regression to estimate odds ratios of *FCGR* SNPs for their association with asthma, BHR, and atopy, adjusting for age and sex. The association between log-transformed IgE levels and *FCGR* genotypes were analyzed using one-way analysis of variance (ANOVA) in addition to the nonparametric t-test (Mann-Whitney test). In both FBAT and regression analysis, an additive model was assumed for SNP genotypes. To correct for multiple hypothesis tests, the Bonferroni method was used and the null hypothesis was reject at 0.05/number of tests.

Results

The Fc γ RIIA SNP is Associated with Atopy

As shown in Table 2, the *FCGR2A* SNP (Fc γ RIIA-131His>Arg, rs1801274) is significantly associated with atopy in the family-based association test (FBAT) ($P=0.003$). The *FCGR2A* SNP is also associated with asthma and BHR in FBAT ($P<0.05$). Conditional logistic regression analysis estimated an OR of 0.732 ($P=0.027$, 95% CI: 0.555–0.965) for *FCGR2A* SNP with atopy, suggesting a protective role against atopy for carriers of the Fc γ RIIA-131His allele (population allele frequency = 0.488). Although the *FCGR2A* SNP is significantly associated with asthma and BHR in FBAT ($P<0.05$), the association were not significant in logistic regression analyses adjusted for age and sex. Further validation may be needed to confirm our findings. Furthermore, the functional SNPs of the other three activating Fc γ Rs (Fc γ RIIA, Fc γ RIIB, and Fc γ RIIC) were not associated with asthma, BHR, and atopy ($P>0.05$) (Table 3).

Table 1. Primers and probes of TaqMan *FCGR* gene SNP assays.

Gene (SNP)	Gene-specific primers (5' to 3')	TaqMan Primers and Probes (5' to 3')
<i>FCGR2A</i> (rs1801274)	TGCCATAAGAGAATGCTCACA	CCAGAATGGAAAATCCCAGAAA
	TCAAAGTGAACAACAGCCTGACT	TTTGCTGTGGGATGGAGAAG
		FAM-TCTCCC <i><u>A</u></i> TTTGGATC Vic-TCTCCC <i><u>A</u></i> TTTGGATCC
<i>FCGR2B</i> (rs1050501)	CTAAGAGGAGCCCTCCCTATGT	CCCTAGTCCCAGCTCTTCA
	AATACGGGCTAGATCTGAATGTG	TGCAGTAGATCAAGGCCACTACA
		FAM-TCACTGGGA <i><u>T</u></i> CGT Vic-CACTGGGA <i><u>T</u></i> CGT
<i>FCGR2C</i> (rs10917661)	CTGCATATGTTGCCCTGTGTGCTAAAT	TCAGCAGTCCCCAAAG
	AACATGAGAGAGAAAAAGAGAGGCAGGG-	CGGCATGTCAGAGTACAGAGT
	AGGGAGCTTA	FAM-AAACTCGAGCCC <i><u>A</u></i> GTG Vic-CTCGAGCCC <i><u>A</u></i> GTGG
<i>FCGR3A</i> (rs396991)	CTGGTGTTCACATTGAGTTCTC	AAGACAGCGGCTCCTACTTCTG
	CTGATTCTGGAGGCTGTTCTACA	GTTACAGTCTCTGAAGACACATTTT
		FAM-AGGGGGCTT <i><u>T</u></i> T Vic-AGGGGGCTT <i><u>T</u></i> TG

Italic and underlined nucleotides are SNP sites in respective *FCGR* genes.
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The Inhibitory FcγRIIB SNP is Associated with Atopy and IgE Production

Although the *FCGR2B* SNP (FcγRIIB-187Ile>Thr, rs1050501) is not associated with asthma, BHR, and atopy in FBAT analyses, conditional logistic regression analyses showed that *FCGR2B* SNP is significantly associated with atopy and that the FcγRIIB-187Thr (allele frequency = 0.088) is a risk allele for atopy ($P=0.003$, OR 1.758, 95% CI: 1.209–2.556) (Table 2). Because immunoglobulin E (IgE) play an important role in allergic diseases and elevated

total IgE is frequently considered as a diagnostic criterion for allergic diseases [3], we subsequently analyzed whether the FcγRIIB SNP is associated with IgE levels in human subjects. As shown in Figure 1A, FcγRIIB genotypes are significantly associated with the serum IgE levels. The human subjects carrying rare FcγRIIB-Thr allele produced significantly more IgE ($P=0.0002$ for 187Ile/Thr heterozygous subjects and $P=0.0004$ for 187Thr/Thr homozygous subjects) than those homozygous (187Ile/Ile) subjects carrying the common allele, suggesting that the functional FcγRIIB SNP may have a role in allergy through IgE production. On the other hand, FcγRIIA SNP is not associated with IgE production in humans (Figure 1B).

