

# Dietary Omega-3 Polyunsaturated Fatty Acids Alter the Fatty Acid Composition of Hepatic and Plasma Bioactive Lipids in C57BL/6 Mice: A Lipidomic Approach

Kayode A. Balogun<sup>1</sup>, Carolyn J. Albert<sup>2</sup>, David A. Ford<sup>2</sup>, Robert J. Brown<sup>1</sup>, Sukhinder K. Cheema<sup>1\*</sup>

<sup>1</sup> Department of Biochemistry, Memorial University, St. John's, NL, Canada, <sup>2</sup> Department of Biochemistry and Molecular Biology, and Center for Cardiovascular Research, Saint Louis University, St. Louis, Missouri, United States of America

## Abstract

**Background:** Omega (n)-3 polyunsaturated fatty acids (PUFA) are converted to bioactive lipid components that are important mediators in metabolic and physiological pathways; however, which bioactive compounds are metabolically active, and their mechanisms of action are still not clear. We investigated using lipidomic techniques, the effects of diets high in n-3 PUFA on the fatty acid composition of various bioactive lipids in plasma and liver.

**Methodology and Principal Findings:** Female C57BL/6 mice were fed semi-purified diets (20% w/w fat) containing varying amounts of n-3 PUFA before mating, during gestation and lactation, and until weaning. Male offspring were continued on their mothers' diets for 16 weeks. Hepatic and plasma lipids were extracted in the presence of non-naturally occurring internal standards, and tandem electrospray ionization mass spectrometry methods were used to measure the fatty acyl compositions. There was no significant difference in total concentrations of phospholipids in both groups. However, there was a significantly higher concentration of eicosapentaenoic acid containing phosphatidylcholine (PC), lysophosphatidylcholine (LPC), and cholesteryl esters (CE) ( $p < 0.01$ ) in the high n-3 PUFA group compared to the low n-3 PUFA group in both liver and plasma. Plasma and liver from the high n-3 PUFA group also had a higher concentration of free n-3 PUFA ( $p < 0.05$ ). There were no significant differences in plasma concentrations of different fatty acyl species of phosphatidylethanolamine, triglycerides, sphingomyelin and ceramides.

**Conclusions/Significance:** Our findings reveal for the first time that a diet high in n-3 PUFA caused enrichment of n-3 PUFA in PC, LPC, CE and free fatty acids in the plasma and liver of C57BL/6 mice. PC, LPC, and unesterified free n-3 PUFA are important bioactive lipids, thus altering their fatty acyl composition will have important metabolic and physiological roles.

**Citation:** Balogun KA, Albert CJ, Ford DA, Brown RJ, Cheema SK (2013) Dietary Omega-3 Polyunsaturated Fatty Acids Alter the Fatty Acid Composition of Hepatic and Plasma Bioactive Lipids in C57BL/6 Mice: A Lipidomic Approach. PLoS ONE 8(11): e82399. doi:10.1371/journal.pone.0082399

**Editor:** Michael Müller, Wageningen University, Netherlands

**Received:** June 3, 2013; **Accepted:** November 1, 2013; **Published:** November 21, 2013

**Copyright:** © 2013 Balogun et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

**Funding:** This research was supported by funds from the Natural Sciences and Engineering Research Council (SKC), an Ignite R&D grant from the Research & Development Corporation of Newfoundland and Labrador (RJB), and National Institutes of Health Grants HL-074214 and HL-111906 (DAF). The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

**Competing interests:** The authors have declared that no competing interests exist.

\* E-mail: skaur@mun.ca

## Introduction

Essential polyunsaturated fatty acids (PUFA) of the omega-3 (n-3) and n-6 classes are important in the regulation of metabolic processes. N-3 PUFA such as docosahexaenoic acid (DHA; 22:6) and eicosapentaenoic acid (EPA; 20:5) have attracted a lot of attention in the past years as a result of their potential health benefits [1-3]. N-3 PUFA has been shown to prevent atherosclerosis [4,5], regulate nuclear transcription factors involved in gene expression of inflammatory markers, and stimulate cognitive development [6,7]. Markers of cardiovascular disease such as high triglycerides (TG),

endothelial dysfunction, cardiac arrhythmia, and inflammation are also reduced by the administration of n-3 PUFA [1,8-11].

*In vivo*, n-3 PUFA can either exist as free products of enzyme hydrolysis or bound to phospholipids (PL) or TG. Reports have shown that dietary n-3 PUFA are preferentially incorporated into PL compared to TG [12]. PL are important constituents of the cellular membrane bilayer, with phosphatidylcholine (PC) and phosphatidylethanolamine (PE) being the most abundant [13]. Membrane fluidity can be influenced by the incorporation of dietary long chain n-3 PUFA into membrane PL, which enhances the functions of transmembrane proteins and their interactions with

extracellular ligands [14–16]. This indirectly affects signalling pathways and other physiological functions of the membrane. PC has been implicated in an array of physiological functions, and their functional properties depend on their fatty acyl chains.

The liver is principally involved in the metabolism and release of PC into circulation. PC is metabolized by the enzyme phospholipase A<sub>2</sub> releasing the fatty acid at the *sn*-2 position, and lysophosphatidylcholine (LPC) into the plasma pool for distribution to extrahepatic tissues [17,18]. Upon hydrolysis of the *sn*-2 fatty acid of PC, the released free fatty acids (FFA) are further metabolized to form bioactive compounds with either pro-inflammatory or anti-inflammatory properties [18]. Arachidonic acid (AA; 20:4), a long chain *n*-6 PUFA released from the metabolism of PC, is further metabolized by the cyclooxygenase or lipoxygenase enzyme pathways to produce the inflammatory series-2 prostaglandins or series-4 leukotrienes, respectively [19]. On the contrary, the cyclooxygenase and lipoxygenase metabolic products of EPA, an *n*-3 PUFA, are generally anti-inflammatory [20]. Furthermore, protectins and resolvins, the products of oxygenation of DHA and EPA, have been reported to exhibit anti-inflammatory properties [21–23].

LPC, the other product of enzyme hydrolysis of PC, is an important lipid mediator involved in cellular metabolism. Direct hepatic secretion is an important source of the abundant unsaturated LPC found in the plasma [24]. LPC has been controversially linked with the development of atherosclerosis [25–27]. However, there are burgeoning evidences suggesting that the biological properties of LPC depend on the acyl chain of the molecule [28,29]. Studies have reported an increase in plasma concentration of saturated LPC in diseases conditions such as obesity, diabetes and rheumatoid arthritis [30]. In another line of evidence, polyunsaturated LPC significantly reversed saturated LPC induced inflammation [31]. However, the importance of specific LPC fatty acyl chain in metabolic regulation is still not clear.

Most studies to date have explored the health benefits of *n*-3 PUFA esterified to ethyl esters or TG [32–34]. Moreover, the majority of the reported findings that support the pro-inflammatory and the atherogenic actions of LPC were established using saturated or monounsaturated LPC [25,26,35–37]. Cells, tissues, and biological fluids consist of numerous bioactive lipid mediators involved in cellular processes, which are likely altered by dietary *n*-3 PUFA. The aim of this study was to employ high throughput lipidomic techniques to evaluate the effect of diets high or low in *n*-3 PUFA content on the fatty acid composition of various lipids in the plasma and liver of C57BL/6 mice. Our findings reveal for the first time that diets high in *n*-3 PUFA caused enrichment of *n*-3 PUFA in PC, LPC, cholesteryl esters (CE) and FFA in the plasma and liver of C57BL/6 mice, which will likely have important physiological roles.

## Materials and Methods

### Ethics statement

All experimental procedures were approved by Memorial University Animal Care Committee in accordance with the

principles and guidelines of Canadian Council on Animal Care (approval no: 10-09-SK).

### Animals and diets

Seven week old male and female C57BL/6 mice were purchased from Charles River Laboratories (MA, USA), and were housed in separate cages under controlled temperature (21± 1°C) and humidity (35 ± 5%) conditions with a 12-hour light/12-hour dark period cycle. Mice were kept on standard rodent chow pellets (Prolab RMH 3000) purchased from PMI nutrition (MO, USA) for one week acclimatization period. After this period, female mice were randomly divided into two groups. Each group was fed either of the two experimental diets that differed only in their *n*-3 PUFA composition, and designated as “high *n*-3” and “low *n*-3” diets for two weeks before mating. The experimental diets were made from a base semi-synthetic diet (MP Biomedicals, OH, USA), which allows for the control of fat level at 20% w/w. Menhaden oil was obtained from Sigma-Aldrich (MO, USA); lard, safflower oil and extra-virgin olive oil were used to prepare two different oil mixtures containing 10% (high *n*-3) and 2% (low *n*-3) *n*-3 PUFA of the total fat, giving an *n*-6 to *n*-3 PUFA ratio of 5:1 and 30:1 respectively. The fatty acid composition of the diets was analyzed by gas chromatography as per our previous publication [38] and is given in Table 1. Females were continued on the experimental diets throughout gestation, lactation, and until weaning. Just before weaning, breast milk samples were collected from the mothers, to ascertain that the dietary essential fatty acid is reflected in the breast milk. There was a positive correlation between dietary and breast milk fatty acids composition (data not shown). At weaning, male offspring (*n*=6) were continued on their mothers’ designated diet for 16 weeks (i.e. offspring obtained from mother’s fed a high *n*-3 PUFA diet continued on high *n*-3 PUFA diet). Only male offspring were used in this study in order to prevent hormonal interference. Female sex hormones have been shown to affect tissue plasma and tissue *n*-3 PUFA content [39]; and it is well known that there is gender specificity in lipid metabolism, possibly due to the effect of hormonal differences [39–42]. Animals were provided with water and fresh food *ad libitum*, every other day. Body weights were recorded once a week, and food intake was recorded every other day. No significant differences were observed in both body weight and food intake (Table S1 in File S1). At 16 weeks, male offspring were fasted overnight and sacrificed using isoflurane. Blood was collected by cardiac puncture in tubes containing EDTA (4.5 mM, pH 7.4), and plasma was separated immediately. Tissues were removed and weighed at the time of sacrifice, snap frozen in liquid nitrogen and stored at -80°C until further analyses.

