

The PPAR γ Locus Makes Long-Range Chromatin Interactions with Selected Tissue-Specific Gene Loci during Adipocyte Differentiation in a Protein Kinase A Dependent Manner

Scott E. LeBlanc¹, Qiong Wu¹, A. Rasim Barutcu¹, Hengyi Xiao^{1,2}, Yasuyuki Ohkawa^{1,3}, Anthony N. Imbalzano^{1*}

1 Department of Cell and Developmental Biology, University of Massachusetts Medical School, Worcester, Massachusetts, United States of America, **2** Laboratory of Aging Research, State Key Laboratory of Biotherapy, West China Hospital, West China Medical School, Sichuan University, Chengdu, China, **3** Department of Advanced Medical Initiatives, JST-CREST, Faculty of Medicine, Kyushu University, Fukuoka, Japan

Abstract

Differentiation signaling results in reprogramming of cellular gene expression that leads to morphological changes and functional specialization of a precursor cell. This global change in gene expression involves temporal regulation of differentiation-specific genes that are located throughout the genome, raising the idea that genome structure may also be re-organized during cell differentiation to facilitate regulated gene expression. Using in vitro adipocyte differentiation as a model, we explored whether gene organization within the nucleus is altered upon exposure of precursor cells to signaling molecules that induce adipogenesis. The peroxisome proliferator-activated receptor gamma (PPAR γ) nuclear hormone receptor is a master determinant of adipogenesis and is required for adipose differentiation. We utilized the chromosome conformation capture (3C) assay to determine whether the position of the PPAR γ locus relative to other adipogenic genes is changed during differentiation. We report that the PPAR γ 2 promoter is transiently positioned in proximity to the promoters of genes encoding adipokines and lipid droplet associated proteins at 6 hours post-differentiation, a time that precedes expression of any of these genes. In contrast, the PPAR γ 2 promoter was not in proximity to the EF1 α promoter, which drives expression of a constitutively active, housekeeping gene that encodes a translation elongation factor, nor was the PPAR γ 2 promoter in proximity to the promoter driving the expression of the C/EBP α regulatory protein. The formation of the long-range, intergenic interactions involving the PPAR γ 2 promoter required the regulatory factor C/EBP β , elevated cyclic AMP (cAMP) levels, and protein kinase A (PKA) signaling. We conclude that genome organization is dynamically remodeled in response to adipogenic signaling, and we speculate that these transient inter-genic interactions may be formed for the purposes of selecting some of the transcriptionally silent tissue-specific loci for subsequent transcriptional activation.

Citation: LeBlanc SE, Wu Q, Barutcu AR, Xiao H, Ohkawa Y, et al. (2014) The PPAR γ Locus Makes Long-Range Chromatin Interactions with Selected Tissue-Specific Gene Loci during Adipocyte Differentiation in a Protein Kinase A Dependent Manner. PLoS ONE 9(1): e86140. doi:10.1371/journal.pone.0086140

Editor: Dmitry I. Nurminky, University of Maryland School of Medicine, United States of America

Received: May 16, 2013; **Accepted:** December 5, 2013; **Published:** January 20, 2014

Copyright: © 2014 LeBlanc et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Funding: This work was supported by National Institutes of Health grants DK079239, DK084278 and GM56244 to ANI and F32DK082263 to SEL. ANI is a member of the UMass Medical School Diabetes and Endocrine Research Center, supported by the National Institutes of Health grant DK32520. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing Interests: The authors have declared that no competing interests exist.

* E-mail: Anthony.imbalzano@umassmed.edu

Introduction

Maturation of a pre-adipocyte into a mature adipocyte involves significant changes in cellular structure and organization. Major changes in the expression of cytoskeletal structural proteins occur during adipogenesis of both cultured and primary cells; these changes have been linked to the notable transformation in cell morphology that occurs when preadipocytes differentiate into adipocytes [1–3]. Recent studies also report structural alterations that occur in the nuclear lamina and in the connections between the nucleus and cytoplasmic filaments as a function of adipogenic differentiation [4]. Furthermore, expression of the lamin associated polypeptide 2 α (LAP2 α) or reduction in the expression of lamins or of the lamin associated protein emerlin modulates adipocyte differentiation [5–7], while mutations in nuclear lamina proteins have been associated with lipodystrophy syndromes [8,9].

Other reports document organizational changes within the nucleus. These include reports of differentiation-dependent changes in chromosome territories [10], relocalization of adipogenic gene positioning within the nucleus relative to the nuclear lamina during differentiation [11,12], and preferential association with SC-35 domains [13], which are nuclear structures that are enriched in factors involved in pre-mRNA metabolism and that may act to spatially link gene expression and mRNA processing [14,15].

The pivotal controller of adipogenesis is PPAR γ 2, which is both necessary and sufficient for adipogenic differentiation [16–18]. While PPAR γ 2 expression does not occur until an intermediate stage in the differentiation program [19], it is well-established that the PPAR γ 2 promoter is marked by early binding of C/EBP β and early changes in histone modification patterns and DNase I

hypersensitivity [20–23] well in advance of PPAR γ 2 expression. This suggests the possibility that the early modulation of factor binding and chromatin structure at the PPAR γ 2 promoter may serve as a focal point for differentiation-dependent changes in higher order chromatin structure. Given these considerations, we wondered whether there was any spatial relationship between the PPAR γ 2 locus and the loci encoding other genes expressed during adipogenesis.

Most adipocyte-specific gene expression involves regulation by PPAR γ 2 [24]. PPAR γ 2 target genes are diverse and include genes encoding adipokines, which are hormones secreted by adipose that contribute to the regulation of energy balance of the organism and guide a number of physiological and pathological processes [25,26]. Classic adipokines include adiponectin and leptin. Adiponectin levels are directly correlated with insulin sensitivity. Leptin is an appetite-sensitizing hormone for providing feedback for satiety, and both adipokines are believed to have high therapeutic potential [27,28]. Another class of PPAR γ 2 target genes mediates lipid accumulation and storage. Lipid droplets are coated in proteins called perilipins, including perilipin 1 and adipose differentiation-related protein (ADRP), also called perilipin 2, which serve to protect the lipids contained within from adipose lipases [29,30].

In this study, we document interactions between the PPAR γ 2 promoter and the promoters controlling the adipocyte hormone genes and perilipins 1 and 2 in differentiating 3T3-L1 adipocytes and in differentiating C3H10T1/2 mesenchymal cells using the chromosome conformation capture (3C) assay. Interestingly, the observed interactions between the PPAR γ 2 promoter and adipokine and perilipin promoters were most frequent at 6 hr following the addition of the adipogenic cocktail and decreased significantly at later time points. Thus the formation of these intergenic interactions involving the PPAR γ 2 promoter preceded PPAR γ 2 gene expression and PPAR γ 2 target gene expression. Interactions were dependent upon the C/EBP β transcriptional regulator. Additional experiments revealed that cAMP signaling was required for the formation of these long-range, inter-genic interactions, and that these interactions were also dependent on elevated cAMP levels and on the presence of protein kinase A.

Materials and Methods

Cell Culture and Retroviral Infections

3T3-L1 pre-adipocyte cells and C3H10T1/2 mesenchymal cells were obtained from ATCC and cultured in Dulbecco's modified Eagle's medium (DMEM) containing 10% calf serum or 10% fetal calf serum (FCS). For adipogenic differentiation, 2 day post-confluent cells were differentiated with a standard adipogenic cocktail (1 μ g/ml insulin, 0.25 μ g/ml dexamethasone, 0.5 mM isobutylmethylxanthine (IBMX) with 10% FCS) or with the indicated subset of components. Where indicated, forskolin (10 μ M, Calbiochem) was added to serum-containing differentiation media lacking the differentiation cocktail components for the first two days of the differentiation process. 3T3-L1 cells were also pre-treated for 1 hour prior to administration of differentiation cocktail with either 10 μ M H89 (Sigma) or 10 μ M myr-PKI (Calbiochem). BOSC23 retroviral packaging cells were cultured in DMEM containing 10% FCS and were cycled through selective media every 1–2 months as described [31]. pSuperior-retro-puro (OligoEngine, Seattle, WA), pSuperior-retro-puro-PKAC1 α , pSuperior-retro-puro-PKAC1 β , and pSuperior-retro-puro-C/EBP β [23] viral packaging was achieved by transfection of the plasmid into BOSC23 cells using Fugene6 (Roche) as described [23].

Collection of viral supernatant and infection of cells was also described previously [23].