Table 2. *FCGR2A* and *FCGR2B* SNPs are associated with atopy.

Genes/ Traits	FBAT		Logistic regression adjusted for age & sex	
	Z Score	P	P	OR (95% CI)
<i>FCGR2A</i>				
Asthma	2.542	0.011	0.187	1.229 (0.906–1.671)
BHR	2.498	0.012	0.207	1.214 (0.898–1.642)
Atopy	2.981	0.003	0.027	0.732 (0.555–0.965)
<i>FCGR2B</i>				
Asthma	0.692	0.489	0.476	0.870 (0.594–1.275)
BHR	0.822	0.411	0.906	0.978 (0.676–1.410)
Atopy	0.341	0.733	0.003	1.758 (1.209–2.556)

FCGR2A SNP (FcγRIIA-131His>Arg, rs1801274) is significantly associated with atopy in family-based association tests (FBAT). Logistic regression analysis also demonstrated that *FCGR2A* SNP is significantly associated with atopy and that the FcγRIIA-131His (allele frequency: 0.488) is a protective allele against atopy ($P=0.027$, OR 0.732, 95%CI: 0.555–0.965). The *FCGR2A* SNP is also associated with asthma ($P=0.011$) and BHR ($P=0.012$) in FBAT. The *FCGR2B* SNP (FcγRIIB-187Ile>Thr, rs1050501) is significantly associated with atopy ($P=0.003$, OR 1.758, 95%CI: 1.209–2.556) in logistic regression analyses adjusted for age and sex. The *FCGR2B* SNP is not associated with asthma and BHR ($P>0.05$).

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Discussion

This study shows the association of two functional SNPs (the activating FcγRIIA-131His>Arg and the inhibitory FcγRIIB-187Ile>Thr) with human atopy. Furthermore, we demonstrated an association between FcγRIIB SNP and IgE production. Our

Table 3. Functional SNPs of *FCGR3A*, *FCGR3B*, and *FCGR2C* are not associated with asthma, BHR, and atopy.

Gene	MAF	Asthma		BHR		Atopy	
		Z	P	Z	P	Z	P
<i>FCGR3A</i>	0.378	0.359	0.7194	0.618	0.5363	0.303	0.7615
<i>FCGR3B</i>	0.362	0.984	0.3253	1.129	0.2590	1.500	0.1336
<i>FCGR2C</i>	0.157	0.159	0.8740	0.141	0.8875	0.563	0.5734

SNPs of *FCGR3A* SNP (FcγRIIA-158Val>Phe, rs396991), *FCGR3B* allele (FcγRIIB-NA1/NA2), and *FCGR2C* SNP (FcγRIIC-13Gln>Stop, rs10917661) are not associated with asthma, BHR, and atopy in family-based association test (FBAT) analyses ($P>0.05$) and logistic regression analyses adjusted for age and sex ($P>0.05$, data not listed).

MAF: minor allele frequency.

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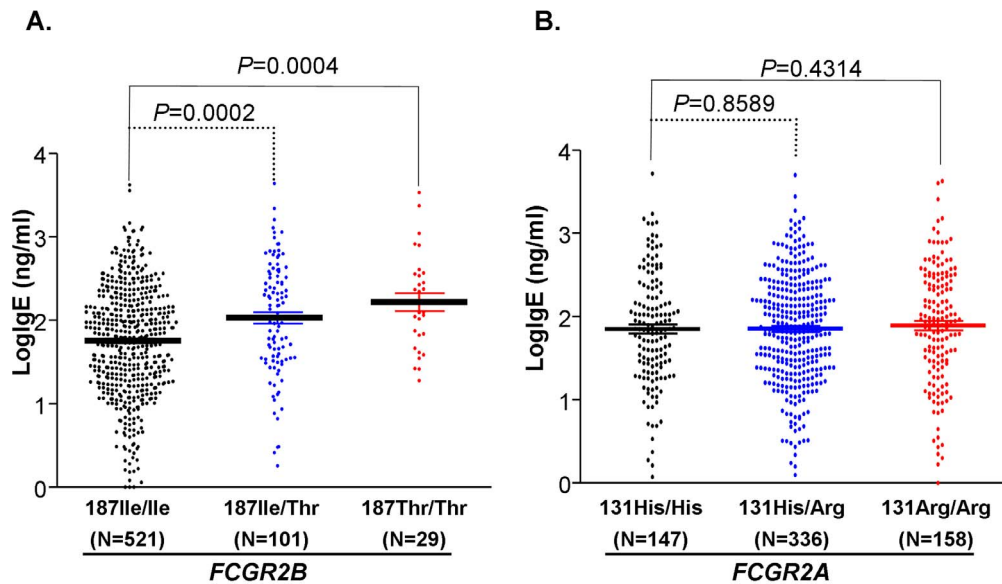