### Lipidomic analysis

**Lipid extraction, standards, and solvents.** Plasma or liver samples were flash frozen at the temperature of liquid nitrogen at collection. 10 µl plasma was directly extracted into organic solvent, while 100 mg liver tissue was pulverized and homogenized and then immediately lipids were extracted into organic solvent. Lipid extraction into organic solvent was performed using the Bligh-Dyer method [43] with high

**Table 1.** Fatty acid composition of the experimental diets.

Fatty Acid	High <i>n</i> -3	Low <i>n</i> -3
C14:0	1.26	0.11
C16:0	8.71	6.32
C18:0	2.67	5.35
<b>ΣSFA</b>	12.64	11.77
C16:1n7	2.41	0.36
C18:1n9 + C18:1n7	25.14	27.82
C20:1n9	0.61	ND
<b>ΣMUFA</b>	28.16	28.18
C18:2n6	47.86	57.73
C20:4n6	0.23	0.11
C18:3n6	0.10	0.04
C22:4n6	0.54	0.09
<b>ΣOmega-6</b>	48.90	57.92
C18:3n3	0.78	0.55
C20:5n3	3.64	0.31
C22:6n3	3.19	0.39
C18:4n3	0.87	0.15
C22:5n3	0.63	0.46
C20:4n3	0.66	0.08
<b>ΣOmega-3</b>	9.76	1.93
<b>ΣPUFA</b>	59.38	60.00
<b>ΣOmega-6/Omega-3</b>	5.00	30.01

Lipids were extracted from the diets and the fatty acid composition was determined by gas chromatography. Values are given as % area of each fatty acid peak. Abbreviations: ND= Not detected, ΣSFA= sum of saturated fatty acids, ΣMUFA= sum of monounsaturated fatty acids, Σ PUFA= sum of polyunsaturated fatty acids, ΣOmega-6= sum of omega-6 fatty acids, ΣOmega-3= sum of omega-3 fatty acid.  
doi: 10.1371/journal.pone.0082399.t001

performance liquid chromatography-mass spectrometry grade solvents in the presence of non-naturally occurring internal lipid standards. The standards used were Δ9-trans-triheptadecenoic (tri-17:1 TG), 1, 2-diarachidoyl-*sn*-glycero-3-phosphocholine (di-20:0 PC), N-heptadecanoyl-D-*erythro*-sphingosylphosphorylcholine (17:0 sphingomyelin), 1-heptadecanoyl-2-hydroxy-*sn*-glycero-3-phosphocholine (17:0 LPC), 1, 2-dimyristoyl-*sn*-glycero-3-phosphoethanolamine (di-14:0 PE), cholesteryl heptadecanoate (17:0 CE), and N-heptadecanoyl-D-*erythro*-sphingosine (17:0 ceramide). The extracted lipids in the chloroform phase were dried down under gentle stream of N<sub>2</sub> gas, and re-suspended in 500 μl of chloroform. 50 μl aliquot of the suspension was then mixed with 200 μl methanol and 2 μl of 10 mM methanolic NaOH and injected into a Thermo Fisher TSQ Quantum Ultra tandem electrospray ionization mass spectrometry (ESI-MS) system for lipid analyses. High pressure liquid chromatography-mass spectrometry grade methanol and chloroform were used for all extractions; these solvents were purchased from Burdick and Jackson (NJ, USA).

**Electrospray ionization-mass spectrometry (ESI-MS).** Samples were analysed using direct-infusion ESI-MS in positive or negative ion mode using a Thermo Fisher TSQ Quantum Ultra system with XCalibur data acquisition software [44]. The tune parameters for sample analyses were optimized

and set as follows: spray voltage = 3500 V, flow rate = 3 μl/min, ion sweep gas pressure = 0.2 (arbitrary units), sheath gas = 12 (arbitrary units), auxiliary gas pressure = 6 (arbitrary units), and capillary temperature = 270°C. The collision energies for the analyses of PC, LPC, and sphingomyelins (SM) were set at 28 eV. The collision energies for analyses of cholesteryl esters (CE) was set at 25 eV, and for ceramide (CER) was set at 32 eV. CEs were detected in positive ion mode by scanning for neutral loss (NL) of cholestadane (*m/z* 368.5). Sodiated species of SM, PC, and LPC were detected in positive ion mode by scanning for the NL of choline (*m/z* 59.1). FFA and PE were identified in negative ion mode by survey scan for [M-H]<sup>-</sup> between *m/z* 200 and 900. Sodiaded species of TG were identified in positive ion mode by survey scan for [M+Na]<sup>+</sup> between *m/z* 800 and 1000. All data analyzed were corrected for <sup>13</sup>C isotope effects as described by Han et al. [44].

### Statistical analysis

Data were analysed using GraphPad Prism software (version 5.0). Statistical significance for differences between groups was determined by unpaired t-test. Results are expressed as mean ± standard deviation (SD). Differences were considered to be statistically significant if the associated *P* value was < 0.05.

## Results

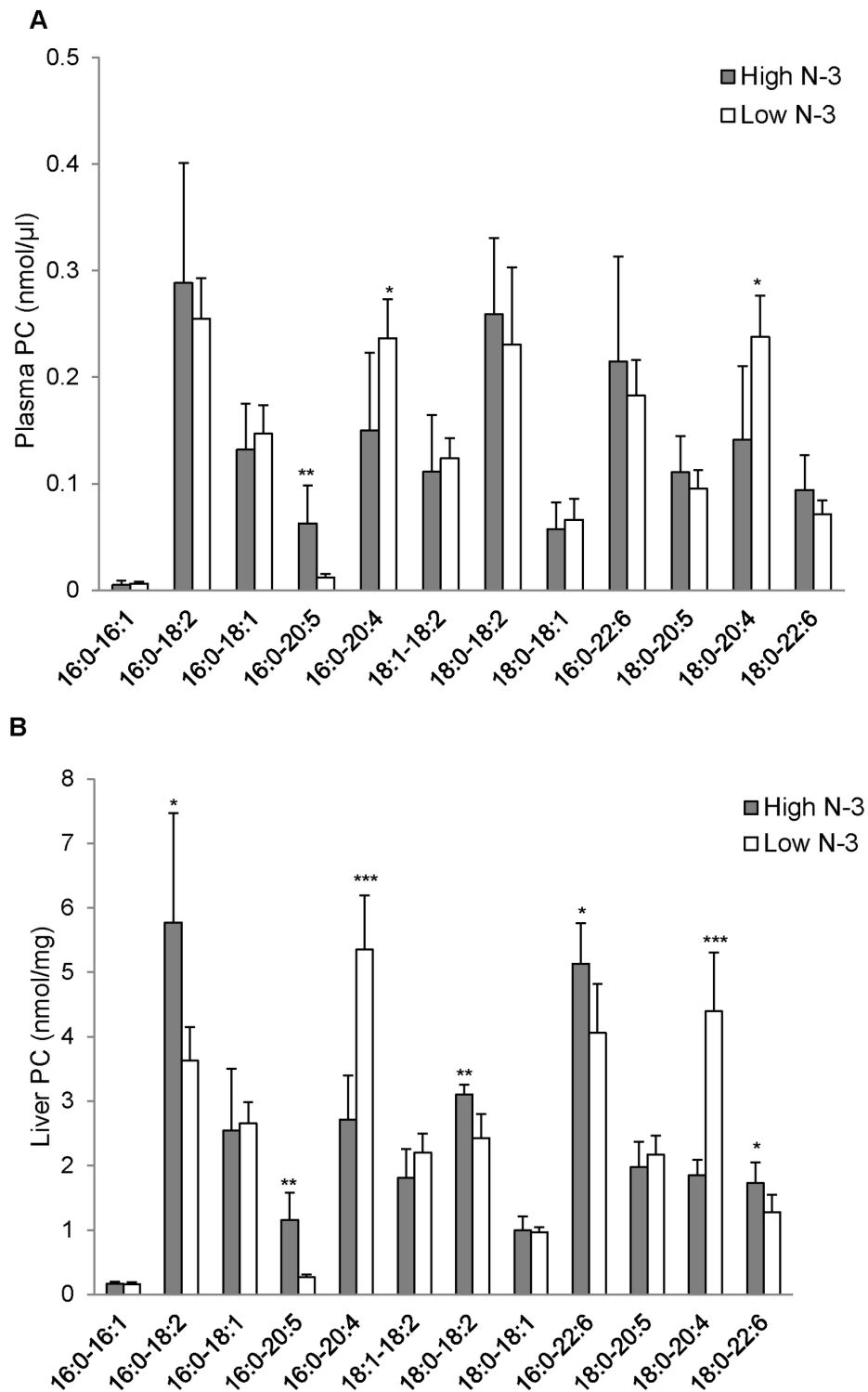
### PC fatty acyl composition of mice fed high and low *n*-3 PUFA

There were no significant differences in the total concentrations of phospholipids in both plasma and liver between high and low *n*-3 PUFA fed mice (Table S2 in File S1). However, there was a significantly higher concentration of 16:0-20:5 PC in the plasma (*p* < 0.01; Figure 1A) and liver (*p* < 0.05; Figure 1B) of the high *n*-3 PUFA group compared to the low *n*-3 PUFA group. Interestingly, 20:4 containing PC (16:0-20:4 and 18:0-20:4) increased significantly in the plasma of the low *n*-3 PUFA group compared to the high *n*-3 PUFA group (*p* < 0.05; Figure 1A). A similar effect was observed in the liver, with 16:0-20:4 and 18:0-20:4 showing a significant increase in the low *n*-3 PUFA group (*p* < 0.001; Figure 1B). This signifies a higher incorporation of EPA and a lower incorporation of AA into plasma PC in the high *n*-3 PUFA group. The high *n*-3 PUFA group also showed higher incorporation of DHA into hepatic PC compared to the low *n*-3 PUFA group (*p* < 0.05; Figure 1B); however, there were no differences in plasma DHA containing PC in both groups (Figure 1A).

### FFA of mice fed high and low *n*-3 PUFA

There was a significant accretion of AA in the plasma of the low *n*-3 PUFA group compared to the high *n*-3 PUFA group (*p* < 0.05; Figure 2A). The plasma concentration of EPA was considerably higher in the high *n*-3 PUFA group compared to the low *n*-3 PUFA group (*p* < 0.05; Figure 2A). However, no significant difference was observed in the plasma concentration of DHA in both groups. Livers of the mice fed a low *n*-3 PUFA

**Figure 1**



**Figure 1. Phosphatidylcholine (PC) fatty acyl composition of mice fed high and low *n*-3 PUFA enriched diets.** Plasma (A) and liver (B) PC species were quantified by measuring the sodiated adducts of PC using ESI-MS in positive ion mode by NL scanning for *m/z* 59.1, as described in the “Methods”. Data were corrected for <sup>13</sup>C isotopic effects. Data are presented as mean (*n*=6) ± SD and *P*-values calculated using unpaired *t*-test. \**P* < 0.05; \*\**P* < 0.01; \*\*\**P* < 0.001.

doi: 10.1371/journal.pone.0082399.g001

diet were significantly enriched with 18:1 and 20:4 FFA ( $p < 0.01$ ; Figure 2B) compared to the high  $n-3$  PUFA group. Similarly, the liver concentrations of EPA ( $p < 0.01$ ; Figure 2B) and DHA ( $p < 0.05$ ; Figure 2B) were significantly higher in the high  $n-3$  PUFA group compared to the low  $n-3$  PUFA group.

#### LPC fatty acyl composition of mice fed high and low $n-3$ PUFA

Similar to the PC data, the concentration of 20:5 LPC was significantly higher in the high  $n-3$  PUFA group compared to the low  $n-3$  PUFA group, in both plasma ( $p < 0.01$ ; Figure 3A) and liver ( $p < 0.01$ ; Figure 3B). However, there was a significant accretion of 20:4 LPC in the plasma ( $p < 0.01$ ; Figure 3A) and liver ( $p < 0.05$ ; Figure 3B) of the low  $n-3$  PUFA group compared to the high  $n-3$  PUFA group. Irrespective of the differences observed in LPC, there was no significant difference in the total concentration of LPC in both groups (Table S2 in File S1).