Chromatin Conformation Capture (3C) Assay

The 3C protocol was adapted from published methods [32–34]. Cells were cross-linked with 1% formaldehyde for 10 minutes at RT and quenched with 0.125M glycine for 5 min. Samples were harvested in PBS containing protease inhibitor cocktail (Sigma). Pellets were lysed in lysis buffer (10 mM Tris HCl pH 8.0, 10 mM NaCl, 0.5% Nonidet P-40) containing protease inhibitors, incubated on ice for 15 min, and dounced 10 times using pestle B, followed by another 15 minute incubation on ice. After removal of supernatant, nuclei pellets were re-suspended in restriction endonuclease buffer 2 (NEB; 50 mM NaCl, 10 mM Tris-HCl, 10 mM MgCl₂, 1 mM Dithiothreitol) and washed once in NEB buffer 2. SDS was added to a final concentration of 0.3%, followed by a 60 min incubation at 65 degrees C, followed by SDS sequestration using 1.8% Triton X-100 for 60 minutes at 37 degrees C. Samples were incubated with 350 units each of restriction enzymes StuI and PvuII (NEB) and were incubated overnight at 37 degrees C. After SDS-mediated inactivation of the enzymes for 30 minutes at 65°C (1.6% final concentration) and SDS-sequestration with 1% Triton X-100 at 37 degrees C for 60 min, 2X Takara Mighty Mix was added to each sample and incubated for 1 hr at 16 degrees C. Ligated samples were treated overnight with proteinase K (20 mg/mL) and RNase A (10 mg/mL), and DNA fragments were purified with a DNeasy Blood and Tissue kit (Qiagen). Samples were analyzed by realtime PCR using the Promega GoTaq Master Mix on a DNA Opticon (MJ Research) or an ABI StepOne Plus (Applied Biosystems) and interaction frequencies were normalized to the values of intra-genic interactions occurring at the endogenous TFIIF (ERCC3) locus [35] or a gene desert region located on chromosome 3 [36].

Primer Sequences for 3C Assays

PPAR γ 2 promoter StuI site 1 (–132 bp from TSS) forward: 5'-GTAATGTACCAAGTCTTGCCAAAGCAGCAG

Leptin PvuII Forward: 5'-TCACAGGATGAAATGAGAC-GACTGTTCTTCG

Adiponectin PvuII Forward: 5'-TAGAGAATGGC-CAAAGCCTGGAAACAGGA

Perilipin-1 PvuII Forward: 5'-TGTTTCTGAGGAGGA-GACCTT

Perilipin-2 PvuII Forward: 5'-AATGAGTGGCCCCA-GAGTC

C/EBP α StuI Forward: 5'-CTAGTTGCTGGTCCAAAG-CAGTCTCCAAC

EF1 α StuI Forward: 5'-GTCGCCTTGACGTTCTTTT
ERCC3 –0.9 Forward: 5'-TGTAGTTCTGTGGCTA-GAACCTG

ERCC3 3.7 Forward: 5'-TGATGCAGAAGGATGCTGAC
Chromosome 3 Gene desert region:

GDR StuI Forward 5'-CCATTTAGAATCGTGGCAGGTC-TAT

GDR PvuII Forward 5'-ATACATTGAAATGAGTCCTCCA-CAT

Protein Extracts and Western Blots

Whole cell extracts were prepared and quantified and used for western blotting as described [37]. Primary antibodies were PKAC1 α (Santa Cruz; sc-903), PKAC1 β (Santa Cruz; sc-904), C/EBP β (Santa Cruz; sc-150), and p85 phosphatidylinositol-3-kinase (Millipore; 06–195).

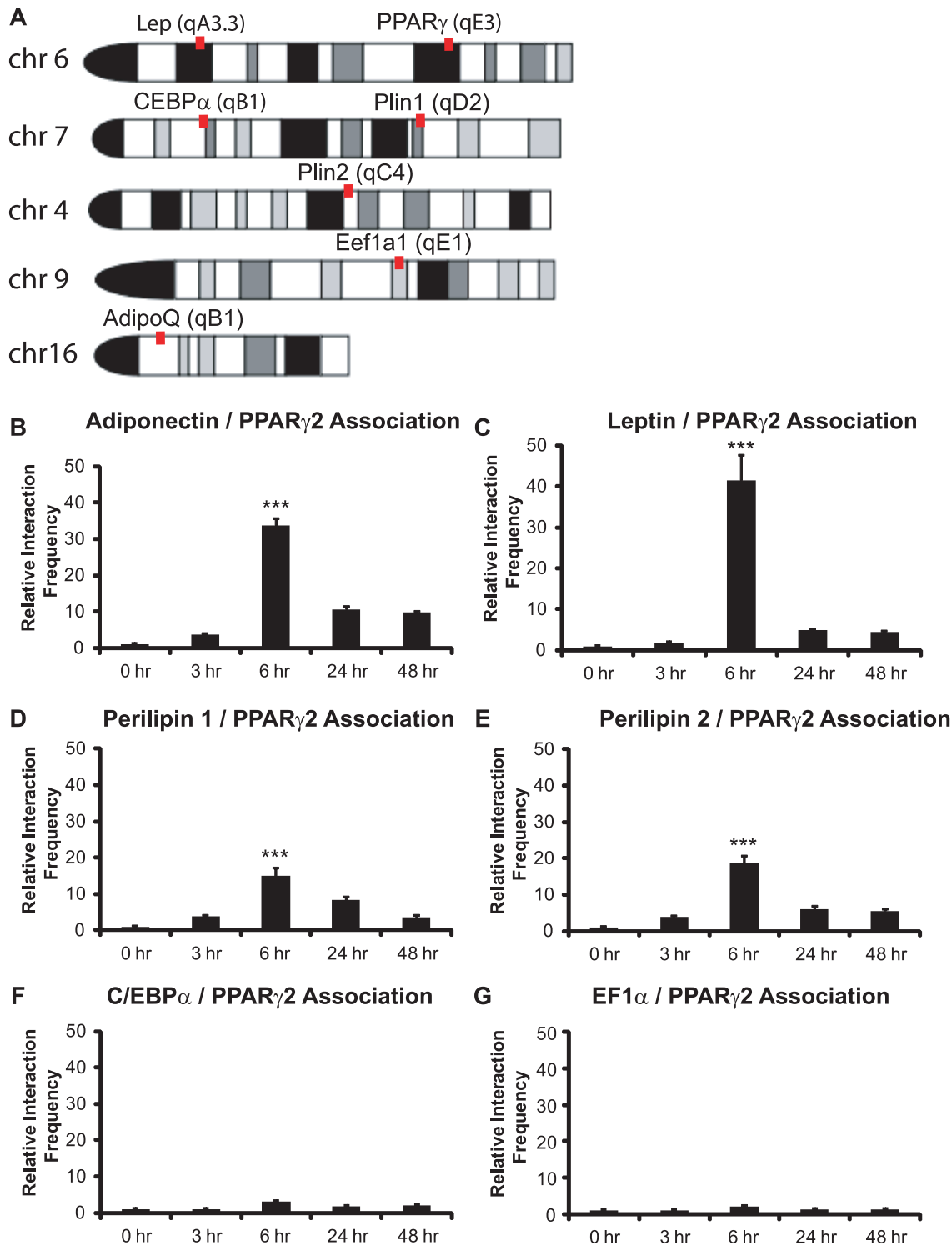


Figure 1. The PPAR γ 2 promoter maximally interacts with adipokine and perilipin gene promoters in 3T3-L1 cells 6 hours post-differentiation. (A) Schematic map of chromosomal locations of loci of interest. Lep: leptin; Plin1: perilipin 1; Plin2: perilipin 2; Eef1a1: EF1 α ; AdipoQ: adiponectin. 3C assays were performed at the indicated times to measure long-range, intergenic interactions between the PPAR γ 2 promoter and (B) the adiponectin promoter, (C) the leptin promoter, (D) the perilipin 1 promoter, (E) the perilipin 2/ADRP promoter, (F) the C/EBP α promoter, or (G) the EF1 α promoter. Interaction frequencies are presented relative to the intragenic interaction frequency observed between different regions of the TFIIH locus, which served as a normalization control. The interaction frequency at 0 h was set to 1; the interaction frequencies for the remaining time points are presented relative to that value. Results indicate the average of three independent experiments \pm standard deviation. Values for the timepoint of maximum interaction frequency (6 h) were compared to values at time 0 by one-tailed t-test. *** p <0.001. doi:10.1371/journal.pone.0086140.g001

