

Review

Review of Climate, Landscape, and Viral Genetics as Drivers of the Japanese Encephalitis Virus Ecology

Guillaume Le Flohic^{1*}, Vincent Porphyre², Philippe Barbazan^{3†}, Jean-Paul Gonzalez^{1,3,4,5}

1 UR Ecologie et Santé, Centre International de Recherches Médicales de Franceville, CIRMF, BP 769, Franceville, Gabon, **2** Centre de Coopération Internationale en Recherche Agronomique pour le Développement, CIRAD, Montpellier, France, **3** Institut de Recherche pour le Développement, IRD, Marseille, France, **4** Ministère des Affaires Étrangères et Européennes, Mission de Coopération, Libreville, Gabon, **5** Metabiota, Washington, District of Columbia, United States of America

Abstract: The Japanese encephalitis virus (JEV), an arthropod-borne *Flavivirus*, is the major cause of viral encephalitis, responsible for 10,000–15,000 deaths each year, yet is a neglected tropical disease. Since the JEV distribution area has been large and continuously extending toward new Asian and Australasian regions, it is considered an emerging and reemerging pathogen. Despite large effective immunization campaigns, Japanese encephalitis remains a disease of global health concern. JEV zoonotic transmission cycles may be either wild or domestic: the first involves wading birds as wild amplifying hosts; the second involves pigs as the main domestic amplifying hosts. *Culex* mosquito species, especially *Cx. tritaeniorhynchus*, are the main competent vectors. Although five JEV genotypes circulate, neither clear-cut genotype-phenotype relationship nor clear variations in genotype fitness to hosts or vectors have been identified. Instead, the molecular epidemiology appears highly dependent on vectors, hosts' biology, and on a set of environmental factors. At global scale, climate, land cover, and land use, otherwise strongly dependent on human activities, affect the abundance of JEV vectors, and of wild and domestic hosts. Chiefly, the increase of rice-cultivated surface, intensively used by wading birds, and of pig production in Asia has provided a high availability of resources to mosquito vectors, enhancing the JEV maintenance, amplification, and transmission. At fine scale, the characteristics (density, size, spatial arrangement) of three landscape elements (paddy fields, pig farms, human habitations) facilitate or impede movement of vectors, then determine how the JEV interacts with hosts and vectors and ultimately the infection risk to humans. If the JEV is introduced in a favorable landscape, either by live infected animals or by vectors, then the virus can emerge and become a major threat for human health. Multidisciplinary research is essential to shed light on the biological mechanisms involved in the emergence, spread, reemergence, and genotypic changes of JEV.

Introduction

The incidence of zoonotic diseases, transmitted to humans from wild or domestic animals, has noticeably increased during the past few decades and currently represents 70% or more of emerging diseases [1]. Japanese encephalitis virus (JEV), an arbovirus of the *Flavivirus* genus, family *Flaviviridae*, is transmitted by mosquitoes from animals to humans. In humans, this zoonotic disease is the largest worldwide cause of epidemic viral encephalitis [2,3]. The pathogenesis of the JEV and the clinical manifestations of the disease, including severe neurological syndromes, depend on several factors that have been deeply reviewed elsewhere [4,5].

The incubation period ranges from 5 to 15 days; JE infections are lethal in about 25–30% of cases, mostly in infants, and lead to permanent sequelae in about 50% of cases.

JE was first described in 1871 in Japan, and first characterized in 1935 [6]. Despite an effective vaccine developed in 1941, and the subsequent national immunization campaigns that have greatly reduced the incidence of JE in several countries, sporadic cases continue to be reported, and JEV continues to spread widely in South, East, and Southeast Asia and Australasia [7]. Currently, more than three billion people live in JE-endemic countries [5]. There, though people of all ages may be exposed to JEV, the JE incidence is higher in children because most adults are immune [8]. Now, about 68,000 JE cases are estimated to occur annually, causing at least 10,000–15,000 deaths in more than 20 Australasian countries [8,9].

Aims and Methodology

This review aims at offering an overview of the factors affecting JEV epidemiology. It discusses the current state of knowledge on JEV diversity, molecular epidemiology, and ecology, and examines how environmental variables and modifications, especially those associated with agriculture, affect the landscape characteristics and JEV ecology. The literature review was achieved by using both general web search engines and scientific web search engines such as PubMed, Springerlink, ScienceDirect, and Web of Science. It focused on the literature available on virology, molecular and spatial epidemiology, entomology, and landscape and behavioral ecology.

Citation: Le Flohic G, Porphyre V, Barbazan P, Gonzalez J-P (2013) Review of Climate, Landscape, and Viral Genetics as Drivers of the Japanese Encephalitis Virus Ecology. *PLoS Negl Trop Dis* 7(9): e2208. doi:10.1371/journal.pntd.0002208

Editor: Michael A. Johansson, Centers for Disease Control and Prevention, United States of America

Published: September 12, 2013

Copyright: © 2013 Le Flohic et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Funding: The present work as been funded by the Research Unit 190 on “Emerging diseases,” Health Department, Institut de Recherche pour le Développement, Le Sextant, 44, bd de Dunkerque, CS 90009, 13572 Marseille cedex 02. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing Interests: The authors have declared that no competing interests exist.

* E-mail: leflohg@yahoo.fr

†: Deceased.