Figure 1. Association of SNP *FCGR2B*-187Ile>Thr with IgE levels. **A.** The genotypes of *FCGR2B*-187Ile>Thr were significantly associated with IgE levels in ANOVA ($P < 0.0001$). The rare *FCGR2B*-187Thr carriers (187Ile/Thr and 187Thr/Thr genotypes) produced significantly more IgE than the homozygous subjects for common allele (187Ile) in Mann-Whitney tests. **B.** The genotypes of *FCGR2B*-131His>Arg were not associated with IgE levels in ANOVA ($P = 0.817$). No significant differences were found between *FCGR2A*-131Arg carriers (131His/Arg and 131Arg/Arg genotypes) and the 131His homozygous subjects in Mann-Whitney tests ($P > 0.05$). doi:10.1371/journal.pone.0089196.g001

data indicate a role for IgG Fc receptors in the development of allergy.

Fc γ RIIA is expressed on the surface of various immune cells including mast cells, basophils, neutrophils, monocytes, dendritic cells, macrophages, and platelets [19,20]. The Fc γ RIIA-131His>Arg SNP significantly affects receptor binding affinity and specificity for IgG subclasses [21,22]. Although both Fc γ RIIA-131His and 131Arg alleles bind IgG1 and IgG3, the Fc γ RIIA-131His allele displays a higher binding affinity for IgG3 and is capable of binding IgG2 most effectively as compared to the Fc γ RIIA-131Arg allele [21,22]. The functional Fc γ RIIA-131His>Arg SNP affects the receptor binding affinity for IgG and thus influences the clinical phenotype in response to infectious diseases and inflammation [21]. The Fc γ RIIA-131His>Arg SNP affects the functions of bacterial phagocytosis [23,24] and immune complex handling [22,25,26]. The Fc γ RIIA-131His>Arg SNP has been reported to be associated with ulcerative colitis [27], Kawasaki diseases [28], systemic lupus erythematosus [29], and chronic inflammatory disorders such as periodontitis [30,31] and Guillain-Barré syndrome [32]. In addition, Fc γ RIIA-131His>Arg polymorphism is associated with infections including recurrent bacterial respiratory tract infections [33], bacteremic pneumococcal pneumonia [34], severe acute respiratory syndrome [35], severe sepsis [36], HIV [37], and EB virus infection [38]. IgG2 is produced primarily in response to polysaccharide/carbohydrate antigens commonly found in allergens. The protective effect of the Fc γ RIIA-131His allele on atopy is possibly due to the increased capacity of this allele to efficiently internalize and destroy allergen-IgG2 immune complexes. The role of Fc γ RIIA in allergy was also demonstrated in transgenic mouse models [39]. Therefore, Fc γ RIIA likely contributes to allergy development in humans. Although the Fc γ RIIA-131His>Arg SNP is associated with atopy, the SNP is not associated with IgE production (Figure 1B), suggesting that Fc γ RIIA likely affects allergy through pathways of immune complex clearance and receptor-mediated cell activation. Future studies are required to reveal whether IgG2 levels are

associated with the asthma or atopy in the context of Fc γ RIIA SNP and whether Fc γ RIIA-mediated functions (immune complex clearance and phagocytosis of allergens) are different between asthmatic and non-asthmatic human subjects.

