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RESEARCH ARTICLE

Can Ivermectin kill *Sarcoptes scabiei* during the molting process?

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Abstract

Background

Sarcoptes scabiei is a permanent obligate ectoparasite that lives and reproduces in the epidermis of humans and other mammals worldwide. There is a lack of information on the molting process of Sarcoptes scabiei. Ivermectin is widely used to treat Sarcoptes infection in humans and animals, while the survival of molting Sarcoptes mites in the presence of ivermectin is unknown. The aim of the present study is to investigate the molting process of Sarcoptes mites and assess the activity of ivermectin during the molting process of Sarcoptes mites.

Methodology/Principal findings

molting *Sarcoptes* mites were incubated at 35°C and 80% relative humidity and observed hourly until complete molt. Of the 192 molting mites recorded, the longest molt periods for larvae and nymphs were 23 and 30 h, respectively. The activity of ivermectin on molting *Sarcoptes* mites was also assessed using two concentrations of the drug (0.1 and 0.05 mg/ml). The exposure time for molting mites was determined by 100% mortality of female mites exposed to the solution of ivermectin. While all female mites were killed after exposure to 0.1 mg/ml ivermectin for 2 h and and 0.05 mg/ml for 7 h, 32% and 36% of molting mites survived and successfully molted, respectively.

Conclusions/significance

The present study demonstrated that molting *Sarcoptes* mites are less susceptible to ivermectin than active mites. As a consequence, mites may survive after two doses of ivermectin given 7 days apart due not only to hatching eggs but also to the resistance of mites during their molting process. Our results provide insight into the optimal therapeutic regimens for scabies and highlight the need for further research on the molting process of *Sarcoptes* mites.

Author summary

Sarcoptes scabiei is a permanent obligate ectoparasite causing contagious disease in humans and other mammals worldwide. *Sarcoptes* mites undergo three molts in their lifetime, however, there is limited information on their molting process. Oral ivermectin is widely used for the treatment of *Sarcoptes* infection in humans and animals. To investigate whether ivermectin is active against molting *Sarcoptes* mites, we first explored the molting process of *Sarcoptes* mites *in vitro*, and then assess the activity of ivermectin. We demonstrated that the average molt period lasts for about 2 days. When exposed to ivermectin, one-third of molting mites survived whereas all active mites died. The present study suggests that mites may survive after two doses of ivermectin 7 days apart due to the resistance of mites during the molting process.

Introduction

Sarcoptes scabiei is a permanent obligate ectoparasite that lives and reproduces in the epidermis of humans and other mammals worldwide [1]. The life cycle of *Sarcoptes scabiei* includes eggs, larvae, protonymphs, tritonymphs and adult males or females [2]. During their development, *Sarcoptes* mites undergo three molts: from larvae to protonymphs, from protonymphs to tritonymphs and from tritonymphs to adults. The molting process of *Sarcoptes scabiei* is still unknown and there is no information about the susceptibility of molting *Sarcoptes* mites to acaricides [3].

Ivermectin is commonly used for the treatment of *Sarcoptes* infection in both humans [4,5] and animals [6]. Studies have demonstrated its effectiveness against various mite species, including *Psoroptes cuniculi* [7], *Cheyletiella* mites [8], *Chorioptes* mites [9] and *Demodex* mites [10]. Ivermectin has been shown to be active against the motile stages of *Sarcoptes scabiei*, with *in vitro* tests revealing an LC₅₀ of 1.8 μ M at 24h. However, ivermectin is not effective against all stages of *Sarcoptes* mites [11], and suboptimal responses in some cases [12,13] indicate a need for more detailed investigation in mite biology. A study proved that the eggs of *Sarcoptes* mites cannot be effectively killed by Ivermectin [14].

In the present study, we collected molting *Sarcoptes* mites to explore their molting process and to assess the activity of two concentrations of ivermectin.