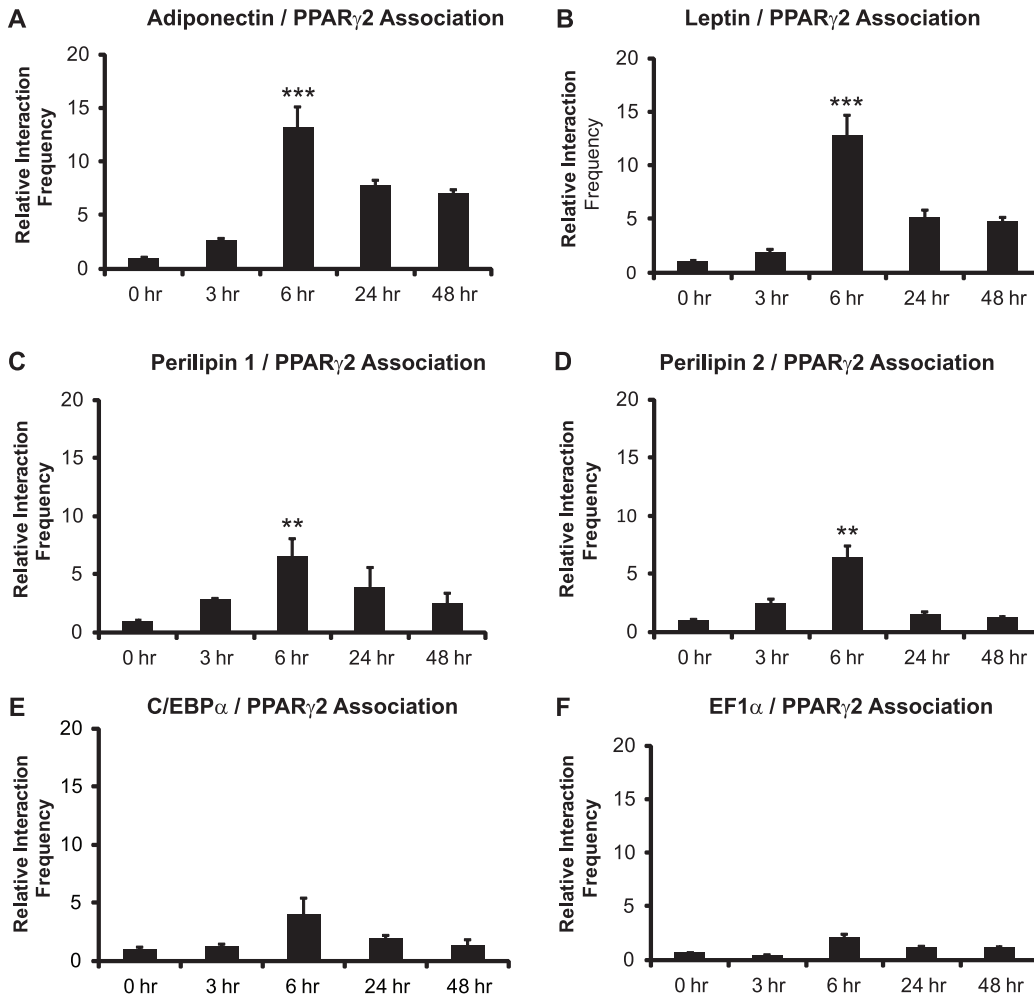


Figure 2. The PPAR γ 2 promoter maximally interacts with adipokine and perilipin gene promoters in C3H10T1/2 cells 6 hours post-differentiation. 3C assays were performed at the indicated times to measure long-range, intergenic interactions between the PPAR γ 2 promoter and (A) the adiponectin promoter, (B) the leptin promoter, (C) the perilipin 1 promoter, (D) the perilipin 2/ADRP promoter, (E) the C/EBP α promoter, or (F) the EF1 α promoter. Interaction frequencies are presented relative to the intragenic interaction frequency observed between different regions of the TFIIH locus, which served as a normalization control. The interaction frequency at 0 h was set to 1; the interaction frequencies for the remaining time points are presented relative to that value. Results indicate the average of three independent experiments \pm standard deviation. Values for the timepoint of maximum interaction frequency (6 h) were compared to values at time 0 by one-tailed t-test. *** p <0.001; ** p <0.01. doi:10.1371/journal.pone.0086140.g002

Statistics

Results are indicated as the average of three independent experiments \pm standard deviation. Data were statistically analyzed by a one-tailed t-test.

Results

The PPAR γ 2 Promoter Makes Long-range Interactions with Adipokine and Perilipin Promoters in Differentiating Adipocytes

Numerous molecular changes occur at the PPAR γ 2 promoter within hours following the onset of adipogenic differentiation signaling. Restriction endonuclease accessibility is increased within 30 minutes, c-fos binds within 1 hour, C/EBP β binds within 2 hours, and DNase I hypersensitivity was reported by 2 hours [21,23]. All of these events occur well before the onset of PPAR γ 2 mRNA synthesis [38].

We hypothesized that the onset of differentiation signaling might cause rearrangement of genomic chromatin structure. Because of the requirement for and central role of the PPAR γ 2 protein in the adipogenic differentiation process, we investigated whether the PPAR γ locus might be a region where changes in genomic organization occur. We utilized 3C methodology [32] to assess relative changes in gene organization. The 3C technique utilizes cross-linking to identify regions of chromatin that are in close juxtaposition to each other and has allowed identification of inter-chromosomal interactions as well as long-range interactions between non-contiguous sequences on the same chromosome [39–41].

For initial 3C experiments, we sought to determine whether the PPAR γ locus is in close physical proximity to other adipogenic loci in undifferentiated pre-adipocytes or in pre-adipocytes that have been exposed to differentiation signaling. We designed primers to the PPAR γ 2 promoter and to promoters from four known PPAR γ 2 target genes, those controlling expression of the

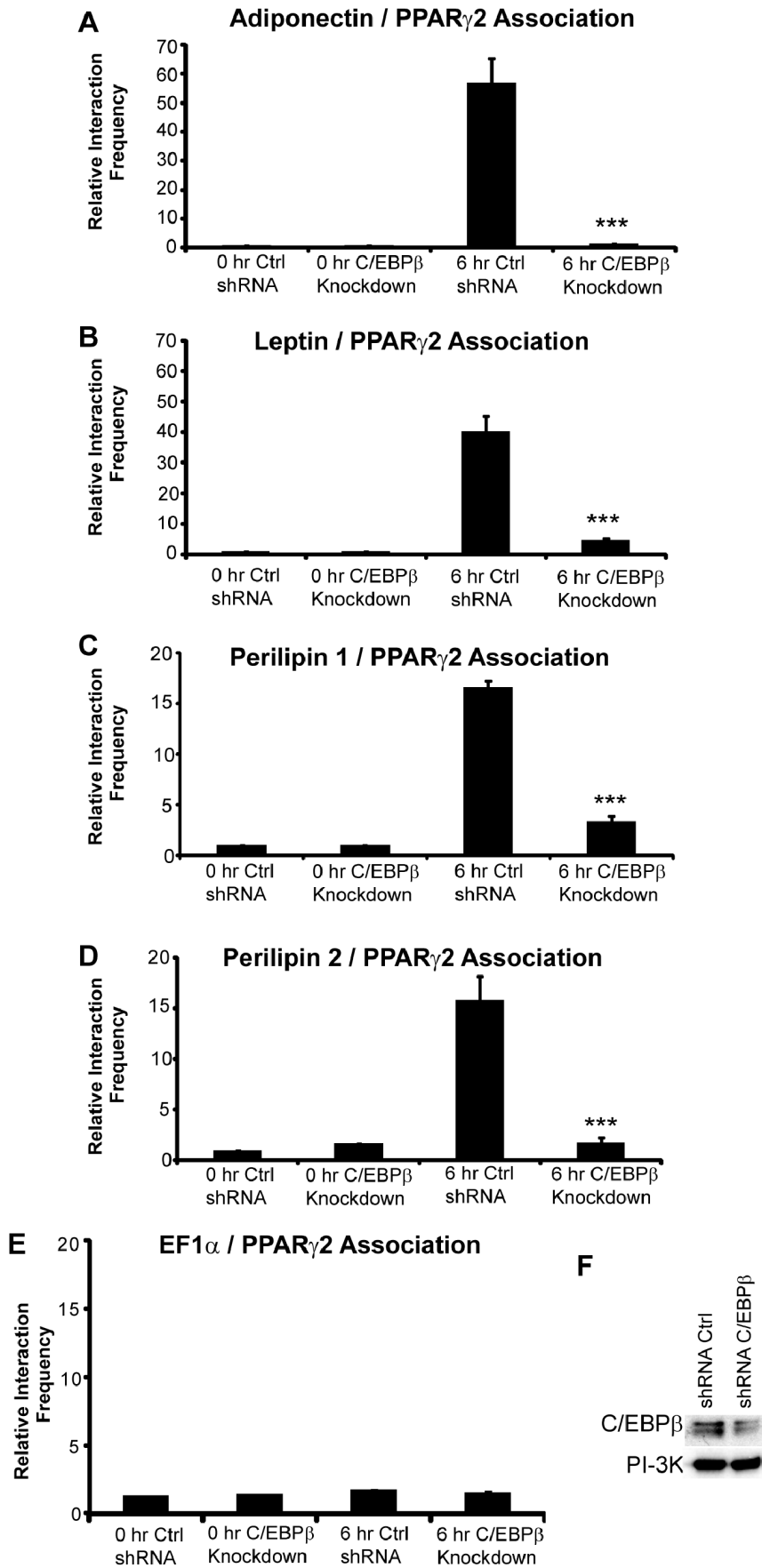


Figure 3. C/EBP β is required for the formation of long-range, inter-genic interactions between the PPAR γ 2 promoter and the indicated promoters. Cells exposed to control shRNA or shRNA targeting C/EBP β were assayed at 0 h and 6 h post-differentiation. Interaction frequencies between the PPAR γ 2 promoter and (A) the adiponectin promoter, (B) the leptin promoter, (C) the perilipin 1 promoter, (D) the perilipin 2/ADRP promoter, or (E) the EF1 α promoter are presented relative to the intragenic interaction frequency observed between different regions of the TFIID locus, which served as a normalization control. The interaction frequency at 0 h in the control shRNA treated cells was set to 1; the interaction frequencies for the other samples are presented relative to that value. Results indicate the average of three independent experiments \pm standard deviation. Values for the 6 h control shRNA treated samples were compared to values for the samples treated with C/EBP β shRNA by one-tailed t-test. *** $p < 0.001$. (F) Western blots indicating C/EBP β protein levels in control and C/EBP β shRNA treated cells. PI-3 kinase levels were monitored as a loading control.
doi:10.1371/journal.pone.0086140.g003

adipokines, leptin and adiponectin, and those regulating the expression of two genes involved in lipid metabolism and storage, perilipin 1 and perilipin 2/ADRP. We also decided to test for interactions between the PPAR γ 2 promoter and the C/EBP α promoter, which is not bound by PPAR γ 2 but is regulated by PPAR γ 2 protein via displacement of co-repressor proteins [42]. Finally we examined interactions between the PPAR γ 2 promoter and the promoter of the EF1 α gene, which is a constitutively active housekeeping gene encoding the eukaryotic translation initiation factor 1 alpha 1. The 3C assay is limited by the choice of restriction sites within range of the promoter, so we took advantage of a StuI restriction site within 150 bp of the transcription start site (TSS) of PPAR γ 2 and thus could assess interactions between genomic sites cut with Stu I and with PvuII, another blunt end cutter. The PPAR γ locus is in band 6qE3 whereas the leptin locus is situated at 6qA3.3 on chromosome 6, 85 Mbp away. The adiponectin locus is located on chromosome 16, while the perilipin 1 and perilipin 2/ADRP loci are located at chromosomes 7 (7qD2) and 4, respectively. The C/EBP α and the EF1 α loci are located on chromosomes 7 (7qB1) and 9, respectively (Fig. 1A). Thus any interactions observed would be long-range within the same chromosome or would be inter-chromosomal.