Fc γ RIIB, mainly expressed on B cells and myeloid cells, is a classical inhibitory IgG Fc receptor [40,41,42]. Cross-linking of Fc γ RIIB by immune complexes leads to the down-regulation of B cell activation and antibody production, which is an important feedback mechanism to maintain the homeostasis of immune responses [5,40,43]. Therefore, Fc γ RIIB overexpression (or enhanced Fc γ RIIB functions) reduces the immunoglobulin production in T-dependent immune responses [44]. In humanized mouse models of immunoglobulin production, co-engagement of IgE B-cell receptor with Fc γ RIIB drastically inhibited human IgE production [45]. Fc γ RIIB-187Ile>Thr SNP (rs1050501) is located within the receptor transmembrane segment and the Fc γ RIIB-187Thr allele is less efficient in mediating inhibitory signals than the Fc γ RIIB-187Ile allele [46,47,48]. We observed that the low function Fc γ RIIB-187Thr allele is significantly associated with elevated IgE levels (Figure 1), suggesting that the reduced Fc γ RIIB function may promote IgE antibody production by B cells in humans. Interestingly, the low function Fc γ RIIB-187Thr allele is also associated with protection against malaria [49], signifying Fc γ RIIB functions play important roles in controlling the immune response to parasites [50]. Nevertheless, Fc γ RIIB-187Ile>Thr SNP may also be in linkage equilibrium with SNPs of the *FCER1A* gene encoding for the alpha chain of the high affinity receptor for IgE (Fc ϵ RIA) because a GWAS identified the *FCER1A* functional variants strongly associated with total IgE levels [51].

Fc γ RIIB on immune cells also inhibits cellular functions including phagocytosis, ADCC, degranulation, and cytokine release [40]. Mast cells from Fc γ RIIB^{-/-} mice are highly sensitive to IgG-triggered degranulation compared to those from the wild-type mice. Fc γ RIIB-deficient mice have an enhanced passive cutaneous anaphylaxis reaction, as a result of the decreased threshold for mast-cell activation through activating Fc receptors

[52]. Fc γ RIIB negatively regulates cell activation triggered by high-affinity IgE receptors (Fc ϵ R1) [53]. Fc γ RIIB binds to the Fc domains of IgE and IgG with similar low affinity [54,55]. Mast cells and basophils could be regulated by immune complexes of allergen-IgG or allergen-IgE. Fc γ RIIB-deficient mice developed more severe eosinophilia compared to wild-type mice, suggesting an important regulatory role for Fc γ RIIB in the onset of allergic diseases [56]. Fc γ RIIB-knockout mice developed the exacerbated lung inflammation [57]. Taken together, Fc γ RIIB seems to play a critical role in allergic inflammations. In the current study, the dysfunctional Fc γ RIIB-187Thr allele was found to be a risk factor for atopy. A decreased activation threshold for immune cells carrying Fc γ RIIB-187Thr allele may be responsible for the increased sensitivity to allergens that trigger the allergic responses, which may explain the association between the defective Fc γ RIIB allele and atopy.

On the other hand, the functional SNPs of the other three activating Fc γ Rs (Fc γ RIIA, Fc γ RIIB, and Fc γ RIIC) were not associated with asthma, BHR, and atopy, suggesting that functions of the restrictively expressed activating Fc γ Rs (Fc γ RIIA, Fc γ RIIB, and Fc γ RIIC) may not play prevailing roles in the development of allergy. Our current study had more than 80% power to detect an association between a *FCGR* SNP and atopy with an OR of 1.75.