Methods

Ethics statement

Mites were obtained from naturally infected New Zealand white rabbits in a rabbit farm in Nanning, Guangxi Province, China, with ethical approval from the ethics committee of Guangxi University (approval no: GXU2019-019).

Sarcoptes mites and compounds

Heavily-infected crusts were placed in Petri dishes and transported to the laboratory within a half-hour. *Sarcoptes* mites were isolated one by one with a needle under a stereomicroscope (SMZ745; Nikon Corp., Tokyo, Japan; 2xmagnification). Mites that were covered with an outer cuticle and remained motionless were considered as molting mites (Fig 1).

Ivermectin (\geq 95%) was purchased from Beijing Solarbio Science & Technology Co., Ltd. Paraffin was purchased from Shanghai Macklin biochemical company, Shanghai, China. We

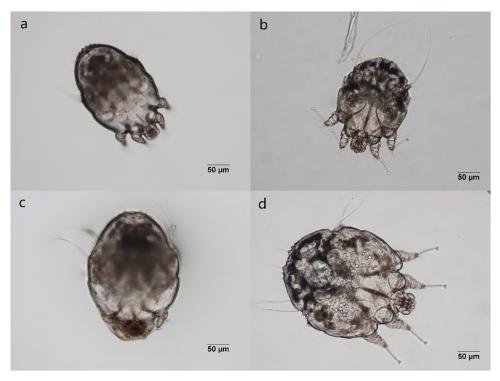


Fig 1. The molting process of *Sarcoptes scabiei* (200x). a: a molting male mite; b: the molted male mite; c: a molting female mite; d: the molted female mite.

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used 10% Medium-chain triglycerides (Medium-chain triglycerides:Paraffin = 1:9) as a solvent for ivermectin, which was purchased from the Nisshin OilliO Group, Ltd.

Molting process of Sarcoptes mites

Molting *Sarcoptes* mites were placed on glass slides. All slides were incubated at 35°C and 80% relative humidity. The status of molting mites was examined under a stereomicroscope (Eclipse 80i; Nikon Corp.; 20×magnification) at hourly intervals until molt.

Assessment of ivermectin activity

To test the efficacy of ivermectin on molting *Sarcoptes* mites, we first exposed female mites to 0.1 and 0.05mg/ml ivermectin diluted in 10% Medium-chain triglycerides (MTC). Acaricidal sensitivity assays were performed using methods previously described [15]. To be brief, female mites (n = 10) were exposed to each concentration of ivermectin with five replicates. In each Petri dish, 1 ml of the solution was included in direct contact with the mites. A control Petri dish was inoculated with 1 ml of 10% MTC. Mites were considered dead when no movement was seen even after touching them with a needle. The exposure time for molting mites was determined by 100% mortality of female mites exposed to the solution of ivermectin. Molting mites (n = 10) were then exposed to 0.1 and 0.05mg/ml ivermectin, respectively. Molting mites were washed and incubated in a humidity chamber (35° C, 80% relative humidity) for the subsequent molting process. Five replicates were performed for each concentration. Negative controls (n = 10) consisted of molting mites exposed to 10% MTC. Mortality and the status of molting mites were measured after 48 h. Five replicates were performed for each concentration of ivermectin. The molting *Sarcoptes* mites were considered dead if they failed to molt after 48 h since no more molting *Sarcoptes* mites molt after this time point.



Fig 2. a: a mite bites through the old cuticle. b: a mite goes out from the old cuticle (200x). https://doi.org/10.1371/journal.pntd.0011337.g002

Data analysis

Data were analyzed by SPSS software version 23.0. The molting time of *Sarcoptes* mites was analyzed with mean ± standard deviation. Survival of molting *Sarcoptes* mites and female mites exposed to ivermectin was compared by one-way analysis of variance (ANOVA) followed by LSD (Least Significant Difference). Values of p < 0.05 were considered significant. The median Lethal Concentration (LC₅₀) was calculated by probit regression analysis. Columns were plotted by Graphpad Prism 9.0 software.