JEV Genotypes

Five JEV genotypes have been distinguished by nucleotide sequencing of the *C/PrM* and *E* (capsid, precursor membrane, and envelope) genes [10]. Genotypes I, II, and III are distributed throughout Asia, while genotype IV is restricted to Eastern Indonesia [11]. Genotype V was long restricted to Malaysia but isolates were recently found in China and in the Republic of Korea [12,13]. Genotypes IV and V form the oldest JEV lineage, which appears to have originated from an ancestral virus in the Indonesian-Malaysian region. JEV therefore probably originally spread from this region [4,11]. Genotypes I, II, and III are the most prevalent, having accounted for 98% of the strains isolated from 1935 to 2009 [14].

JEV Molecular Epidemiology

Until recently, genotypes I and III were mostly associated with epidemic diseases in temperate regions of Asia, while genotypes II and IV were associated with endemic diseases in tropical regions. It was therefore postulated that the JEV epidemiology differed with genotypes [15]. However, more recent observations show that genotypes can be found indifferently within epidemic or endemic areas. Indeed, genotype I has been isolated in Australasia within the tropical region [16], genotype II has been shown to have circulated in the Republic of Korea within the temperate region before the 1970s [17,18], and genotype III, the only one in India so far, has circulated both in the south Indian peninsula (characterized by endemic activity) and in the north (characterized by epidemic activity) [7,19].

Differences in the JEV genome might explain phenotypic variations of the disease. Experimental changes in the *E* gene have indeed been shown to be associated with loss or gain of neuroinvasiveness and neurovirulence in mice and monkeys [20–23]. Even so, experimental studies of the neurovirulence of wild JEV strains in mice showed no clear-cut genotype-phenotype relationship [24], and there is still no firm evidence that the JEV genotypes circulating differ in their virulence (Figure 1). However, the dengue virus type 2, another closely related *Flavivirus*, has been shown to possess interesting genotype-dependent characteristics such as human viral pathogenicity or midgut viral replication in *Aedes aegypti* [25,26]. The JEV molecular markers previously and currently studied might not be sufficient and need further investigation.

Rather than by viral genetic determinants solely, the JEV epidemiological pattern appears to be actually influenced by a complex set of variables, including several environmental, ecological, and immunological factors [7] (Figure 1).

JEV Spatial Epidemiology

The distribution area of JEV has consistently enlarged. Sometimes, the genotypes of JEV isolates changed locally. In northern Vietnam and Thailand, a shift from genotype II to genotype I was reported [27,28]. In northern temperate regions (i.e., Japan, South Korea, and Northeast China), genotype I progressively replaced genotype III and became the main genotype [29]. It is now very probably the most widespread genotype in Asia, which was otherwise found to be lethal for humans [9,29–33]. However, to date, vaccines are only derived from genotype III strains; whereas protective levels of cross-reactive neutralizing antibodies of these vaccines were found against the various circulating genotypes, variations between genotypes call for further studies [34,35]. In particular, since the

immune response against genotype I was less pronounced, its duration should be addressed [34].

JEV Ecology: Vector- and Host-Virus Interactions

Vectors

Over 30 mosquito species (family Culicidae) belonging to the *Aedes*, *Anopheles*, *Armigeres*, *Culex*, and *Mansonia* genera are recognized to potentially carry JEV, but not all are equally competent for virus transmission [7]. Susceptibility and competence vary among and within mosquito species [36], but also possibly covariate with genotypes, as has been shown for dengue [26,37] and Chikungunya virus strains [38]. However, no variations in vector competence and transmissibility by various mosquito species for different JEV genotypes have yet been demonstrated.

Culex species are the most competent JEV vectors, in the same way that *Aedes* species are the most competent Dengue viruses vectors [39]. This low plasticity of the virus to vectors may be explained by the vector ecology, especially host-feeding preferences, as few species feed on JEV reservoir-amplifying hosts. For example, in mainland Australia *Cx. annulirostris*, a competent JEV vector, feeds predominantly on marsupials, and thus remains outside the JEV transmission cycle [40,41]. JEV is transmitted by *Cx. annulirostris*, *Cx. annulus*, *Cx. fuscocephala*, *Cx. gelidus*, *Cx. sitiens*, *Cx. vishnui* species complex, and the rice field-breeding mosquitoes *Culex tritaeniorhynchus* [42]. *Cx. tritaeniorhynchus* is the major JEV vector because it is highly competent and is largely distributed all over the JEV-endemic regions [3,43]. JEV vectors are highly zoophilic, tending to prefer wading birds, cattle, and pigs to humans [42,44,45]. Therefore, vector trophic preferences are a major ecological factor affecting the fitness of the virus.

Variations of competence may be due to differences in the susceptibility to infection of mosquito midgut and salivary glands, and then to differences in the efficiency of virus secretion and transmission [46]. Susceptibility to JEV infection is partly dependent on the *E* gene sequence of the virus. This gene encodes a protein involved first in virus cellular entry through attachment to membrane receptors and second in the fusion of both JEV and host-cell membranes [47]. However, despite some study on the subject [48], no data demonstrates there are important variations in vector competence and transmissibility between various mosquito species for different JEV genotypes.

