

Analysis of Gender Differences in Genetic Risk: Association of TNFAIP3 Polymorphism with Male Childhood-Onset Systemic Lupus Erythematosus in the Japanese Population

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Abstract

Background: Systemic lupus erythematosus (SLE) is a systemic multisystem autoimmune disorder influenced by genetic background and environmental factors. Our aim here was to replicate findings of associations between 7 of the implicated single nucleotide polymorphisms (SNPs) in *IRF5*, *BLK*, *STAT4*, *TNFAIP3*, *SPP1*, *TNIP1* and *ETS1* genes with susceptibility to childhood-onset SLE in the Japanese population. In particular, we focused on gender differences in allelic frequencies.

Methodology/Principal Findings: The 7 SNPs were genotyped using TaqMan assays in 75 patients with childhood-onset SLE and in 190 healthy controls. The relationship between the cumulative number of risk alleles and SLE manifestations was explored in childhood-onset SLE. Logistic regression was used to test the effect of each polymorphism on susceptibility to SLE, and Wilcoxon rank sum testing was used for comparison of total risk alleles. Data on rs7574865 in the STAT4 gene and rs9138 in SPP1 were replicated for associations with SLE when comparing cases and controls (corrected P values ranging from 0.0043 to 0.027). The rs2230926 allele of TNFAIP3 was associated with susceptibility to SLE in males, but after Bonferroni correction there were no significant associations with any of the other four SNPs in IRF5, BLK, TNIP1 and ETS1 genes. The cumulative number of risk alleles was significantly increased in childhood-onset SLE relative to healthy controls (P = 0.0000041). Male SLE patients had a slightly but significantly higher frequency of the TNFAIP3 (rs2230926G) risk allele than female patients (odds ratio [OR] = 4.05, 95% confidence interval [95%CI] = 1.46–11.2 P<0.05).

Conclusions: Associations of polymorphisms in *STAT4* and *SPP1* with childhood-onset SLE were confirmed in a Japanese population. Although these are preliminary results for a limited number of cases, *TNFAIP3* rs2230926G may be an important predictor of disease onset in males. We also replicated findings that the cumulative number of risk alleles was significantly increased in childhood-onset SLE.

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1

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Introduction

Systemic lupus erythematosus (SLE) is an autoimmune connective tissue disease which presents with various clinical symptoms and disorders in many internal organs. Disease onset in about 60% of patients occurs between the ages of 16 and 55 years, and approximately 15–20% are diagnosed in adolescence [1]. SLE occurs predominantly in women, with a female-to-male ratio of 10:1. However, for childhood-onset SLE, this ratio is lower, at about 3:1.

Childhood-onset SLE represents 10–20% of all SLE cases, and is associated with greater disease severity than adult-onset SLE, including more rapid renal damage. The majority of childhood-

onset SLE patients will have developed renal damage within 5–10 years of disease onset [2]. Other studies of clinical characteristics indicated that men younger than 33 at the time of diagnosis were more likely to develop lupus nephritis, and that patients of Asian descent tend to have more severe disease, including the renal manifestations [3]. Recent large-scale association studies revealed a relationship between genetic risk factors and patient age at disease onset [4]. Childhood-onset SLE is associated with more frequent occurrence of the various SLE symptoms than seen in adult-onset disease, and has a more serious disease course [1]. Relative to adult-onset SLE, there is a higher genetic risk in childhood-onset SLE; this influences age of disease onset, which is an important predictor of disease severity [4]. Therefore, we

hypothesize that gender-specific genetic effects of childhood-onset SLE are involved in its etiology. To test this hypothesis, here we explore the distribution of known susceptibility loci in a cohort of Japanese childhood-onset SLE patients.