Results

To determine the duration of the molting period, 192 molting *Sarcoptes* mites were collected and examined. Mites during the molting process were quiescent (Fig 1). When they start to molt, they bite through the old cuticle using capitulum (Fig 2). Of the 192 molting mites recorded, the average duration of the molting period was 12.6 \pm 5.9 h for 51 larvae/protonymphs, 17.8 \pm 6.9h for 32 males (including protonymphs/tritonymphs and tritonymphs/ males), and 15.1 \pm 5.7h for 106 females (including protonymphs/tritonymphs and tritonymphs/females). The longest molting periods observed were 23, 30, and 24 h for protonymphs, males (including protonymphs/tritonymphs/males) and females (including protonymphs/tritonymphs and tritonymphs/males). Three molting mites died during the incubation.

When exposed to 0.1 and 0.05 mg/ml of ivermectin, all female mites were dead in 2 and 7 h, respectively. After 2 h exposure to 0.1mg/ml ivermectin, $32 \pm 10.4\%$ of mites were still able to molt successfully. After 7 h of exposure to 0.05 mg/ml ivermectin, $36 \pm 14.2\%$ of mites were still able to molt successfully (Fig 4). Significant differences (p<0.01) were observed between the survival rates of females and molting mites exposed to 0.1 mg/ml and 0.05 mg/ml of

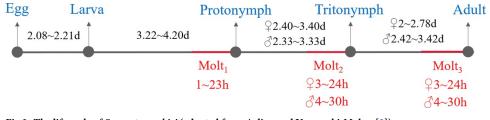
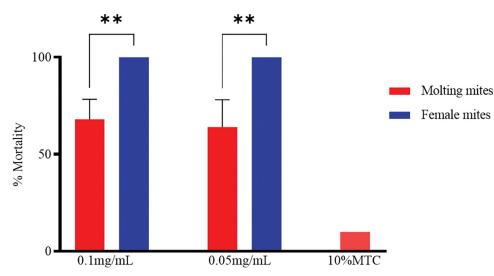
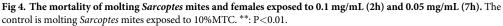


Fig 3. The life cycle of Sarcoptes scabiei (adapted from Arlian and Vyszenski-Moher [2]).

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ivermectin for 2 and 7 h. No significant difference (p>0.05) were observed between the molting *Sarcoptes* mites exposed to 0.1 mg/ml and 0.05mg/ml ivermectin for 2 and 7 h.

Discussion

The present investigation demonstrated for the first time how long it takes for *Sarcoptes* mites to molt. During the 10–13 day-duration of the life cycle of *Sarcoptes* mites [2,16], the average molt period could last for 2 days. Since it is not possible to incubate the molting mites from the start of the molting process, the duration of one molt period could last over 30 h.

Our findings suggest that molting mites are less susceptible to ivermectin than motile mites. This might be attributed to the outer cuticle which offers protection to the mites [16,17]. In the present study, the LC₅₀ for ivermectin against female mites at 1h was 84.1 μ M, which is higher than the values reported in previous studies (50.5 μ M and 45.1 μ M) [18,19]. This could be due to mite resistance since the rabbits had been treated with ivermectin before the investigation.

The molt stage is a crucial point in the life cycle of arthropods, during which the old exoskeleton is shed, and a new one is formed. Currently, there is limited information available on the molting process of parasitic mites. Our findings may enhance the understanding of the life cycle of other parasitic mites.