Initial experiments were performed as a time course using 2-day confluent 3T3-L1 preadipocytes (time 0) and cells treated with the standard adipogenic differentiation cocktail of insulin, dexamethasone, and IBMX for the indicated number of hours. Interaction frequencies were normalized to the interaction frequencies observed in each sample for TFIID locus interactions, and were presented as relative values with the frequency observed at time 0 set to 1. The results indicate a spike in interaction frequency between the PPAR γ 2 promoter and the adiponectin promoter at 6 hours post-differentiation, which precedes PPAR γ 2 gene expression, with interaction frequencies at 24 and 48 hours greatly reduced in comparison to the 6 hour timepoint (Fig. 1B). Nearly identical results were observed for the interaction frequency between the PPAR γ 2 promoter and the leptin promoter (Fig. 1C). Interaction frequencies between the PPAR γ 2 promoter and the perilipin 1 and perilipin 2/ADRP promoters showed a similar result (Fig. 1D–E), though the relative interaction frequencies for these promoters was reduced by \sim 40–60%. In contrast, interactions between the PPAR γ 2 and C/EBP α promoters were minimal (Fig. 1F). Interactions between the PPAR γ 2 promoter and the constitutively active EF1 α promoter were also at background level (Fig. 1G). The temporal nature of the interactions between the PPAR γ 2 promoter and the two adipokine and the two lipid droplet associated protein gene promoters, along with the lack of interactions between the PPAR γ 2 promoter and the C/EBP α or EF1 α promoters, indicates a level of specificity for the observed long-range genomic interactions.

The data in Figure 1 and in all subsequent figures were normalized to interaction frequencies detected by 3C at the TFIID locus. The TFIID locus was chosen to control for interactions because it exhibits a specific three-dimensional structure that

appears to be invariant between different cell types and under different conditions. Thus the interaction frequency between sequences from different parts of the TFIID locus are expected to be equivalent in all samples and can be used to control for sample to sample variation, as previously demonstrated [35]. As an additional control, we normalized the data in Figure 1 to the interaction frequency determined by 3C analysis of a gene desert region on chromosome 3, as done by others [36]. The data obtained by normalization to the interaction frequencies detected at the gene desert region is indistinguishable from the data normalized to TFIID locus interactions (Fig. S1).

To confirm the results, we utilized a different cellular model for adipocyte differentiation. C3H10T1/2 cells are mesenchymal precursor cells that are capable of differentiating along the adipocyte lineage in response to the same cocktail used to induce 3T3-L1 cell differentiation [43]. The promoters for the adipokines as well as the perilipin genes associated with the PPAR γ 2 promoter maximally at the 6 hr timepoint (Fig. 2A–D), while interactions with the C/EBP α and EF1 α promoters were minimal (Fig. 2E–F). The relative interaction frequencies between interacting promoters in the differentiating C3H10T1/2 cells were lower than those observed in the 3T3-L1 cells. The reason(s) for these differences is not known, though it is unrelated to the efficiency of differentiation, as Oil Red O staining of plates of both cell types maintained in differentiation conditions for 4 days showed equivalent levels of Oil Red O stained cells (data not shown).

Long-range Inter-genic Interactions Involving the PPAR γ 2 Promoter Require C/EBP β

C/EBP β is a critical regulator of adipogenesis, factoring into the activation of both C/EBP α and PPAR γ 2 gene expression, which are essential for adipogenic differentiation [44,45]. Since C/EBP β binds to the PPAR γ 2 promoter within the first 2 hours of differentiation and is required to maintain open chromatin structure at the PPAR γ 2 promoter [23,46], we asked whether C/EBP β was similarly required for the long-range chromatin interactions involving the PPAR γ 2 locus at 6 h post-differentiation. 3T3-L1 cells were infected with retrovirus expressing a control shRNA or shRNA previously shown to reduce C/EBP β levels [23] and subsequently differentiated for 6 h. As indicated in Fig. 3A–E, the interaction frequency for PPAR γ 2 with each of its interacting loci was almost completely eliminated. Control western blots demonstrated that C/EBP β protein levels were reduced in cells expressing the shRNA against C/EBP β (Fig. 3F). These results suggest that C/EBP β and the initial events that occur in the first few hours of PPAR γ 2-driven gene activation during adipogenesis are required for subsequent formation of long-range chromatin interactions involving the PPAR γ 2 locus.

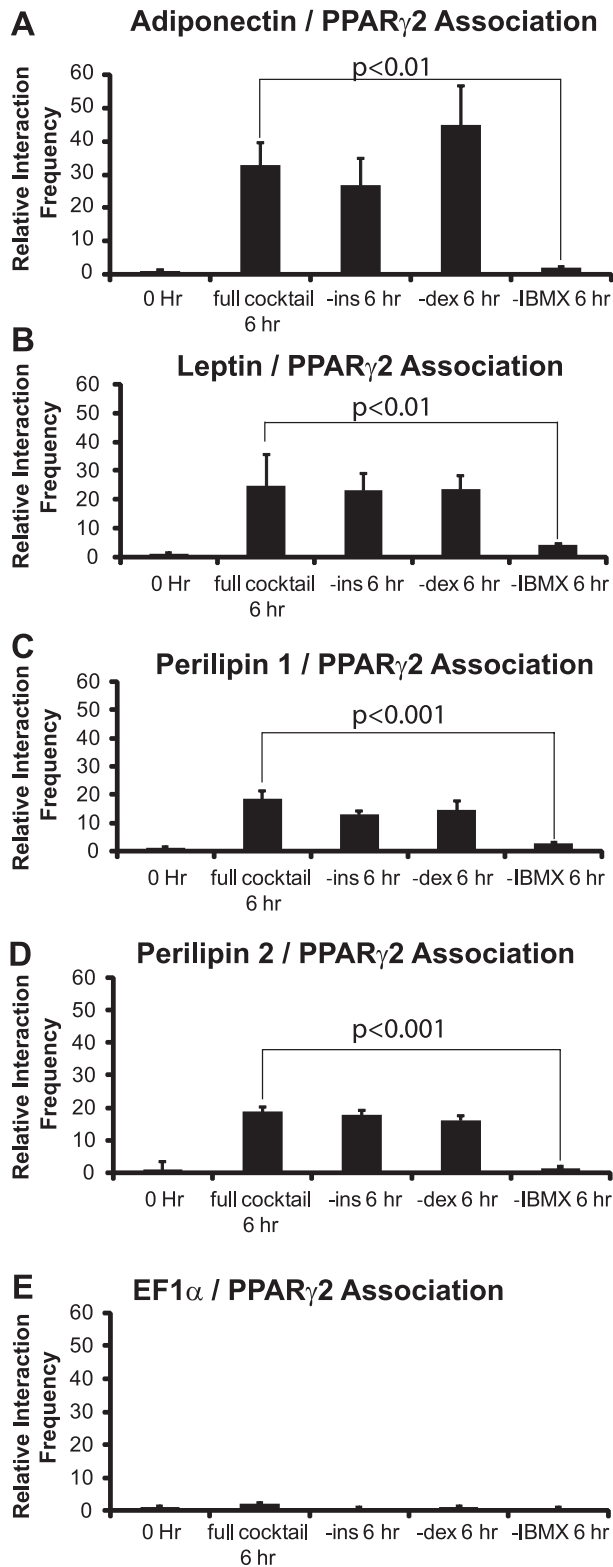


Figure 4. Elevation of cAMP levels is required for the formation of long-range, inter-genic interactions between the PPAR γ 2 promoter and the indicated promoters. The EF1 α promoter is shown as a control. 3C assays were performed on 3T3-L1 cells at 0 or at 6 h post-differentiation in the presence of complete differentiation cocktail or in cocktail lacking the indicated component. Interaction frequencies were quantified as described in the legends for Figures 1–2. Results indicate the average of three independent experiments \pm standard deviation.

standard deviation. Values for the 6 h samples differentiated in complete cocktail were compared to values for the samples differentiated in cocktail lacking IBMX by one-tailed t-test. p values are indicated on the graph. ins; insulin. dex; dexamethasone. doi:10.1371/journal.pone.0086140.g004

Long-range Inter-genic Interactions Involving the PPAR γ 2 Promoter Require PKA Signaling

The cocktail used to induce differentiation contains activators of different signaling pathways. These include dexamethasone, a synthetic glucocorticoid agonist, IBMX, a phosphodiesterase inhibitor that elevates cellular cAMP levels, and insulin, a well-known modulator of glucose uptake. To determine whether one or more of these signaling molecules were required for the formation of interactions between the PPAR γ 2 promoter and the other promoters, we performed a series of dropout experiments where a series of differentiation cocktails, each lacking one component, were used to stimulate differentiation. Cocktails lacking dexamethasone or insulin had no effect on the interaction frequencies observed between the PPAR γ 2 promoter and the adipokine and the perilipin promoters 6 hours post-differentiation (Fig. 4). As expected under these conditions, adipogenic differentiation was inhibited (data not shown). However, removal of IBMX blocked the formation of the long-range inter- and intra-genic interactions at this time point (Fig. 4), along with differentiation into mature adipocytes.