References

- Duffy DL, Martin NG, Battistutta D, Hopper JL, Mathews JD (1990) Genetics of asthma and hay fever in Australian twins. *Am Rev Respir Dis* 142: 1351–1358.
- Nieminen MM, Kaprio J, Koskenvuo M (1991) A population-based study of bronchial asthma in adult twin pairs. *Chest* 100: 70–75.
- Galli SJ, Tsai M (2012) IgE and mast cells in allergic disease. *Nat Med* 18: 693–704.
- Torgerson DG, Ampleford EJ, Chiu GY, Gauderman WJ, Gignoux CR, et al. (2011) Meta-analysis of genome-wide association studies of asthma in ethnically diverse North American populations. *Nat Genet* 43: 887–892.
- Ravetch JV, Bolland S (2001) IgG Fc receptors. *Annu Rev Immunol* 19: 275–290.
- Borucho AM, Heller G, Veri MC, Bonvini E, Ravetch JV, et al. (2005) Activating and inhibitory IgG Fc receptors on human DCs mediate opposing functions. *J Clin Invest* 115: 2914–2923.
- Takai T (2002) Roles of Fc receptors in autoimmunity. *Nat Rev Immunol* 2: 580–592.
- Chen JY, Wang CM, Ma CC, Luo SF, Edberg JC, et al. (2006) Association of a transmembrane polymorphism of Fc γ receptor IIb (FCGR2B) with systemic lupus erythematosus in Taiwanese patients. *Arthritis Rheum* 54: 3908–3917.
- Edberg JC, Langefeld CD, Wu J, Moser KL, Kaufman KM, et al. (2002) Genetic linkage and association of Fc γ receptor IIIA (CD16A) on chromosome 1q23 with human systemic lupus erythematosus. *Arthritis Rheum* 46: 2132–2140.
- Morgan AW, Griffiths B, Ponchel F, Montague BM, Ali M, et al. (2000) Fc γ receptor type IIIA is associated with rheumatoid arthritis in two distinct ethnic groups. *Arthritis Rheum* 43: 2328–2334.
- Wu J, Edberg JC, Redecha PB, Bansal V, Guyre PM, et al. (1997) A novel polymorphism of Fc γ RIIA (CD16) alters receptor function and predisposes to autoimmune disease. *J Clin Invest* 100: 1059–1070.
- (1997) A genome-wide search for asthma susceptibility loci in ethnically diverse populations. The Collaborative Study on the Genetics of Asthma (CSGA). *Nat Genet* 15: 389–392.
- Reilly C, Miller MB, Liu Y, Oetting WS, King R, et al. (2007) Linkage analysis of a cluster-based quantitative phenotype constructed from pulmonary function test data in 27 multigenerational families with multiple asthmatic members. *Hum Hered* 64: 136–145.
- Qiu WQ, de Bruin D, Brownstein BH, Pearce R, Ravetch JV (1990) Organization of the human and mouse low-affinity Fc gamma R genes: duplication and recombination. *Science* 248: 732–735.
- Chen JY, Wang CM, Ma CC, Hsu LA, Ho HH, et al. (2008) A transmembrane polymorphism in Fc γ RIIB (FCGR2B) is associated with the production of anti-cyclic citrullinated peptide autoantibodies in Taiwanese RA. *Genes Immun* 9: 680–688.
- Ory PA, Clark MR, Kwoh EE, Clarkson SB, Goldstein IM (1989) Sequences of complementary DNAs that encode the NA1 and NA2 forms of Fc receptor III on human neutrophils. *J Clin Invest* 84: 1688–1691.
- Ory PA, Goldstein IM, Kwoh EE, Clarkson SB (1989) Characterization of polymorphic forms of Fc receptor III on human neutrophils. *J Clin Invest* 83: 1676–1681.