Studying the molting process of mites could lead to the development of new drugs that target the molt stage. A class of molecules known as benzoylphenyl ureas (BPUs) can disrupt chitin synthesis in arthropod. This disruption prevents the formation of the exoskeleton necessary for molting, ultimately leading mites to death [20]. BPUs, such as diflubenzuron and flufenoxuron, have been found to be effective against several species of mites, including *Tetranychus urticae* [21] and *Panonychus citri* [22]. Juvenile hormone analogs mimic the effects of juvenile hormone and can disrupt the normal molting process in mites, preventing them from maturing [23]. Methoprene and fenoxycarb have been found to be effective against house dust mite (*Dermatophagoides farina*) [24,25]. These drugs offer a notable benefit as they target arthropod-specific physiological and biochemical mechanisms that are not present in vertebrates, thus rendering them practically non-toxic to mammals [26]. Oral ivermectin has become the first-line treatment option for both classical and crusted scabies [27]. It is the only systemic treatment currently available, at a dose of $150-200 \mu g/kg$, to be repeated after 7 days [28]. After oral administration, the maximum concentration (C_{max}) in plasma of ivermectin in humans was 20.2–81.0 ng/ml [29]. A study of pigs measured the C_{max} of ivermectin in the skin as 54.8 ng/ml and the $t_{1/2}$ as one day [30]. Concerning the short half-life of ivermectin, mites during the molting process may survive. For example, if an infested individual receives oral ivermectin, a larva during the molting process may remain on the individual and molt into a protonymph. This protonymph could survive a second dose of ivermectin after 7 days, since it may have progressed to the tritonymph- adult molt stage and eventually develop into an adult, thus continuing the cycle of infection. This may explain why the pigs under heavy infection couldn't be cured completely after two doses of ivermectin, while hatched egg should have been killed by the second treatment [30,31]. Therefore, to clear the infection successfully, a second treatment may be required after the mites have completed their molting process.

One limitation of the current study is that only two concentrations of ivermectin were tested. Collecting molting mites in large quantities is extremely difficult since they remain quiescent, typically located deep within the crust, leading them complicated to work with and even more difficult than mites' eggs.

The molting process of *Sarcoptes* mites has been disregarded due to the challenges associated with finding molting mites and the absence of an in vitro culture system. Our discovery of the molt stage offers promising directions for future research. Additional concentrations of ivermectin must be tested to determine the concentration required to kill 50% of molting mites. The efficacy of other acaricides, such as permethrin or moxidectin, should be evaluated against molting *Sarcoptes* mites. Furthermore, it is worth investigating molecules that have been proven to target the molt stage of arthropods to gain a better understanding of their potential for controlling parasitic mite infestations.

In conclusion, this work provided complementary and original information regarding the life cycle of *Sarcoptes scabiei*. It clearly demonstrated that molting *Sarcoptes* mites are less susceptible to ivermectin than motile mites. This finding could be useful in refining the treatment approach for scabies.

Supporting information

S1 Video. A mite about to molt. (MP4)

S1 Table. Raw experimental data. (XLSX)

Author Contributions

Conceptualization: Fang Fang. Funding acquisition: Fang Fang. Investigation: Shenrui Feng, Minmin Shi, Zhijuan Yin. Methodology: Shenrui Feng. Supervision: Fang Fang. Writing – original draft: Shenrui Feng, Fang Fang. Writing – review & editing: Wenda Di, Jacques Guillot.

References

- Leung AKC, Lam JM, Leong KF. Scabies: A Neglected Global Disease. Curr Pediatr Rev. 2020; 16: 33–42. https://doi.org/10.2174/1573396315666190717114131 PMID: 31544694
- Arlian LG, Vyszenski-Moher DL. Life cycle of Sarcoptes scabiei var. canis. J Parasitol. 1988; 74: 427. https://doi.org/10.2307/3282050 PMID: 3132547
- Arlian LG. Biology, host relations, and epidemiology of Sarcoptes scabiei. Annu Rev Entomol. 1989; 34: 139–161. https://doi.org/10.1146/annurev.en.34.010189.001035 PMID: 2494934
- 4. Rowe ML, Whiteley PL, Carver S. The treatment of sarcoptic mange in wildlife: a systematic review. Parasit Vectors. 2019; 12: 99. https://doi.org/10.1186/s13071-019-3340-z PMID: 30867019
- Bernigaud C, Samarawickrama GR, Jones MK, Gasser RB, Fischer K. The Challenge of Developing a Single-Dose Treatment for Scabies. Trends Parasitol. 2019; 35: 931–943. <u>https://doi.org/10.1016/j.pt.</u> 2019.08.002 PMID: 31474559
- Seaman JT, Thompson DR, Barrick RA. Treatment with ivermectin of sarcoptic mange in pigs. Aust Vet J. 1993; 70: 307–8. https://doi.org/10.1111/j.1751-0813.1993.tb07981.x PMID: 8216098
- Bowman DD, Fogelson ML, Carbone LG. Effect of ivermectin on the control of ear mites (Psoroptes cuniculi) in naturally infested rabbits. Am J Vet Res. 1992; 53: 105–109. PMID: 1539901
- Wagner R, Stallmeister N. Cheyletiella dermatitis in humans, dogs and cats. Br J Dermatol. 2000; 143: 1110–1112. https://doi.org/10.1046/j.1365-2133.2000.03869x
- Rehbein S, Visser M, Winter R, Trommer B, Matthes H-F, Maciel AE, et al. Productivity effects of bovine mange and control with ivermectin. Vet Parasitol. 2003; 114: 267–284. <u>https://doi.org/10.1016/s0304-4017(03)00140-7 PMID: 12809753</u>
- Holzchuh FG, Hida RY, Moscovici BK, Villa Albers MB, Santo RM, Kara-Jose N, et al. Clinical Treatment of Ocular Demodex folliculorum by Systemic Ivermectin. Am J Ophthalmol. 2011; 151: 1030– 1034. https://doi.org/10.1016/j.ajo.2010.11.024 PMID: 21334593
- Goldust M, Rezaee E, Hemayat S. Treatment of scabies: Comparison of permethrin 5% versus ivermectin. J Dermatol. 2012; 39: 545–547. <u>https://doi.org/10.1111/j.1346-8138.2011.01481.x</u> PMID: 22385121
- Mounsey KE, Holt DC, McCarthy JS, Currie BJ, Walton SF. Longitudinal evidence of increasing in vitro tolerance of scabies mites to ivermectin in scabies-endemic communities. Arch Dermatol. 2009; 145: 840–841. https://doi.org/10.1001/archdermatol.2009.125 PMID: 19620572
- Fujimoto K, Kawasaki Y, Morimoto K, Kikuchi I, Kawana S. Treatment for crusted scabies: limitations and side effects of treatment with ivermectin. J Nippon Med Sch. 2014; 81: 157–163. https://doi.org/10. 1272/jnms.81.157 PMID: 24998962
- Bernigaud C, Fernando DD, Lu H, Taylor S, Hartel G, Guillot J, et al. In vitro ovicidal activity of current and under-development scabicides: which treatments kill scabies eggs? Br J Dermatol. 2020; 182: 511–513. https://doi.org/10.1111/bjd.18517 PMID: 31508811
- Fang F, Candy K, Melloul E, Bernigaud C, Chai L, Darmon C, et al. In vitro activity of ten essential oils against Sarcoptes scabiei. Parasit Vectors. 2016; 9: 594. <u>https://doi.org/10.1186/s13071-016-1889-3</u> PMID: 27876081
- Arlian LG, Morgan MS, Rider SD. Sarcoptes scabiei: genomics to proteomics to biology. Parasit Vectors. 2016; 9: 380. https://doi.org/10.1186/s13071-016-1663-6 PMID: 27371026
- Cheong SPS, Huang J, Bendena WG, Tobe SS, Hui JHL. Evolution of Ecdysis and Metamorphosis in Arthropods: The Rise of Regulation of Juvenile Hormone. Integr Comp Biol. 2015; 55: 878–890. https:// doi.org/10.1093/icb/icv066 PMID: 26105594
- Mounsey KE, Walton SF, Innes A, Cash-Deans S, McCarthy JS. *In Vitro* Efficacy of Moxidectin versus Ivermectin against Sarcoptes scabiei. Antimicrob Agents Chemother. 2017; 61. https://doi.org/10.1128/ AAC.00381-17 PMID: 28559258
- Al Khoury C, Nemer N, Nemer G, Kurban M, Bernigaud C, Fischer K, et al. *In Vitro* Activity of Beauvericin against All Developmental Stages of *Sarcoptes scabiei*. Antimicrob Agents Chemother. 2020; 64: e02118–19. https://doi.org/10.1128/AAC.02118-19 PMID: 32122897
- Junquera P, Hosking B, Gameiro M, Macdonald A. Benzoylphenyl ureas as veterinary antiparasitics. An overview and outlook with emphasis on efficacy, usage and resistance. Parasite. 2019; 26: 26. https://doi.org/10.1051/parasite/2019026 PMID: 31041897
- Hernández MM, Martínez-Villar E, Peace C, Pérez-Moreno I, Marco V. Compatibility of the entomopathogenic fungus Beauveria bassiana with flufenoxuron and azadirachtin against Tetranychus urticae. Exp Appl Acarol. 2012; 58: 395–405. https://doi.org/10.1007/s10493-012-9594-1 PMID: 22744196

- 22. Xia W-K, Ding T-B, Niu J-Z, Liao C-Y, Zhong R, Yang W-J, et al. Exposure to Diflubenzuron Results in an Up-Regulation of a Chitin Synthase 1 Gene in Citrus Red Mite, Panonychus citri (Acari: Tetranychidae). Int J Mol Sci. 2014; 15: 3711–3728. https://doi.org/10.3390/ijms15033711 PMID: 24590130
- Duke SO, Menn JJ, Plimmer JR, editors. Pest Control with Enhanced Environmental Safety. Washington, DC: American Chemical Society; 1993. https://doi.org/10.1021/bk-1993-0524
- Downing AS, Wright CG, Farrier MH. Population Growth of Dermatophagoides farinae Hughes (Acari: Epidermoptidae) Suppressed by Methoprene and Hydroprene. J Med Entomol. 1993; 30: 531–536. https://doi.org/10.1093/jmedent/30.3.531 PMID: 8510113
- Downing AS, Wright CG, Farrier MH. Effects of five insect growth regulators on laboratory populations of the North American house-dust mite, Dermatophagoides farinae. Exp Appl Acarol. 1990; 9: 123–130. https://doi.org/10.1007/BF01198990 PMID: 2226070
- Sun R, Liu C, Zhang H, Wang Q. Benzoylurea Chitin Synthesis Inhibitors. J Agric Food Chem. 2015; 63: 6847–6865. https://doi.org/10.1021/acs.jafc.5b02460 PMID: 26168369
- Bernigaud C, Fischer K, Chosidow O. The Management of Scabies in the 21st Century: Past, Advances and Potentials. Acta Derm Venereol. 2020; 100: adv00112-234. <u>https://doi.org/10.2340/00015555-</u> 3468 PMID: 32207535
- Salavastru CM, Chosidow O, Boffa MJ, Janier M, Tiplica GS. European guideline for the management of scabies. J Eur Acad Dermatol Venereol. 2017; 31: 1248–1253. https://doi.org/10.1111/jdv.14351 PMID: 28639722
- González Canga A, Sahagún Prieto AM, Diez Liébana MJ, Fernández Martínez N, Sierra Vega M, García Vieitez JJ. The pharmacokinetics and interactions of ivermectin in humans—a mini-review. AAPS J. 2008; 10: 42–46. https://doi.org/10.1208/s12248-007-9000-9 PMID: 18446504
- 30. Bernigaud C, Fang F, Fischer K, Lespine A, Aho LS, Dreau D, et al. Preclinical Study of Single-Dose Moxidectin, a New Oral Treatment for Scabies: Efficacy, Safety, and Pharmacokinetics Compared to Two-Dose Ivermectin in a Porcine Model. Vinetz JM, editor. PLoS Negl Trop Dis. 2016; 10: e0005030. https://doi.org/10.1371/journal.pntd.0005030 PMID: 27732588
- Bernigaud C, