

Lineage-Specific Restraint of Pituitary Gonadotroph Cell Adenoma Growth

Vera Chesnokova¹, Svetlana Zonis¹, Cuiqi Zhou¹, Anat Ben-Shlomo¹, Kolja Wawrowsky¹, Yoel Toledano¹, Yunguang Tong^{1,2,3}, Kalman Kovacs², Bernd Scheithauer³, Shlomo Melmed^{1*}

1 Department of Medicine, Pituitary Center, Cedars-Sinai Medical Center, Los Angeles, California, United States of America, **2** Departments of Pathology, St. Michael's Hospital, Toronto, Canada, **3** Mayo Clinic, Rochester, Minnesota, United States of America

Abstract

Although pituitary adenomas are usually benign, unique trophic mechanisms restraining cell proliferation are unclear. As GH-secreting adenomas are associated with p53/p21-dependent senescence, we tested mechanisms constraining non-functioning pituitary adenoma growth. Thirty six gonadotroph-derived non-functioning pituitary adenomas all exhibited DNA damage, but undetectable p21 expression. However, these adenomas all expressed p16, and >90% abundantly expressed cytoplasmic clusterin associated with induction of the Cdk inhibitor p15 in 70% of gonadotroph and in 26% of somatotroph lineage adenomas ($p=0.006$). Murine L β T2 and α T3 gonadotroph pituitary cells, and α GSU.PTTG transgenic mice with targeted gonadotroph cell adenomas also abundantly expressed clusterin and exhibited features of oncogene-induced senescence as evidenced by C/EBP β and C/EBP δ induction. In turn, C/EBPs activated the clusterin promoter \sim 5 fold, and elevated clusterin subsequently elicited p15 and p16 expression, acting to arrest murine gonadotroph cell proliferation. In contrast, specific clusterin suppression by RNAi enhanced gonadotroph proliferation. FOXL2, a tissue-specific gonadotroph lineage factor, also induced the clusterin promoter \sim 3 fold in α T3 pituitary cells. As nine of 12 pituitary carcinomas were devoid of clusterin expression, this protein may limit proliferation of benign adenomatous pituitary cells. These results point to lineage-specific pathways restricting uncontrolled murine and human pituitary gonadotroph adenoma cell growth.

Citation: Chesnokova V, Zonis S, Zhou C, Ben-Shlomo A, Wawrowsky K, et al. (2011) Lineage-Specific Restraint of Pituitary Gonadotroph Cell Adenoma Growth. PLoS ONE 6(3): e17924. doi:10.1371/journal.pone.0017924

Editor: Alfredo Fusco, Consiglio Nazionale delle Ricerche (CNR), Italy

Received: December 22, 2010; **Accepted:** February 15, 2011; **Published:** March 25, 2011

Copyright: © 2011 Chesnokova et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Funding: This work was supported by grants from the National Institutes of Health, CA75979 (SM) and T32 DK07770 (SM to YT), and the Doris Factor Molecular Endocrinology Laboratory. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing Interests: The authors have declared that no competing interests exist.

* E-mail: Melmed@csmc.edu

Introduction

Pituitary tumors arise from highly specialized cell types expressing the respective pituitary polypeptide hormones. Thus, tumors derived from somatotrophs secrete growth hormone (GH), lactotrophs, prolactin (PRL), thyrotrophs, thyrotropin (TSH), and corticotrophs, adrenocorticotropin (ACTH). In contrast, non-functioning pituitary tumors usually arise from non-secreting cells of gonadotroph origin [1]. Clinically inapparent pituitary tumors are identified in 25% of autopsy specimens with a population prevalence of \sim 77 cases/10⁵. Pituitary tumors are usually benign neoplasms (adenomas), however, they may also exhibit invasive or recurrent growth. Rarely encountered malignant pituitary carcinomas comprise 0.02% of all pituitary tumors, proliferate rapidly and show extracranial metastases [2,3,4]. Although most aggressive pituitary adenomas persistently exhibit low mitotic activity [3], mechanisms underlying these unique growth properties are largely elusive. We postulate that intrinsic cell-specific trophic properties as well as the lineage-origin of highly differentiated and specialized pituitary cells underlies constrained adenoma proliferation.

Cellular senescence is characterized by irreversible proliferative arrest, while cells remain viable and metabolically active. Proliferation arrest may occur as a result of age-related telomere shortening, and also in response to oxidative or genotoxic stress,

DNA damage, aneuploidy or chromosomal instability, as well as oncogene activation [5,6]. Thus, oncogenic RAS causes stable proliferative arrest rather than transformation in diploid fibroblasts [7]. BRAF in benign skin nevi elicits an initial increased proliferation followed by DNA stress and cellular senescence [8]. As cellular senescence appears to be bypassed in advanced malignancies [8,9,10], this antiproliferative mechanism may represent an initial impediment against oncogenic development [11]. Senescence is mediated by activation of p53/p21 and other Cdk inhibitors including p15 and p16 [5]. Oncogene-induced senescence is also associated with a senescence-messaging secretome, enabling senescence responses [11].

Clusterin (CLU), a highly conserved cellular and circulating protein [12], is also known as apolipoprotein J (ApoJ), sulphated glycoprotein 2 (SGP-2), testosterone-repressed prostate message 2 (TRPM-2), or serum protein-40 (SP-40). Intracellular clusterin forms include a partially glycosylated uncleaved pre-secretory protein, a secretory/intracellular glycosylated α - β chain heterodimer, and an anti-apoptotic nuclear form [13,14]. Nuclear clusterin is induced in response to DNA damage evoked by chemotherapeutic agents, and protects some tumorous cells from apoptosis [15,16]. In contrast, intracellular clusterin was also shown to inhibit prostate cancer cell proliferation, and clusterin knockout induced highly aggressive transgenic mouse prostate tumors [17].

Clusterin thus functions as a tumor suppressor gene [18], inhibits cell proliferation [19,20], promotes experimental skin carcinoma differentiation [21], and exhibits features of a cellular stress responder [14].

Pituitary tumor transforming gene (*PTTG*) induction is a hallmark of human pituitary tumors [22,23,24]. *PTTG* was isolated from pituitary tumor cells [25] and *PTTG* abundance also correlates with breast, thyroid, endometrial, esophageal and colorectal tumor invasiveness [26]. *PTTG* facilitates cell cycle progression [22,25] and when over-expressed, causes cell transformation [22] and promotes tumor formation *in vivo* [27]. Transgenic *PTTG* over-expression targeted to pituitary gonadotroph cells results in focal pituitary adenoma formation [28], while *Pttg* deletion abrogates murine pituitary tumor development [29]. *PTTG* is the index mammalian securin [30], and both *Pttg* deletion [31] as well as overexpression [27] result in aneuploidy and chromosomal instability highlighting the requirement for intracellular securin equilibrium to maintain chromosomal stability [32,33].

In most human GH-producing pituitary adenomas *PTTG* overexpression is associated with DNA damage and p21-dependent senescence [34], however pathways restraining growth and transformation of the more commonly encountered non-functioning pituitary adenomas are not known. We show here that similar to GH-cell adenomas, tumors arising from the gonadotroph lineage exhibit high *PTTG* levels and DNA damage.

However, unlike GH-cell adenomas, p53/p21 senescence markers are not activated in non-functioning adenomas, which do, however, selectively express abundant cytoplasmic clusterin. High clusterin levels restrain cell proliferation by triggering Cdk inhibitors p15, p16 and p27, while suppression of clusterin expression enhanced pituitary gonadotroph cell proliferation. Thus, we identify a novel role for clusterin in enabling pituitary gonadotroph tumor cell proliferation arrest. FOXL2, a transcription factor specifically expressed in pituitary gonadotroph cells [35] stimulates the clusterin promoter, further highlighting a differential lineage-specific pathway restricting pituitary cell cycle progression, acting to buffer non-functioning pituitary adenomas from unrestrained growth.

Results

DNA damage and senescence markers are induced in human pituitary adenomas

Immunoreactive *PTTG* was induced in all 36 gonadotroph cell adenomas analyzed, but not in normal pituitary tissue (Figure 1A), confirming previous reports [22,23,24]. Markers of DNA damage and aneuploidy including γ H2A.X foci and phosphorylated kinase mutated in ataxia telangiectasia (pATM) [36,37] were not detected by fluorescent immunohistochemistry in two non-tumorous human pituitary specimens. In contrast, all of 12 human pituitary adenomas analyzed (2 GH cell, 10 non-secreting

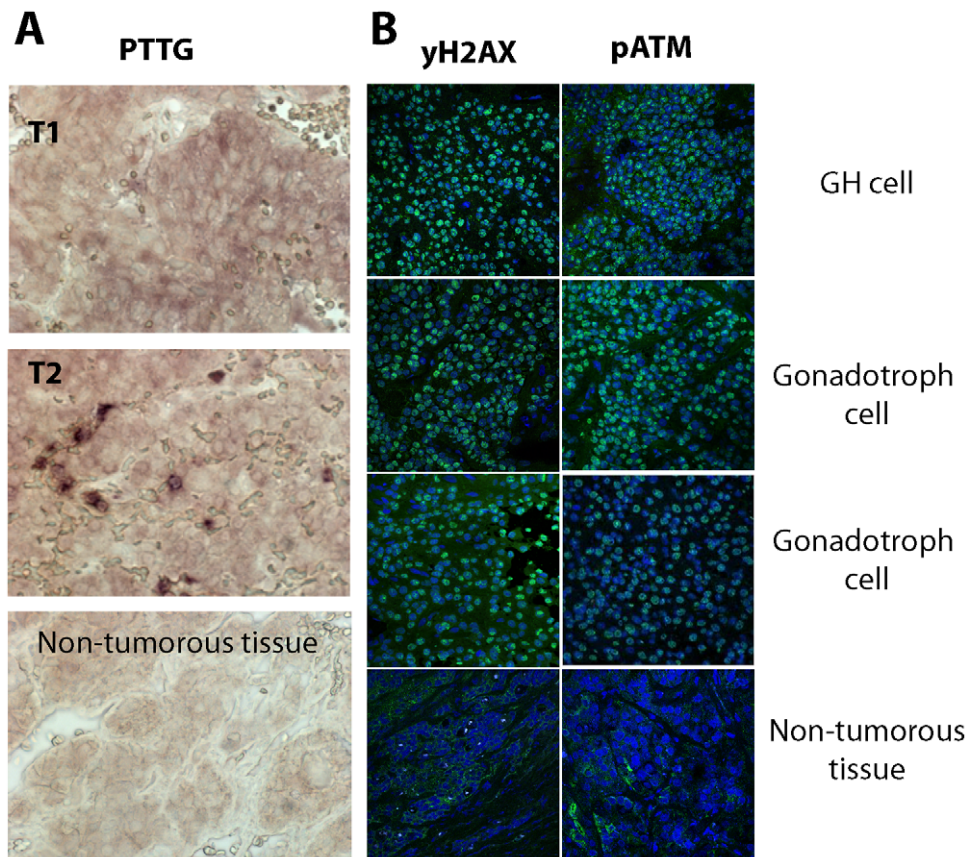


Figure 1. PTTG and DNA damage in human pituitary adenomas. **A**) PTTG immunoreactivity (brown signal, both intranuclear and cytoplasmic) in non-tumorous pituitary and human pituitary gonadotroph adenomas (T-1,2); **B**) DNA damage in human pituitary adenoma. Confocal image of human pituitary adenoma and non-tumorous pituitary tissue specimens labeled with γ H2A.X or pATM antibodies (green). Both proteins are expressed in the nucleus. Specimens here and shown below were counterstained with DNA-specific dye ToPro3 (blue). doi:10.1371/journal.pone.0017924.g001

gonadotroph cell) invariably expressed both γ H2A.X and pATM, reflecting activated DNA damage signaling (Figure 1B).