- Laird NM, Horvath S, Xu X (2000) Implementing a unified approach to family-based tests of association. *Genet Epidemiol* 19 Suppl 1: S36–42.
- Takai T (2005) Fc receptors and their role in immune regulation and autoimmunity. *J Clin Immunol* 25: 1–18.
- Rascu A, Repp R, Westerdaal NA, Kalden JR, van de Winkel JG (1997) Clinical relevance of Fc gamma receptor polymorphisms. *Ann N Y Acad Sci* 815: 282–295.
- Bruhns P, Iannascoli B, England P, Mancardi DA, Fernandez N, et al. (2009) Specificity and affinity of human Fc γ receptors and their polymorphic variants for human IgG subclasses. *Blood* 113: 3716–3725.
- Warmerdam PA, van de Winkel JG, Vluga A, Westerdaal NA, Capel PJ (1991) A single amino acid in the second Ig-like domain of the human Fc gamma receptor II is critical for human IgG2 binding. *J Immunol* 147: 1338–1343.
- Sanders LA, Feldman RG, Voorhorst-Ogink MM, de Haas M, Rijkers GT, et al. (1995) Human immunoglobulin G (IgG) Fc receptor IIA (CD32) polymorphism and IgG2-mediated bacterial phagocytosis by neutrophils. *Infect Immun* 63: 73–81.
- Bredius RG, de Vries CE, Troelstra A, van Alphen L, Weening RS, et al. (1993) Phagocytosis of *Staphylococcus aureus* and *Haemophilus influenzae* type B opsonized with polyclonal human IgG1 and IgG2 antibodies. Functional hFc gamma RIIa polymorphism to IgG2. *J Immunol* 151: 1463–1472.
- Salmon JE, Edberg JC, Brogle NL, Kimberly RP (1992) Allelic polymorphisms of human Fc gamma receptor IIA and Fc gamma receptor IIIB. Independent mechanisms for differences in human phagocyte function. *J Clin Invest* 89: 1274–1281.
- Salmon JE, Millard S, Schachter LA, Arnett FC, Ginzler EM, et al. (1996) Fc gamma RIIA alleles are heritable risk factors for lupus nephritis in African Americans. *J Clin Invest* 97: 1348–1354.
- Asano K, Matsushita T, Umeno J, Hosono N, Takahashi A, et al. (2009) A genome-wide association study identifies three new susceptibility loci for ulcerative colitis in the Japanese population. *Nat Genet* 41: 1325–1329.
- Khor CC, Davila S, Breunis WB, Lee YC, Shimizu C, et al. (2011) Genome-wide association study identifies FCGR2A as a susceptibility locus for Kawasaki disease. *Nat Genet* 43: 1241–1246.
- Harley JB, Alarcon-Riquelme ME, Criswell LA, Jacob CO, Kimberly RP, et al. (2008) Genome-wide association scan in women with systemic lupus erythematosus identifies susceptibility variants in ITGAM, PTK, KIAA1542 and other loci. *Nat Genet* 40: 204–210.
- Yamamoto K, Kobayashi T, Grossi S, Ho AW, Genco RJ, et al. (2004) Association of Fc γ receptor IIA genotype with chronic periodontitis in Caucasians. *J Periodontol* 75: 517–522.
- Chai L, Song YQ, Leung WK (2012) Genetic polymorphism studies in periodontitis and Fc γ receptors. *J Periodontol Res* 47: 273–285.
- van der Pol WL, van den Berg LH, Scheepers RH, van der Bom JG, van Doorn PA, et al. (2000) IgG receptor IIA alleles determine susceptibility and severity of Guillain-Barre syndrome. *Neurology* 54: 1661–1665.