Since IBMX was necessary for the formation of the interactions between PPAR γ 2 and the other gene promoters, we next decided to test whether IBMX was sufficient to form the interactions. 3C assays were performed on 3T3-L1 cells following 6 hours of exposure to the complete cocktail or to each of the signaling components alone. Neither insulin alone nor dexamethasone alone supported the formation of the observed long-range interactions whereas exposure to IBMX alone did support the formation of the interactions between the PPAR γ 2 and the adipokine and perilipin promoters (Fig. 5). Since IBMX elevates cAMP levels, we sought to confirm that cAMP elevation was sufficient to promote intergenic interactions. Exposure of 3T3-L1 cells to serum-containing media containing forskolin, an activator of adenylyl cyclase [47], but lacking the other differentiation cocktail components promoted interactions between PPAR γ 2 and the adipokine and perilipin promoters 6 hours later (Fig. 5). Collectively, these findings indicate that enhanced cAMP levels are critical for the formation of the long-range interactions between PPAR γ 2 and other adipogenic genes.

The cAMP-dependent protein kinase, also known as protein kinase A (PKA), is activated by elevated levels of cAMP. PKA consists of a tetrameric holoenzyme [48] that contains two catalytic subunits, C α and C β [49]. To assess the requirement for PKA in the formation of the long-range genomic interactions involving the PPAR γ 2 promoter, we inhibited PKA activity by treating cells with PKI, a competitive, synthetic peptide inhibitor of PKA derived from the active domain of the naturally occurring PKA inhibitory protein also called PKI [50], or with H89, an isoquinolinesulfonamide selected for potent and specific PKA inhibition [51]. PKA inhibition by either compound inhibited long-range interactions between PPAR γ 2 promoter and the perilipin, leptin, and adiponectin promoters (Fig. 6). These experiments demonstrate the requirement for PKA in mediating long-range chromatin interactions involving the PPAR γ 2 promoter.

To directly determine whether PKA is involved in the formation of long-range interactions between the PPAR γ 2 and PPAR γ 2-target gene promoters, we used shRNAs against either the C α or

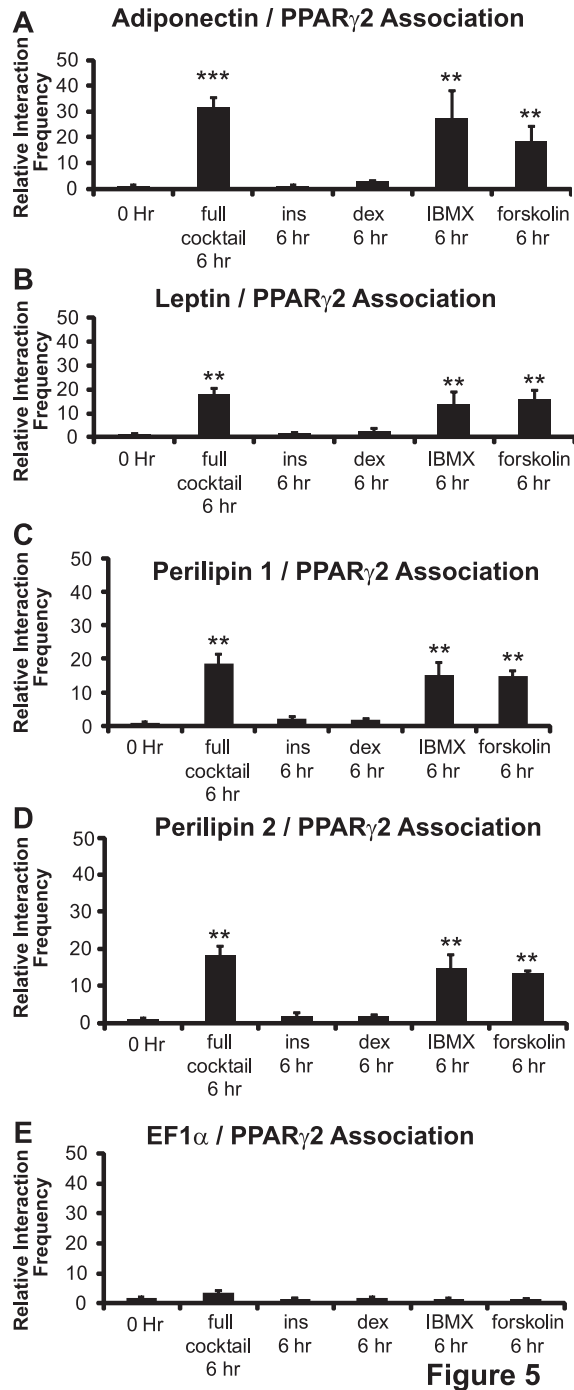


Figure 5

Figure 5. Elevation of cAMP levels is sufficient for the formation of long-range, inter-genic interactions between the PPAR γ 2 promoter and promoters. The EF1 α promoter is shown as a control. 3C assays were performed on 3T3-L1 cells at 0 or at 6 h post-differentiation in the presence of complete differentiation cocktail or in serum-containing media containing only the indicated component. Interaction frequencies were quantified as described in the legends for Figures 1–2. Results indicate the average of three independent experiments \pm standard deviation. Values for the 6 h samples differentiated in complete cocktail were compared to values for the samples differentiated for 6 h in cocktail containing serum and IBMX or serum and forskolin by one-tailed t-test. *** p <0.001; ** p <0.01. ins; insulin. dex; dexamethasone. doi:10.1371/journal.pone.0086140.g005

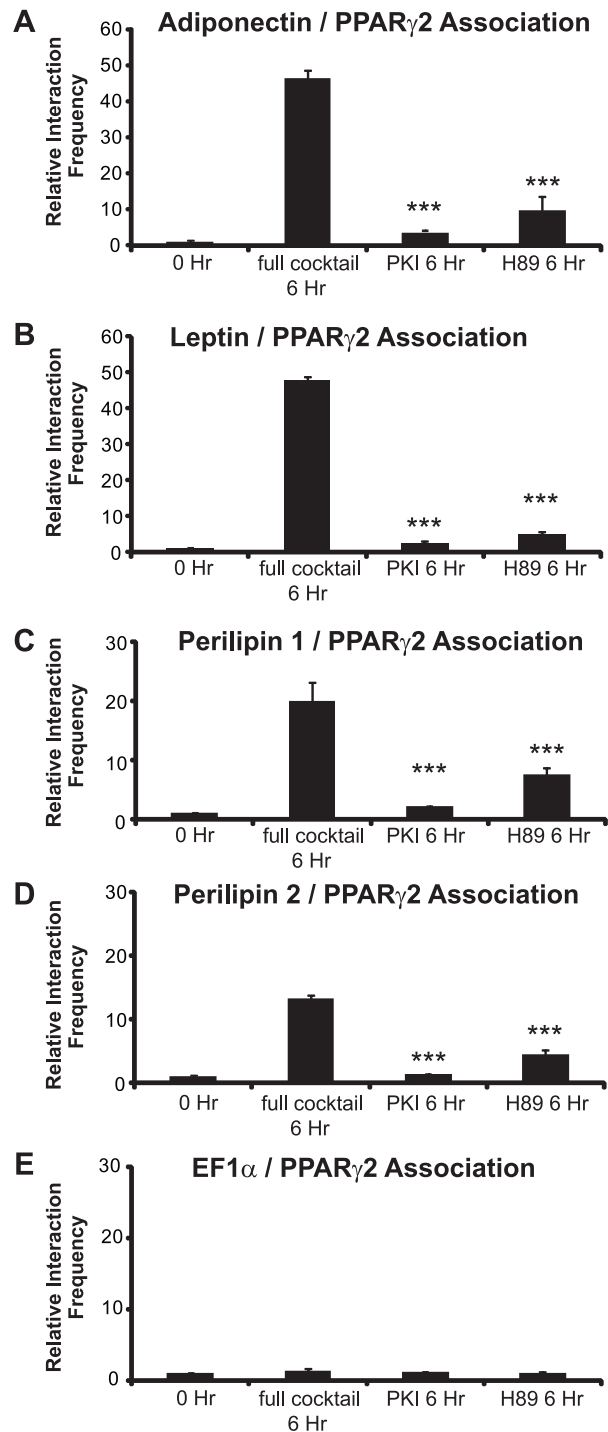


Figure 6. PKA inhibitors prevent the formation of long-range, inter-genic interactions between the PPAR γ 2 promoter and (A) the adiponectin promoter, (B) the leptin promoter, (C) the perilipin 1 promoter, (D) the perilipin 2/ADRP promoter. The EF1 α promoter (E) is shown as a control. Interaction frequencies were quantified as described in the legends for Figures 1–2. Results indicate the average of three independent experiments \pm standard deviation. Values for the 6 h samples differentiated in complete cocktail were compared to values for the samples differentiated for 6 h in complete cocktail containing the PKI or H89 inhibitors by one-tailed t-test. *** p <0.001. doi:10.1371/journal.pone.0086140.g006