We showed earlier that human GH-secreting adenomas, but not carcinomas, abundantly express intra-nuclear p21, an end-point inhibitor of cell proliferation in the senescence pathway [29,34]. In contrast, gonadotroph cell adenomas did not express p21 (Figure 2). However, because these tumors also do not (with exceedingly rare exceptions) evolve to malignancy, we analyzed additional pathways restraining pituitary tumor cell proliferation. p16 and p15 of the ARF/INK Cdk inhibitor family act to restrain cellular proliferation in response to activated oncogenes [8,10], and were strongly expressed in gonadotroph adenomas. Seventy percent of gonadotroph adenomas expressed high levels of p15, as compared to 26% of GH-secreting adenomas ($p = 0.006$, Table 1). Thus, both DNA damage pathways and senescence markers were expressed in gonadotroph cell-derived adenomas (Figure 2).

Clusterin is expressed predominantly in gonadotroph cell adenomas

As Affymetrix datasets (www.oncomine.org) showed that clusterin mRNA expression was low in advanced metastatic cancers [18], we measured clusterin immunofluorescence in non-tumorous pituitary tissue, and in pituitary adenoma cell types (Figure 3). Of 4 non-tumorous pituitary tissues, 2 exhibited intermediate (<50% positivity), while 2 cases exhibited very low (0–5%) clusterin levels. Only 1 of 17 GH/PRL-secreting adenomas (6%) expressed high clusterin levels, 3 expressed moderate levels, while this protein was minimally detectable in 13 such adenomas. In contrast, 33 of 36 (92%) of the more commonly encountered non-functioning adenomas exhibited abundant clusterin cytoplasmic immunopositivity. We also analyzed 12 pituitary carcinomas (9 corticotroph cell and 3 PRL cell). None of PRL cell carcinomas express clusterin, only 1 corticotroph

cell carcinoma (8%) showed high clusterin levels, 2 (16%) exhibited moderate clusterin levels, while clusterin was undetectable in 6 corticotroph cell carcinoma samples. Thus, 9 of 12 (76%) carcinomas did not express clusterin (Figure 3 and Table 1). Three different adenoma types (GH-, PRL- and gonadotroph) were each analyzed for co-localization of clusterin with respective pituitary hormone markers. Moderately expressed clusterin in GH-secreting adenomas did not co-localize with GH, nor with PRL in PRL cell adenomas. In contrast, clusterin strongly co-localized with α GSU in all 3 gonadotroph cell adenomas analyzed (Figure 3B).

α GSU.PTTG mice exhibit features of oncogene-induced pituitary senescence

We recapitulated human gonadotroph tumors in an *in vivo* transgenic murine model of gonadotroph *PTTG* expression driven by the α GSU promoter [28]. α GSU.*PTTG* pituitary glands express up-regulated gonadotroph *PTTG* with pituitary hyperplasia starting from 4 months of age leading to development of focal pituitary adenomas expressing LH. Other transgenic lines also expressed GH and PRL [38]. In accordance with evidence supporting proto-oncogenic properties of *PTTG* [26], hyperplastic pre-tumorous pituitary glands derived from transgenic animals were shown to already express markers of increased pituitary proliferation as evidenced by increased BrdU incorporation (Figure 4A), and elevated levels of pro-proliferative proteins including PCNA and E2F1 *in vivo* (Figure 4B). However, as these animals developed penetrant pituitary tumors only after 10 months, and these invariably remain small, we tested whether pituitary *PTTG* overexpression also affects anti-proliferative pathways in these transgenic mice.

DNA damage was already evident in pre-tumorous transgenic pituitary glands overexpressing *PTTG* as evidenced by enhanced pituitary γ H2A.X and pATM levels, accompanied by induced

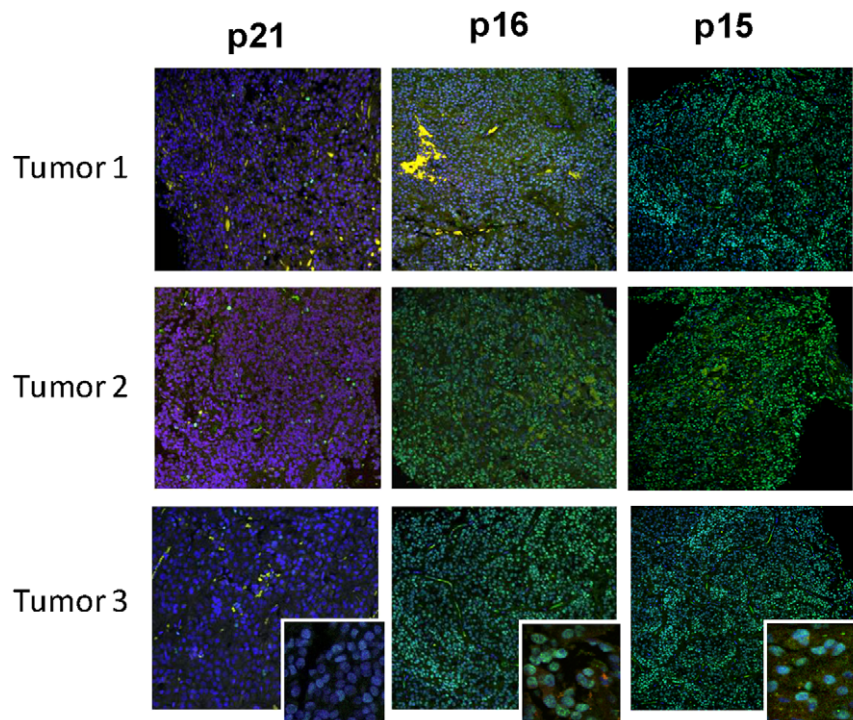


Figure 2. Cdk inhibitor expression in human pituitary adenomas. Confocal image of human gonadotroph pituitary adenoma specimens labeled with p21, p16 or p15 antibodies (green, intranuclear). doi:10.1371/journal.pone.0017924.g002

Table 1. p15 and clusterin expression in human pituitary adenomas.

Intensity (%)	p15 n (%)			Clusterin n (%)				
	0-5	6-50	51-100	0-5	6-50	51-100		
Tumor Types	<i>n</i>			<i>n</i>				
GH/PRL	27	16(59)	4(15)	7(26)	17	13(76)	3(18)	1(6)
Gonadotroph	43	8(18)	5(12)	30(70)	36	1(3)	2(6)	33(92)
ACTH	14	2(14)	1(7)	11(79)	10	1(10)	0(0)	9(90)
Carcinoma	N/A				12	9(76)	2(16)	1(8)
Normal pituitary	5	1(20)	2(40)	2(40)	4	2(50)	2(50)	0(0)

Protein distribution differences between GH/PRL and gonadotroph tumors, $p=0.0006$ for p15, and $p=0.0001$ for clusterin (Wilcoxon Rank Sum Test).
N/A- not available.

doi:10.1371/journal.pone.0017924.t001

DNA damage repair proteins including MSH2, MLH1 and Rad51, as well as tumor suppressors including p19 and p53 (Figure 4C). The Cdk inhibitor p27, a marker of DNA damage [39], was also induced in pre-tumorous α GSU.PTTG pituitary glands, as were the cell cycle suppressor proteins p15 and p16 (Figure 4D). Two intracellular forms of pituitary clusterin, a mature glycosylated ~76 kDa secretory form and ~60 kDa pre-secretory form were up-regulated in the pretumorous transgenic pituitary gland (Figure 4D).

These results were confirmed by fluorescent immunostaining. Although only modest cytoplasmic clusterin, and intra-nuclear p15 and p16 expression were observed in WT murine pituitary

glands, expression of these 3 proteins was enhanced in the transgenic pre-tumorous pituitary, and further induced in α GSU.PTTG pituitary tumors (Figure 4E). Thus, features of oncogene-induced senescence in the α GSU.PTTG pituitary included induction of the p19/p53/p27 DNA damage pathway, and both p15 and p16 Cdk inhibitors [7,40]. In the pre-tumorous hyperplastic α GSU.PTTG pituitary gland the observed increased SA- β galactosidase activity supported the presence of cellular senescence (Figure 4F).

Pttg over-expression in L β T2 cells results in a senescent phenotype

To recapitulate *in vivo* effects of pituitary *Pttg* over-expression, we transiently transfected murine gonadotroph-derived L β T2 cells with a plasmid expressing murine *Pttg*, and also isolated L β T2 cells stably overexpressing *Pttg*. As shown in Figure 5, *Pttg* overexpression lead to induction of clusterin and p15 in both gonadotroph cell transfectants, similar to *in vivo* patterns observed in the α GSU.PTTG pituitary (Figure 4D).

Clones of stably transfected L β T2 cells sorted and selected for high PTTG expression, showed lower rates of BrdU incorporation as compared to control vector-expressing cells, reflecting decreased proliferation (Figure 5C). These transfectants were spread-out, and larger in size with giant aneuploid nuclei, consistent with a senescent phenotype (Figure 5D). High *Pttg* expression resulted in decreased apoptosis as detected by TUNEL assay (Figure 5E), and these cells also exhibited increased SA- β -galactosidase activity (Figure 5 F,G). Thus, constitutively high gonadotroph cell *Pttg* expression resulted in premature cellular senescence similar to the *in vivo* pituitary phenotype observed in α GSU.PTTG mice.

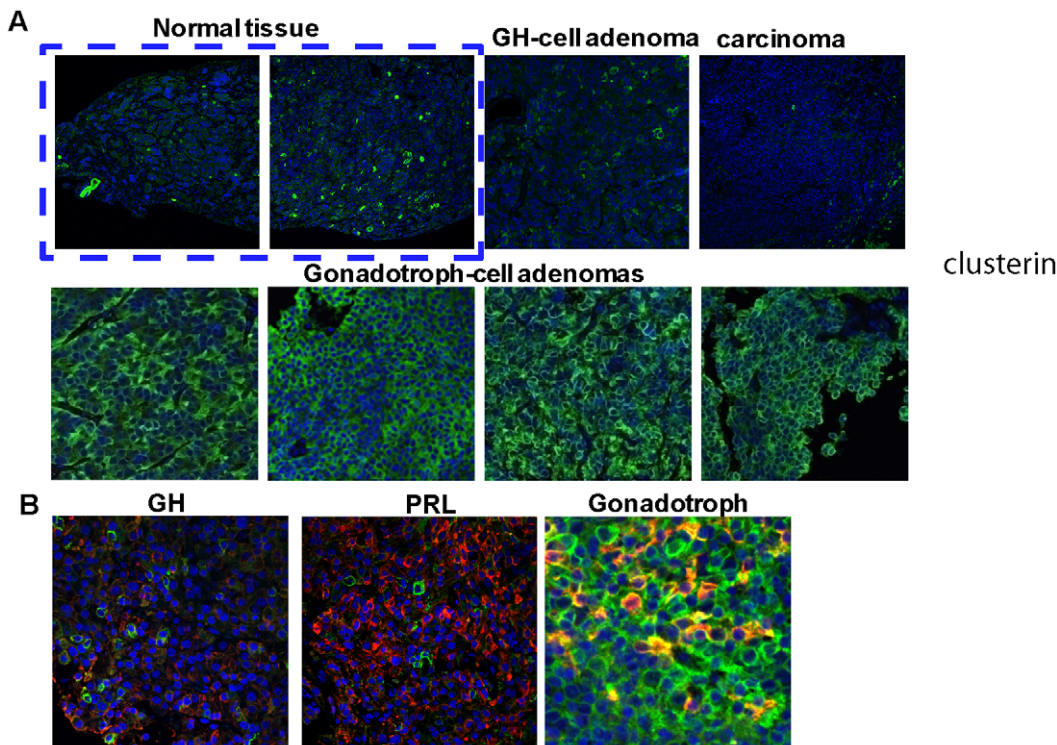


Figure 3. Clusterin in human pituitary adenomas. Confocal image of **A**) human pituitary adenoma and non-tumorous pituitary tissue specimens showing clusterin (green) expressed exclusively in the cytoplasm; **B**) Co-localization of clusterin with GH, PRL and α GSU in respective human pituitary adenoma specimens (clusterin green, respective hormones red).

doi:10.1371/journal.pone.0017924.g003

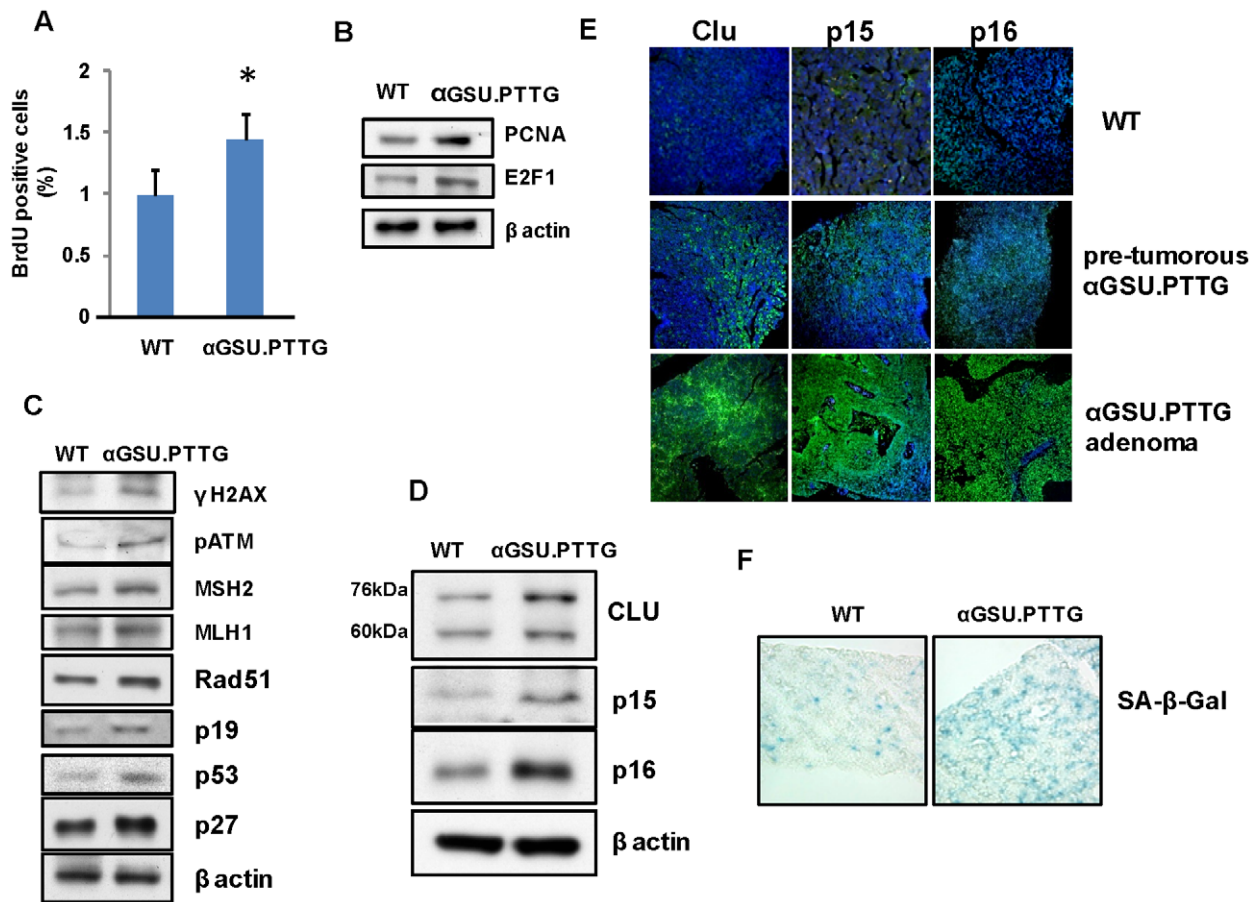


Figure 4. Pituitary proliferation, DNA damage and senescence markers in the α GSU.PTTG pituitary gland. **A)** *In vivo* BrdU incorporation. Mice were injected with BrdU (50 μ g/g BW), and pituitary sections stained for BrdU. One thousand cells/section, 3 sections/animal, $n=3$ animals/genotype were analyzed. *, $p<0.05$; Western blot analysis of **B)** proliferation markers; **C)** DNA damage, DNA repair and p53-dependent senescence markers, and **D)** oncogene-induced senescence markers; **E)** Confocal image showing immunofluorescent cytoplasmic clusterin, and intranuclear p15 and p16 expression (green) in WT and in pre-tumorous α GSU.PTTG pituitary glands, and in α GSU.PTTG pituitary adenomas; **F)** Pituitary SA- β -galactosidase enzymatic activity (blue) in WT and in pre-tumorous α GSU.PTTG pituitary gland. Three pituitary cryosections/animal were analyzed from 3 animals/genotype, and a representative image shown. Western blots here and elsewhere were repeated 3 times with similar results and representative blots shown.
doi:10.1371/journal.pone.0017924.g004

C/EBPs induce pituitary cell clusterin

C/EBP transcription factors are involved in cellular proliferation and differentiation [41,42,43]. In the pre-tumorous hyperplastic α GSU.PTTG pituitary, C/EBP β was induced both in α GSU, and in GH- and PRL-secreting cells (Figure 6A). In L β T2 cells stably expressing *Pttg*, several C/EBP β and C/EBP δ isoforms [41] were also up-regulated (Figure 6B).

We therefore assessed whether C/EBPs activate the clusterin promoter in L β T2 cells, and also in a murine gonadotroph-derived α T3 cell line. pGL3-*mClu*-luc reporter plasmid co-transfected with full length murine C/EBP β or with C/EBP δ constructs (Figure 6C) resulted in induced luciferase activity in both cell types. However, the plasmid encoding C/EBP α did not induce the clusterin promoter, indicating the specificity of C/EBP β and δ effects. Accordingly, clusterin protein levels were also found to be up-regulated in cells transfected with pcDNA3-C/EBP β and δ respectively (Figure 6D,E).

As L β T2 *mPttg* cells exhibit increased clusterin levels, we treated these cells with siRNAs directed against either C/EBP β or C/EBP δ . However, separate suppression of either of these genes resulted in compensatory increase of the other protein (data not

shown), while simultaneous suppression of C/EBP β and δ with 3 nM of each RNAi lead to decreased clusterin protein levels in L β T2 *mPttg* cells after 48 hours. These experiments were conducted with two different siRNA combinations directed against both C/EBP β and δ , and a representative Western blot is shown in Figure 6F. The results confirmed that both C/EBP proteins act to regulate gonadotroph cell clusterin expression.

Clusterin restrains pituitary cell proliferation

As high clusterin expression was observed in benign gonadotroph adenomas and in small slow-growing α GSU.PTTG pituitary tumors (Figures 3 and 4E), and also in L β T2 cells overexpressing *mPttg* (Figure 5A,B), we analyzed the effects of altering intracellular clusterin levels. Transient transfection with *mClu*-pIRES2-ZsGreen1 resulted in increased p15, p16, and p27 expression in L β T2, and p16 in the α T3 cell line. In contrast, levels of phosphorylated histone H3 (pH 3), a specific marker for S and M phases were attenuated in both cell types after clusterin transfection (Figure 7A,B).

Senescence-associated heterochromatic foci (SAHF) mark concentrated spots of transcriptionally silenced DNA [44]. This

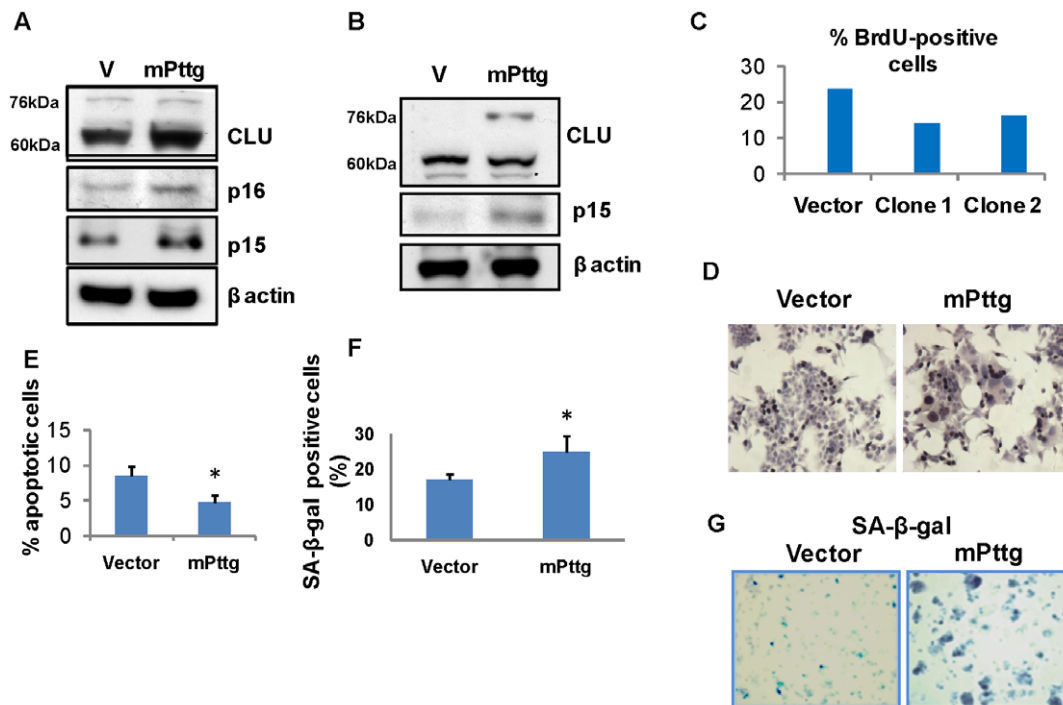


Figure 5. Senescence markers in gonadotroph-derived L β T2 cells transfected with mPttg. Western blot analysis of senescence markers in **A)** L β T2 cells transiently transfected with mPttg; **B)** in L β T2 cells stably transfected with mPttg; **C)** Percent BrdU positive cells in two selected clones stably transfected with mPttg. Duplicate samples were pulsed with BrdU for 30 min and analyzed by flow cytometry; **D)** Senescent morphology of L β T2 cells stably transfected with mPttg. Brown dots depict incorporated BrdU; **E)** Percent apoptotic cells stably transfected with mPttg. Cells were fixed, and one thousand cells/field counted in three randomly chosen visual fields; **F)** Percent SA- β -galactosidase positivity in cells stably transfected with Pttg was assessed in 6-well plates in triplicate. One thousand cells/field were counted in three fields/well. **G)** SA- β -galactosidase enzymatic activity (blue) in cells stably transfected with mPttg. *, $p < 0.05$. doi:10.1371/journal.pone.0017924.g005

localization is accompanied by focal accumulation of specific heterochromatin-associated modified histone. Methylation of lysine 9 of histone H3 (K9M-H3) [8,44] is associated with restraint of cellular proliferation [44]. Using a specific antibody in L β T2 cells transiently transfected with mClu, we observed 30% increase in H3-K9M-positive cells (Figure 7 C,D), concordant with decreased murine gonadotroph cell proliferation observed when clusterin was overexpressed (Figure 7E).

After synchronization of α T3 cells stably overexpressing mClu, and adding 10% FBS, cells were pulsed with BrdU, and flow cytometry demonstrated that BrdU incorporation was decreased, reflecting attenuated DNA synthesis (Figure 7F).

Next we suppressed clusterin expression in L β T2 mPttg stable transfectants, and also tested α T3 cells, where endogenous clusterin levels were relatively high. Both cell lines were transfected with 6 nM of two different siRNAs directed against clusterin, and the pRS shClu-GFP RNA expressing plasmid. Treatments with siClu#2 and transfection with shClu-GFP both depleted clusterin mRNA by 80% as measured by real time PCR (data not shown). Figure 8 (A,B) shows Western blots of representative experiments using siClu#2 where both clusterin protein forms were down-regulated, p15 and p16 levels were low, and pH 3 protein levels induced in both cell lines. In L β T2mPttg cells and in α T3 cells, clusterin suppression led to increased numbers of cells incorporating BrdU respectively ($p < 0.05$) (Figure 8C). Depletion of clusterin by shClu RNA resulted in 30 and 26% increase in BrdU-positive L β T2mPttg cells and α T3 cells respectively (data not shown).

The results show that in pituitary gonadotroph cells, induced clusterin restrains proliferation associated with up-regulated p15

and p16, while clusterin depletion led to decreased p15 and p16, accompanied by increased cell proliferation. Indeed, suppression of p15 and p16 transcription by respective siRNAs markedly increased the number of BrdU-incorporated L β T2mPttg cells 48 hours after transfection, reflective of increased cell proliferation (Figure 8 D,E).

FOXL2 activates clusterin promoter in gonadotroph pituitary cells

Forkhead box gene transcription factor L2 (FOXL2) is a cell-specific factor for pituitary gonadotroph differentiation and triggers α GSU expression [35]. FOXL2 is abundantly expressed in human pituitary gonadotroph and null cell adenomas [45], and in normal pituitary co-localizes with LH, FSH and α GSU [35]. As distribution of FOXL2 appeared to mirror clusterin expression in pituitary adenomas, we tested whether FOXL2 stimulates clusterin in gonadotroph cells. pGL3-mClu-luc reporter plasmid was co-transfected with the murine FOXL2 (pcDNA3-His-Foxl2) in α T3 cells (Figure 9A), and luciferase activity was induced ~3 fold ($p < 0.01$) indicating the stimulatory effect of FOXL2 on the clusterin promoter. Clusterin protein levels were also enhanced in cells transfected with pcDNA3-His-Foxl2 (Figure 9B). In contrast, clusterin was not induced in ACTH-secreting AtT20 murine corticotroph pituitary cells transfected with pcDNA3-His-Foxl2 (data not shown), indicating cell specificity of FOXL2 action on the clusterin promoter.

We then tested whether FOXL2 is recruited to the endogenous clusterin promoter. Lysates derived from α T3 pituitary gonadotrophs were isolated and chromatin immunoprecipitation (ChIP)

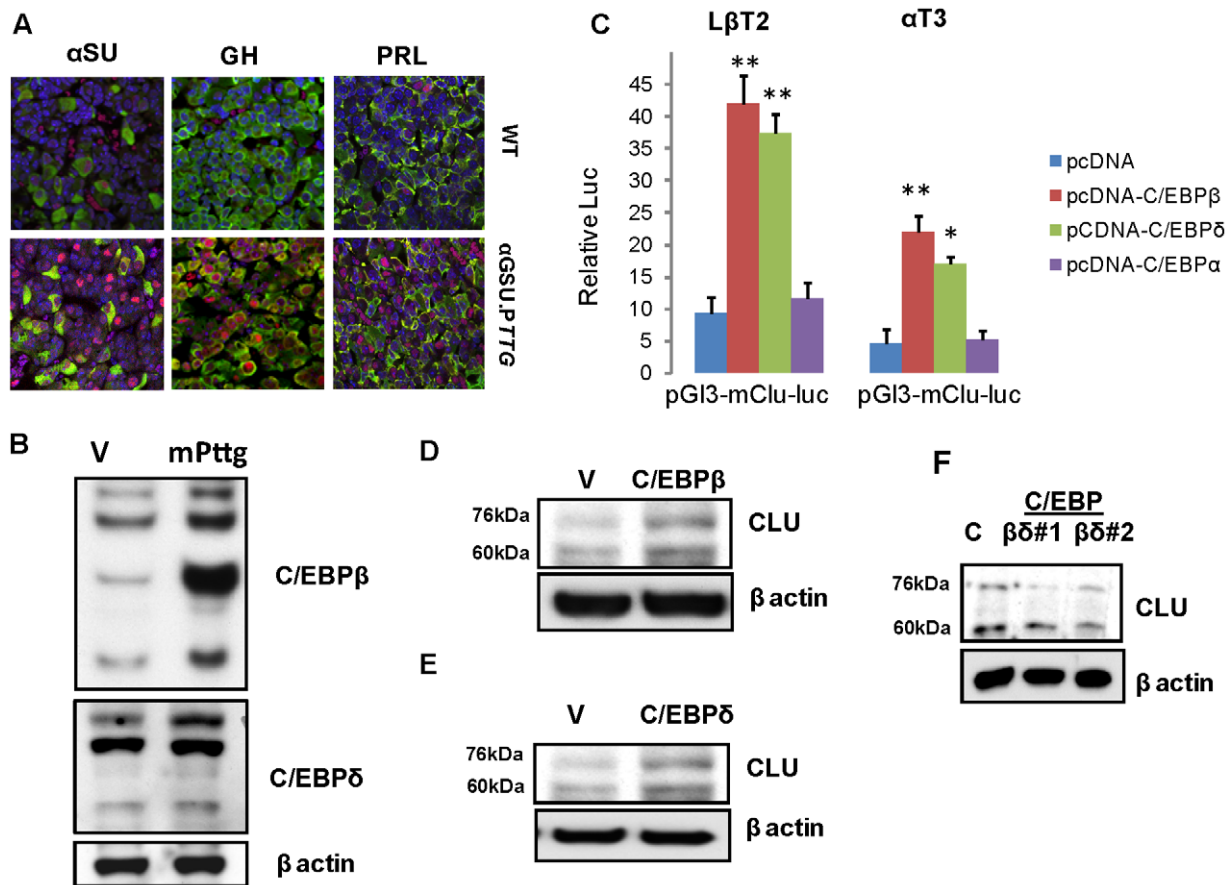


Figure 6. C/EBPs induce clusterin. **A)** C/EBP β is up-regulated in the α GSU.PTTG pituitary. Confocal image showing C/EBP β co-localization with α GSU-positive, GH-positive and PRL-positive cells in WT and pre-tumorous α GSU.PTTG pituitary glands. (Hormones-green, cytoplasmic, C/EBP β -red, intranuclear); **B)** Western blot analysis of C/EBP β and δ isoforms induced in L β T2 cells stably transfected with mPttg; **C)** Effects of C/EBPs on the clusterin promoter in L β T2 and α T3 cells 24 h after transfection. Cells were co-transfected with 200 ng murine pGL3-luc-mClu reporter plasmid and 800 ng murine pCDNA3-C/EBP α , β or δ . The ratio of luciferase to co-transfected β -galactosidase control reporter vector was normalized to pCDNA3-null expression vector. SEM was calculated from triplicate assays, and experiments repeated three times with similar results. Results of a representative experiment are shown. *, $p < 0.05$, **, $p < 0.01$; **D)** Western blot analysis of clusterin expression in gonadotroph-derived α T3 cells 24 hours after transfection with pCDNA3-C/EBP β or **E)** pCDNA3-C/EBP δ ; **F)** Western blot analysis of clusterin expression in L β T2 mPttg cells 48 hours after simultaneous transfection with siC/EBP β and siC/EBP δ (3 nM each). Two different combinations of siRNAs were used. doi:10.1371/journal.pone.0017924.g006

assays performed with a polyclonal FOXL2 antibody. FOXL2 was shown to bind the clusterin promoter, spanning -700 – 1339 nucleotides upstream from the transcription start site (Figure 9C), but did not bind the -1468 – 1865 or $+1$ – 700 promoter regions (not shown). Enrichment of specific -700 – 1339 clusterin promoter sequences in the precipitate indicated FOXL2 association with the clusterin promoter *in vivo*.

Discussion

Pituitary lineage specificity determines highly distinct peripheral pituitary trophic hormone actions as exemplified by differentiated GH, PRL, ACTH, TSH or gonadotroph hormone functions. Cell-specific pituitary hormone synthesis is regulated by specific hypothalamic, intrapituitary and peripheral hormone signals [46,47,48,49,50]. Pituitary cells are also sensitive to aneuploidy, DNA damage or oncogene overexpression. In response to these insults, we show here that cell-type specific trophic pathways are activated, with the common end-point of pituitary cell proliferation arrest.

GH secreting tumors exhibit high PTTG, features of aneuploidy, chromosomal instability, activation of DNA damage responses,

p21-dependent cell proliferation restraint and senescence [34]. Although high PTTG levels are also observed in gonadotroph pituitary adenomas [26], unlike GH-cell adenomas, we now show that gonadotroph adenomas do not express p21, but abundantly express clusterin in a cell specific manner. In contrast, only modest clusterin expression was observed in non-tumorous pituitary glands and in GH/PRL-secreting tumors, while clusterin was undetectable in 76% of the very rarely encountered pituitary carcinomas.

The results in human tumors were validated using *in vivo* and *in vitro* models of gonadotroph cell adenomas with high PTTG expression. Reflecting pro-proliferative properties of excess PTTG, α GSU.PTTG pituitary glands exhibit microadenoma formation [38], also evidenced by increased *in vivo* pituitary BrdU incorporation, and up-regulated proliferation markers including PCNA and E2F1. Similar to human pituitary adenomas, in the α GSU.PTTG pituitary gland, high PTTG levels result in aneuploidy and chromosomal instability, also evidenced by DNA damage, pATM induction and activation of p53/p27 pathways known to arrest cell proliferation in the course of continuing DNA damage [39]. Concordantly, both the pre-tumorous α GSU.PTTG

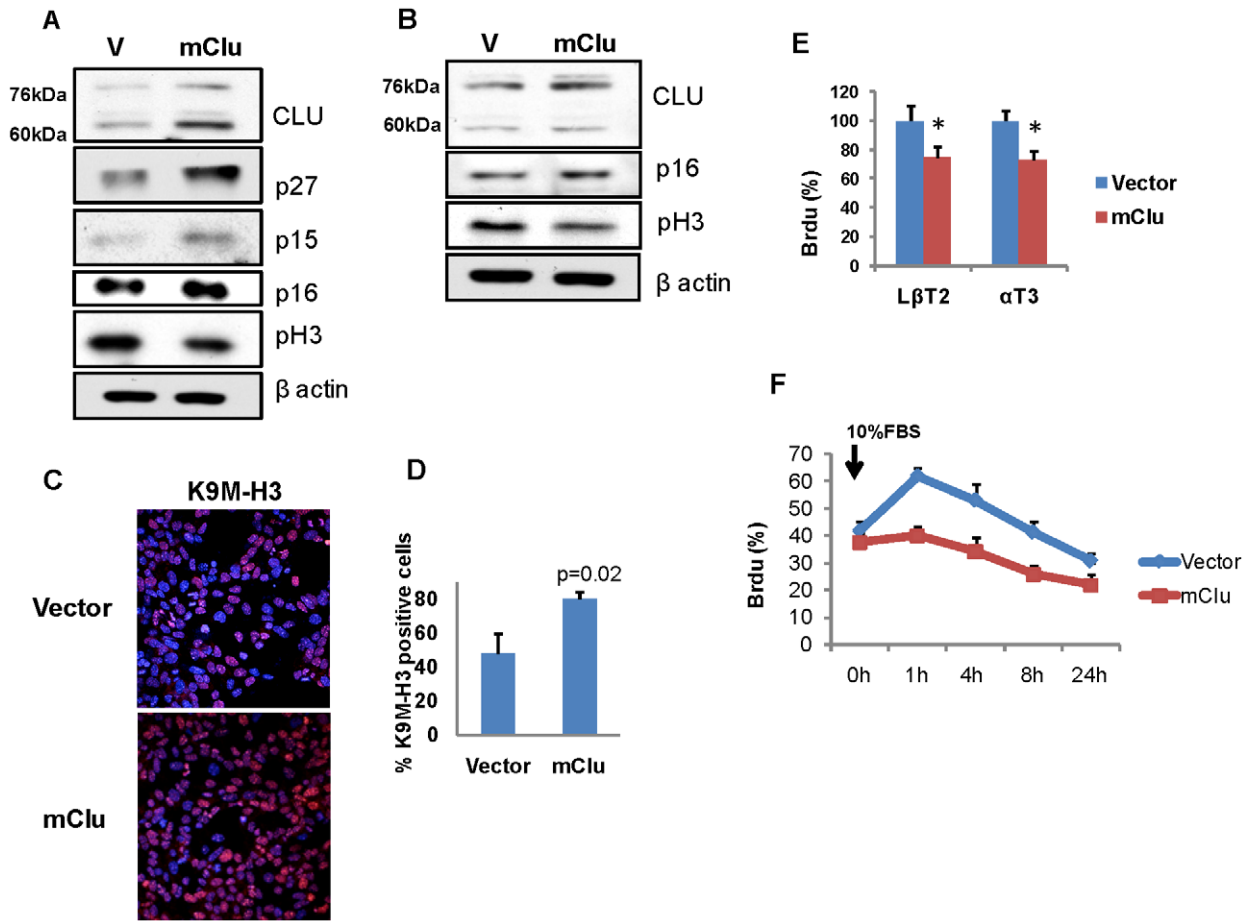


Figure 7. Clusterin restrains pituitary cell proliferation by inducing Cdk inhibitors. Western blot analysis of Cdk inhibitors and proliferation markers **A**) in LβT2 cells, **B**) in αT3 cells 48 h after transfection with mClu; **C**) Confocal images of immunofluorescence of histone H3 methylation on lysine 9 (H3-K9M) (red) in vector and Clu-expressing αT3 cells 48 hours after transfection; **D**) Quantification of positive H3-K9M foci. Cells were fixed, stained with H3-K9M antibody, and one thousand cells/field counted in three randomly chosen visual fields; **E**) Percentage of BrdU positive cells 48 h after transfection with mClu. Triplicate samples were pulsed with BrdU for 30 min and analyzed by flow cytometry, *, p<0.05; **F**) αT3 cells stably overexpressing mClu or vector were synchronized in 0.1% fetal bovine serum for 18 hours, and then cultured in 10% fetal bovine serum. At the indicated times, duplicate samples were pulsed with BrdU for 30 min, analyzed by flow cytometry, and cells in S-phase identified by staining with BrdU antibodies.

doi:10.1371/journal.pone.0017924.g007

pituitary gland and pituitary tumors express high levels of clusterin, p15 and p16.

The results suggest a biphasic response to transforming effects of excess PTTG *in vivo*: Abundant PTTG is apparently sufficient to trigger an initial proliferative burst leading to hyperplasia and tumor initiation, however, inability of these pituitary tumors to undergo persistent further growth, is likely due to proliferation-restraining pathways activated by PTTG overexpression. These findings underscore the observed SA-β-galactosidase activation in pre-tumorous αGSU.PTTG glands and in LβT2 cells constitutively expressing Pttg.

High clusterin levels were observed in the αGSU.PTTG pituitary gland and in LβT2 cells stably and transiently transfected with mPttg, suggesting that in human gonadotroph adenomas clusterin might be also induced by high PTTG levels. In gonadotroph cells, PTTG-overexpression is accompanied by C/EBP induction. C/EBP proteins uniquely regulate cell-type specific growth and differentiation [41,42]. C/EBPβ is associated with oncogene-induced senescence [51], while C/EBPδ triggers growth arrest and cell differentiation [43,52,53]. Both C/EBPβ and C/EBPδ are shown here to activate the clusterin promoter,

and induced clusterin protein expression was evident in gonadotroph cells and pituitary tissue overexpressing PTTG. Furthermore, a specific gonadotroph cell lineage transcription factor FOXL2, independently activates the clusterin promoter in these cells.

We show that forced clusterin expression in LβT2 and αT3 pituitary gonadotroph cells triggers a lineage-specific cytostatic response, inducing p15, p16, or p27; decreased cell proliferation was also evidenced by lower expression of pH 3, similar to observations in prostate cancer cells [54]. Accordingly, when either p15 or p16 gene expression were suppressed, pituitary cell proliferation was enhanced. These results are in accordance with those showing that TGFβ-induced p15 decreases proliferation and induces cell cycle arrest in rat GH₃ pituitary cells [55]. Thus, clusterin-triggered p15 and p16 likely restrain pituitary cell proliferation in αGSU.PTTG pituitary tumors and in LβT2 gonadotroph-derived cells. Induced p15 in human gonadotroph adenomas might therefore limit growth of these tumors.

Clusterin function in tumorigenesis is unclear. Clusterin expression is enhanced in human prostate cancer, and antisense oligonucleotides targeting clusterin inhibit prostate tumorigenesis

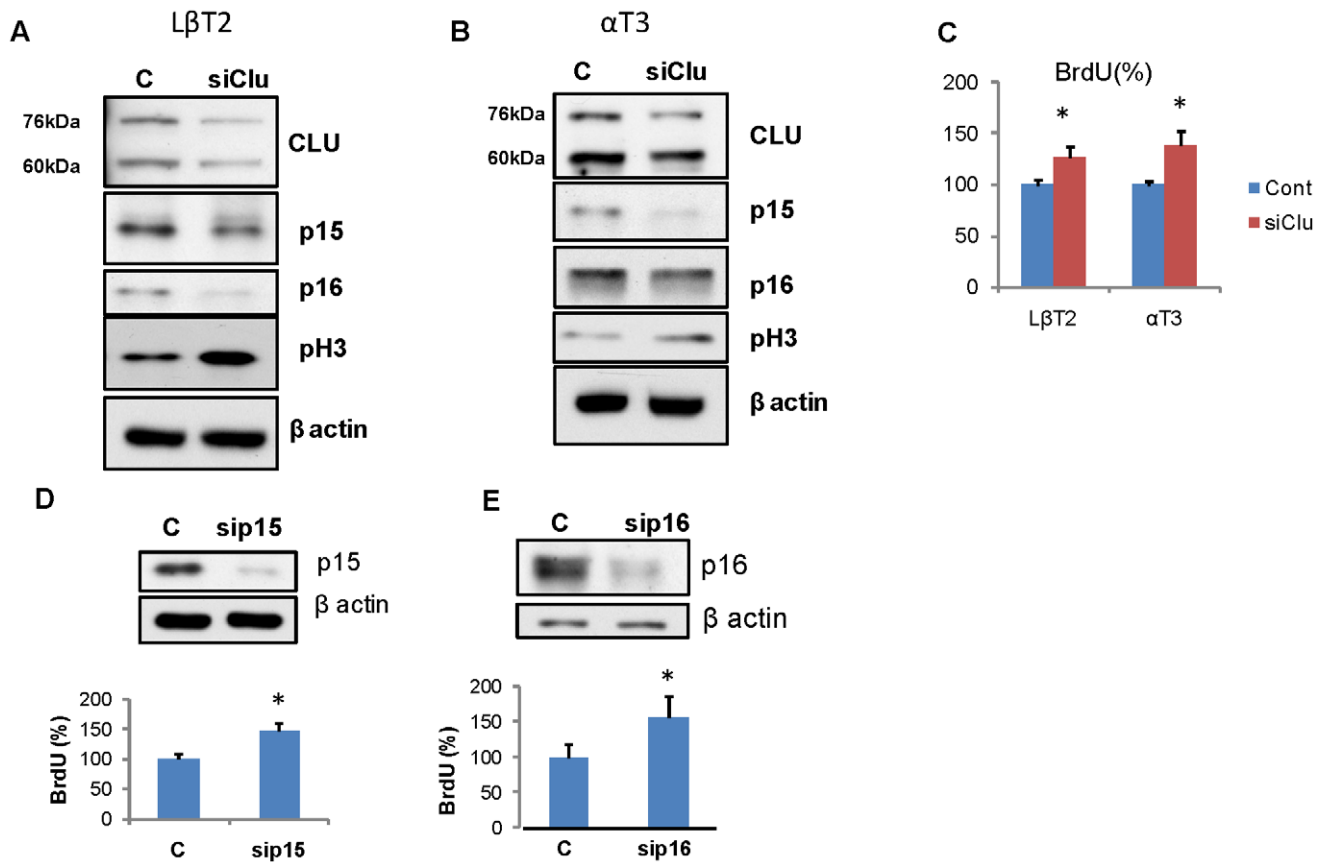


Figure 8. Clusterin attenuation promotes proliferation. Western blot analysis of Cdk inhibitors and proliferation markers **A**) in L β T2 cells, **B**) in α T3 cells; **C**) Percentage of BrdU positive cells 48 h after transfection with siClu. **D**) Upper panel, Western blot confirms p15 down-regulation, Lower panel, Percentage of BrdU positive L β T2 cells 48 h after transfection with sip15. **E**) Upper panel, Western blot confirms p16 down-regulation, Lower panel, Percentage of BrdU positive L β T2 cells 48 h after transfection with sip16. For BrdU detection, cells were fixed, stained with BrdU antibody and one thousand cells/field in three randomly chosen fields counted. *, $p < 0.05$. doi:10.1371/journal.pone.0017924.g008

[56]. Clusterin also induces breast cancer cell growth and metastatic progression [57] and is associated with human lung adenocarcinoma cell growth [58]. The nuclear anti-apoptotic form of clusterin is induced in late stage cancers following chemotherapy, hormonal ablation or radiotherapy, thus protecting tumor cells undergoing damaging stress [59]. As pituitary carcinomas are rarely treated with radiation or chemotherapy before surgery, we did not observe clusterin expression in human carcinoma specimens, as expected. Several lines of evidence also point to the role of clusterin as a tumor suppressor protein. Thus, clusterin was down-regulated, and its expression inversely proportional to tumor grade/or metastatic stage [16,18,60]. Patients with clusterin-positive lung cancer have enhanced disease-free survival [61]. Moreover, *Clu*^{-/-} mice are more prone to oncogene-induced tumorigenesis [17,21]. Although clusterin restrains proliferation of untransformed epithelial cells [21] and acts primarily as a tumor-suppressor during early stages of carcinogenesis [16,21], when re-expressed in advanced cancers, clusterin might promote tumor growth.

Based on the results presented here, we propose that in benign pituitary tumor cells of gonadotroph origin, the role of clusterin is to restrain proliferation. Similar affects were demonstrated for TGF β 1, which functions as a tumor suppressor in normal epithelial cells and during early stages of tumor development [62]. In late-stage tumors TGF β 1 exhibits features of a tumor

promoter, modulating vascular and immune compartments of the tumor stroma [63]. Similarly, in normal fibroblasts [32] and in benign pituitary tumor cells (here and [34]), high PTTG restrains the cell cycle and leads to senescence. In transformed tumor cell lines and in malignant tumors, overexpressed PTTG triggers production of FGF-2 and VEGF-A, cell cycle progression and angiogenesis, and in malignant tumors, high PTTG levels correlate with tumor invasiveness and serve as a marker of poor prognosis (reviewed in [26]). Thus, the cellular environment appears to determine end-point effects of these proteins.

Our hypothesis is outlined in Figure 10. Human gonadotroph tumors express both FOXL2 and PTTG. The results shown here indicate that FOXL2 directly activates the clusterin promoter, while PTTG triggers clusterin via C/EBPs. Both C/EBPs and clusterin are also induced by DNA damage (data not shown and [59]). High clusterin, in turn, provokes p15, p16 and p27 expression *in vivo* and cell-specifically *in vitro* thus restraining gonadotroph cell proliferation. These observations point to the existence of intrinsic lineage-specific pathways restricting pituitary cell cycle progression. Activation of these pathways should be considered as a contributing factor underlying the overwhelmingly benign nature of pituitary adenomas, enabling maintenance of vital pituitary homeostatic and metabolic functions, while protecting the hormone-secreting gland from destruction by malignancy.

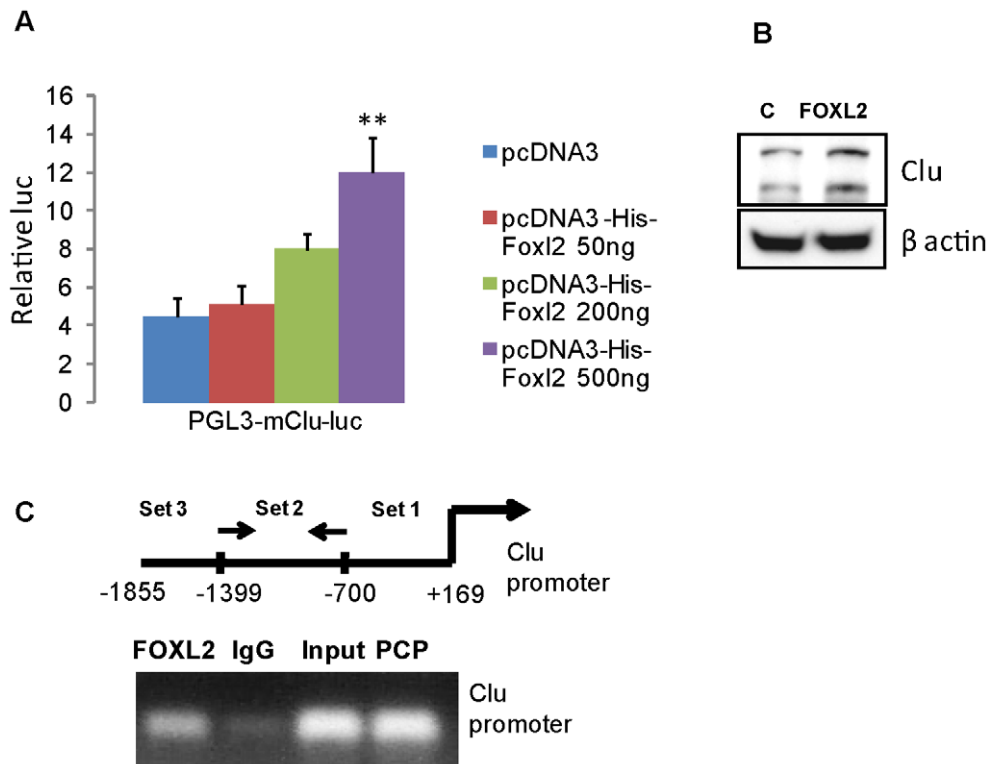


Figure 9. FOXL2 stimulates the clusterin promoter. **A)** Effects of FOXL2 on the clusterin promoter in α T3 cells 24 h after transfection. Cells were co-transfected with 200 ng murine pGL3-luc-mClu reporter plasmid and indicated amounts of pcDNA3-His-mFoxl2. The ratio of luciferase to co-transfected β -galactosidase control reporter vector was normalized to pcDNA3-null expression vector. SEM was calculated from triplicate assays, and experiments repeated three times with similar results. Results of a representative experiment are shown; **, $p < 0.01$; **B)** Western blot analysis of clusterin expression in α T3 cells 24 hours after transfection with pcDNA3-His-mFoxl2; **C)** ChIP assay was performed in nuclear fractions derived from α T3 cell lysates. Top, schematic of the approximate location of primers used in the PCR reactions. Enrichment of specific clusterin promoter sequences was obtained with primer Set 2. FOXL2, specific antibody, IgG, nonspecific antibody, PCP, positive control primers. The experiment was repeated twice, and results of a representative assay shown.
doi:10.1371/journal.pone.0017924.g009

Methods

Human tissue samples

Pituitary tumors were freshly collected at transsphenoidal surgery according to an approved Cedars Sinai and Mayo Clinic Institutional Review Board protocols. Written informed consent were obtained from all participants. Samples were formalin-fixed and paraffin-embedded for immunohistochemistry. Diagnosis of individual tumors was established on the basis of clinical features, histology, and pituitary hormone immunohistochemistry. Non-functioning tumors exhibiting gonadotroph-cell markers, including alpha-glycoprotein subunit (α GSU), LH or FSH were selected for study. Normal anterior pituitary tissue controls were freshly obtained at surgery.

Animals

Experiments were approved by the Cedars Sinai Institutional Animal Care and Use Committee (protocol # 2683). Mice in a B6C3 genetic background harboring the α GSU-*PTTG1*-IRES-eGFP (α GSU.*PTTG*) transgene were previously described [28]. To obtain WT and α GSU.*PTTG* mice from the same breeding, we crossbred α GSU.*PTTG*^{+/−} males and females, and genotyped by PCR.

BrdU incorporation in vivo

Mice were injected i.p. with BrdU (50 μ g/g BW, Sigma-Aldrich, St Louis, MO) three times at 3 hour intervals, sacrificed

24 h after the first injection and pituitary sections stained (5-BrdU Labeling and Detection Kit, Roche, Palo Alto, CA). Three randomly chosen visual fields (1000 cells/per field) were counted, and three sections/per animal were derived from three of each genotype analyzed.

SA- β -galactosidase activity

Senescence-associated (SA)- β -galactosidase enzymatic activity was detected in pituitary cryosections (10 μ m) using a β -galactosidase staining kit (Senescence Cell Staining Kit, Sigma-Aldrich). Only senescent cells stain at pH 6.0. SA- β -galactosidase activity *in vitro* was assayed in 6-well plates in triplicate [29]. Three randomly chosen visual fields/per well were identified, and 1000 cells/per field counted.

Protein analysis

Pituitary tissues or cells were processed (Immunoprecipitation Kit, Roche Diagnostics, Indianapolis, IN) for Western blot analysis, proteins separated by SDS-PAGE, electroblotted onto Millipore membranes (Millipore, Temecula, CA), and incubated overnight with antibodies, followed by corresponding secondary antibodies (Sigma-Aldrich, St. Louis, MO). Antibodies purchased from Santa Cruz (CA): PCNA, E2F1; and MSH2, MLH1, Rad51, p27, p16, Clu. PTTG, Ki67, p53, C/EBP β , C/EBP δ were obtained from Abcam (Cambridge, MA). We also used antibodies to p15 (Biosource); phosphoHistone 3 (ser10) (Cell Signaling

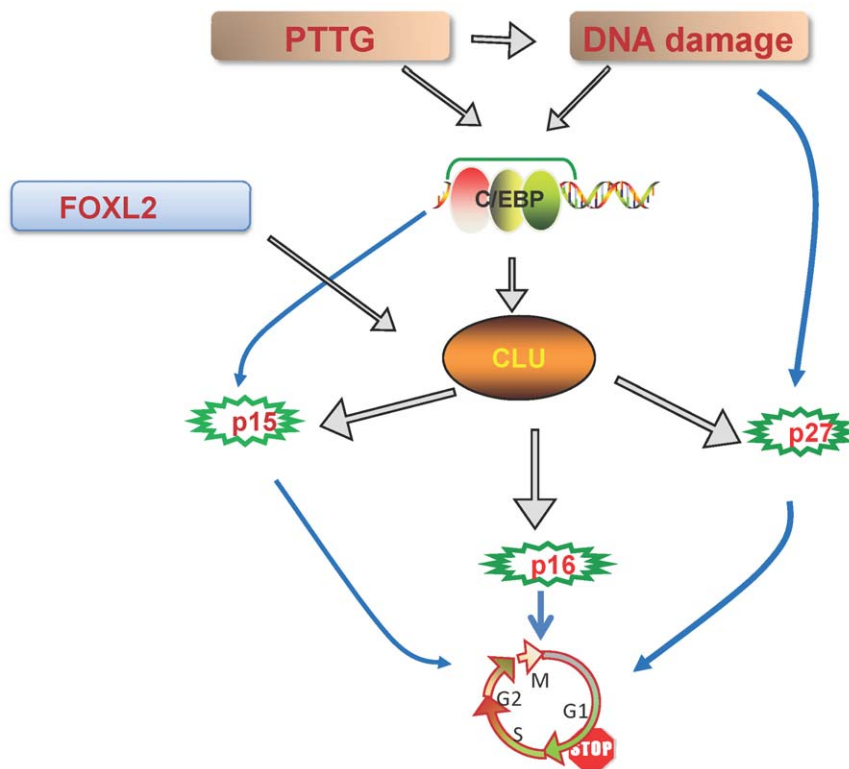


Figure 10. Proliferation restricting pathways in the pituitary gonadotroph cell lineage. FOXL2 directly activates the clusterin promoter, while *Pttg* overexpression results in proliferation, DNA damage and stimulation of C/EBP β and δ ; C/EBPs activate the clusterin promoter. High levels of secretory clusterin trigger expression of Cdk inhibitors p15, p16 and p27, and C/EBP β also cooperates to induce p15. Up-regulated tumor suppressor proteins likely underlie proliferation restraint preventing uncontrolled growth of benign pituitary adenomas of gonadotroph cell origin.
doi:10.1371/journal.pone.0017924.g010

Technology, Danvers, MA), and β actin (Sigma-Aldrich, St. Louis, MO).

For immunofluorescence analysis of human tissue we used antibodies to γ H2A.X (ser139), CLU and H3-K9M antibody, all from Millipore (Billerica, MA); phosphoATM (ser1981, Upstate Biotech (Millipore)); p15, p16, all from Abcam; and p21 (Cell Signaling) antibodies followed by corresponding secondary antibodies conjugated with Alexa 488 or with Alexa 568 fluorescent dye (both Molecular Probe, Carlsbad, California). Antigen retrieval was performed in 10 mM sodium citrate, and control reactions were devoid of primary antibodies or stained with blocking antibodies. Samples were imaged with a Leica TCS/SP spectral confocal scanner (Leica Microsystems, Mannheim, Germany) in dual emission mode to distinguish autofluorescence from specific staining.

Human pituitary adenoma PTTG was detected by immunohistochemistry with the same antibodies as for Western blot, and using an avidin-biotin-peroxidase kit (Vector Laboratories, Burlingame, CA).

Cells, constructs, plasmids and transfections

Mouse pituitary gonadotroph L β T2 and α T3 cell lines were generously provided by Dr. Pamela Mellon (UC San Diego). These cells, immortalized with SV40 T-antigen [64], are the only functional gonadotroph cell lines available.

Murine testis mRNA was used as a source for *Pttg1*. Primers were designed as follows: forward, GGAATTCATGGC-TACTCTTATCTT, reverse CGGGATCCCCGAATATCTG-CATCGT. The Expand High Fidelity PCR system (Roche Diagnostics) was used for amplification reactions. PCR products

were double digested (EcoR1/BamH1), purified and ligated (DNA ligation Kit, Takara Bio, Japan) into pIRES2-ZsGreen1 vector (Clontech, Mountain View, CA) to generate cells that co-express *Pttg* and a ZsGreen tag.

Mouse clusterin expressing plasmids were amplified from pCMv6-*mClu* (Origene, Rockville, MD) by using TaKaRa LA Taq (Takara), and cloned into the pIRES2-ZsGreen1 vector (Clontech). The following primers were used for PCR: clusterin forward: 5' CGGAATTCATGAAGATTCTCCTGCTGTGCGT 3'; reverse: 5' CGG GAT CCTCATTCCGCACGGC-TTTTCCT 3'.

Mouse clusterin promoter fragment (−1855 to +169) was amplified from mouse genomic DNA using TaKaRa LA Taq, inserted into the pGL3-Basic luciferase reporter vector (Promega, San Luis Obispo, CA) and the following primers used for PCR: forward 5' GGG GTA CCA CAT TCC TCC AAG TTT CTG 3', and reverse 5' CGG GAT CCA TGG GCT CTA GTC ACC TC 3'.

L β T2 and α T3 cells were stably transfected with m*Pttg*-pIRES2-ZsGreen1 or with m*Clu*-pIRES2-ZsGreen1 to create L β T2m*Pttg* cells or α T3m*Clu* cells respectively, or with pZsGreen1-N1 alone (vector). Cells were grown in the presence of 400 μ g/ml geneticin (Invitrogen, Carlsbad, CA). Fourth- and fifth-generation enriched stable L β T2m*Pttg* or α T3m*Clu* or vector cells were used for experiments.

Short hairpin RNA expressing vector targeting murine clusterin sh*Clu*-pGFP-V-RS (sh*Clu*) were purchased from OriGene (Rockville, MD). Small interfering RNAs targeting murine clusterin, p15 and p16 (si*Clu*, 1 and 2; sip15, 1 and 2; sip16, 1 and 2) and scrambled siRNAs as negative controls are from Ambion (Foster

City, CA). Small interfering RNAs targeting murine C/EBP β , C/EBP δ and scrambled siRNA are from Qiagen (Valencia, CA). Murine pcDNA3.1(-)C/EBP β was from Addgene (plasmid12557), and murine pcDNA3 C/EBP δ was a generous gift from Dr. Koeffler (Cedars Sinai Medical Center). As fetal bovine serum contains clusterin, cell transfections with m*Clu*, sh*Clu* and si*Clu* were conducted in 0.5% fetal bovine serum (FBS). Transfections were performed using Lipofectamine 2000 (Invitrogen, Carlsbad, CA).

pcDNA3-His-mFoxl2 overexpressing plasmid was a generous gift from Dr. Wei-Hsiung Yang (Mercer University School of Medicine, Savannah, GA) with the permission of Dr. Buffy S. Ellsworth (Southern Illinois University School of Medicine, Carbondale, IL).

Luciferase Assays

L β T2 and α T3 cells were transfected with 800 ng pGL3-luc basic vector or pGL3-luc-m*Clu* reporter plasmid and co-transfected with pcDNA3 or pcDNA3 encoding murine C/EBP β or δ in 12-well plates. Twenty four later cells were harvested and monitored for luciferase activity (Promega, Madison, WI). Light emission was evaluated by luminometer, and normalized to a β -galactosidase luciferase reference plasmid.

Chromatin Immunoprecipitation Assay (ChIP)

ChIP was performed (Chip-IT Express kit, Active Motif, Carlsbad, CA) using α T3 pituitary gonadotroph cells which exhibit abundant clusterin expression. Cells were cross-linked with formaldehyde, harvested, sonicated, nuclear fraction isolated, and chromatin immunoprecipitation performed with polyclonal FOXL2 antibody (Abcam) as well as non-specific mouse IgG. DNA released from precipitated complexes was amplified by PCR with 3 pairs of specific primers spanning 1855 nucleotides upstream from the murine clusterin transcription start site. Primer set 1 (+169–700): F 5'CTTTCCTACCCAGCGCCGC-3', R 5'-ACCCTGCGCAGCTTCCACC-3'; Primer set 2 (–700–1399): F 5'-GGACAGCAGAGCCCTTCGGGA-3', R 5'-GGGCTGCTTGCTGGTCCCTTG-3'; Primer set 3 (–1400–1855): F 5'-AGGCCAAGGCAGAGTGGT-3', R 5'-ACACAGTTTG-GGTGGCAGGCC-3'. PCR products were resolved in 1% agarose gel and visualized by ethidium bromide.

References

- Kovacs K, Horvath E, Vidal S (2001) Classification of pituitary adenomas. *J Neurooncol* 54: 121–127.
- Fernandez A, Karavitaki N, Wass JA (2010) Prevalence of pituitary adenomas: a community-based, cross-sectional study in Banbury (Oxfordshire, UK). *Clin Endocrinol (Oxf)* 72: 377–382.
- Melmed S (2003) Mechanisms for pituitary tumorigenesis: the plastic pituitary. *J Clin Invest* 112: 1603–1618.
- Scheithauer BW, Gaffey TA, Lloyd RV, Sebo TJ, Kovacs KT, et al. (2006) Pathobiology of pituitary adenomas and carcinomas. *Neurosurgery* 59: 341–353; discussion 341–353.
- Sharpless NE, DePinho RA (2004) Telomeres, stem cells, senescence, and cancer. *J Clin Invest* 113: 160–168.
- Serrano M, Blasco MA (2001) Putting the stress on senescence. *Curr Opin Cell Biol* 13: 748–753.
- Serrano M, Lin AW, McCurrach ME, Beach D, Lowe SW (1997) Oncogenic ras provokes premature cell senescence associated with accumulation of p53 and p16INK4a. *Cell* 88: 593–602.
- Michaloglou C, Vredeveld LC, Soengas MS, Denoyelle C, Kuilman T, et al. (2005) BRAF600-associated senescence-like cell cycle arrest of human naevi. *Nature* 436: 720–724.
- Collado M, Gil J, Efeyan A, Guerra C, Schuhmacher AJ, et al. (2005) Tumour biology: senescence in premalignant tumours. *Nature* 436: 642.
- Braig M, Lee S, Lodenkemper C, Rudolph C, Peters AH, et al. (2005) Oncogene-induced senescence as an initial barrier in lymphoma development. *Nature* 436: 660–665.

Apoptosis

The rate of apoptosis was assessed *in vitro* using the *In Situ* Cell Death Detection Kit, AP (TUNEL assay, Roche Diagnostics, Indianapolis, IN). Three randomly chosen visual fields, each containing 1000 cells were counted.

Cell proliferation assay

Asynchronous cells were pulsed with 10 μ M BrdU (Sigma, St Louis, Missouri) in phosphate buffered saline for 30 min at 37°C. Cells were washed, harvested, fixed in 75% ethanol/50 mM glycine at pH 2.0 and analyzed by FACScan (Becton Dickinson, Mountain View, California). Experiments were performed in triplicate. In separate experiments α T3m*Clu* or α T3vector cells were synchronized in 0.1% FBS for 18 hours, and then stimulated by addition of 10% FBS. At the indicated times, duplicate samples were pulsed with BrdU for 30 min, analyzed by flow cytometry, and S-phase cells identified by staining with BrdU antibodies.

Statistics

Clusterin and p15 protein expression were compared between human GH/PRL- and gonadotroph cell pituitary tumors using Wilcoxon Rank Sum Test. Cellular labeling indices were analyzed using ANOVA followed by non-parametric t-test (Mann-Whitney) or Student t-test. Probability of $p < 0.05$ was considered significant.

Acknowledgments

The authors are grateful to Drs. Mark Greene, Yukko and Masahide Tone for helpful discussions and to Dr. Serguei Bannykh for help with human tumor specimens.

Author Contributions

Conceived and designed the experiments: VC SM. Performed the experiments: VC SZ CZ KW Y. Toledano Y. Tong. Analyzed the data: VC AB-S KW KK BS SM. Contributed reagents/materials/analysis tools: KW Y. Toledano Y. Tong. Wrote the paper: VC SM.

- effect on vascular smooth muscle cells and cytoprotective effect on endothelial cells. *Arterioscler Thromb Vasc Biol* 29: 1558–1564.
20. Sivamurthy N, Stone DH, Logerfo FW, Quist WC (2001) Apolipoprotein J inhibits the migration, adhesion, and proliferation of vascular smooth muscle cells. *J Vasc Surg* 34: 716–723.
 21. Thomas-Tikhonenko A, Viard-Leveugle I, Dews M, Wehrli P, Sevignani C, et al. (2004) Myc-transformed epithelial cells down-regulate clusterin, which inhibits their growth in vitro and carcinogenesis in vivo. *Cancer Res* 64: 3126–3136.
 22. Zhang X, Horwitz GA, Heaney AP, Nakashima M, Prezant TR, et al. (1999) Pituitary tumor transforming gene (PTTG) expression in pituitary adenomas. *J Clin Endocrinol Metab* 84: 761–767.
 23. Salehi F, Kovacs K, Scheithauer BW, Lloyd RV, Cusimano M (2008) Pituitary tumor-transforming gene in endocrine and other neoplasms: a review and update. *Endocr Relat Cancer* 15: 721–743.
 24. Filippella M, Galland F, Kujas M, Young J, Faggiano A, et al. (2006) Pituitary tumour transforming gene (PTTG) expression correlates with the proliferative activity and recurrence status of pituitary adenomas: a clinical and immunohistochemical study. *Clin Endocrinol (Oxf)* 65: 536–543.
 25. Pei L, Melmed S (1997) Isolation and characterization of a pituitary tumor-transforming gene (PTTG). *Mol Endocrinol* 11: 433–441.
 26. Vlotides G, Eigler T, Melmed S (2007) Pituitary tumor-transforming gene: physiology and implications for tumorigenesis. *Endocr Rev* 28: 165–186.
 27. Heaney AP, Horwitz GA, Wang Z, Singson R, Melmed S (1999) Early involvement of estrogen-induced pituitary tumor transforming gene and fibroblast growth factor expression in prolactinoma pathogenesis. *Nat Med* 5: 1317–1321.
 28. Abbud RA, Takumi I, Barker EM, Ren SG, Chen DY, et al. (2005) Early multipotential pituitary focal hyperplasia in the alpha-subunit of glycoprotein hormone-driven pituitary tumor-transforming gene transgenic mice. *Mol Endocrinol* 19: 1383–1391.
 29. Chesnokova V, Zonis S, Rubinek T, Yu R, Ben-Shlomo A, et al. (2007) Senescence mediates pituitary hypoplasia and restrains pituitary tumor growth. *Cancer Res* 67: 10564–10572.
 30. Zou H, McGarry TJ, Bernal T, Kirschner MW (1999) Identification of a vertebrate sister-chromatid separation inhibitor involved in transformation and tumorigenesis. *Science* 285: 418–422.
 31. Wang Z, Yu R, Melmed S (2001) Mice lacking pituitary tumor transforming gene show testicular and splenic hypoplasia, thymic hyperplasia, thrombocytopenia, aberrant cell cycle progression, and premature centromere division. *Mol Endocrinol* 15: 1870–1879.
 32. Hsu YH, Liao LJ, Yu CH, Chiang CP, Jhan JR, et al. (2010) Overexpression of the pituitary tumor transforming gene induces p53-dependent senescence through activating DNA damage response pathway in normal human fibroblasts. *J Biol Chem* 285: 22630–22638.
 33. Bernal JA, Roche M, Mendez-Vidal C, Espina A, Tortolero M, et al. (2008) Proliferative potential after DNA damage and non-homologous end joining are affected by loss of securin. *Cell Death Differ* 15: 202–212.
 34. Chesnokova V, Zonis S, Kovacs K, Ben-Shlomo A, Wawrowsky K, et al. (2008) p21(Cip1) restrains pituitary tumor growth. *Proc Natl Acad Sci U S A* 105: 17498–17503.
 35. Ellsworth BS, Egashira N, Haller JL, Butts DL, Cocquet J, et al. (2006) FOXL2 in the pituitary: molecular, genetic, and developmental analysis. *Mol Endocrinol* 20: 2796–2805.
 36. Sedelnikova OA, Horikawa I, Zimonjic DB, Popescu NC, Bonner WM, et al. (2004) Senescing human cells and ageing mice accumulate DNA lesions with unrepairable double-strand breaks. *Nat Cell Biol* 6: 168–170.
 37. Shen KC, Heng H, Wang Y, Lu S, Liu G, et al. (2005) ATM and p21 cooperate to suppress aneuploidy and subsequent tumor development. *Cancer Res* 65: 8747–8753.
 38. Donangelo I, Gutman S, Horvath E, Kovacs K, Wawrowsky K, et al. (2006) Pituitary tumor transforming gene overexpression facilitates pituitary tumor development. *Endocrinology* 147: 4781–4791.
 39. Cuadrado M, Gutierrez-Martinez P, Swat A, Nebreda AR, Fernandez-Capetillo O (2009) p27Kip1 stabilization is essential for the maintenance of cell cycle arrest in response to DNA damage. *Cancer Res* 69: 8726–8732.
 40. Collado M, Blasco MA, Serrano M (2007) Cellular senescence in cancer and aging. *Cell* 130: 223–233.
 41. Ramji DP, Foka P (2002) CCAAT/enhancer-binding proteins: structure, function and regulation. *Biochem J* 365: 561–575.
 42. Nerlov C (2007) The C/EBP family of transcription factors: a paradigm for interaction between gene expression and proliferation control. *Trends Cell Biol* 17: 318–324.
 43. Gery S, Tanosaki S, Hofmann WK, Koppel A, Koeffler HP (2005) C/EBPdelta expression in a BCR-ABL-positive cell line induces growth arrest and myeloid differentiation. *Oncogene* 24: 1589–1597.
 44. Narita M, Nunez S, Heard E, Lin AW, Hearn SA, et al. (2003) Rb-mediated heterochromatin formation and silencing of E2F target genes during cellular senescence. *Cell* 113: 703–716.
 45. Egashira N, Takedoshi S, Takei M, Teramoto A, Osamura R (2010) Expression of FOXL2 in human normal pituitaries and pituitary adenomas. *Modern Pathology*. pp 1–9.
 46. Swanson SM, Kopchick JJ (2007) Nuclear localization of growth hormone receptor: another age of discovery for cytokine action? *Sci STKE* 2007: pe69.
 47. Ciccone NA, Xu S, Lacza CT, Carroll RS, Kaiser UB (2010) Frequency-dependent regulation of follicle-stimulating hormone beta by pulsatile gonadotropin-releasing hormone is mediated by functional antagonism of bZIP transcription factors. *Mol Cell Biol* 30: 1028–1040.
 48. Kleinberg DL, Wood TL, Furth PA, Lee AV (2009) Growth hormone and insulin-like growth factor-I in the transition from normal mammary development to preneoplastic mammary lesions. *Endocr Rev* 30: 51–74.
 49. Goffin V, Binart N, Touraine P, Kelly PA (2002) Prolactin: the new biology of an old hormone. *Annu Rev Physiol* 64: 47–67.
 50. Janovick JA, Conn PM (2010) Salt bridge integrates GPCR activation with protein trafficking. *Proc Natl Acad Sci U S A* 107: 4454–4458.
 51. Kulman T, Peeper DS (2009) Senescence-messaging secretome: SMS-ing cellular stress. *Nat Rev Cancer* 9: 81–94.
 52. Thangaraju M, Rudelius M, Bierie B, Raffeld M, Sharan S, et al. (2005) C/EBPdelta is a crucial regulator of pro-apoptotic gene expression during mammary gland involution. *Development* 132: 4675–4685.
 53. Cao Z, Umek RM, McKnight SL (1991) Regulated expression of three C/EBP isoforms during adipose conversion of 3T3-L1 cells. *Genes Dev* 5: 1538–1552.
 54. Rizzi F, Caccamo AE, Belloni L, Bettuzzi S (2009) Clusterin is a short half-life, poly-ubiquitinated protein, which controls the fate of prostate cancer cells. *J Cell Physiol* 219: 314–323.
 55. Frost SJ, Simpson DJ, Farrell WE (2001) Decreased proliferation and cell cycle arrest in neoplastic rat pituitary cells is associated with transforming growth factor-beta1-induced expression of p15/INK4B. *Mol Cell Endocrinol* 176: 29–37.
 56. Chi KN, Zoubeidi A, Gleave ME (2008) Custirsen (OGX-011): a second-generation antisense inhibitor of clusterin for the treatment of cancer. *Expert Opin Investig Drugs* 17: 1955–1962.
 57. Flanagan L, Whyte L, Chatterjee N, Tenniswood M (2010) Effects of clusterin over-expression on metastatic progression and therapy in breast cancer. *BMC Cancer* 10: 107.
 58. Chou TY, Chen WC, Lee AC, Hung SM, Shih NY, et al. (2009) Clusterin silencing in human lung adenocarcinoma cells induces a mesenchymal-to-epithelial transition through modulating the ERK/Slug pathway. *Cell Signal* 21: 704–711.
 59. Criswell T, Beman M, Araki S, Leskov K, Cataldo E, et al. (2005) Delayed activation of insulin-like growth factor-1 receptor/Src/MAPK/Egr-1 signaling regulates clusterin expression, a pro-survival factor. *J Biol Chem* 280: 14212–14221.
 60. Bettuzzi S, Scorcioni F, Astancolle S, Davalli P, Scaltriti M, et al. (2002) Clusterin (SGP-2) transient overexpression decreases proliferation rate of SV40-immortalized human prostate epithelial cells by slowing down cell cycle progression. *Oncogene* 21: 4328–4334.
 61. Panico F, Rizzi F, Fabbri LM, Bettuzzi S, Luppi F (2009) Clusterin (CLU) and lung cancer. *Adv Cancer Res* 105: 63–76.
 62. Wakefield LM, Roberts AB (2002) TGF-beta signaling: positive and negative effects on tumorigenesis. *Curr Opin Genet Dev* 12: 22–29.
 63. Nam JS, Terabe M, Kang MJ, Chae H, Voong N, et al. (2008) Transforming growth factor beta subverts the immune system into directly promoting tumor growth through interleukin-17. *Cancer Res* 68: 3915–3923.
 64. Alarid ET, Windle JJ, Whyte DB, Mellon PL (1996) Immortalization of pituitary cells at discrete stages of development by directed oncogenesis in transgenic mice. *Development* 122: 3319–3329.