Recent studies have identified the following genes as affecting susceptibility to SLE in Japanese: human leukocyte antigen DRB1 (HLA-DRBI) *15:01, interferon regulatory factor 5 (IRF5) rs41298401C, signal transducer and activator of transcription-4 (STAT4) rs7574865T, B lymphoid tyrosine kinase (BLK) rs13277113A, tumor necrosis factor alpha-induced protein 3 (TNFAIP3) rs2230926G and TNFAIP3-interacting protein 1 (TNIPI) rs7708392C [5–10]. The same risk alleles are associated with SLE in the Caucasian and Chinese populations [11–14]. We previously evaluated associations of polymorphisms in IRF5, BLK, STAT4, TNFAIP3 and TNIP1 genes with susceptibility to SLE in Japanese children [7,9].

Osteopontin (SPP1: secreted phosphoprotein 1) located on 4q22, also called early T lymphocyte activation 1, is a bone matrix mediator with important roles in inflammation and immunity [15]. An association between SPP1 genetic polymorphisms and increased osteopontin protein levels has been reported [16]. Two SPP1 single nucleotide polymorphisms (SNPs) (rs1126772 and rs9138) in the 3' untranslated region were found to be associated with autoimmune/lymphoproliferative syndrome (ALPS). Osteopontin serum levels are associated both with these polymorphisms and an elevated risk of developing ALPS [17]. The same investigators reported that rs7687316 and rs9138 were significantly associated with SLE in an Italian population [18]. Another report implicated rs1126616 and rs9138 in conferring a significantly higher risk of SLE on men than women [19]. Therefore, it was suggested that rs9138 in the SPP1 gene is a specific autosomal gene in male SLE.

E26 transformation-specific 1 (ETSI) is a member of the E26 transformation-specific family of transcription factors. ETSI plays a role in the negative regulation of Th17 cells and in B cell differentiation [20]. Recently, SLE susceptibility genes have been identified in two genome-wide association studies (GWAS) in Asian populations. It was found that genetic variants in ETSI were associated with susceptibility to SLE; reduced expression of ETSI may increase differentiation and activity of both plasma cells and Th17 cells [21,22]. Two SNPs in ETSI (rs6590330 and rs4937333) were found to be significantly related to SLE, and reported to be associated with age of onset [11].

Recently, an association of the cumulative number of risk alleles and SLE has been reported in the United States and Japan [4,23]. Koga et al. analyzed the cumulative number of risk alleles at eight established susceptibility loci, namely *HLA-DRB1*, *IRF5*, *STAT4*, *BLK*, *TNFAIP3*, *TNIP1*, *FCGR2B* and *TNFSF13*, in a Japanese population. Patients were found to carry more risk alleles than healthy controls. The odds ratio (OR) for SLE was significantly increased in individuals carrying 10 or more risk alleles. In contrast, subjects with 4 or less risk alleles had significantly decreased SLE occurrence.

The present study was conducted to determine whether the 7 candidate genes *IRF5*, *BLK*, *STAT4*, *TNFAIP3*, *SPP1*, *TNIP1* and *ETS1* were associated with childhood-onset SLE in the Japanese population. We investigated sex-specific differences in allelic frequencies and the cumulative number of risk alleles. We further examined the relationship between genetic risk for SLE and many clinical manifestations of the disease.

Materials and Methods

Ethics Statement

This study was performed in accordance with the Declaration of Helsinki and approved by the Ethics Committee of Yokohama City University School of Medicine. Written informed consent was given by parents/guardians for all children that participated in this study.

Subjects

The aim of this study was to explore associations between SNPs in the seven genes *IRF5*, *BLK*, *STAT4*, *TNFAIP3*, *SPP1*, *TNIP1* and *ETS1* and susceptibility to childhood-onset SLE in a Japanese population. The distribution of SNPs in these genes confirmed that they acted as SLE susceptibility loci (Table S1) [6–11,19]. We analyzed the genotype data at the seven risk SNPs and investigated associations between cumulative number of risk alleles and susceptibility to SLE. In addition, we examined gender differences in allelic frequencies and cumulative risk for SLE. Moreover, we investigated associations between these SNPs and the different clinical characteristics of the patients.