Supporting Information

Table S1 Distribution of *FCGR2A* SNP (rs1801274) in atopy⁺ and atopy⁻ subjects. (DOC)

Table S2 Distribution of *FCGR2B* SNP (rs1050501) in atopy⁺ and atopy⁻ subjects. (DOC)

Author Contributions

Conceived and designed the experiments: JW WSO PS MNB. Performed the experiments: JW RL JH WG WSO MNB. Analyzed the data: JW RL JH WG WSO PS MNB. Contributed reagents/materials/analysis tools: WG WSO MNB. Wrote the paper: JW WG WSO MNB.

33. Sanders LA, van de Winkel JG, Rijkers GT, Voorhorst-Ogink MM, de Haas M, et al. (1994) Fc gamma receptor IIa (CD32) heterogeneity in patients with recurrent bacterial respiratory tract infections. *J Infect Dis* 170: 854–861.
34. Yee AM, Phan HM, Zuniga R, Salmon JE, Musher DM (2000) Association between FcgammaRIIa-R131 allotype and bacteremic pneumococcal pneumonia. *Clin Infect Dis* 30: 25–28.
35. Yuan FF, Tanner J, Chan PK, Biffin S, Dyer WB, et al. (2005) Influence of FcgammaRIIA and MBL polymorphisms on severe acute respiratory syndrome. *Tissue Antigens* 66: 291–296.
36. Endeman H, Cornips MC, Grutters JC, van den Bosch JM, Ruven HJ, et al. (2009) The Fc gamma receptor IIA-R/R131 genotype is associated with severe sepsis in community-acquired pneumonia. *Clin Vaccine Immunol* 16: 1087–1090.
37. Forthal DN, Landucci G, Bream J, Jacobson LP, Phan TB, et al. (2007) FcgammaRIIa genotype predicts progression of HIV infection. *J Immunol* 179: 7916–7923.
38. Diamantopoulos PT, Kalotychoy V, Polonyfi K, Sofotasiou M, Anastasopoulou A, et al. (2013) Correlation of Fc-gamma RIIA polymorphisms with latent Epstein-Barr virus infection and latent membrane protein 1 expression in patients with low grade B-cell lymphomas. *Leuk Lymphoma* 54: 2030–2034.
39. Jonsson F, Mancardi DA, Zhao W, Kita Y, Iannascoli B, et al. (2012) Human FcgammaRIIA induces anaphylactic and allergic reactions. *Blood* 119: 2533–2544.
40. Ravetch JV, Lanier LL (2000) Immune inhibitory receptors. *Science* 290: 84–89.
41. Muta T, Kurosaki T, Misulovin Z, Sanchez M, Nussenzweig MC, et al. (1994) A 13-amino-acid motif in the cytoplasmic domain of Fc gamma RIIB modulates B-cell receptor signalling. *Nature* 368: 70–73.
42. Xiang Z, Cutler AJ, Brownlie RJ, Fairfax K, Lawlor KE, et al. (2007) FcgammaRIIb controls bone marrow plasma cell persistence and apoptosis. *Nat Immunol* 8: 419–429.
43. Cohen-Solal JF, Cassard L, Fridman WH, Sautes-Fridman C (2004) Fc gamma receptors. *Immunol Lett* 92: 199–205.
44. Brownlie RJ, Lawlor KE, Niederer HA, Cutler AJ, Xiang Z, et al. (2008) Distinct cell-specific control of autoimmunity and infection by FcgammaRIIb. *J Exp Med* 205: 883–895.
45. Chu SY, Horton HM, Pong E, Leung IW, Chen H, et al. (2012) Reduction of total IgE by targeted coengagement of IgE B-cell receptor and FcgammaRIIb with Fc-engineered antibody. *J Allergy Clin Immunol* 129: 1102–1115.
46. Li X, Wu J, Carter RH, Edberg JC, Su K, et al. (2003) A novel polymorphism in the Fc gamma receptor IIB (CD32B) transmembrane region alters receptor signaling. *Arthritis Rheum* 48: 3242–3252.
47. Kono H, Kyogoku C, Suzuki T, Tsuchiya N, Honda H, et al. (2005) Fc{gamma}RIIB Ile232Thr transmembrane polymorphism associated with human systemic lupus erythematosus decreases affinity to lipid rafts and attenuates inhibitory effects on B cell receptor signaling. *Hum Mol Genet* 14: 2881–2892.
48. Floto RA, Clatworthy MR, Heilbronn KR, Rosner DR, Macary PA, et al. (2005) Loss of function of a lupus-associated Fc gamma RIIB polymorphism through exclusion from lipid rafts. *Nat Med* 11: 1056–1058.
49. Willcocks LC, Carr EJ, Niederer HA, Rayner TF, Williams TN, et al. (2010) A defuncting polymorphism in FCGR2B is associated with protection against malaria but susceptibility to systemic lupus erythematosus. *Proc Natl Acad Sci U S A* 107: 7881–7885.
50. Clatworthy MR, Willcocks L, Urban B, Langhorne J, Williams TN, et al. (2007) Systemic lupus erythematosus-associated defects in the inhibitory receptor Fc gamma RIIB reduce susceptibility to malaria. *Proc Natl Acad Sci U S A* 104: 7169–7174.
51. Weidinger S, Gieger C, Rodriguez E, Baurecht H, Mempel M, et al. (2008) Genome-wide scan on total serum IgE levels identifies FCER1A as novel susceptibility locus. *PLoS Genet* 4: e1000166.
52. Takai T, Ono M, Hikida M, Ohmori H, Ravetch JV (1996) Augmented humoral and anaphylactic responses in Fc gamma RII-deficient mice. *Nature* 379: 346–349.
53. Malbec O, Attal JP, Fridman WH, Daeron M (2002) Negative regulation of mast cell proliferation by Fc gamma RIIB. *Mol Immunol* 38: 1295–1299.
54. Takizawa F, Adamczewski M, Kinet JP (1992) Identification of the low affinity receptor for immunoglobulin E on mouse mast cells and macrophages as Fc gamma RII and Fc gamma RIII. *J Exp Med* 176: 469–475.
55. Ujike A, Ishikawa Y, Ono M, Yuasa T, Yoshino T, et al. (1999) Modulation of immunoglobulin (Ig)E-mediated systemic anaphylaxis by low-affinity Fc receptors for IgG. *J Exp Med* 189: 1573–1579.
56. Watanabe T, Okano M, Hattori H, Yoshino T, Ohno N, et al. (2004) Roles of Fc gamma RIIB in nasal eosinophilia and IgE production in murine allergic rhinitis. *Am J Respir Crit Care Med* 169: 105–112.
57. Dharajiya N, Vaidya SV, Murai H, Cardenas V, Kurosky A, et al. (2010) Fc gamma RIIB inhibits allergic lung inflammation in a murine model of allergic asthma. *PLoS One* 5: e9337.