Hosts also play a central role in virus ecology, according to their viraemia and availability within the immediate vector flight range. Ultimately, two types of JEV transmission cycle can be drawn: a bird-associated wild cycle and a pig-associated rural domestic cycle.

Bird-Associated Wild Cycle

Although in the wild a wide range of animals (bats, birds, rodents, snakes, wild boars...) may be infected by JEV, with high seroprevalence rates in several mammal and bird species, most of them are dead-end hosts and are unable to infect mosquitoes [2,3]. However, over 90 bird species are known to be amplifying and reservoir hosts of JEV. Among them, wild wading birds, in particular egrets (*Egretta garzetta*) and herons (*Nycticorax nycticorax*) of the Ardeidae family, are highly susceptible to JEV infection. They represent the primary effective animal hosts, since they have a high virus titer, and are an outstanding source of infection for mosquitoes [3,7,9]. As many of them are widely distributed and migratory, ardeid species are suspected of being responsible for several virus introductions, from China to Japan, and from the

intensity of circulation in farms depends on the amount of pigs reared by farm, their age at slaughter, their reproduction rate, and the vaccination effort [59]. Subsequently, the epidemiology of JEV in the domestic reservoir host follows a complex multifactorial pattern that needs to be brought to light.

The mosquito vector species present in the wild transmission cycle are also greatly implicated in the rural domestic cycle. Although most mosquito species usually consume bird, cattle, and pig blood [61], *Cx. quinquefasciatus*, whereas opportunistic, seems also very anthropophilic on account of its use of various domestic water collections and wastewater from pig farms as breeding sites [2,45,55,62,63]. Because it has been shown to naturally carry the JEV and to be experimentally capable of transmitting it [48,63], *Cx. quinquefasciatus* is another potential vector of the JEV domestic cycle. It represents an important potential source of infection for humans, so much the more it has emerged in areas where it was absent before following urbanization and human expansion, and increases in number in urban households with human density and pig keeping [64,65].

Evolutionary Ecology of the JEV

Until recently, the factors influencing the historical and geographical emergence and spread of JEV genotypes were not clearly understood. In fact, genotypes' emergence and spread appear to have a complex pattern relevant with the role of environmental factors. Some emerge in areas where they were absent before (e.g., genotype V in Tibet, a cold and high-altitude region that was considered free of JEV [12,66], and genotype I in the Australasian region [16]), whereas some replace the dominant genotype (e.g., genotype III to genotype I shift).

It is thought that these changes are due to the JEV fitness to a new competent vector (e.g., genotype V, besides being isolated from *Cx. tritaeniorhynchus*, seems associated with a new JEV vector in the Republic of Korea: *Cx. bitaeniorhynchus* [13]) or to new host availability (e.g., increasing pig farming in Tibet [12]). More likely, environmental factors (e.g., mosquito dispersal by wind in Australia), animal ecology (e.g., migration of animals), human activity (e.g., pig trade introducing genotype I in Australia [16]), and climatic changes [67] are variables independent to JEV genetics that may modify the distribution area of JEV and its genotypes.

Environmental Factors: Drivers of JEV Epidemiology

The (re)emergence of vector-borne and zoonotic diseases as well as variations in disease risk and incidence are strongly driven by environmental factors. Among the most relevant factors are climate but also land cover and land use, respectively the biophysical attributes of the earth's surface and the human purpose or intent applied to these attributes [68]. However, these variables occur at several different geographic ranges, and hence multi-scale studies are a major stake for the understanding of the spatial epidemiology of diseases [69].

For a long time, JEV epidemiology has exclusively been broached at the global scale and primarily on the basis of climatic variables (i.e., rainfall and temperature) because they indeed strongly affect vector density [67]. Recently, Miller et al. [70] identified an optimal range of temperature during the wet season that is favorable to the *Cx. tritaeniorhynchus* biology. As they also found that most of JE cases were located in areas of high probability of vectors, they underlined the link between climatic covariables, vector ecology, and human health. Moreover, temperature might affect the competence of a

vector-genotype couple over another, as shown for the West Nile virus [71].

In our previous study [14], six Asian and Australasian regions were identified according to anthropogenic, biological, geographic, and physical factors including not only climatic conditions, but also biomes and land use. The regional history of JEV genotypes was analyzed (i.e., the number of JEV isolates and the proportion of each genotype). Then we emphasized the two regions of intensive JEV circulation: the Palearctic biogeographic realm [72] comprising Eastern Russia, Japan, Korea, and Northern China, and a tropical region characterized by large cover of rice fields comprising the Indochinese peninsula (Cambodia, Lao PDR, Myanmar, Thailand, and Vietnam), South China, and Taiwan [73].

Actually, land cover and land use changes have been driving the JE risk and incidence. In Asia, paddy field surfaces have constantly extended since the early 1960s (<http://faostat3.fao.org/>). Given paddy fields provide long-term *Culex* sp. breeding sites and attract many wading birds for foraging and resting, they enhance the circulation and expansion of mosquito and wading bird populations [74]. Thus, in areas where rice-irrigated farming is widespread, the JEV transmission might become less dependent on rainfall [75,76]. Likewise, because the amount of live pig heads has increased since the 1960s (<http://faostat3.fao.org/>), both backyard and industrial pig farming have provided a continuous increasing, outstanding potential source of blood meals for mosquitoes.