Study Population and Clinical Data

A total of 75 children, 15 males (20.0%) and 60 females (80.0%), enrolled in this study were followed-up at Yokohama City University Hospital between December 2007 and December 2010. The mean age of patients was about 12 years at onset of SLE (Table 1). All patients fulfilled the American College of Rheumatology (ACR) Criteria score for SLE of 4 or higher [24,25]. We explored gender differences in the clinical manifestations (malar rash, arthritis, serositis, thrombocytopenia, proteinuria and anti-dsDNA antibodies, according to the ACR classification criteria).

Genotyping

SNPs in the *BLK* (rs13277113), *STAT4* (rs7574865), *TNFAIP3* (rs2230926), *SPP1* (rs9138), *TNIP1* (rs7708392) and *ETS1* (rs4937333) genes were selected based on previous research [7–10,21,26]. Seventy five patients with SLE and 190 healthy controls were genotyped. Genomic DNA was isolated from peripheral blood using the QIAamp DNA Mini kit (Qiagen K.K, Tokyo, Japan). Genotyping was performed using TaqMan SNP Genotyping Assays (Applied Biosystems, Foster, CA, USA assay ID: C__1886916_10 for rs13277113, C_29882391_10 for rs7574865, C__7701116_10 for rs2230926, C__8826997_10 for rs9138, C_29349759_10 for rs7708392, C__278298_10 for rs4937333). We developed a custom-made assay for rs41298401 in *IRF5* (forward primer, 5'-GCTGCGCCTGGAAAGC-3'; reverse

Table 1. Demographic characteristics of the SLE population studied.

Age		yrs.	
	Range	3–18	
	Mean	11.9	
Gender		n (%)	
	Males	15 (20.0)	
	Females	60 (80.0)	

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primer, 5'-GGGAGGCGCTTTGGAAGT-3'; probes, VIC-CTGCTGTAGGCACCC and FAM-CTGCTGTAGCCACCC; Custom TaqMan Applied Biosystems, Foster, CA, USA). To confirm the accuracy of genotyping, we conducted bidirectional sequencing on 6 randomly selected individuals for each genotype at rs41298401 and undertook direct sequencing of the SNP. No genotyping errors were found. These SNPs were analyzed by real-time PCR using the AB7500 Real Time PCR system (Applied Biosystems, Foster, CA, USA) under the conditions recommended by the manufacturer.

Statistical Analysis

Hardy-Weinberg Equilibrium (HWE) was measured by chisquared testing for the control groups of each study (significance set at P<0.05). All analyses were performed using IBM SPSS software (SPSS for Windows, Version 19.0, Chicago, IL, USA). The multiple comparison P-values were corrected by the Bonferroni method, in which P values were multiplied by the number of tested SNPs, and P<0.05 considered significant (2sided test).

We compared the clinical manifestations of SLE cases between male and female patients using Fisher's exact probability test. Logistic regression was used to test the effect of each of the 7 SNPs on differences between SLE cases and healthy controls. A genetic additive model (with values: AA = 2, Aa = 1 and aa = 0, respectively) was considered. We analyzed the correlation between the risk for each SNP using a dominant or recessive model in logistic regression analysis. Dominant (risk allele homozygotes = 1, heterozygotes = 1 and nonrisk allele homozygotes = 0) and recessive (risk allele homozygotes = 1, heterozygotes = 0 and nonrisk allele homozygotes = 0) models were tested. Results are expressed as odds ratios (OR) with 95% confidence intervals (CI). Comparison of male and female patients used logistic regression [27], which was also used to test the effect of each polymorphism separately. Gender-specific associations were assessed by comparing the additive model between cases and controls for each gender separately.

The cumulative number of risk alleles was determined in each patient. Wilcoxon rank sum testing was used for comparisons of groups.

Results

Study Population and Clinical Data

We studied 75 cases and 190 controls. The median age at SLE diagnosis was 11.9 years (Table 1). There were 15 male and 60 female patients. All patients and controls (male n=91, female n=99) were Japanese. Female gender was a risk factor for SLE susceptibility (P<0.001, Fisher's exact test). However, proteinuria was higher in males than females with SLE (P=0.026, Fisher's exact test) (Table 2), while malar rash, arthritis, serositis, thrombocytopenia and the presence of anti-dsDNA antibodies were not significantly difference between the sexes.

Associations between the 7 Tested SNPs and Susceptibility to SLE

We conducted an analysis of Hardy-Weinberg equilibrium and genotypic and allelic associations of 7 SNPs with susceptibility to SLE. The genotype frequencies for the SNPs in both patients with SLE and healthy controls were in Hardy-Weinberg equilibrium (P>0.05).

A significant difference in allele distribution of STAT4 rs7574865T between SLE patients and controls (OR = 1.77, 95%CT = 1.20–2.62, P = 0.027) was observed (Table 3). The

genotype frequency of rs9138 in SPP1 was also significantly different between SLE patients and controls (OR = 3.31, 95%CI = 1.67–6.56, P = 0.0043) in the recessive model (AA vs AC/CC) (Table 3). The genotype AA was significantly higher in patients than in controls.

Clinical Subsets

Next, we explored whether these risk alleles were associated with the development of particular clinical manifestations. Frequencies of the rs9138 A-allele in SPP1 (OR = 0.48, 95%CI = 0.25–0.92, P = 0.028) and the rs4937333 T-allele in ETS1 (OR = 2.28, 95%CI = 1.03–5.04, P = 0.042) were significantly increased in SLE patients with proteinuria as compared to those without. There were no significant associations between TNFAIP3 rs2230926G and proteinuria in SLE patients. Malar rash was associated with STAT4 rs7574865T (OR = 3.08, 95%CI = 1.21–7.85, P = 0.019), whereas no significant associations were observed in the other subsets of patients with arthritis, serositis, thrombocytopenia and anti-dsDNA antibodies.

The Cumulative Number of Risk Alleles at the 7 SNPs and Gender Differences

We had selected these 7 established SLE susceptibility alleles due to their reported significance in Asian populations or because they conferred a higher risk of SLE to males than females (Table S1). The cumulative risk was calculated in each patient by summing the risk scores. It was found that the cumulative number of the risk alleles was significantly increased in patients with childhood-onset SLE relative to the healthy controls (P=0.0000041, Wilcoxon rank sum test). In contrast, there were no significant associations with gender itself (P=0.085, Wilcoxon rank sum test).

Secondly, we evaluated the effect of these SNPs separately on the gender difference in SLE susceptibility. A significant gendergene interaction was observed for rs2230926, a polymorphism located in TNFAIP3. The frequency of the risk allele in TNFAIP3 (rs2230926G) was slightly but significantly higher in men than women with SLE (OR = 4.05, 95%CI = 1.46–11.23, P<0.05) (Table 4), whereas there were no differences between men (n = 91) and women (n = 99) in the controls. To further assess the effect of TNFAIP3 (rs2230926), we calculated associations with SLE in gender-specific case-control analysis. Rs2230926 was associated with SLE in males (case n = 15, control n-91) but not in females (case n = 60, control n = 99) (OR male = 6.17, 95%CI = 2.10–18.09, P = 0.0064 corrected; OR female = 1.25, 95%CI = 0.52–3.02, P = 0.62 uncorrected) (Table 5).

Discussion

SLE is a systemic multisystem autoimmune disorder, susceptibility to which is influenced by a complex genetic background and potentially by environmental exposures. Because chronologically, childhood has less time to accumulate environmental exposures, the impact of genetic factors may be easier to disentangle than in adult-onset SLE. Recent studies identified many susceptibility genes relevant to SLE. We have replicated results for 3 of these (STAT4, TNFAIP3 and SPP1) in the present study, but we were unable to replicate the findings of susceptibility for four others (IRF5, BLK, ETS1 and TNIP1). The frequencies of the risk alleles IRF5, BLK, ETS1 and TNIP1 in the present study are similar to those published in other recent Asian studies [6,8,10,11]. We may not have seen a significant difference because of the small size of our study.

Table 2. The clinical and serological manifestations of patients with SLE.