#### PE fatty acyl composition of mice fed high and low $n-3$ PUFA

No significant differences were observed in the fatty acyl species of PE in the plasma (Figure 4A) of the high  $n-3$  PUFA and low  $n-3$  PUFA groups. The liver showed no differences amongst PE species containing EPA; however, there was a significant increase of hepatic 18:0-22:6 PE in the high  $n-3$  PUFA group compared to the low  $n-3$  PUFA group ( $p < 0.05$ ; Figure 4B).

#### Sphingolipid and ceramide fatty acyl composition of mice fed high and low $n-3$ PUFA

Treatment with high  $n-3$  PUFA did not significantly modify the fatty acyl species of SM in the plasma (Figure 5A) and liver (Figure 5B) of mice fed diet high in  $n-3$  PUFA. Plasma of mice from high  $n-3$  PUFA group showed an increase in 16:0 CER compared to the low  $n-3$  PUFA group ( $p < 0.05$  Figure 6A); however, there was no effect of diet on different fatty acyl species of CER in the liver (Figure 6B).

#### CE and TG fatty acyl composition of mice fed high and low $n-3$ PUFA

The high  $n-3$  PUFA group showed a significant increase in plasma 20:5 CE compared to the low  $n-3$  PUFA group ( $p < 0.05$ ; Figure 7A). Conversely, there was a high accumulation of plasma 20:4 CE in the low  $n-3$  PUFA group compared to the high  $n-3$  PUFA group ( $p < 0.05$ ; Figure 7A). The liver of the high  $n-3$  PUFA group was enriched in 20:5 CE ( $p < 0.001$ ; Figure 7B) and 22:6 CE ( $p < 0.01$ ; Figure 7B) compared to the low  $n-3$  PUFA group. The liver concentration of 20:4 CE was significantly higher in the low  $n-3$  PUFA group ( $p < 0.01$ ; Figure 7B) compared to the high  $n-3$  PUFA group.

No differences were observed in the concentrations of the different fatty acyl species of TG found in the plasma in both experimental groups (Figure 8A). Nevertheless, the liver showed a distinctive difference in the concentration of fatty acyl species profile for TG. The concentrations of 54:6, 56:7, and 58:8 TGs were significantly greater in the high  $n-3$  PUFA group

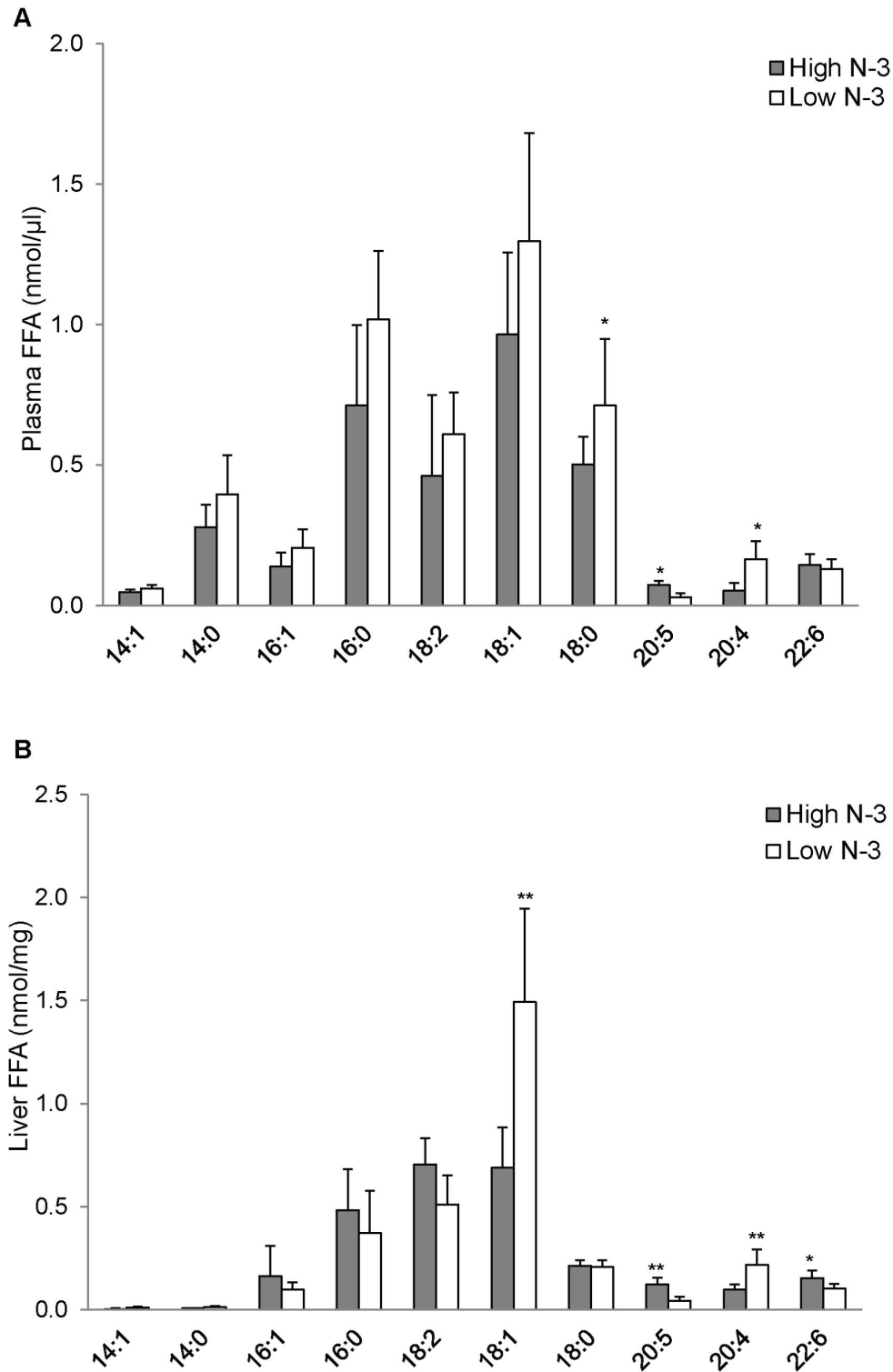
compared to the low  $n-3$  PUFA group ( $p < 0.01$ ; Figure 8B). The low  $n-3$  PUFA group showed a significantly higher 54:3 and 56:5 TG species compared to the high  $n-3$  PUFA group ( $p < 0.05$ ; Figure 8B).

## Discussion

$n-3$  PUFA have been shown to reduce plasma TG [8,45], prevent atherosclerosis [46], and alleviate inflammation [47]. It has been proposed that the beneficial effects of  $n-3$  PUFA are mediated through the actions of bioactive lipid components; however which bioactive lipids are metabolically active, and their mechanisms of action are still not clear. We performed lipidomic analyses on plasma and liver samples of high and low  $n-3$  PUFA fed male C57BL/6 mice. There was an evident distinction in the fatty acid profiles of different lipids present in the plasma and liver of the two dietary groups. Feeding a high  $n-3$  PUFA diet led to a marked reduction of AA containing PC, with a concomitant increase of EPA containing PC. Although plasma samples from mice fed a high or low  $n-3$  PUFA diet showed no difference in DHA containing PC, there was a trend towards an increase in the high  $n-3$  PUFA group. Browning et al. reported that habitual supplementation of fish oil led to an increased incorporation of EPA in plasma PC [48]; however, they did observe a significant incorporation of DHA into PC after dietary supplements of DHA [48]. A plausible explanation for the disparity in our findings is that DHA containing PC is rapidly converted to LPC for delivery to extrahepatic tissues. The liver PC fatty acyl composition is consistent with changes in plasma PC, where high dietary  $n-3$  PUFA increased the concentrations of hepatic EPA and DHA species of PC. On the contrary, there was a significant reduction of hepatic AA containing PC in mice fed high  $n-3$  PUFA diet. PC is an important phospholipid involved in lipid metabolism. PC has been shown to play a major role in cellular proliferation, degeneration, and membrane fluidity and functions [13]. The physiological properties of PC are heavily dependent on their fatty acid composition and  $n-3$  PUFA has been shown to be preferentially incorporated into PC [48]. PC from marine sources are rich in  $n-3$  PUFA and have been shown to significantly reduce markers of inflammation [49], by the inhibition of TNF- $\alpha$  induced activity of NF- $\kappa$ B [50]. In addition to their anti-inflammatory effects,  $n-3$  PUFA rich PC are also known to possess lipid lowering effects [51-53].

Plasma PC is rapidly metabolised by phospholipase A<sub>2</sub> to produce FFA and LPC. These bioactive metabolites of PC can also elicit beneficial effects depending on the type, chain length, and degree of unsaturation of their fatty acids. Feeding diets enriched in  $n-3$  PUFA caused an increase in the plasma concentration of EPA and a decrease in plasma concentration of AA. PC is an important source of FFA in plasma [48]. We found a high concentration of EPA containing PC in the high  $n-3$  PUFA group; it is thus reasonable to assume that the high plasma concentration of EPA found in the high  $n-3$  PUFA fed mice is partly a product of enzyme hydrolysis of EPA rich PC. This is in line with previous findings that support the incorporation of  $n-3$  PUFA into the plasma pool when given as dietary supplements [48,54]. We observed that the dietary