Additionally, a finer-scale spatial analysis (i.e., local or regional landscape) helps to better picture how landscape structure affect the JEV epidemiology. At fine scale, JEV ecology is indeed largely affected by both land cover and land use as these factors affect vectors' and hosts' ecology (Figure 1). Once the JEV is introduced, its ecological sustainability indeed relies on its growth within the susceptible host population and on its transmission between hosts through vectors. Then the risk of incidental infection to humans depends on the likelihood of disease transmission within the surrounding landscape [77]. To be transmitted to humans, JEV needs: 1) competent mosquito vectors, 2) reservoir-amplifying hosts, and 3) nonimmune humans within the range of the JEV-competent vector and animal hosts. In this process, the three major landscape elements are paddy fields, pig farms, and human habitations. In the landscape, they are distributed in habitat patches. Their density, size, and spatial arrangement are key factors determining the degree to which the landscape facilitates or impedes movement of vectors among them (i.e., the landscape connectivity) [78] (Figure 1). Within a given landscape, this biological connectivity may strongly affect the JEV transmission dynamics.

Nonenvironmental Factors of JEV Epidemiology

Vaccination is the most effective way to reduce the incidence of JE in humans, yet it has no effect on the JEV transmission cycle. Vaccination of livestock, especially pigs, would in contrast reduce the amplification of the virus, the rate of mosquito infection, and subsequently the risk of transmission to humans. Yet, vaccination of pigs is generally not used to prevent JE because it is costly, hardly feasible logistically, and not necessarily effective in piglets (they must be immunized after the disappearance of maternal antibodies) [6,79]. Moreover, pigs represent a relevant sentinel model, the surveillance of which could predict a potential JE outbreak in a human population nearby [54]. Immunizing sentinel pigs would also impede the detection of such a threat. Other factors have strong effects on the JEV and genotype circulation

Five Key Articles in the Field

1. van den Hurk AF, Ritchie SA, Mackenzie JS (2009) Ecology and geographical expansion of Japanese encephalitis virus. *Annu Rev Entomol* 54: 17–35.
2. Weaver SC, Barrett ADT (2004) Transmission cycles, host range, evolution and emergence of arboviral disease. *Nat Rev Microbiol* 2: 789–801.
3. Mackenzie JS, Gubler DJ, Petersen LR (2004) Emerging flaviviruses: the spread and resurgence of Japanese encephalitis, West Nile and dengue viruses. *Nat Med* 10: S98–109.
4. Solomon T (2006) Control of Japanese encephalitis — within our grasp? *N Engl J Med* 355: 869–871.
5. Solomon T, Ni H, Beasley DWC, Ekkelenkamp M, Cardosa MJ, et al. (2003) Origin and evolution of Japanese encephalitis virus in Southeast Asia. *J Virol* 77: 3091–3098.

within infected landscapes and their emergence in nonendemic landscapes and regions. There are, among them, the movements of either naive or infected animals such as the migrations of flying vertebrates and trading and transportation of domestic animals. These movements modify human exposure to JEV and thus require the particular attention of the health system. In southeastern Asia, trade of live animals occurs between farms, local markets, and more importantly within the Indochinese peninsula and China, and also to Hong Kong and Singapore [80]. Viremic birds, for instance, may have been responsible for JEV spread and introductions [4,27] into India [81], Taiwan [82], or Papua New Guinea [83]. Over long distances, migratory birds are the most likely spreader of the JEV as some species, such as *Egretta garzetta* and *Nycticorax nycticorax*, have a complex migration system over a large geographical area (<http://maps.iucnredlist.org>). However, the large-scale movement patterns of the main wading birds species implicated in JEV transmission are little known. Additionally, JEV may disperse through wind-blown infected mosquitoes, as is assumed to be the cause of the JEV introduction from Papua New Guinea to north Australia [84] or from China to Japan [85].

Moreover, there is no evidence that JEV requires adaptation to shift between the bird-associated wild cycle and the pig-associated rural domestic cycle [2,3]. The JEV may easily be transferred from one to another if competent vectors feeding on both wild and domestic hosts are present in JEV-infected areas where both hosts are in close proximity.

Conclusion

For more than a half century, human activities have led to changes of land use and land cover and subsequent deep

References

1. Kuiken T, Leighton FA, Fouchier RA, LeDuc JW, Peiris JS, et al. (2005) Public health. Pathogen surveillance in animals. *Science* 309: 1680–1681.
2. Weaver SC, Barrett ADT (2004) Transmission cycles, host range, evolution and emergence of arboviral disease. *Nat Rev Microbiol* 2: 789–801.
3. van den Hurk AF, Ritchie SA, Mackenzie JS (2009) Ecology and geographical expansion of Japanese encephalitis virus. *Annu Rev Entomol* 54: 17–35. doi:10.1146/annurev.ento.54.110807.090510.
4. Mackenzie JS, Gubler DJ, Petersen LR (2004) Emerging flaviviruses: the spread and resurgence of Japanese encephalitis, West Nile and dengue viruses. *Nat Med* 10: S98–109.
5. Ghosh D, Basu A (2009) Japanese encephalitis—a pathological and clinical perspective. *PLoS Negl Trop Dis* 3: e437. doi:10.1371/journal.pntd.0000437.

Key Learning Points

1. The Japanese encephalitis virus epidemiology is strongly dependant on the ecology of both amplifying hosts and mosquito vectors, which themselves are affected not only by natural environmental factors but also by human activities.
2. The spatial epidemiology of the JEV needs to be investigated at global and landscape scales, as human-induced land use and land cover have effects on the species distribution area, population density, species community structure, and interactions between vectors, hosts, and humans.
3. Though the fitness of the virus to hosts and vectors appears little dependent on genetic factors (genotypes), knowledge of other closely related arboviruses suggests that the epidemiological implications of genotypes might be underestimated.
4. The introduction, spread, emergence, and persistence of JEV zoonotic cycles in a new region may be predicted according to the characteristics of the landscape at various spatial scales.

environmental changes of habitats. These modifications have modified the risk of infection for humans. The ecology of arthropod-borne diseases, noticeably JE, remains complex since they are highly dependent on various biotic and abiotic environmental factors and on the spatial scale of study. To assess and predict JE emergence therefore remains difficult. Multidisciplinary research focusing on virology, molecular and spatial epidemiology, landscape and behavioral ecology, history, and socioeconomic studies are essential to shed light on the biological mechanisms involved in the emergence, spread, reemergence, and genotypic changes of JEV.

Acknowledgments

We are grateful to Dr. Christophe Paupy from IRD and CIRMF and Dr. François Rouet from CIRMF for reviewing the manuscript and providing a great deal of useful advice. Special thanks to Heidi Lançon who revised the article.

Obituary. Philippe BARBAZAN, Ph.D. (1950–2009) was a medical entomologist working for the French Institute of Research for Development (IRD). For more than a decade he worked as a researcher, and was a mentor for the students, at Mahidol University, Center for Vaccine Development and Center for Vectors and Vector-borne Diseases (Thailand). He dedicated his career to epidemiology of arthropod-borne viral diseases, and spatial and environmental factors affecting their transmission among several developing countries around the world. Sadly, Philippe passed away in June 2009 after months of courage fighting against cancer. The authors, colleagues and friends of Philippe, such a great scientist and humanist, would like to convey their sympathy to his kids, Camille and Arthur, and his family.

6. Erlanger TE, Weiss S, Keiser J, Utzinger J, Wiedenmayer K (2009) Past, present, and future of Japanese encephalitis. *Emerg Infect Dis* 15: 1–7.
7. Mackenzie J, Williams D, Smith D (2006) Japanese encephalitis virus: the geographic distribution, incidence, and spread of a virus with a propensity to emerge in new areas. *Perspect Med Virol* 16: 201–268.
8. Campbell GL, Hills SL, Fischer M, Jacobson JA, Hoke CH, et al. (2011) Estimated global incidence of Japanese encephalitis: a systematic review. *Bull World Health Organ* 89: 766–774.
9. Solomon T (2006) Control of Japanese encephalitis — within our grasp? *N Engl J Med* 355: 869–871.
10. Chen W-R, Tesh RB, Rico-Hesse R (1990) Genetic variation of Japanese encephalitis virus in nature. *J Gen Virol* 71: 2915–2922.

11. Solomon T, Ni H, Beasley DWC, Ekkelenkamp M, Cardosa MJ, et al. (2003) Origin and evolution of Japanese encephalitis virus in Southeast Asia. *J Virol* 77: 3091–3098. doi:10.1128/JVI.77.5.3091.
12. Li Y-X, Li M-H, Fu S-H, Chen W-X, Liu Q-Y, et al. (2011) Japanese encephalitis, Tibet, China. *Emerg Infect Dis* 17: 934–936. doi:10.1093/bib/5.2.150.
13. Takhampunya R, Kim H-C, Tippayachai B, Kengluocha A, Klein TA, et al. (2011) Emergence of Japanese encephalitis virus genotype V in the Republic of Korea. *Virol J* 8: 449.
14. Le Flohic G, Gonzalez J (2011) When Japanese encephalitis virus invaded eastern hemisphere – the history of the spread of virus genotypes. In: Růžek D, editor. *Flavivirus encephalitis*. Volume I. InTech. pp. 405–426.
15. Williams DT, Wang LF, Daniels PW, Mackenzie JS (2000) Molecular characterization of the first Australian isolate of Japanese encephalitis virus, the FU strain. *J Gen Virol* 81: 2471–2480.
16. Pyke A, Williams D, Nisbet D (2001) The appearance of a second genotype of Japanese encephalitis virus in the Australasian region. *Am J Trop Med Hyg* 65: 747–753.
17. Schuh AJ, Li L, Tesh RB, Innis BL, Barrett ADT (2010) Genetic characterization of early isolates of Japanese encephalitis virus: genotype II has been circulating since at least 1951. *J Gen Virol* 91: 95–102.
18. Schuh AJ, Tesh RB, Barrett ADT (2011) Genetic characterization of Japanese encephalitis virus genotype II strains isolated from 1951 to 1978. *J Gen Virol* 92: 516–527.
19. Reuben R, Gajanana A (1997) Japanese encephalitis in India. *Indian J Pediatr* 64: 243–251.
20. Lee E, Lobigs M (2000) Substitutions at the putative receptor-binding site of an encephalitic *Flavivirus* alter virulence and host cell tropism and reveal a role for glycosaminoglycans in entry. *J Virol* 74: 8867–8875.
21. Monath TP, Arroyo J, Levenbook I, Zhang Z-X, Catalan J, et al. (2002) Single mutation in the *Flavivirus* envelope protein hinge region increases neurovirulence for mice and monkeys but decreases viscerotropism for monkeys: relevance to development and safety testing of live, attenuated vaccines. *J Virol* 76: 1932–1943.
22. Tajima S, Nerome R, Nukui Y, Kato F, Takasaki T, et al. (2010) A single mutation in the Japanese encephalitis virus E protein (S123R) increases its growth rate in mouse neuroblastoma cells and its pathogenicity in mice. *Virology* 396: 298–304.
23. Ni H, Barrett A D (1996) Molecular differences between wild-type Japanese encephalitis virus strains of high and low mouse neuroinvasiveness. *J Gen Virol* 77 (Pt 7): 1449–1455.
24. Solomon T (2003) Recent advances in Japanese encephalitis. *J Neurovirol* 9: 274–283.
25. Anderson JR, Rico-Hesse R (2006) *Aedes aegypti* vectorial capacity is determined by the infecting genotype of dengue virus. *Am J Trop Med Hyg* 75: 886–892.
26. Cox J, Brown HE, Rico-Hesse R (2011) Variation in vector competence for dengue viruses does not depend on mosquito midgut binding affinity. *PLoS Negl Trop Dis* 5: e1172. doi:10.1371/journal.pntd.0001172.
27. Nga PT, Parquet C, Cuong VD, Ma S, Hasebe F, et al. (2004) Shift in Japanese encephalitis virus (JEV) genotype circulating in northern Vietnam: implications for frequent introductions of JEV from Southeast Asia to East Asia. *J Gen Virol* 3: 1625–1631. doi:10.1099/vir.0.79797-0.
28. Nitapattana N, Dubot-Pères A (2008) Change in Japanese encephalitis virus distribution, Thailand. *Emerg Infect Dis* 14: 1762–1765.
29. Pan X-L, Liu H, Wang H-Y, Fu S-H, Liu H-Z, et al. (2011) Emergence of genotype I of Japanese encephalitis virus as the dominant genotype in Asia. *J Virol* 85: 9847–9853.
30. Ma S-P, Yoshida Y, Makino Y, Tadano M, Ono T, et al. (2003) Short report: a major genotype of Japanese encephalitis virus currently circulating in Japan. *Am J Trop Med Hyg* 69: 151–154.
31. Wang HY, Takasaki T, Fu SH, Sun XH, Zhang HL, et al. (2007) Molecular epidemiological analysis of Japanese encephalitis virus in China. *J Gen Virol* 88: 885–894. doi:10.1099/vir.0.82185-0.
32. Teng M, Luo J, Fan J-M, Chen L, Wang X-T, et al. (2013) Molecular characterization of Japanese encephalitis viruses circulating in pigs and mosquitoes on pig farms in the Chinese province of Henan. *Virus Genes* 46: 170–174.
33. Wang L, Fu S, Zhang H, Ye X, Yu D, et al. (2010) Identification and isolation of Genotype-I Japanese encephalitis virus from encephalitis patients. *Virol J* 7: 345.
34. Erra EO, Askling HH, Yoksan S, Rombo L, Riutta J, et al. (2013) Cross-protective capacity of Japanese encephalitis (JE) vaccines against circulating heterogeneous JE virus genotypes. *Clin Infect Dis* 56: 267–270.
35. Fan Y-C, Chen J-M, Chiu H-C, Chen Y-Y, Lin J-W, et al. (2012) Partially neutralizing potency against emerging genotype I virus among children received formalin-inactivated Japanese encephalitis virus vaccine. *PLoS Negl Trop Dis* 6: e1834. doi:10.1371/journal.pntd.0001834.
36. Goddard J (2008) Dynamics of arthropod-borne diseases. In: Goddard J, editor. *Infectious diseases and arthropods*. Totowa, NJ: Humana Press. pp. 19–28.
37. Hanley KA, Nelson JT, Schirtzinger EE, Whitehead SS, Hanson CT (2008) Superior infectivity for mosquito vectors contributes to competitive displacement among strains of dengue virus. *BMC Ecol* 8: 1.
38. Tssetsarkin KA, Vanlandingham DL, McGee CE, Higgs S (2007) A single mutation in chikungunya virus affects vector specificity and epidemic potential. *PLoS Pathog* 3: e201. doi:10.1371/journal.ppat.0030201.
39. Weaver SC (2006) Evolutionary influences in arboviral disease. *Curr Top Microbiol Immunol* 299: 285–314.
40. van den Hurk A, Johansen C, Zborowski P, Paru R, Foley P, et al. (2003) Mosquito host-feeding patterns and implications for Japanese encephalitis virus transmission in northern Australia and Papua New Guinea. *Med Vet Entomol* 17: 403–411.
41. Hemmerter S, Šlapeta J, van den Hurk AF, Cooper RD, Whelan PI, et al. (2007) A curious coincidence: mosquito biodiversity and the limits of the Japanese encephalitis virus in Australasia. *BMC Evol Biol* 11: 100.
42. Reuben R, Tewari S, Hiriyani J, Akiyama J (1994) Illustrated keys to species of *Culex* (*Culex*) associated with Japanese Encephalitis in Southeast Asia (Diptera: Culicidae). *Mosquito Systematics* 26: 75–96.
43. Impoinvil D, Solomon T, Schluter W (2011) The spatial heterogeneity between Japanese encephalitis incidence distribution and environmental variables in Nepal. *PLoS ONE* 6: e22192. doi:10.1371/journal.pone.0022192.
44. Mitchell CJ, Chen PS, Boreham PF (1973) Host-feeding patterns and behaviour of 4 *Culex* species in an endemic area of Japanese encephalitis. *Bull World Health Organ* 49: 293–299.
45. Burke D, Leake C (1988) Japanese encephalitis. In: Monath T, editor. *The arboviruses: epidemiology and ecology*. Volume 3. pp. 63–92.
46. Takahashi M (1982) Differential transmission efficiency for Japanese encephalitis virus among colonized strains of *Culex tritaeniorhynchus*. *Japanese Journal of Sanitary Zoology* 33: 325–333.
47. Solomon T, Dung NM, Kneen R, Gainsborough M, Vaughn DW, et al. (2000) Japanese encephalitis. *J Neurol Neurosurg Psychiatry* 68: 405–415.
48. Rosen L, Roseboom L, Gubler D (1985) Comparative susceptibility of mosquito species and strains to oral and parenteral infection with dengue and Japanese encephalitis viruses. *Am J Trop Med Hyg* 34: 603–615.
49. See E, Cheng H, Wang D, Eong E, Lee M (2002) Presence of hemagglutination inhibition and neutralization antibodies to Japanese encephalitis virus in wild pigs on an offshore island in Singapore. *Acta Tropica* 81: 233–236.
50. Ohno Y, Sato H, Suzuki K, Yokoyama M, Uni S, et al. (2009) Detection of antibodies against Japanese encephalitis virus in raccoons, raccoon dogs and wild boars in Japan. *J Vet Med Sci* 71: 1035–1039.
51. van den Hurk AF, Smith CS, Field HE, Smith IL, Northill JA, et al. (2009) Transmission of Japanese Encephalitis virus from the Black Flying Fox, *Pteropus alecto*, to *Culex annulirostris* mosquitoes, despite the absence of detectable viremia. *Am J Trop Med Hyg* 81: 457–462.
52. Wang J, Pan X, Zhang H, Fu S, Wang H, et al. (2009) Japanese Encephalitis viruses from bats in Yunnan, China. *Emerg Infect Dis* 15: 939–942. doi:10.3201/eid1506.081525.
53. Chaves LF, Harrington LC, Keogh CL, Nguyen AM, Kitron UD (2010) Blood feeding patterns of mosquitoes: random or structured? *Front Zool* 7: 3.
54. Nitapattana N, Le Flohic G, Thongchai P, Nakgoi K, Palabodeewat S, et al. (2011) Elevated Japanese encephalitis virus activity monitored by domestic sentinel piglets in Thailand. *Vector Borne Zoonotic Dis* 11: 391–394.
55. Weaver S (2005) Host range, amplification and arboviral disease emergence. In: Peters CJ, Calisher CH, editors. *Infectious diseases from nature: mechanisms of viral emergence and persistence*. New York: Springer Wien. pp. 33–44.
56. Gibson JP, Bishop SC (2005) Use of molecular markers to enhance resistance of livestock to disease: a global approach. *Rev Sci Tech* 24: 343–353.
57. Roy BA, Kirchner JW (2000) Evolutionary dynamics of pathogen resistance and tolerance. *Evolution* 54: 51–63.
58. Gibson JP (2002) Role of genetically determined resistance of livestock to disease in the developing world: potential impacts and researchable issues. In: Perry BD, Randolph TF, McDermott JJ, Sones KR, Thornton PK, editors. *Investing in animal health research to alleviate poverty*. Nairobi, Kenya. p. 148.
59. Rodhain F (2010) Japanese encephalitis: a fast-changing viral disease. *Bull Soc Pathol Exot* 103: 135–154.
60. Sota T, Hayamizu E, Mogi M (1991) Distribution of biting *Culex tritaeniorhynchus* (Diptera: Culicidae) among pigs: effects of host size and behavior. *J Med Entomol* 28: 428–433.
61. Hasegawa M, Tuno N, Yen NT, Nam VS, Takagi M (2008) Influence of the distribution of host species on adult abundance of Japanese Encephalitis vectors —*Culex vishnu* subgroup and *Culex gelidus*— in a rice-cultivating village in Northern Vietnam. *Am J Trop Med Hyg* 78: 159–168.
62. Samuel PP, Arunachalam N, Hiriyani J, Thenmozhi V, Gajanana A, et al. (2004) Host-feeding pattern of *Culex quinquefasciatus* Say and *Mansonia annulifera* (Theobald) (Diptera: Culicidae), the major vectors of filariasis in a rural area of south India. *J Med Entomol* 41: 442–446.
63. Nitapattana N, Apiwathnasorn C, Barbazan P, Leemingsawatt S, Yoksan S, et al. (2005) First isolation of Japanese encephalitis from *Culex quinquefasciatus* in Thailand. *Southeast Asian J Trop Med Public Health* 36: 875–878.
64. Fonseca DM, Smith JL, Wilkerson RC, Fleischer RC (2006) Pathways of expansion and multiple introductions illustrated by large genetic differentiation among worldwide populations of the southern house mosquito. *Am J Trop Med Hyg* 74: 284–289.
65. Lindahl J, Chirico J, Boqvist S, Thu HTV, Magnusson U (2012) Occurrence of Japanese encephalitis virus mosquito vectors in relation to urban pig holdings. *Am J Trop Med Hyg* 87: 1076–1082.
66. Li M-H, Fu S-H, Chen W-X, Wang H-Y, Guo Y-H, et al. (2011) Genotype V Japanese encephalitis virus is emerging. *PLoS Negl Trop Dis* 5: e1231. doi:10.1371/journal.pntd.0001231.

67. McMichael A, Woodruff R, Hales S (2006) Climate change and human health: present and future risks. *Lancet* 367: 859–869.
68. Lambin EF, Turner BL, Geist HJ, Agbola SB, Angelsen A, et al. (2001) The causes of land-use and land-cover change: moving beyond the myths. *Glob Environ Change* 11: 261–269.
69. Ostfeld RS, Glass GE, Keesing F (2005) Spatial epidemiology: an emerging (or re-emerging) discipline. *Trends Ecol Evol* 20: 328–336.
70. Miller RH, Masuoka P, Klein TA, Kim H-C, Somer T, et al. (2012) Ecological niche modeling to estimate the distribution of Japanese encephalitis virus in Asia. *PLoS Negl Trop Dis* 6: e1678. doi:10.1371/journal.pntd.0001678.
71. Kilpatrick AM, Meola MA, Moudy RM, Kramer LD (2008) Temperature, viral genetics, and the transmission of West Nile virus by *Culex pipiens* mosquitoes. *PLoS Pathog* 4: e1000092. doi:10.1371/journal.ppat.1000092.
72. Udvardy MD (1975) A classification of the biogeographical provinces of the world. Morges (Switzerland): International Union for Conservation of Nature and Natural Resources. 48 pp.
73. Xiao X, Boles S, Frohling S, Li C, Babu JY, et al. (2006) Mapping paddy rice agriculture in South and Southeast Asia using multi-temporal MODIS images. *Remote Sens Environ* 100: 95–113. doi:10.1016/j.rse.2005.10.004.
74. Elphick CS, Baicich P, Parsons KC, Fasola M, Mugica L (2010) The future for research on waterbirds in rice fields. *Waterbirds* 33: 231–243.
75. Keiser J, Maltese MF, Erlanger TE, Bos R, Tanner M, et al. (2005) Effect of irrigated rice agriculture on Japanese encephalitis, including challenges and opportunities for integrated vector management. *Acta Tropica* 95: 40–57. doi:10.1016/j.actatropica.2005.04.012.
76. Richards EE, Masuoka P, Brett-major D, Smith M, Klein TA, et al. (2010) The relationship between mosquito abundance and rice field density in the Republic of Korea. *Int J Health Geogr* 9: 32.
77. Lambin EF, Tran A, Vanwambeke SO, Linard C, Soti V (2010) Pathogenic landscapes: interactions between land, people, disease vectors, and their animal hosts. *Int J Health Geogr* 9: 54. doi:10.1186/1476-072X-9-54.
78. Tischendorf L, Fahrig L (2000) On the usage and measurement of landscape connectivity. *Oikos* 90: 7–19.
79. Wada Y (1988) Strategies for control of Japanese encephalitis in rice production systems in developing countries. Vector-borne disease control in humans through rice agroecosystem management: proceedings. International Rice Research Institute in collaboration with the WHO/FAO/UNEP Panel of Experts on Environmental Management for Vector Control.
80. Di Nardo A, Knowles N, Paton D (2011) Combining livestock trade patterns with phylogenetics to help understand the spread of foot and mouth disease in sub-Saharan Africa, the Middle East and Southeast Asia. *Rev Sci Tech* 30: 63–85.
81. Fulmali PV, Sapkal GN, Athawale S, Gore MM, Mishra AC, et al. (2011) Introduction of Japanese encephalitis virus genotype I, India. *Emerg Infect Dis* 17: 319–321.
82. Huang J-H, Lin T-H, Teng H-J, Su C-L, Tsai K-H, et al. (2010) Molecular epidemiology of Japanese encephalitis virus, Taiwan. *Emerg Infect Dis* 16: 876–878.
83. Johansen C, van den Hurk A, Ritchie SA, Zborowski P, Nisbet DJ, et al. (2000) Isolation of Japanese encephalitis virus from mosquitoes (Diptera: Culicidae) collected in the Western Province of Papua New Guinea, 1997–1998. *Am J Trop Med Hyg* 62: 631–638.
84. Chapman HF, Hughes JM, Ritchie SA, Kay BH (2003) Population structure and dispersal of the freshwater mosquitoes *Culex annulirostris* and *Culex palpalis* (Diptera: Culicidae) in Papua New Guinea and northern Australia. *J Med Entomol* 40: 165–169.
85. Nabeshima T, Loan HTK, Inoue S, Sumiyoshi M, Haruta Y, et al. (2009) Evidence of frequent introductions of Japanese encephalitis virus from south-east Asia and continental east Asia to Japan. *J Gen Virol* 90: 827–832.