		All	Male (n = 15)	Female (n = 60)	P value*
		N (% of group)	N (% of group)	N (% of group)	
Malar rash	+	49 (76.6)	11 (91.7)	38 (73.1)	0.27
	-	15 (23.4)	1 (8.3)	14 (26.9)	
Arthritis	+	39 (60.0)	6 (50.0)	33 (62.3)	0.52
	-	26 (40.0)	6 (50.0)	20 (37.7)	
Serositis	+	4 (6.3)	1 (10.0)	3 (5.7)	0.51
	-	59 (93.7)	9 (90.0)	50 (94.3)	
Thrombocytopenia	+	19 (27.5)	3 (23.1)	16 (28.6)	1.00
	-	50 (72.5)	10 (76.9)	40 (71.4)	
Proteinuria	+	47 (65.3)	13 (92.9)	34 (58.6)	0.026
	-	25 (34.7)	1 (7.1)	24 (41.4)	
Anti-dsDNA	+	55 (93.2)	11 (91.7)	44 (93.6)	1.00
	-	4 (6.8)	1 (8.3)	3 (6.4)	

*Fisher's exact p value.

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The cumulative number of risk alleles is reported to be significantly increased in SLE relative to healthy controls in the Japanese population [23]. We did replicate this association in

Japanese childhood-onset SLE. A recent study of cumulative genetic risk scores for SLE in men and women concluded that there is a higher genetic risk factor in men [28] and that male

Table 3. Associations of the seven SNPs with SLE in a Japanese population.

						Allele association		Dominant model		Recessive model	
	SNPs	Genotype			Allele	OR (95% CI)	P*	OR (95% CI)	P*	OR (95% CI)	P *
IRF5	rs41298401C	сс	CG	GG	С						
	Case (n = 75)	56 (74.7)	18 (24.0)	1 (1.3)	130 (86.7)	1.21 (0.73–2.02)	n.s.	4.11 (0.52–32.7)	n.s.	1.08 (0.59–1.99)	n.s.
	Control (n = 190)	139 (73.2)	41 (21.6)	10 (5.3)	319 (83.9)						
BLK	rs13277113A	AA	AG	GG	A						
	Case (n = 75)	43 (57.3)	28 (37.3)	4 (5.3)	114 (76.0)	1.54 (0.98–2.42)	0.41	1.63 (0.53–5.05)	n.s.	1.70 (0.99–2.91)	0.39
	Control (n = 190)	84 (44.2)	90 (47.4)	16 (8.4)	258 (67.9)						
STAT4	rs7574865T	TT	TG	GG	Т						
	Case (n = 75)	16 (21.3)	39 (52.0)	20 (26.7)	71 (47.3)	1.77 (1.20–2.62)	0.027	2.27 (1.27–4.09)	0.042	1.97 (0.97–3.98)	0.41
	Control (n = 190)	23 (12.1)	81 (42.6)	86 (45.3)	127 (33.4)						
TNFAIP	rs2230926G	GG	GT	TT	G						
	Case (n = 75)	3 (4.0)	12 (16.0)	60 (80.0)	18 (12.0)	2.02 (1.07-3.81)	0.21	1.82 (0.89–3.71)	0.70	INF**	
	Control (n = 190)	0 (0.0)	23 (12.1)	167 (87.9)	23 (6.1)						
SPP1	rs9138A	AA	AC	cc	Α						
	Case (n = 75)	21 (28.0)	29 (38.7)	25 (33.3)	71 (47.3)	1.61 (1.10-2.38)	0.11	1.28 (0.73–2.24)	n.s.	3.31 (1.67–6.56)	0.0043
	Control (n = 190)	20 (10.5)	96 (50.5)	74 (38.9)	136 (35.8)						
TNIP1	rs7708392C	сс	CG	GG	c						
	Case (n = 75)	40 (53.3)	35 (46.7)	0 (0.0)	115 (76.7)	1.06 (0.66-1.70)	n.s.	INF**		0.89 (0.52–1.52)	n.s.
	Control (n = 190)	107 (56.3)	74 (38.9)	9 (4.7)	288 (75.8)						
ETS1	rs4937333T	TT	TC	cc	Т						
	Case (n = 75)	24 (32.0)	40 (53.3)	11 (14.7)	88 (58.7)	1.40 (0.95-2.04)	n.s.	2.08 (1.02-4.25)	0.32	1.28 (0.72–2.30)	n.s.
	Control (n = 190)	51 (26.8)	89 (46.8)	50 (26.3)	191 (50.3)						

*P values calculated by logistic regression analysis and then corrected by the Bongerroni criterion.

**infinity.

Genotype and allele frequencies are shown in parentheses (%).

n.s., not significant.

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Table 4. Comparison between male and female patients with SLE.

						Allele association			
SNPs		Genotype		Allele	OR (95% CI)	<i>P</i> value	Corrected P value*		
IRF5	rs41298401C	сс	CG	GG					
	Male $(n = 15)$	9 (60.0)	6 (40.0)	0 (0.0)	24 (80.0)	0.51 (0.17–1.53)	0.23	n.s.	
	Female $(n = 60)$	47 (78.3)	12 (20.0)	1 (1.7)	106 (88.3)				
BLK	rs13277113A	AA	AG	GG					
	Male (n = 15)	8 (53.3)	6 (40.0)	1 (6.7)	22 (73.3)	0.83 (0.33-2.10)	0.70	n.s.	
	Female (n = 60)	35 (58.3)	22 (36.7)	3 (5.0)	92 (76.7)				
STAT4	rs7574865T	TT	TG	GG					
	Male (n = 15)	5 (33.3)	9 (60.0)	1 (6.7)	19 (63.3)	2.40 (1.00-5.81)	0.051	0.36	
	Female (n = 60)	11 (18.3)	30 (50.0)	19 (31.7)	52 (43.3)				
TNFAIP3	rs2230926G	GG	GT	TT					
	Male (n = 15)	2 (13.3)	5 (33.3)	8 (53.3)	9 (30.0)	4.05 (1.46-11.23)	0.0071	<0.05	
	Female (n = 60)	1 (1.7)	7 (11.7)	52 (86.7)	9 (75.0)				
SPP1	rs9138A	AA	AC	cc					
	Male (n = 15)	2 (13.3)	7 (46.7)	6 (40.0)	11 (36.7)	0.64 (0.30-1.36)	0.24	n.s.	
	Female (n = 60)	19 (31.7)	22 (36.7)	19 (31.7)	60 (50.0)				
TNIP1	rs7708392C	cc	CG	GG					
	Male (n = 15)	10 (66.7)	5 (33.3)	0 (0.0)	25 (83.3)	2.00 (0.61-6.55)	0.25	n.s.	
	Female (n = 60)	30 (50.0)	30 (50.0)	0 (0.0)	90 (75.0)				
ETS1	rs4937333T	TT	TC	сс					
	Male (n = 15)	7 (46.7)	7 (46.7)	1 (6.7)	21 (70.0)	2.00 (0.79-5.00)	0.14	n.s.	
	Female (n = 60)	17 (28.3)	33 (55.0)	10 (16.7)	67 (55.8)				

*P values calculated by logistic regression analysis and then corrected by the Bongerroni criterion. Genotype and allele frequencies are shown in parentheses (%).

n.s., not significant.

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patients had a higher frequency of clinical manifestations than women [29]. In the present study, we found no significant differences in the number of SLE risk alleles in male and female patients. Female predominance in SLE is clearly evident in general, but the male: female ratio is lower in children with prepubertal onset and onset recently after puberty than at reproductive ages [30]. We suggest that genetic susceptibility is more clearly associated with childhood-onset SLE than adult-onset SLE. We did not find a significant difference in the cumulative number of risk alleles between male and female SLE patients. Therefore, the increased frequency of adult-onset SLE among women may be attributed to differences in environmental factors and the metabolism of sex hormones in particular.