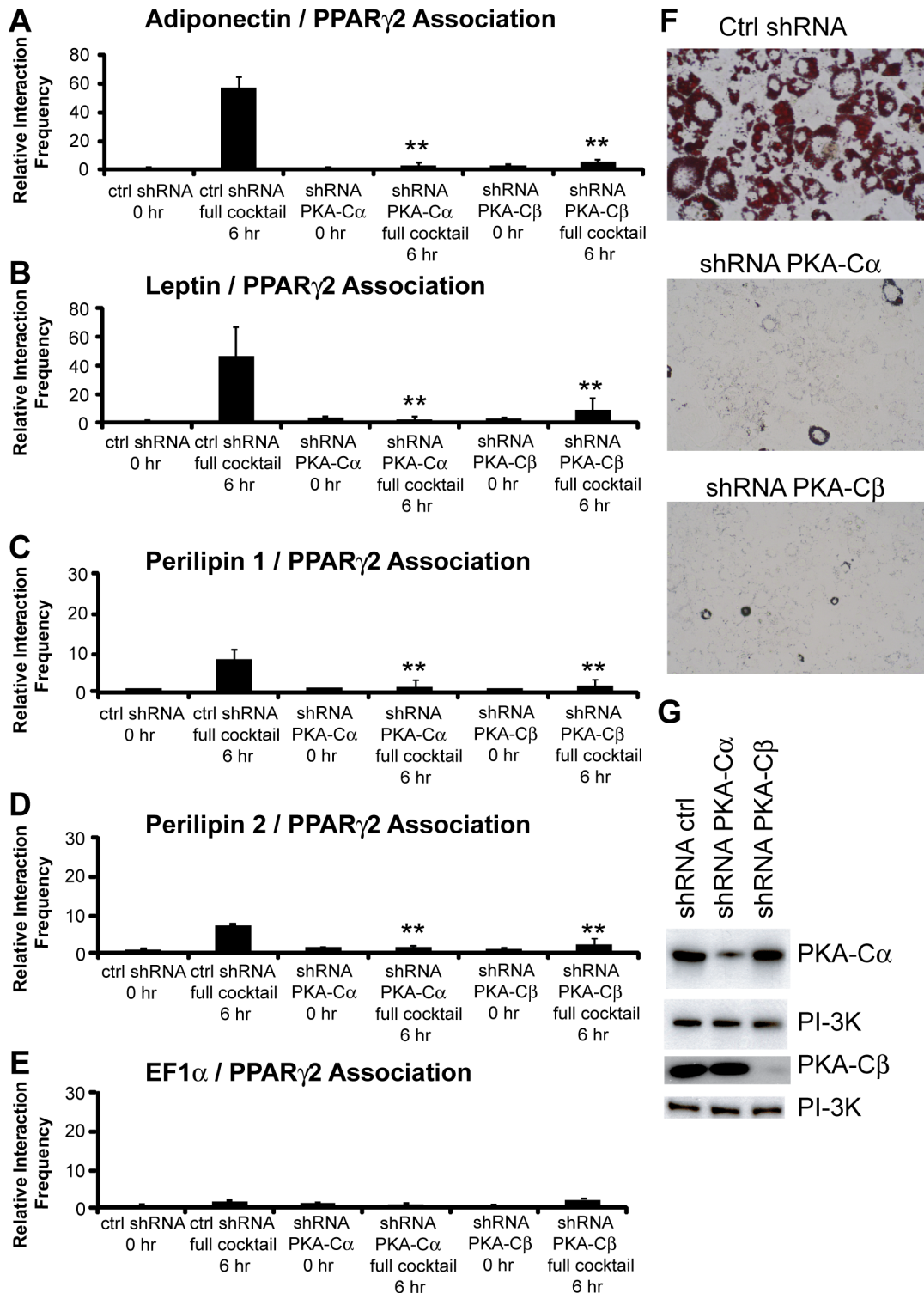


Figure 7. PKA is required for the formation of long-range, inter-genic interactions between the PPAR γ 2 promoter and (A) the adiponectin promoter, (B) the leptin promoter, (C) the perilipin 1 promoter, (D) the perilipin 2/ADRP promoter. The EF1 α promoter (E) is shown as a control. 3T3-L1 cells infected with retroviral vectors expressing control shRNA or shRNA targeting the PKA-C α or PKA-C β subunits were generated (see Methods). 3C assays were performed on these cells at 0 or at 6 h post-differentiation in the presence of complete differentiation cocktail. Interaction frequencies were quantified as described in the legends for Figures 1–2. Results indicate the average of three independent experiments \pm standard deviation. Values for the 6 h samples treated with control shRNA were compared to values for the 6 h samples treated with PKA-C α or PKA-C β shRNA by one-tailed t-test. $**p < 0.01$. (F) Oil Red O staining of infected 3T3-L1 cells allowed to differentiate for 4 days. (G) Western blots indicating the levels of the PKA-C α or PKA-C β subunits in the infected cells. PI-3K levels were monitored as a loading control. doi:10.1371/journal.pone.0086140.g007

C β subunit to reduce PKA protein levels in 3T3-L1 preadipocytes. Previously we demonstrated that knockdown of either catalytic subunit blocked 3T3-L1 cell differentiation, in part because chromatin accessibility changes at the PPAR γ 2 promoter were inhibited [23]. Knockdown of either subunit inhibited inter-chromosomal interactions between the PPAR γ 2 promoter and the adiponectin and the perilipin promoters along with the intra-chromosomal interaction between the PPAR γ 2 promoter and the leptin promoter (Figure 7A–E). Oil Red O staining confirmed that knockdown of the PKA catalytic subunit inhibited differentiation and western blotting demonstrated that the PKA catalytic subunit protein levels were, in fact, reduced (Figure 7F–G). We conclude that PKA is required for PPAR γ 2 promoter involvement in long-range interactions with other promoters.

Discussion

In this study, we provide evidence that long-range chromatin interactions occur between the promoter controlling the expression of the master regulator of adipogenesis, PPAR γ 2, and promoters controlling the expression of adipokines and lipid-droplet associated proteins in a transient manner that requires C/EBP β and protein kinase A prior to the onset of the expression of these genes. While we do not understand the functional ramifications of such associations, we speculate that they may relate to early events in adipogenesis that prepare or mark tissue-specific loci for subsequent gene activation.

Cytological evidence supports the idea of chromosomal domains in interphase cells, however, the relative positions of chromosomal domains can vary by cell type and by the activation of signaling and/or gene induction pathways [52,53]. The interfaces between chromosomal domains provide an environment in which inter-chromosomal interactions may form. Inter-chromosomal interactions have been documented previously. Such associations have often been correlated with the regulation of gene expression [54–61]. The transient nature of the observed long-range interactions suggests that if the interactions are part of the process through which gene activation is initiated, they could reflect the localization of the regulatory sequences with machinery that modifies the local chromatin structure. Obvious candidates for such machinery would be enzymes that post-translationally modify the promoter histones. Alternatively, or in addition, promoter sequences could have nucleosomal composition altered by addition or removal of histone variants, linker histones, or non-histone chromatin proteins. Recent work has suggested that deposition of the variant histone H3.3 marks skeletal muscle-specific genes for activation in myoblasts even before differentiation is initiated [62]. Precise time course experiments would be required to determine whether H3.3 deposition or the addition or removal of any histone or histone modification tracks with the formation of long-range interactions between the PPAR γ 2 and other promoters. This model hypothesizes that the enzymatic modifications occur at one or more specific sites in the nucleus. Determining whether such a site corresponds to a recognizable nuclear domain (*e.g.* – PML body, splicing speckle) or not would require immuno-FISH or similar approaches. Regardless, if fixed sites of enzymatic modification of genomic DNA exist, it would necessitate localization of the promoter sequences to that site. The transient nature of the interactions between the adipogenic gene promoters would then suggest that any chromatin modifications or structural changes that were made would be reasonably stable and not subjected to a dynamic cycle of addition/removal of the specific mark and/or histone. Whether the observed inter-genic associations relate to the formation or maintenance of transcrip-

tion factories, where multiple genes are co-localized with pol II and active transcription [63–65], requires additional investigation.

The possibility of a link between the promoter interactions between PPAR γ 2 and the selected adipokine and perilipin genes and functions related to subsequent adipocyte-specific gene expression is further complicated by the selectivity of the interactions that were observed. The relative interaction frequencies observed between PPAR γ 2 and the C/EBP α promoter were minimal and equivalent to those observed between the PPAR γ 2 and EF1 α promoters, which represent background levels. Clearly, the C/EBP α gene is activated during adipogenesis, yet unlike the other gene promoters examined, regulation of C/EBP α expression by PPAR γ 2 seems to be limited to post-transcriptional regulation in which the PPAR γ 2 protein functions to release HDAC1 from the C/EBP α promoter as part of the activation process [42]. Analyses of ChIP-seq data from 3T3-L1 cells and adipose tissue do not reveal significant binding of PPAR γ 2 or C/EBP α in the proximal promoter region of the C/EBP α gene [66,67]. Perhaps different mechanisms occur for controlling the organization and activation of the C/EBP α gene during differentiation. Alternatively, perhaps C/EBP α participates in similar interactions with other adipogenic genes, but gene associations involving the C/EBP α promoter do not overlap with interactions involving the PPAR γ 2 locus. These possibilities remain to be tested; a more genome-wide approach would likely be more productive than the candidate gene approach taken here.

Regardless of the functional consequences of these long-range interactions, their existence raises the interesting question of how genomic DNA “moves” around the nucleus. The concept of rapid repositioning of genomic sequences is controversial. There are previous reports in the literature indicating that rapid, locus-specific genomic rearrangements occur [55,68,69]. Interestingly, the movement of the loci analyzed has been linked to the activity of nuclear actin and myosin as well as to dynein light chain [55,68]. However, the reported rapid repositioning of estrogen-responsive genes upon estrogen treatment [55] has been questioned [70], and other work indicates that movement of chromosome territories is limited during interphase [71,72]. Clearly there is a need for substantial additional work to better understand the dynamics of locus-specific chromatin organization and re-organization during physiological processes such as cell differentiation.

Figure 4, Figure 5, Figure 6, Figure 7 demonstrate that formation of the long-range interactions required elevated cAMP levels and the catalytic subunits of protein kinase A. Adipogenic signaling in both the 3T3-L1 and the C3H10T1/2 cell culture models requires addition of IBMX, which is a phosphodiesterase inhibitor that elevates cellular cAMP levels, which in turn activates PKA. During the early stages of adipogenic differentiation, PKA phosphorylates CREB, the cAMP regulatory element-binding protein, which contributes to the transcriptional activation of C/EBP β [73,74], at least in part by binding to the C/EBP β promoter within an hour of the induction of differentiation [73]. At the PPAR γ 2 promoter, chromatin accessibility and c-fos binding occur within an hour of the induction of differentiation, and C/EBP β binding occurs two hours post-differentiation. All of these events are dependent on elevation of cAMP levels and PKA function [23]. An additional intriguing finding is that PKA has the potential to form a complex with dynein [75], which as previously stated, was implicated as a molecular motor that is necessary for gene-specific organizational changes in estrogen treated cells [55]. Thus a potential relationship between PKA and a factor linked to genomic organization exists that might explain the observed formation of long-range intergenic interactions between selected

adipogenic promoters. Though the connections between PKA, C/EBP β , the molecular events at the PPAR γ 2 promoter, and the involvement of the PPAR γ 2 promoter in long-range intergenic interactions require further clarification, it seems clear that signaling through PKA is intricately involved in modulating the gene organization and transcriptional potential of tissue-specific gene expression during adipocyte differentiation.

Supporting Information

Figure S1 The PPAR γ 2 promoter maximally interacts with adipokine and perilipin gene promoters in 3T3-L1 cells 6 hours post-differentiation. 3C interaction frequencies between PPAR γ 2 and the indicated promoters that were

presented in Figure 1 were re-calculated based on normalization to interaction frequencies measured in a gene desert region on chromosome 3 [36] instead of to interaction frequencies measured at the TFIID locus. Results indicate the average of three independent experiments \pm standard deviation. Values for the timepoint of maximum interaction frequency (6 h) were compared to values at time 0 by one-tailed t-test. ***** p <0.001.** (EPS)

Author Contributions

Conceived and designed the experiments: SEL HX YO ANI. Performed the experiments: SEL QW ARB HX YO. Analyzed the data: SEL ARB HX YO ANI. Wrote the paper: SEL ANI.

References

- Sidhu RS (1979) Two-dimensional electrophoretic analyses of proteins synthesized during differentiation of 3T3-L1 preadipocytes. *J Biol Chem* 254: 11111–11118.
- Spiegelman BM, Green H (1980) Control of specific protein biosynthesis during the adipose conversion of 3T3 cells. *J Biol Chem* 255: 8811–8818.
- Spiegelman BM, Farmer SR (1982) Decreases in tubulin and actin gene expression prior to morphological differentiation of 3T3 adipocytes. *Cell* 29: 53–60.
- Verstraeten VL, Renes J, Ramaekers FC, Kamps M, Kuijpers HJ, et al. (2011) Reorganization of the nuclear lamina and cytoskeleton in adipogenesis. *Histochem Cell Biol* 135: 251–261.
- Naito M, Omoteyama K, Mikami Y, Takagi M, Takahashi T (2012) Suppression of lamin A/C by short hairpin RNAs promotes adipocyte lineage commitment in mesenchymal progenitor cell line, ROB-C26. *Histochem Cell Biol* 137: 235–247.
- Tilgner K, Wojciechowski K, Jahoda C, Hutchison C, Markiewicz E (2009) Dynamic complexes of A-type lamins and emerin influence adipogenic capacity of the cell via nucleocytoplasmic distribution of beta-catenin. *J Cell Sci* 122: 401–413.
- Dorner D, Vleck S, Foeger N, Gajewski A, Makolm C, et al. (2006) Lamina-associated polypeptide 2alpha regulates cell cycle progression and differentiation via the retinoblastoma-E2F pathway. *J Cell Biol* 173: 83–93.
- Broers JL, Ramaekers FC, Bonne G, Yaou RB, Hutchison CJ (2006) Nuclear lamins: laminopathies and their role in premature ageing. *Physiol Rev* 86: 967–1008.
- Verstraeten VL, Broers JL, Ramaekers FC, van Steensel MA (2007) The nuclear envelope, a key structure in cellular integrity and gene expression. *Curr Med Chem* 14: 1231–1248.
- Kuroda M, Tanabe H, Yoshida K, Oikawa K, Saito A, et al. (2004) Alteration of chromosome positioning during adipocyte differentiation. *J Cell Sci* 117: 5897–5903.
- Kociucka B, Cieslak J, Szczerbal I (2012) Three-dimensional arrangement of genes involved in lipid metabolism in nuclei of porcine adipocytes and fibroblasts in relation to their transcription level. *Cytogenet Genome Res* 136: 295–302.
- Szczerbal I, Foster HA, Bridger JM (2009) The spatial repositioning of adipogenesis genes is correlated with their expression status in a porcine mesenchymal stem cell adipogenesis model system. *Chromosoma* 118: 647–663.
- Szczerbal I, Bridger JM (2010) Association of adipogenic genes with SC-35 domains during porcine adipogenesis. *Chromosome Res* 18: 887–895.
- Hall LL, Smith KP, Byron M, Lawrence JB (2006) Molecular anatomy of a speckle. *Anat Rec A Discov Mol Cell Evol Biol* 288: 664–675.
- Spector DL, Lamond AI (2011) Nuclear speckles. *Cold Spring Harb Perspect Biol* 3.
- Rosen ED, Walkey CJ, Puigserver P, Spiegelman BM (2000) Transcriptional regulation of adipogenesis. *Genes Dev* 14: 1293–1307.
- Rosen ED, Sarraf P, Troy AE, Bradwin G, Moore K, et al. (1999) PPAR gamma is required for the differentiation of adipose tissue in vivo and in vitro. *Mol Cell* 4: 611–617.
- He W, Barak Y, Hevener A, Olson P, Liao D, et al. (2003) Adipose-specific peroxisome proliferator-activated receptor gamma knockout causes insulin resistance in fat and liver but not in muscle. *Proc Natl Acad Sci U S A* 100: 15712–15717.
- Ntambi JM, Young-Cheul K (2000) Adipocyte differentiation and gene expression. *J Nutr* 130: 3122S–3126S.
- Lefterova MI, Lazar MA (2009) New developments in adipogenesis. *Trends Endocrinol Metab* 20: 107–114.
- Siersbaek R, Nielsen R, John S, Sung MH, Baek S, et al. (2011) Extensive chromatin remodelling and establishment of transcription factor 'hotspots' during early adipogenesis. *EMBO J* 30: 1459–1472.
- Steger DJ, Grant GR, Schupp M, Tomaru T, Lefterova MI, et al. (2010) Propagation of adipogenic signals through an epigenomic transition state. *Genes Dev* 24: 1035–1044.
- Xiao H, Leblanc SE, Wu Q, Konda S, Salma N, et al. (2011) Chromatin accessibility and transcription factor binding at the PPARgamma2 promoter during adipogenesis is protein kinase A-dependent. *J Cell Physiol* 226: 86–93.
- Tontonoz P, Spiegelman BM (2008) Fat and beyond: the diverse biology of PPARgamma. *Annu Rev Biochem* 77: 289–312.
- Friedman JM (2000) Obesity in the new millennium. *Nature* 404: 632–634.
- Kadowaki T, Yamauchi T (2005) Adiponectin and adiponectin receptors. *Endocr Rev* 26: 439–451.
- Billyard T, McTernan P, Kumar S (2007) Potential therapies based on antidiabetic peptides. *Best Pract Res Clin Endocrinol Metab* 21: 641–655.
- Falcao-Pires I, Castro-Chaves P, Miranda-Silva D, Lourenco AP, Leite-Moreira AF (2012) Physiological, pathological and potential therapeutic roles of adipokines. *Drug Discov Today* 17: 880–889.
- Paul A, Chan L, Bickel PE (2008) The PAT family of lipid droplet proteins in heart and vascular cells. *Curr Hypertens Rep* 10: 461–466.
- Lafont M, Langin D (2009) Lipolysis and lipid mobilization in human adipose tissue. *Prog Lipid Res* 48: 275–297.
- Pear WS, Nolan GP, Scott ML, Baltimore D (1993) Production of high-titer helper-free retroviruses by transient transfection. *Proc Natl Acad Sci U S A* 90: 8392–8396.
- Dekker J, Rippe K, Dekker M, Kleckner N (2002) Capturing chromosome conformation. *Science* 295: 1306–1311.
- Osborne CS, Chakalova L, Brown KE, Carter D, Horton A, et al. (2004) Active genes dynamically colocalize to shared sites of ongoing transcription. *Nat Genet* 36: 1065–1071.
- Sexton T, Kurukuti S, Mitchell JA, Umlauf D, Nagano T, et al. (2012) Sensitive detection of chromatin coassociations using enhanced chromosome conformation capture on chip. *Nat Protoc* 7: 1335–1350.
- Splinter E, Grosveld F, de Laat W (2004) 3C technology: analyzing the spatial organization of genomic loci in vivo. *Methods Enzymol* 375: 493–507.
- Kagey MH, Newman JJ, Bilodeau S, Zhan Y, Orlando DA, et al. (2010) Mediator and cohesin connect gene expression and chromatin architecture. *Nature* 467: 430–435.
- LeBlanc SE, Konda S, Wu Q, Hu YJ, Osowski CM, et al. (2012) Protein arginine methyltransferase 5 (Prmt5) promotes gene expression of peroxisome proliferator-activated receptor gamma2 (PPARgamma2) and its target genes during adipogenesis. *Mol Endocrinol* 26: 583–597.
- Tontonoz P, Hu E, Spiegelman BM (1994) Stimulation of adipogenesis in fibroblasts by PPAR gamma 2, a lipid-activated transcription factor. *Cell* 79: 1147–1156.
- de Wit E, de Laat W (2012) A decade of 3C technologies: insights into nuclear organization. *Genes Dev* 26: 11–24.
- Cavalli G, Misteli T (2013) Functional implications of genome topology. *Nat Struct Mol Biol* 20: 290–299.
- Cope NF, Fraser P, Eskiw CH (2010) The yin and yang of chromatin spatial organization. *Genome Biol* 11: 204.
- Zuo Y, Qiang L, Farmer SR (2006) Activation of CCAAT/enhancer-binding protein (C/EBP) alpha expression by C/EBP beta during adipogenesis requires a peroxisome proliferator-activated receptor-gamma-associated repression of HDAC1 at the C/ebp alpha gene promoter. *J Biol Chem* 281: 7960–7967.
- Cho YC, Jefcoate CR (2004) PPARgamma1 synthesis and adipogenesis in C3H10T1/2 cells depends on S-phase progression, but does not require mitotic clonal expansion. *J Cell Biochem* 91: 336–353.
- Tang QQ, Lane MD (2012) Adipogenesis: from stem cell to adipocyte. *Annu Rev Biochem* 81: 715–736.
- Farmer SR (2005) Regulation of PPARgamma activity during adipogenesis. *Int J Obes (Lond)* 29 Suppl 1: S13–16.
- Salma N, Xiao H, Imbalzano AN (2006) Temporal recruitment of CCAAT/enhancer-binding proteins to early and late adipogenic promoters in vivo. *J Mol Endocrinol* 36: 139–151.
- Russell TR, Ho R (1976) Conversion of 3T3 fibroblasts into adipose cells: triggering of differentiation by prostaglandin F2alpha and 1-methyl-3-isobutyl xanthine. *Proc Natl Acad Sci U S A* 73: 4516–4520.

48. Cheng X, Ji Z, Tsalkova T, Mei F (2008) Epac and PKA: a tale of two intracellular cAMP receptors. *Acta Biochim Biophys Sin (Shanghai)* 40: 651–662.
49. Sugiyama H, Chen P, Hunter MG, Sitkovsky MV (1997) Perturbation of the expression of the catalytic subunit C alpha of cyclic AMP-dependent protein kinase inhibits TCR-triggered secretion of IL-2 by T helper hybridoma cells. *J Immunol* 158: 171–179.
50. Cheng HC, Kemp BE, Pearson RB, Smith AJ, Misconi L, et al. (1986) A potent synthetic peptide inhibitor of the cAMP-dependent protein kinase. *J Biol Chem* 261: 989–992.
51. Chijiwa T, Mishima A, Hagiwara M, Sano M, Hayashi K, et al. (1990) Inhibition of forskolin-induced neurite outgrowth and protein phosphorylation by a newly synthesized selective inhibitor of cyclic AMP-dependent protein kinase, N-[2-(p-bromocinnamylamino)ethyl]-5-isoquinolinesulfonamide (H-89), of PC12D pheochromocytoma cells. *J Biol Chem* 265: 5267–5272.
52. Geyer PK, Vitalini MW, Wallrath LL (2011) Nuclear organization: taking a position on gene expression. *Curr Opin Cell Biol* 23: 354–359.
53. Cremer T, Cremer M (2010) Chromosome territories. *Cold Spring Harb Perspect Biol* 2: a003889.
54. Apostolou E, Thanos D (2008) Virus Infection Induces NF-kappaB-dependent interchromosomal associations mediating monoallelic IFN-beta gene expression. *Cell* 134: 85–96.
55. Hu Q, Kwon YS, Nunez E, Cardamone MD, Hutt KR, et al. (2008) Enhancing nuclear receptor-induced transcription requires nuclear motor and LSD1-dependent gene networking in interchromatin granules. *Proc Natl Acad Sci U S A* 105: 19199–19204.
56. Lomvardas S, Barnea G, Pisapia DJ, Mendelsohn M, Kirkland J, et al. (2006) Interchromosomal interactions and olfactory receptor choice. *Cell* 126: 403–413.
57. Simonis M, Klous P, Splinter E, Moshkin Y, Willemsen R, et al. (2006) Nuclear organization of active and inactive chromatin domains uncovered by chromosome conformation capture-on-chip (4C). *Nat Genet* 38: 1348–1354.
58. Schoenfelder S, Sexton T, Chakalova L, Cope NF, Horton A, et al. (2010) Preferential associations between co-regulated genes reveal a transcriptional interactome in erythroid cells. *Nat Genet* 42: 53–61.
59. Fullwood MJ, Liu MH, Pan YF, Liu J, Xu H, et al. (2009) An oestrogen-receptor-alpha-bound human chromatin interactome. *Nature* 462: 58–64.
60. Handoko L, Xu H, Li G, Ngan CY, Chew E, et al. (2011) CTCF-mediated functional chromatin interactome in pluripotent cells. *Nat Genet* 43: 630–638.
61. Li G, Ruan X, Auerbach RK, Sandhu KS, Zheng M, et al. (2012) Extensive promoter-centered chromatin interactions provide a topological basis for transcription regulation. *Cell* 148: 84–98.
62. Harada A, Okada S, Konno D, Odawara J, Yoshimi T, et al. (2012) Chd2 interacts with H3.3 to determine myogenic cell fate. *EMBO J* 31: 2994–3007.
63. Sutherland H, Bickmore WA (2009) Transcription factories: gene expression in unions? *Nat Rev Genet* 10: 457–466.
64. Papanonis A, Cook PR (2011) Fixing the model for transcription: the DNA moves, not the polymerase. *Transcription* 2: 41–44.
65. Cook PR (2010) A model for all genomes: the role of transcription factories. *J Mol Biol* 395: 1–10.
66. Schmidt SF, Jorgensen M, Chen Y, Nielsen R, Sandelin A, et al. (2011) Cross species comparison of C/EBPalpha and PPARgamma profiles in mouse and human adipocytes reveals interdependent retention of binding sites. *BMC Genomics* 12: 152.
67. Lefterova MI, Steger DJ, Zhuo D, Qatanani M, Mullican SE, et al. (2010) Cell-specific determinants of peroxisome proliferator-activated receptor gamma function in adipocytes and macrophages. *Mol Cell Biol* 30: 2078–2089.
68. Chuang CH, Carpenter AE, Fuchsova B, Johnson T, de Lanerolle P, et al. (2006) Long-range directional movement of an interphase chromosome site. *Curr Biol* 16: 825–831.
69. Fraser P, Bickmore W (2007) Nuclear organization of the genome and the potential for gene regulation. *Nature* 447: 413–417.
70. Kocanova S, Buytaert E, Matroule JY, Piette J, Golab J, et al. (2007) Induction of heme-oxygenase 1 requires the p38MAPK and PI3K pathways and suppresses apoptotic cell death following hypericin-mediated photodynamic therapy. *Apoptosis* 12: 731–741.
71. Chubb JR, Boyle S, Perry P, Bickmore WA (2002) Chromatin motion is constrained by association with nuclear compartments in human cells. *Curr Biol* 12: 439–445.
72. Walter J, Schermelleh L, Cremer M, Tashiro S, Cremer T (2003) Chromosome order in HeLa cells changes during mitosis and early G1, but is stably maintained during subsequent interphase stages. *J Cell Biol* 160: 685–697.
73. Zhang JW, Klemm DJ, Vinson C, Lane MD (2004) Role of CREB in transcriptional regulation of CCAAT/enhancer-binding protein beta gene during adipogenesis. *J Biol Chem* 279: 4471–4478.
74. Reusch JE, Colton LA, Klemm DJ (2000) CREB activation induces adipogenesis in 3T3-L1 cells. *Mol Cell Biol* 20: 1008–1020.
75. Kashina AS, Semenova IV, Ivanov PA, Potekhina ES, Zaliapin I, et al. (2004) Protein kinase A, which regulates intracellular transport, forms complexes with molecular motors on organelles. *Curr Biol* 14: 1877–1881.