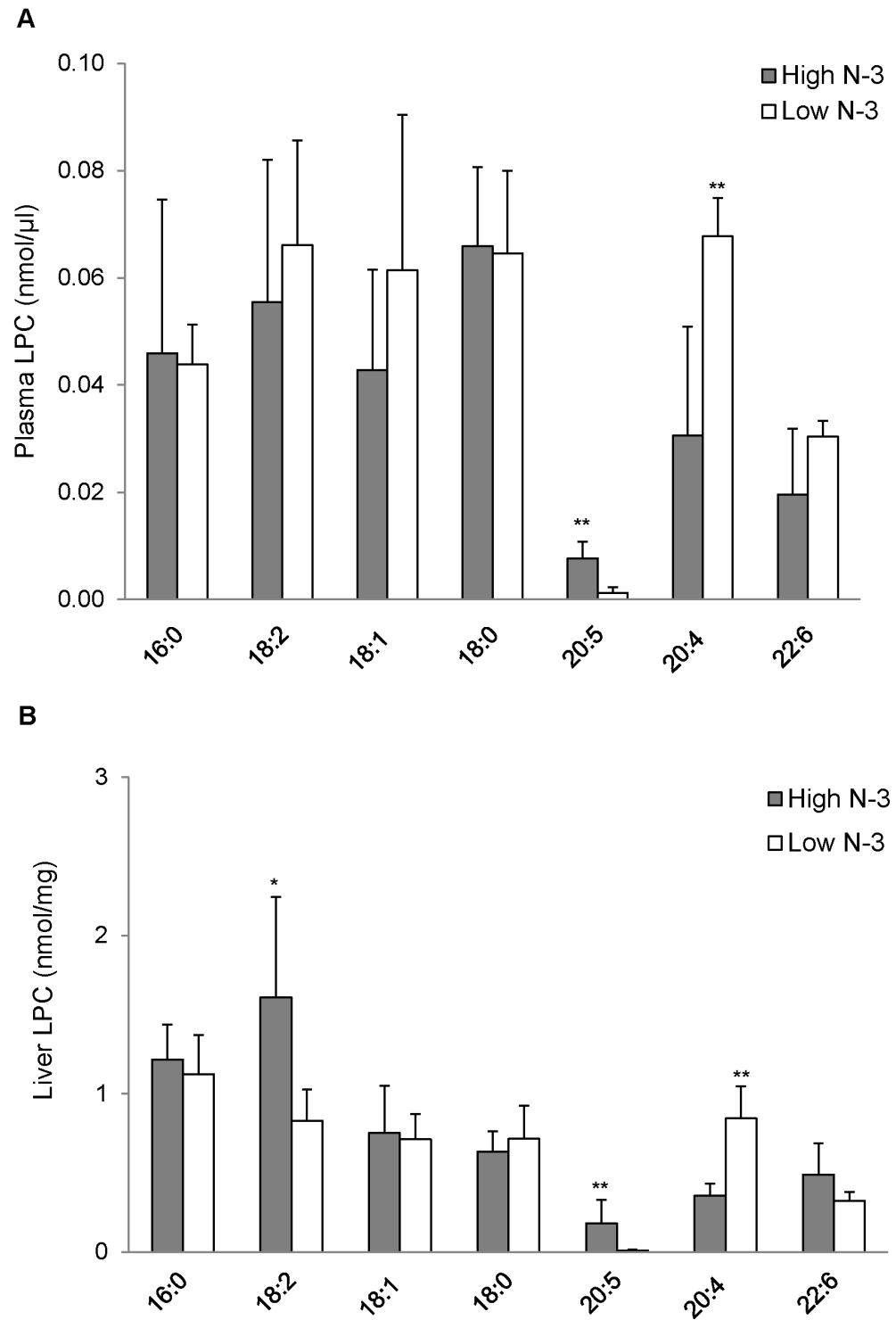
**Figure 2**



**Figure 2. Free fatty acid (FFA) composition of mice fed high and low *n*-3 PUFA enriched diets.** Plasma (A) and liver (B) FFA composition was quantified by measuring [M-H]<sup>-</sup> using ESI-MS in negative ion mode between *m/z* 200 and 900. Data were corrected for <sup>13</sup>C isotopic effects. Data are presented as mean (*n*=6) ± SD and *P*-values calculated using unpaired t-test. \**P* < 0.05; \*\**P* < 0.01; \*\*\**P* < 0.001.

doi: 10.1371/journal.pone.0082399.g002

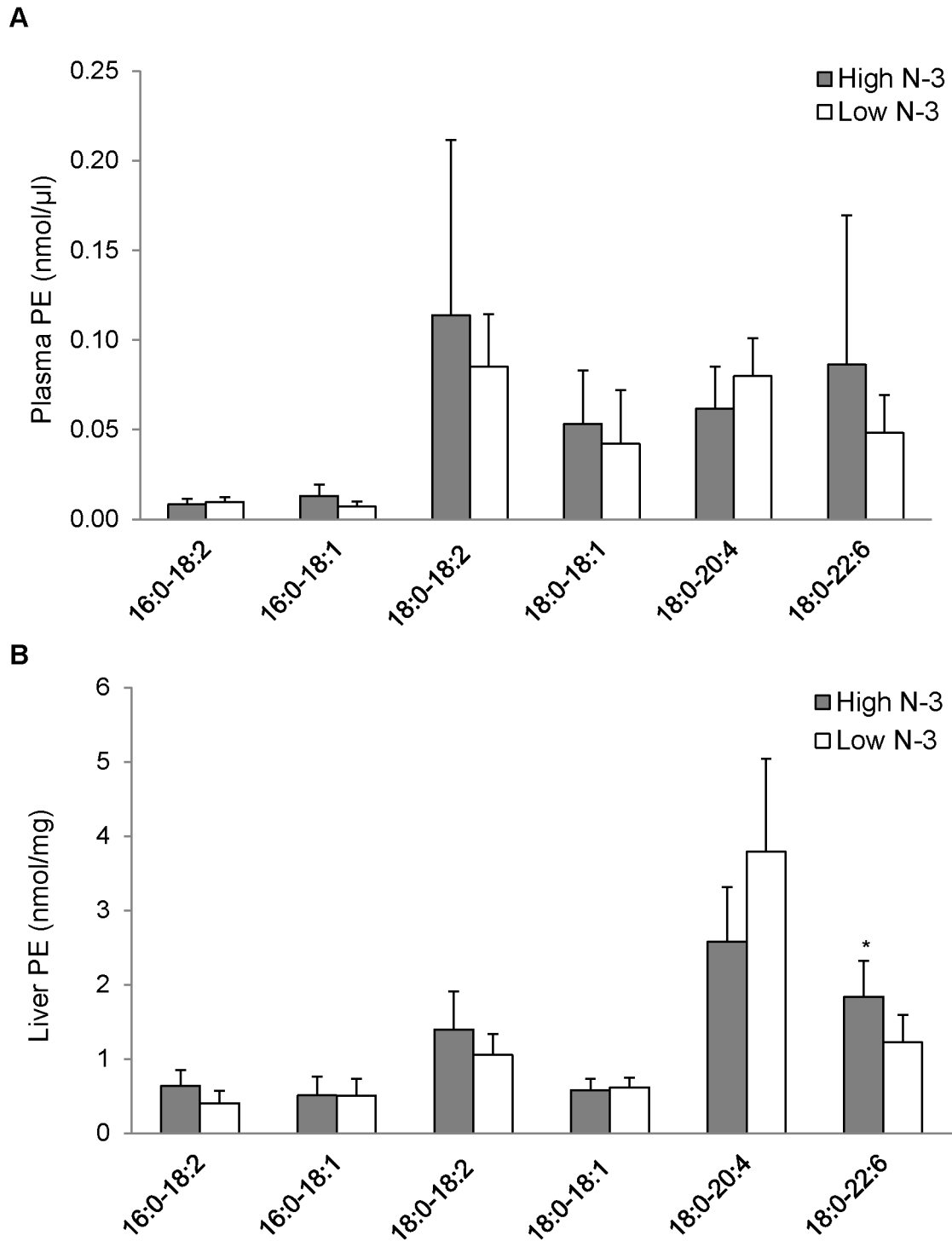
**Figure 3**



**Figure 3. Lysophosphatidylcholine (LPC) fatty acyl composition of mice fed high and low *n*-3 PUFA enriched diets.** Plasma (A) and liver (B) LPC species were quantified by measuring the sodiated adducts of PC using ESI-MS in positive ion mode by NL scanning for *m/z* 59.1, as described in the “Methods”. Data were corrected for <sup>13</sup>C isotopic effects. Data are presented as mean (*n*=6) ± SD and *P*-values calculated using unpaired *t*-test. \**P* < 0.05; \*\**P* < 0.01; \*\*\**P* < 0.001.

doi: 10.1371/journal.pone.0082399.g003

**Figure 4**



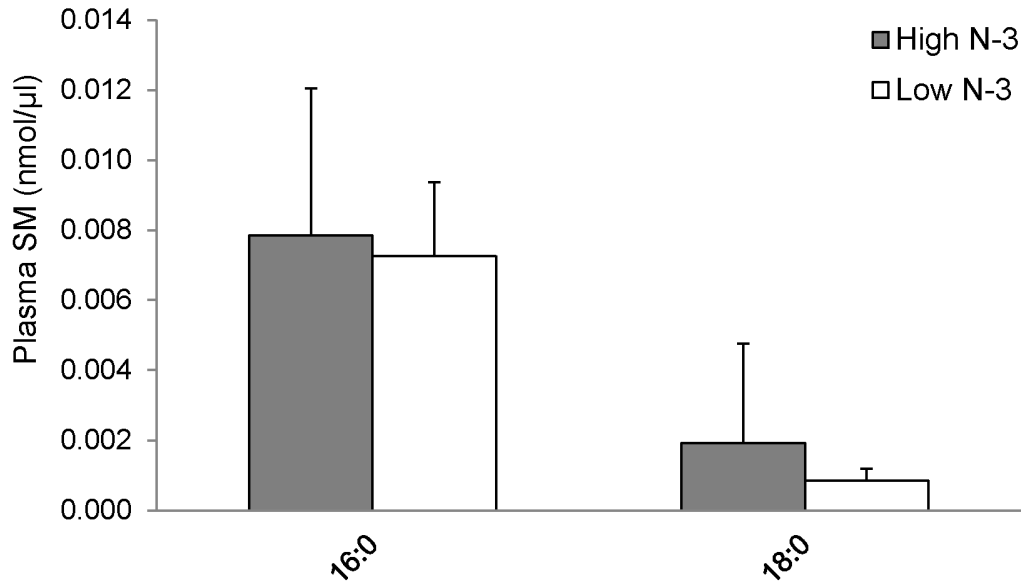
**Figure 4. Phosphatidylethanolamine (PE) fatty acyl composition of mice fed high and low *n*-3 PUFA enriched diets.** Plasma (A) and liver (B) levels of PE species were quantified by measuring [M-H]<sup>-</sup> using ESI-MS in negative ion mode between *m/z* 200 and 900. Data were corrected for <sup>13</sup>C isotopic effects. Data are presented as mean (*n*=6) ± SD and *P*-values calculated using unpaired t-test. \**P* < 0.05; \*\**P* < 0.01; \*\*\**P* < 0.001.

doi: 10.1371/journal.pone.0082399.g004

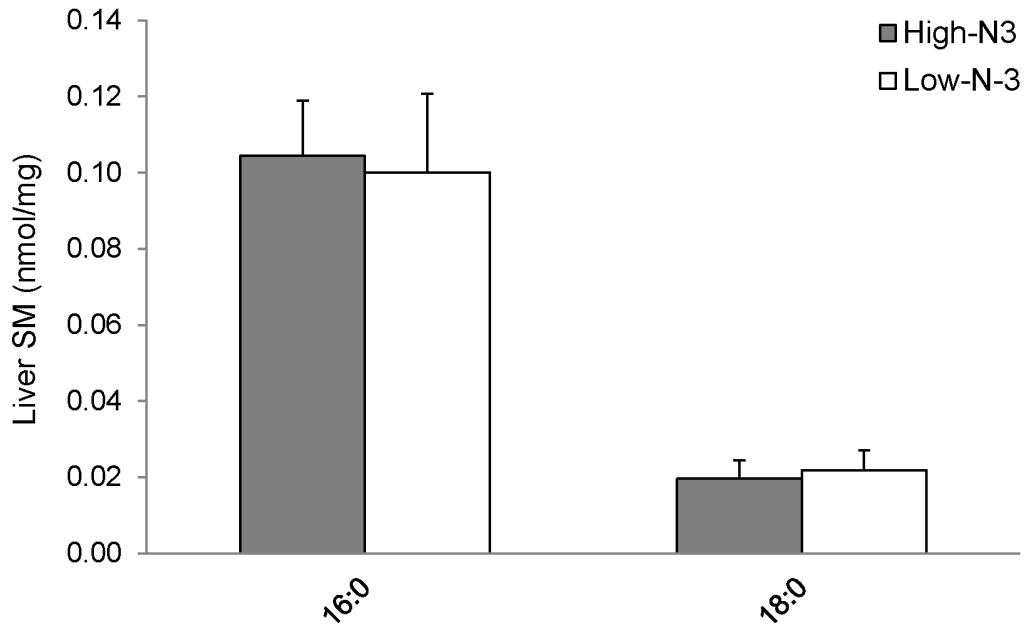


**Figure 5**

**A**



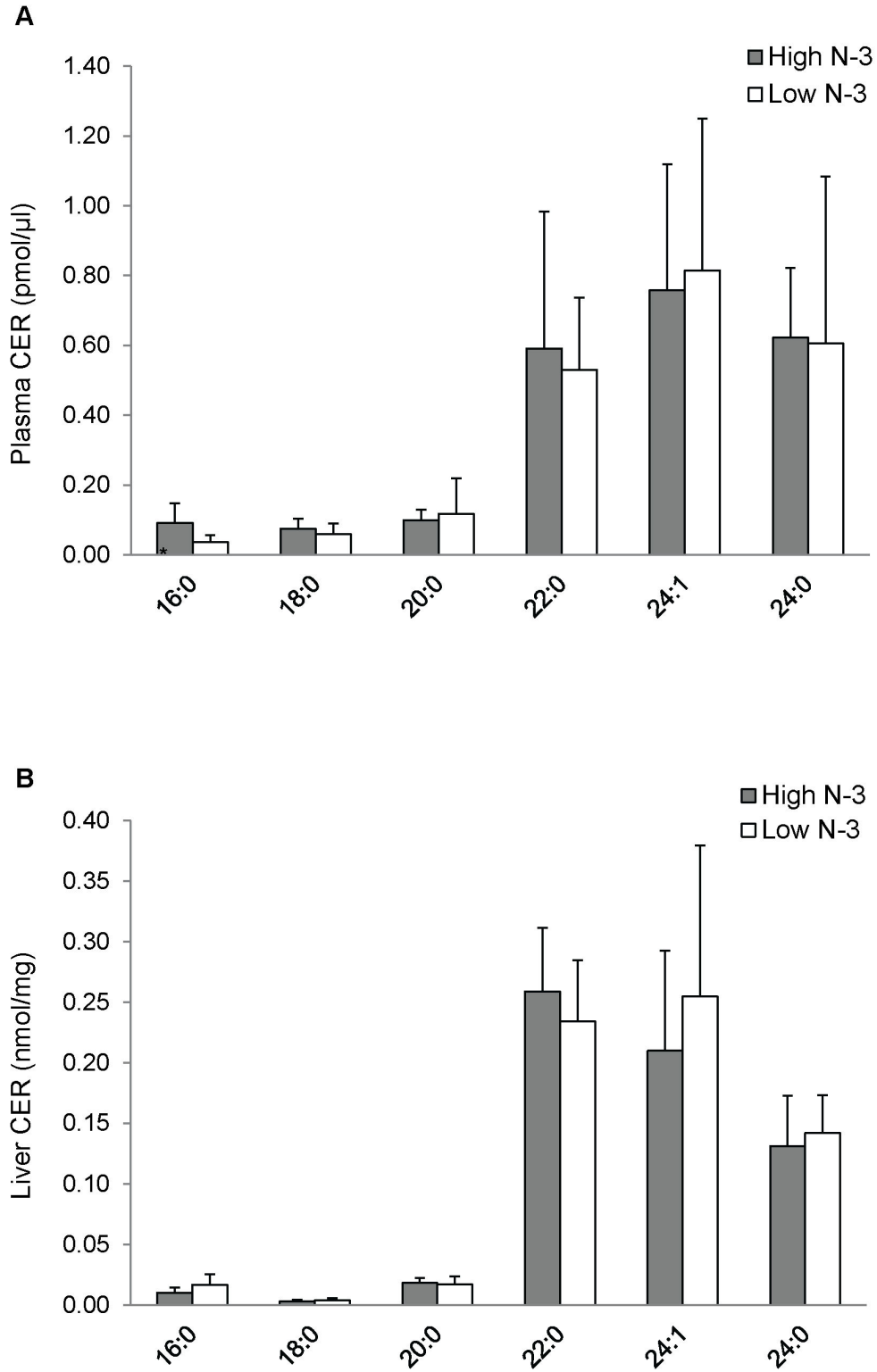
**B**



**Figure 5. Sphingomyelin (SM) fatty acyl composition of mice fed high and low *n*-3 PUFA enriched diets.** Plasma (A) and liver (B) levels of SM species were quantified by measuring the sodiated adducts of PC using ESI-MS in positive ion mode by NL scanning for *m/z* 59.1, as described in the “Methods”. Data were corrected for <sup>13</sup>C isotopic effects. Data are presented as mean (*n*=6) ± SD and *P*-values calculated using unpaired t-test. \**P* < 0.05; \*\**P* < 0.01; \*\*\**P* < 0.001.

doi: 10.1371/journal.pone.0082399.g005

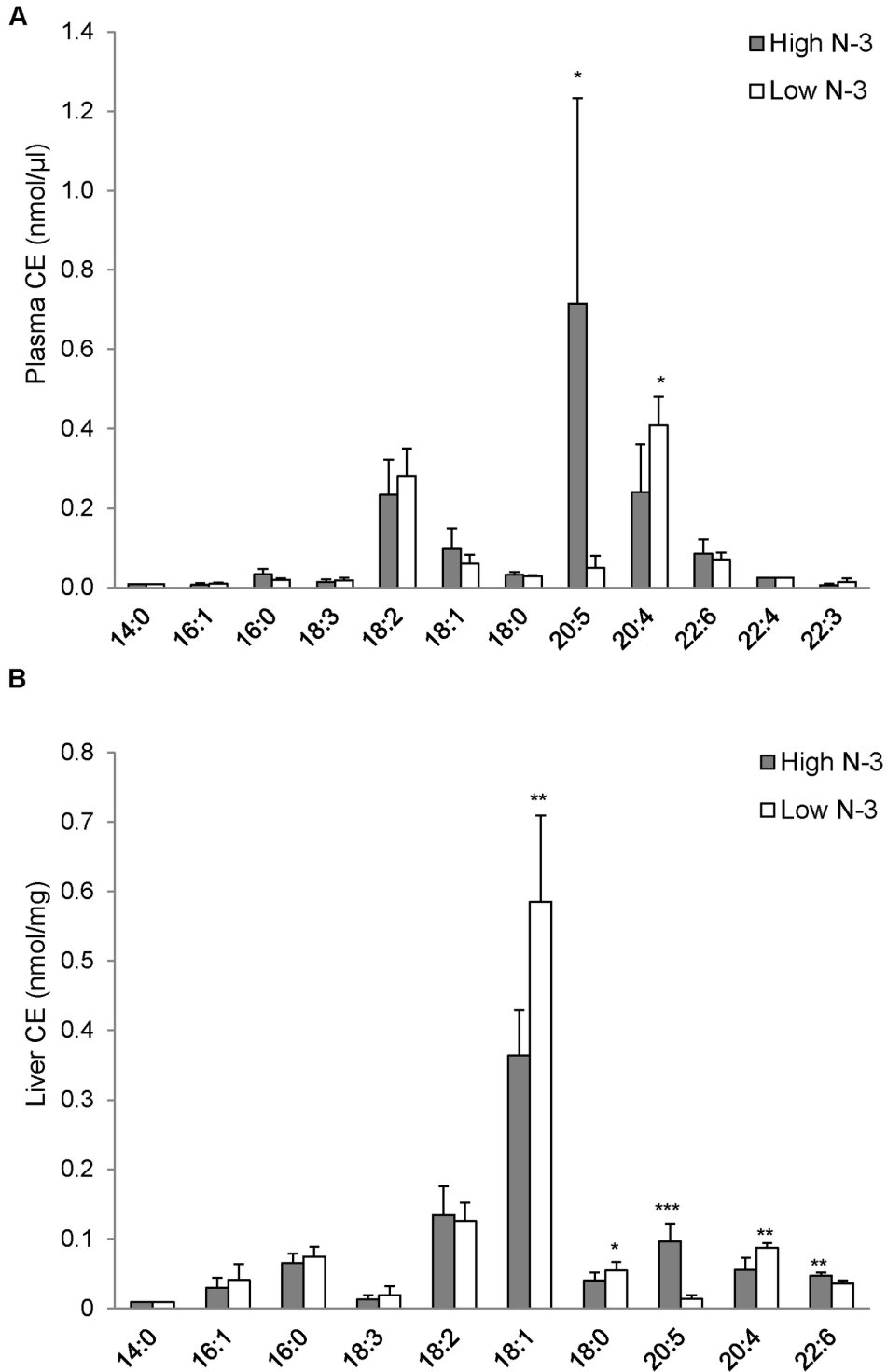
**Figure 6**



**Figure 6. Ceramide (CER) fatty acyl composition of mice fed high and low *n*-3 PUFA enriched diets.** Plasma (A) and liver (B) CER species were quantified using ESI-MS in negative ion mode by NL scanning for *m/z* 256.2. Data were corrected for <sup>13</sup>C isotope effects. Data are presented as mean (*n*=6) ± SD and *P*-values calculated using unpaired t-test. \**P* < 0.05; \*\**P* < 0.01; \*\*\**P* < 0.001.

doi: 10.1371/journal.pone.0082399.g006

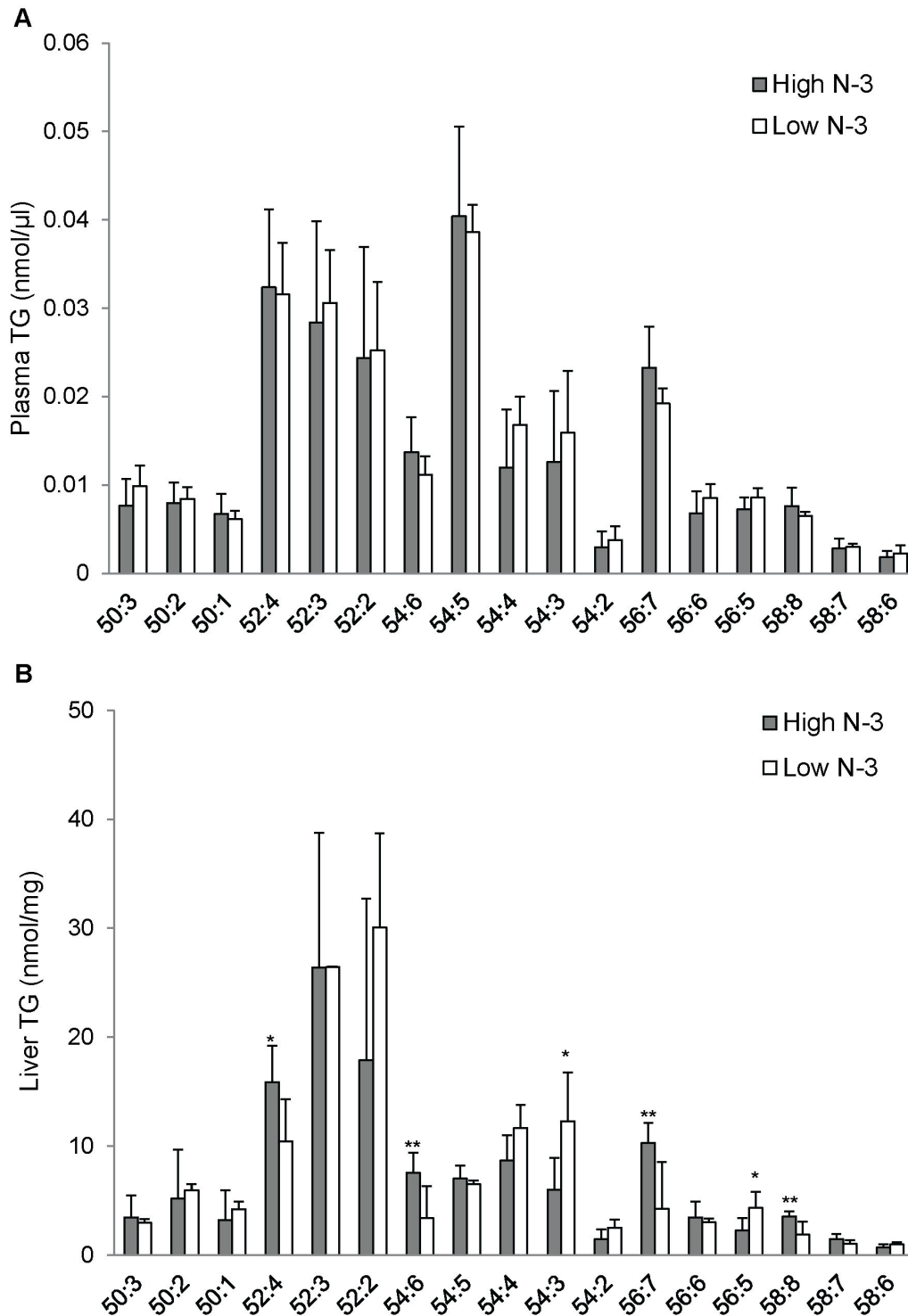
**Figure 7**



**Figure 7. Cholesteryl ester (CE) fatty acyl composition of mice fed high and low *n*-3 PUFA enriched diets.** Plasma (A) and liver (B) CE species were quantified using ESI-MS in positive ion mode by NL scanning of *m/z* 368.5 as described in the “Methods”. Data were corrected for <sup>13</sup>C isotope effects. Data are presented as mean (*n*=6) ± SD and *P*-values calculated using unpaired *t*-test. \**P* < 0.05; \*\**P* < 0.01; \*\*\**P* < 0.001.

doi: 10.1371/journal.pone.0082399.g007

**Figure 8**



**Figure 8. Triglyceride (TG) fatty acyl composition of mice fed high and low *n*-3 PUFA enriched diets.** Plasma (A) and liver (B) TG species were quantified by measuring  $[M+Na]^+$  using ESI-MS in positive ion mode between  $m/z$  800 and 1000 described in the “Methods”. Data were corrected for  $^{13}C$  isotope effects. Data are presented as mean ( $n=6$ )  $\pm$  SD and *P*-values calculated using unpaired t-test. \**P* < 0.05; \*\**P* < 0.01; \*\*\**P* < 0.001.

doi: 10.1371/journal.pone.0082399.g008

supplementation with high *n*-3 PUFA caused an increase in the concentrations of hepatic EPA and DHA, and a decline in the hepatic concentration of AA. Our observation is in line with the study of Lamaziere et al., who reported an increase in hepatic EPA and DHA after rats were administered fish oil for 30 days [55]. In addition to the aforementioned health benefits of *n*-3 PUFA, high hepatic *n*-3 PUFA are also involved in inhibiting lipogenesis thereby preventing the development of non-alcoholic fatty liver disease (NAFLD) [55]. Mechanistically, *n*-3 PUFA has been shown to regulate lipogenesis by inhibiting LXR and SREBP-1c [56,57]. Chronic low-grade inflammation underlies the pathology of most metabolic disorders, and free *n*-3 PUFA has been shown to possess potent anti-inflammatory properties [58]. *N*-3 PUFA alleviates inflammation by directly regulating transcription factors involved in inflammation [59-61] and indirectly by producing series-3 and series-5 eicosanoids [62,63]. In addition to their inflammation resolving properties, free unesterified *n*-3 PUFA have also been shown to improve symptoms of dyslipidaemia [9-11].

Another important bioactive product of enzyme hydrolysis of PC is LPC. We found that feeding a diet high in *n*-3 PUFA considerably increased plasma circulating levels of 20:5 LPC, and drastically reduced plasma concentration of 20:4 LPC. Similar to our plasma data, we found a higher concentration of hepatic 20:5 LPC and a low concentration of hepatic 20:4 LPC in the high *n*-3 group. Interestingly, there was no significant difference in hepatic 22:6 LPC, although there was a trend towards an increase in the high *n*-3 PUFA group. Ottestad et al. recently reported a similar finding where fish oil supplementation to healthy humans significantly increased plasma 20:5 LPC and 22:6 LPC [64]. Another study by Block et al. also found that fish oil supplementation increased EPA and DHA containing LPC [65]. However, we only found a change in plasma EPA containing LPC, while there was no difference in DHA containing LPC between the two groups. LPC has been suggested as the major carrier of DHA to the brain tissues [66]; tracer studies revealed that labelled LPC injected into the blood of rat disappeared within 20 s and were recovered in different organs including the brain [17]. It would thus be logical to believe that the DHA containing LPC was rapidly cleared from circulation.

Studies have controversially linked LPC with the development of atherosclerosis [25-27]. This is possibly due to their association with oxidized LDL, and promotion of inflammation [67,68] by generating reactive oxygen species and nitric oxides in different types of cells [69,70]. However, the studies that linked LPC with the pathogenesis of obesity, diabetes, and rheumatoid arthritis have reported an increase in saturated LPC [30]. Of paramount importance to the biological functions of LPC are acyl length and degree of saturation of their fatty acids [65]. LPC rich in *n*-3 PUFA have been shown to possess beneficial properties. *N*-3 PUFA species of LPC were found to reduce inflammation [71,72], and LPC containing DHA at *sn*-1 position exhibits significantly higher anti-inflammatory properties compared to LPC with either linoleic or arachidonic acid [31,73,74]. It has been proposed that LPC containing *n*-3 PUFA exhibit their anti-inflammatory properties through hydrolytic cleavage of the *n*-3 PUFA moiety, or oxygenation by

15-lipoxygenase (15-LOX) to produce inflammation resolving lipids such as 1-(15-hydroperoxyeicosatetraenyl)-LPC and 1-(17-hydroperoxydocosahexaenyl)-LPC [74]. Most of the available data on the health benefits of *n*-3 PUFA rich LPC were shown using LPC containing *n*-3 PUFA at *sn*-1 position. We have not investigated the position of the acyl group of our LPC, however, it is known that there is rapid isomerization of acyl group from *sn*-2 to a more stable *sn*-1 position in LPC [75]. It has been reported that after separation of blood from plasma, 90% of the unsaturated fatty acids were found at *sn*-1 position in LPC [24]. We can therefore speculate that a significant percentage of the plasma 20:5 LPC contain EPA at *sn*-1 position.

Also noteworthy is the fact that, despite the changes in fatty acyl species of PC and LPC in response to diet, there was no difference in the total concentrations of PC and LPC between the two experimental groups. A similar observation was found by Ottestad et al., where they found no difference in total concentrations of PC and LPC despite the apparent changes in individual fatty acyl species in response to fish oil supplementation [64]. This interesting observation suggests that the functional properties of *n*-3 PUFA involve remodelling and improving the quality of bioactive lipid mediators without affecting their concentrations.

A diet rich in *n*-3 PUFA had no effect on different fatty acyl species of plasma TG, confirming preferential incorporation of *n*-3 PUFA into PC. However, the high *n*-3 PUFA diet led to an increase in plasma and liver concentrations of *n*-3 PUFA CE, and a decrease in 20:4 CE. CE is less polar than free cholesterol and it functions as an inert storage molecule. The high incorporation of *n*-3 PUFA in CE could be explained as a response to the high *n*-3 PUFA in the diet. This would simply indicate that *n*-3 PUFA are stored and will be later released for other physiological functions. A cross sectional study has reported a positive correlation between dietary PUFA and CE [76].

Although we have shown that there is high incorporation of *n*-3 PUFA in the liver and plasma PC and LPC of mice fed high *n*-3 diet, it was also imperative to ascertain the effect of high *n*-3 PUFA diet on PE, which is also abundant in the membranes. There was no difference in plasma PE between the two dietary groups; the only difference detected in the liver was an increase in 18:0-22:6 PE in the high *n*-3 group. Our findings confirm that fish oil supplementation leads to a preferential incorporation of *n*-3 PUFA into PC and LPC compared to other phospholipids [64].

Sphingolipids, such as CER and SM, are important signalling molecules. CER is a sphingolipid linked with inflammation and the pathogenesis of cardiovascular diseases (CVD) [77,78]. There are also speculations on the involvement of CER in insulin signalling, although available information is scanty [79,80]. We found no difference in total concentration of CER in plasma and liver. Our findings are similar to Ottestad et al., who found that fish oil supplement had no effect on plasma concentration of CER. Interestingly, we found an increase in 16:0 CER in the high *n*-3 PUFA group. The functional roles of this CER species are currently unknown and needs to be explored. There was no difference in plasma and liver SM

concentrations between the two groups. The physiological functions of SM have not been extensively studied. However, it is known that SM are involved in the formation of specialized membrane microdomains known as lipid rafts involved in signalling.

In conclusion, our findings have shown that diets high in *n*-3 PUFA alter plasma and liver lipidomic profile of the offspring. We found that *n*-3 PUFA is preferentially incorporated into PC and LPC, and despite the changes in lipidomic profile, the total concentrations of these lipids were not altered. Additionally, we found that dietary *n*-3 PUFA is capable of remodelling the fatty acyl moieties of PC, LPC, and CE, which may have important physiological implications, and needs to be further investigated. Future studies will be undertaken to investigate the mechanism(s) by which *n*-3 PUFA remodelled bioactive lipids regulate metabolic pathways.

## References

- Calder PC (2004) *n*-3 Fatty acids and cardiovascular disease: evidence explained and mechanisms explored. *Clin Sci (Lond)* 107: 1-11. doi: 10.1042/CS20040119. PubMed: 15132735.
- Schmidt EB, Arnesen H, de Caterina R, Rasmussen LH, Kristensen SD (2005) Marine *n*-3 polyunsaturated fatty acids and coronary heart disease. Part I. Background, epidemiology, animal data, effects on risk factors and safety. *Thromb Res* 115: 163-170. doi:10.1016/j.thromres.2004.09.006. PubMed: 15617737.
- Belluzzi A, Boschi S, Brignola C, Munarini A, Cariani G et al. (2000) Polyunsaturated fatty acids and inflammatory bowel disease. *Am J Clin Nutr* 71: 339S-342S. PubMed: 10617993.
- Wassall SR, Stillwell W (2009) Polyunsaturated fatty acid-cholesterol interactions: domain formation in membranes. *Biochim Biophys Acta* 1788: 24-32. doi:10.1016/j.bbame.2008.10.011. PubMed: 19014904.
- Riediger ND, Othman RA, Suh M, Moghadasian MH (2009) A systemic review of the roles of *n*-3 fatty acids in health and disease. *J Am Diet Assoc* 109: 668-679. doi:10.1016/j.jada.2008.12.022. PubMed: 19328262.
- Cottin SC, Sanders TA, Hall WL (2011) The differential effects of EPA and DHA on cardiovascular risk factors. *Proc Nutr Soc* 70: 215-231. doi:10.1017/S0029665111000061. PubMed: 21349231.
- Banni S, Di Marzo V (2010) Effect of dietary fat on endocannabinoids and related mediators: consequences on energy homeostasis, inflammation and mood. *Mol Nutr Food Res* 54: 82-92. doi:10.1002/mnfr.200900516. PubMed: 20013888.
- Harris W (1997) *n*-3 fatty acids and serum lipoproteins: human studies. *Am J Clin Nutr* 65: 1645S-11654.
- Leaf A, Kang JX, Xiao YF, Billman GE (2003) Clinical prevention of sudden cardiac death by *n*-3 polyunsaturated fatty acids and mechanism of prevention of arrhythmias by *n*-3 fish oils. *Circulation* 107: 2646-2652. doi:10.1161/01.CIR.0000069566.78305.33. PubMed: 12782616.
- Goodfellow J, Bellamy MF, Ramsey MW, Jones CJ, Lewis MJ (2000) Dietary supplementation with marine omega-3 fatty acids improve systemic large artery endothelial function in subjects with hypercholesterolemia. *J Am Coll Cardiol* 35: 265-270. doi:10.1016/S0735-1097(99)00548-3. PubMed: 10676668.
- Hirafuji M, Machida T, Hamaue N, Minami M (2003) Cardiovascular protective effects of *n*-3 polyunsaturated fatty acids with special emphasis on docosahexaenoic acid. *J Pharmacol Sci* 92: 308-316. doi: 10.1254/jphs.92.308. PubMed: 12939515.
- Holub BJ (1978) Differential utilization of 1-palmitoyl and 1-stearoyl homologues of various unsaturated 1,2-diacyl-sn-glycerols for phosphatidylcholine and phosphatidylethanolamine synthesis in rat liver microsomes. *J Biol Chem* 253: 691-696. PubMed: 202595.
- Gundermann KJ, Kuenker A, Kuntz E, Drożdżik M (2011) Activity of essential phospholipids (EPL) from soybean in liver diseases. *Pharmacol Rep* 63: 643-659. PubMed: 21857075.
- Hulbert AJ, Turner N, Storlien LH, Else PL (2005) Dietary fats and membrane function: implications for metabolism and disease. *Biol Rev Camb Philos Soc* 80: 155-169. doi:10.1017/S1464793104006578. PubMed: 15727042.
- Leaf A, Xiao YF, Kang JX, Billman GE (2005) Membrane effects of the *n*-3 fish oil fatty acids, which prevent fatal ventricular arrhythmias. *J Membr Biol* 206: 129-139. doi:10.1007/s00232-005-0789-9. PubMed: 16456723.
- Stillwell W, Wassall SR (2003) Docosahexaenoic acid: membrane properties of a unique fatty acid. *Chem Phys Lipids* 126: 1-27. doi: 10.1016/S0009-3084(03)00101-4. PubMed: 14580707.
- Thiéss F, Delachambre MC, Bentejac M, Lagarde M, Lecerf J (1992) Unsaturated fatty acids esterified in 2-acyl-l-lysophosphatidylcholine bound to albumin are more efficiently taken up by the young rat brain than the unesterified form. *J Neurochem* 59: 1110-1116. doi:10.1111/j.1471-4159.1992.tb08353.x. PubMed: 1494901.
- Huwiler A, Pfeilschifter J (2009) Lipids as targets for novel anti-inflammatory therapies. *Pharmacol Ther* 124: 96-112. doi:10.1016/j.pharmthera.2009.06.008. PubMed: 19576246.
- Farooqui AA, Horrocks LA, Farooqui T (2007) Modulation of inflammation in brain: a matter of fat. *J Neurochem* 101: 577-599. PubMed: 17257165.
- Adkins Y, Kelley DS (2010) Mechanisms underlying the cardioprotective effects of omega-3 polyunsaturated fatty acids. *J Nutr Biochem* 21: 781-792. doi:10.1016/j.jnutbio.2009.12.004. PubMed: 20382009.
- Ariel A, Serhan CN (2007) Resolvins and protectins in the termination program of acute inflammation. *Trends Immunol* 28: 176-183. doi: 10.1016/j.it.2007.02.007. PubMed: 17337246.
- Bannenberg G, Serhan CN (2010) Specialized pro-resolving lipid mediators in the inflammatory response: An update. *Biochim Biophys Acta* 1801: 1260-1273. doi:10.1016/j.bbali.2010.08.002. PubMed: 20780999.
- Kohli P, Levy BD (2009) Resolvins and protectins: mediating solutions to inflammation. *Br J Pharmacol* 158: 960-971. doi:10.1111/j.1476-5381.2009.00290.x. PubMed: 19594757.
- Croset M, Brossard N, Polette A, Lagarde M (2000) Characterization of plasma unsaturated lysophosphatidylcholines in human and rat. *Biochem J* 345 (Pt. 1): 61-67. doi:10.1042/0264-6021:3450061. PubMed: 10600639.
- Wu WT, Chen CN, Lin CI, Chen JH, Lee H (2005) Lysophospholipids enhance matrix metalloproteinase-2 expression in human endothelial cells. *Endocrinology* 146: 3387-3400. doi:10.1210/en.2004-1654. PubMed: 15878967.
- Lucas A, Grynberg A, Lacour B, Goirand F (2008) Dietary *n*-3 polyunsaturated fatty acids and endothelium dysfunction induced by lysophosphatidylcholine in Syrian hamster aorta. *Metabolism* 57: 233-240. doi:10.1016/j.metabol.2007.09.006. PubMed: 18191054.
- Schmitz G, Ruebsaamen K (2010) Metabolism and atherogenic disease association of lysophosphatidylcholine. *Atherosclerosis* 208: 10-18. doi:10.1016/j.atherosclerosis.2009.05.029. PubMed: 19570538.
- Hayashi K, Takahashi M, Nishida W, Yoshida K, Ohkawa Y et al. (2001) Phenotypic modulation of vascular smooth muscle cells induced by unsaturated lysophosphatidic acids. *Circ Res* 89: 251-258. doi: 10.1161/hh1501.094265. PubMed: 11485975.
- Yoshida K, Nishida W, Hayashi K, Ohkawa Y, Ogawa A et al. (2003) Vascular remodeling induced by naturally occurring unsaturated

## Supporting Information

**File S1. Table S1, Body weight at sacrifice, and average weekly food intake of mice fed high and low *n*-3 PUFA diets; Table S2, Total concentrations of plasma and liver phospholipids of mice fed high and low *n*-3 PUFA diets. (DOCX)**

## Author Contributions

Conceived and designed the experiments: SKC KAB. Performed the experiments: SKC KAB RJB CJA. Analyzed the data: SKC KAB RJB CJA DAF. Contributed reagents/materials/analysis tools: SKC KAB RJB CJA DAF. Wrote the manuscript: SKC KAB.

- lysophosphatidic acid in vivo. *Circulation* 108: 1746-1752. doi:10.1161/01.CIR.0000089374.35455.F3. PubMed: 14504178.
30. Fuchs B, Schiller J, Wagner U, Hantzschel H, Arnold K (2005) The phosphatidylcholine/lysophosphatidylcholine ratio in human plasma is an indicator of the severity of rheumatoid arthritis: investigations by 31P NMR and MALDI-TOF MS. *Clin Biochem* 38: 925-933. doi:10.1016/j.clinbiochem.2005.06.006.
  31. Hung ND, Sok DE, Kim MR (2012) Prevention of 1-palmitoyl lysophosphatidylcholine-induced inflammation by polyunsaturated acyl lysophosphatidylcholine. *Inflamm Res* 61: 473-483. doi:10.1007/s00011-012-0434-x. PubMed: 22252240.
  32. Schuchardt JP, Neubronner J, Kressel G, Merkel M, von Schacky C et al. (2011) Moderate doses of EPA and DHA from re-esterified triacylglycerols but not from ethyl-esters lower fasting serum triacylglycerols in statin-treated dyslipidemic subjects: Results from a six month randomized controlled trial. *Prostaglandins Leukot Essent Fatty Acids* 85: 381-386. doi:10.1016/j.plefa.2011.07.006. PubMed: 21862301.
  33. Galli C, Maggi FM, Risé P, Sirtori CR (2012) Bioequivalence of two omega-3 fatty acid ethyl ester formulations: a case of clinical pharmacology of dietary supplements. *Br J Clin Pharmacol* 74: 60-65. doi:10.1111/j.1365-2125.2012.04174.x. PubMed: 22242645.
  34. Tang X, Li ZJ, Xu J, Xue Y, Li JZ et al. (2012) Short term effects of different omega-3 fatty acid formulation on lipid metabolism in mice fed high or low fat diet. *Lipids Health Dis* 11: 70. doi:10.1186/1476-511X-11-70. PubMed: 22676394.
  35. Portman OW, Alexander M (1969) Lysophosphatidylcholine concentrations and metabolism in aortic intima plus inner media: effect of nutritionally induced atherosclerosis. *J Lipid Res* 10: 158-165. PubMed: 4238547.
  36. Thukkani AK, McHowat J, Hsu FF, Brennan ML, Hazen SL et al. (2003) Identification of alpha-chloro fatty aldehydes and unsaturated lysophosphatidylcholine molecular species in human atherosclerotic lesions. *Circulation* 108: 3128-3133. doi:10.1161/01.CIR.0000104564.01539.6A. PubMed: 14638540.
  37. Aiyar N, Disa J, Ao Z, Ju H, Nerurkar S et al. (2007) Lysophosphatidylcholine induces inflammatory activation of human coronary artery smooth muscle cells. *Mol Cell Biochem* 295: 113-120. doi:10.1007/s11010-006-9280-x. PubMed: 16896535.
  38. Checchi K, Herzberg GR, Cheema SK (2010) Maternal dietary fat intake during gestation and lactation alters tissue fatty acid composition in the adult offspring of C57Bl/6 mice. *Prostaglandins Leukot Essent Fatty Acids* 83: 97-104. doi:10.1016/j.plefa.2010.06.001. PubMed: 20688254.
  39. Childs CE, Romeu-Nadal M, Burdge GC, Calder PC (2008) Gender differences in the n-3 fatty acid content of tissues. *Proc Nutr Soc* 67: 19-27. doi:10.1017/S0029665108005983. PubMed: 18234128.
  40. Knopp RH, Paramsothy P, Retzlaff BM, Fish B, Walden C et al. (2005) Gender differences in lipoprotein metabolism and dietary response: basis in hormonal differences and implications for cardiovascular disease. *Curr Atheroscler Rep* 7: 472-479. doi:10.1007/s11883-005-0065-6. PubMed: 16256006.
  41. Knopp RH, Zhu X (1997) Multiple beneficial effects of estrogen on lipoprotein metabolism. *J Clin Endocrinol Metab* 82: 3952-3954. doi:10.1210/jc.82.12.3952. PubMed: 9398694.
  42. Morise A, Thomas C, Landrier JF, Besnard P, Hermier D (2009) Hepatic lipid metabolism response to dietary fatty acids is differently modulated by PPARalpha in male and female mice. *Eur J Nutr* 48: 465-473. doi:10.1007/s00394-009-0037-7. PubMed: 19588182.
  43. Bligh EG, Dyer WJ (1959) A rapid method of total lipid extraction and purification. *Can J Biochem Physiol* 37: 911-917. doi:10.1139/o59-099. PubMed: 13671378.
  44. Han X, Gross RW (2005) Shotgun lipidomics: electrospray ionization mass spectrometric analysis and quantitation of cellular lipidomes directly from crude extracts of biological samples. *Mass Spectrom Rev* 24: 367-412. doi:10.1002/mas.20023. PubMed: 15389848.
  45. Hartweg J, Perera R, Montori V, Dinneen S, Neil HA et al. (2008) Omega-3 polyunsaturated fatty acids (PUFA) for type 2 diabetes mellitus. *Cochrane Database Syst Rev*: CD003205. PubMed: 18254017.
  46. von Schacky C (2003) The role of omega-3 fatty acids in cardiovascular disease. *Curr Atheroscler Rep* 5: 139-145. doi:10.1007/s11883-003-0086-y. PubMed: 12573200.
  47. Calder PC (2006) n-3 polyunsaturated fatty acids, inflammation, and inflammatory diseases. *Am J Clin Nutr* 83: 1505S-1519S. PubMed: 16841861.
  48. Browning LM, Walker CG, Mander AP, West AL, Madden J et al. (2012) Incorporation of eicosapentaenoic and docosahexaenoic acids into lipid pools when given as supplements providing doses equivalent to typical intakes of oily fish. *Am J Clin Nutr* 96: 748-758. doi:10.3945/ajcn.112.041343. PubMed: 22932281.
  49. Deutsch L (2007) Evaluation of the effect of Neptune Krill Oil on chronic inflammation and arthritic symptoms. *J Am Coll Nutr* 26: 39-48. doi:10.1080/07315724.2007.10719584. PubMed: 17353582.
  50. Treede I, Braun A, Sparla R, Kühnel M, Giese T et al. (2007) Anti-inflammatory effects of phosphatidylcholine. *J Biol Chem* 282: 27155-27164. doi:10.1074/jbc.M704408200. PubMed: 17636253.
  51. Klimov AN, Konstantinov VO, Lipovetsky BM, Kuznetsov AS, Lozovsky VT et al. (1995) "Essential" phospholipids versus nicotinic acid in the treatment of patients with type IIb hyperlipoproteinemia and ischemic heart disease. *Cardiovasc Drugs Ther* 9: 779-784. doi:10.1007/BF00879871. PubMed: 8850382.
  52. Bunea R, El Farrah K, Deutsch L (2004) Evaluation of the effects of Neptune Krill Oil on the clinical course of hyperlipidemia. *Altern Med Rev* 9: 420-428. PubMed: 15656713.
  53. Taylor LA, Pletschen L, Arends J, Unger C, Massing U (2010) Marine phospholipids—a promising new dietary approach to tumor-associated weight loss. *Support Care Cancer* 18: 159-170. doi:10.1007/s00520-009-0640-4. PubMed: 19404684.
  54. Hodson L, Skeaff CM, Fielding BA (2008) Fatty acid composition of adipose tissue and blood in humans and its use as a biomarker of dietary intake. *Prog Lipid Res* 47: 348-380. doi:10.1016/j.plipres.2008.03.003. PubMed: 18435934.
  55. Lamaziere A, Wolf C, Barbe U, Bausero P, Visioli F (2013) Lipidomics of hepatic lipogenesis inhibition by omega 3 fatty acids. *Prostaglandins Leukot Essent Fatty Acids* 88: 149-154. doi:10.1016/j.plefa.2012.12.001. PubMed: 23313470.
  56. Xu J, Nakamura MT, Cho HP, Clarke SD (1999) Sterol Regulatory Element Binding Protein-1 Expression Is Suppressed by Dietary Polyunsaturated Fatty Acids. A mechanism for the coordinate suppression of lipogenic genes by polyunsaturated fats. *J Biol Chem* 274: 23577-23583. doi:10.1074/jbc.274.33.23577. PubMed: 10438539.
  57. Yoshikawa T, Shimano H, Yahagi N, Ide T, Amemiya-Kudo M et al. (2002) Polyunsaturated fatty acids suppress sterol regulatory element-binding protein 1c promoter activity by inhibition of liver X receptor (LXR) binding to LXR response elements. *J Biol Chem* 277: 1705-1711. doi:10.1074/jbc.M105711200. PubMed: 11694526.
  58. Massaro M, Scoditti E, Carluccio MA, De Caterina R (2008) Basic mechanisms behind the effects of n-3 fatty acids on cardiovascular disease. *Prostaglandins Leukot Essent Fatty Acids* 79: 109-115. doi:10.1016/j.plefa.2008.09.009. PubMed: 18951002.
  59. Jump DB, Clarke SD, Thelen A, Liimatta M (1994) Coordinate regulation of glycolytic and lipogenic gene expression by polyunsaturated fatty acids. *J Lipid Res* 35: 1076-1084. PubMed: 8077846.
  60. Pawar A, Jump DB (2003) Unsaturated fatty acid regulation of peroxisome proliferator-activated receptor alpha activity in rat primary hepatocytes. *J Biol Chem* 278: 35931-35939. doi:10.1074/jbc.M306238200. PubMed: 12853447.
  61. Pawar A, Botolin D, Mangelsdorf DJ, Jump DB (2003) The role of liver X receptor-alpha in the fatty acid regulation of hepatic gene expression. *J Biol Chem* 278: 40736-40743. doi:10.1074/jbc.M307973200. PubMed: 12917410.
  62. von Schacky C, Fischer S, Weber PC (1985) Long-term effects of dietary marine omega-3 fatty acids upon plasma and cellular lipids, platelet function, and eicosanoid formation in humans. *J Clin Invest* 76: 1626-1631. doi:10.1172/JCI112147. PubMed: 2997286.
  63. Wanten GJ, Calder PC (2007) Immune modulation by parenteral lipid emulsions. *Am J Clin Nutr* 85: 1171-1184. PubMed: 17490951.
  64. Ottestad I, Hassani S, Borge GI, Kohler A, Vogt G et al. (2012) Fish oil supplementation alters the plasma lipidomic profile and increases long-chain PUFAs of phospholipids and triglycerides in healthy subjects. *PLOS ONE* 7: e42550. doi:10.1371/journal.pone.0042550. PubMed: 22952598.
  65. Block RC, Duff R, Lawrence P, Kakinami L, Brenna JT et al. (2010) The effects of EPA, DHA, and aspirin ingestion on plasma lysophospholipids and autotaxin. *Prostaglandins Leukot Essent Fatty Acids* 82: 87-95. doi:10.1016/j.plefa.2009.12.005. PubMed: 20106646.
  66. Lagarde M, Bernoud N, Brossard N, Lemaitre-Delaunay D, Thies F et al. (2001) Lysophosphatidylcholine as a preferred carrier form of docosahexaenoic acid to the brain. *J Mol Neurosci* 16: 201-221; discussion: 11478375.
  67. Fuentes L, Hernández M, Fernández-Avilés FJ, Crespo MS, Nieto ML (2002) Cooperation between secretory phospholipase A2 and TNF-receptor superfamily signaling: implications for the inflammatory response in atherogenesis. *Circ Res* 91: 681-688. doi:10.1161/01.RES.0000038341.34243.64. PubMed: 12386144.

68. Shi Y, Zhang P, Zhang L, Osman H, Mohler ER 3rd et al. (2007) Role of lipoprotein-associated phospholipase A2 in leukocyte activation and inflammatory responses. *Atherosclerosis* 191: 54-62. doi:10.1016/j.atherosclerosis.2006.05.001. PubMed: 16765356.
69. Park CH, Kim MR, Han JM, Jeong TS, Sok DE (2009) Lysophosphatidylcholine exhibits selective cytotoxicity, accompanied by ROS formation, in RAW 264.7 macrophages. *Lipids* 44: 425-435.
70. Colles SM, Chisolm GM (2000) Lysophosphatidylcholine-induced cellular injury in cultured fibroblasts involves oxidative events. *J Lipid Res* 41: 1188-1198. PubMed: 10946005.
71. Chen G, Li J, Qiang X, Czura CJ, Ochani M et al. (2005) Suppression of HMGB1 release by stearoyl lysophosphatidylcholine: an additional mechanism for its therapeutic effects in experimental sepsis. *J Lipid Res* 46: 623-627. doi:10.1194/jlr.C400018-JLR200. PubMed: 15687351.
72. Yan JJ, Jung JS, Lee JE, Lee J, Huh SO et al. (2004) Therapeutic effects of lysophosphatidylcholine in experimental sepsis. *Nat Med* 10: 161-167. doi:10.1038/nm989. PubMed: 14716308.
73. Hung ND, Kim MR, Sok DE (2011) Oral administration of 2-docosahexaenoyl lysophosphatidylcholine displayed anti-inflammatory effects on zymosan A-induced peritonitis. *Inflammation* 34: 147-160. doi:10.1007/s10753-010-9218-z. PubMed: 20490641.
74. Hung ND, Kim MR, Sok DE (2011) Mechanisms for anti-inflammatory effects of 1-[15(S)-hydroxyeicosapentaenoyl] lysophosphatidylcholine, administered intraperitoneally, in zymosan A-induced peritonitis. *Br J Pharmacol* 162: 1119-1135. doi:10.1111/j.1476-5381.2010.01117.x. PubMed: 21091644.
75. Plückerthun A, Dennis EA (1982) Acyl and phosphoryl migration in lysophospholipids: importance in phospholipid synthesis and phospholipase specificity. *Biochemistry* 21: 1743-1750. doi:10.1021/bi00537a007. PubMed: 7082643.
76. Ma J, Folsom AR, Lewis L, Eckfeldt JH (1997) Relation of plasma phospholipid and cholesterol ester fatty acid composition to carotid artery intima-media thickness: the Atherosclerosis Risk in Communities (ARIC) Study. *Am J Clin Nutr* 65: 551-559. PubMed: 9022543.
77. Pfeiffer A, Böttcher A, Orsó E, Kapinsky M, Nagy P et al. (2001) Lipopolysaccharide and ceramide docking to CD14 provokes ligand-specific receptor clustering in rafts. *Eur J Immunol* 31: 3153-3164. doi:10.1002/1521-4141(200111)31:11. PubMed: 11745332.
78. de Mello VD, Lankinen M, Schwab U, Kolehmainen M, Lehto S et al. (2009) Link between plasma ceramides, inflammation and insulin resistance: association with serum IL-6 concentration in patients with coronary heart disease. *Diabetologia* 52: 2612-2615. doi:10.1007/s00125-009-1482-9. PubMed: 19669729.
79. Wymann MP, Schneider R (2008) Lipid signalling in disease. *Nat Rev Mol Cell Biol* 9: 162-176. doi:10.1038/nrg2324. PubMed: 18216772.
80. Turpin SM, Lancaster GI, Darby I, Febbraio MA, Watt MJ (2006) Apoptosis in skeletal muscle myotubes is induced by ceramides and is positively related to insulin resistance. *Am J Physiol Endocrinol Metab* 291: E1341-E1350. doi:10.1152/ajpendo.00095.2006. PubMed: 16849630.