Our findings suggest that susceptibility risks conferred by polymorphisms in the three genes STAT4, TNFAIP3, SPP1 participate in the pathogenesis of childhood-onset SLE, but identified the TNFAIP3 rs2230926G allele as the only one significantly different in male and female patients. This finding suggests that TNFAIP3 rs2230926G may be an important predictor of disease in males. However, this should still be considered a tentative conclusion because there were only 15 male patients in our study, and previous large-scale studies of TNFAIP3 have not identified a similar gender effect. Therefore, a multicenter study of the same ethnic group is needed to validate the results from this study. A20 (also known as TNFAIP3) encodes a ubiquitin-editing protein that acts in the negative regulation of nuclear factor-kappaB (NF-kB) signaling via several pathways including those involving tumor necrosis factor (TNF) and Toll-

like receptors (TLR) [31]. Resent genome-wide association studies (GWAS) have revealed associations between *TNFAIP3* and SLE [32]. The SNPs rs2230926 in *TNFAIP3* introduces the amino acid substitution Phe127 to Cys127. Previous studies demonstrated that *TNFAIP3* risk allele rs2230926G leads to reduced NF-κB inhibition. Levels of *TNFAIP3* mRNA are lower in carriers of the rs2230926 G-allele (GT subjects) than in non-carriers (TT), resulting in less protein expression and less A20 inhibitory activity [9,33]. Further research is needed to determine the influence of gene polymorphisms in the *TNFAIP3* gene on the potential role of A20 protein in the pathogenesis of SLE.

A recent review reported that a high a percentage of childhoodonset SLE patients have renal involvement [34]. In the present study, we observed that SPP1 rs9138C was associated with proteinuria in SLE patients (OR = 2.10, 95%CI = 1.09-4.08, P = 0.028). On the other hand, the frequency of SPP1 rs9138C tended to be decreased in the SLE patients studied here (OR = 0.62, 95%CI = 0.42-0.91, P = 0.15). Comparing the data from our Japanese population with those previously published from a Caucasian population, the frequency of the C allele at rs9138 was found to be significantly increased [26]. SPP1 rs9138C was reported to contribute to SLE susceptibility in a population including both African-American and European-American males, but not females [19]. It can be suggested that SPP1 rs9138C is only associated with increased risk of renal disorders in childhood- but not adult-onset SLE patients. Kariuki et al reported age- and gender- dependent effects of SPP1 (rs9138C) on cytokine profiles in SLE patients. The rs9138C allele is associated with higher

Table 5. Associations of SNPs with SLE in male and female groups separately assessed by logistic regression.

		Genotyp	e			Allele		Allele association			
SNPs		Case (male n = 15, female n = 60) Control (male n = 91, female n = 99)							Control	OR (95% CI)	*P value
IRF5	rs41298401C	сс	CG	GG	сс	CG	GG				
	Male	9 (60.0)	6 (40.0)	0 (0.0)	67 (73.6)	15 (16.5)	9 (9.9)	24 (80.0)	149 (81.9)	0.91 (0.40-2.11)	n.s.
	Female	47 (78.3)	12 (20.0)	1 (1.7)	72 (72.7)	26 (26.3)	1 (1.0)	106 (88.3)	170 (85.9)	1.26 (0.63–2.54)	n.s.
BLK	rs13277113A	AA	AG	GG	AA	AG	GG				
	Male	8 (53.3)	6 (40.0)	1 (6.7)	39 (42.9)	43 (47.3)	9 (9.9)	22 (73.3)	121 (66.5)	1.41 (0.58–3.44)	n.s.
	Female	35 (58.3)	22 (36.7)	3 (5.0)	45 (45.5)	47 (47.5)	7 (7.1)	92 (76.7)	137 (69.2)	1.51 (0.88–2.61)	n.s.
STAT4	rs7574865T	TT	TG	GG	TT	TG	GG				
	Male	5 (33.3)	9 (60.0)	1 (6.7)	13 (14.3)	40 (44.0)	38 (41.8)	19 (63.3)	66 (36.3)	3.03 (1.32–6.97)	0.062
	Female	11 (18.3)	30 (50.0)	19 (31.7)	10 (10.1)	41 (41.4)	48 (48.5)	52 (43.3)	61 (30.8)	1.71 (1.06–2.75)	0.19
TNFAIP3	rs2230926G	GG	GT	TT	GG	GT	TT				
	Male	2 (13.3)	5 (33.3)	8 (53.3)	0 (0.0)	11 (12.1)	80 (87.9)	9 (30.0)	11 (60.4)	6.17 (2.10–18.09)	0.0064
	Female	1 (1.7)	7 (11.7)	52 (86.7)	0 (0.0)	12 (12.1)	87 (87.9)	9 (75.0)	12 (60.6)	1.25 (0.52–3.02)	n.s.
SPP1	rs9138A	AA	AC	cc	AA	AC	cc				
	Male	2 (13.3)	7 (46.7)	6 (40.0)	8 (8.8)	46 (50.5)	37 (40.7)	11 (36.7)	62 (34.1)	0.88 (0.38-2.06)	n.s.
	Female	19 (31.7)	22 (36.7)	19 (31.7)	12 (12.1)	50 (50.5)	37 (37.4)	60 (50.0)	74 (37.4)	0.61 (0.39-0.97)	0.24
TNIP1	rs7708392C	cc	CG	GG	cc	CG	GG				
	Male	10 (66.7)	5 (33.3)	0 (0.0)	49 (53.8)	40 (44.0)	2 (2.2)	25 (83.3)	138 (75.8)	1.76 (0.58–5.31)	n.s.
	Female	30 (50.0)	30 (50.0)	0 (0.0)	58 (58.6)	34 (34.3)	7 (7.1)	90 (75.0)	150 (75.8)	0.96 (0.55–1.66)	n.s.
ETS1	rs4937333T	TT	TC	cc	TT	TC	сс				
	Male	7 (46.7)	7 (46.7)	1 (6.7)	26 (28.6)	45 (49.5)	20 (22.0)	21 (70.0)	97 (53.3)	2.06 (0.88–4.79)	0.67
	Female	17 (28.3)	33 (55.0)	10 (16.7)	25 (25.3)	44 (44.4)	30 (30.3)	67 (55.8)	94 (47.5)	1.39 (0.88–2.18)	n.s.

*P values calculated by logistic regression analysis and then corrected by the Bongerroni criterion. Genotype and allele frequencies are shown in parentheses (%).

n.s., not significant.

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serum osteopontin and interferon-alpha in male and female patients \leq 23 years old. [35] These results are consistent with our findings.

Our study is the first to report an association between risk alleles and clinical manifestations in Japanese children with SLE. In particular, both geography and race affect the prevalence of SLE and the frequency and severity of clinical manifestations. Although our study is of a small single-center cohort, this minimizes any discrepancies in management and assessment, and it reflects the homogeneity of the ethnic and socio-demographic background. In addition, because it focuses exclusively on patients with childhoodonset SLE, it is more appropriate for defining detailed genetic susceptibility in this group than cohorts investigating all SLE patients. Our hospital is a pediatric rheumatology center treating many patients with severe SLE. However, we acknowledge that the current study suffers from some shortcomings. The sample size for male SLE patients is small (15 males). In addition, further research is needed to study relationships between adult-onset SLE and childhood-onset patients. Therefore, it is important that the association of TNFAIP3 and SPP1 polymorphisms with childhoodonset SLE susceptibility is confirmed by other groups.

Conclusions

Associations of STAT4 and SPP1 with childhood-onset SLE were confirmed in a Japanese population. For TNFAIP3, this

genetic effect may be confined to males. We replicated previous findings that the cumulative number of risk alleles of seven known SNPs was significantly increased in childhood-onset SLE relative to controls, while there was no association with the observed gender difference.

Supporting Information

Table S1 Single nucleotide polymorphisms (SNPs) representing the 7 systemic lupus erythematosus (SLE) susceptibility loci genotyped in our study. (XLS)

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Author Contributions

Conceived and designed the experiments: KK MM MY SY MK. Performed the experiments: KK MM MY TM TH T. Kanetaka TN RH TI. Analyzed the data: KK MM MY T. Kaneko. Contributed reagents/materials/analysis tools: KK MM MY. Wrote the paper: KK MM MY T. Kaneko SY.

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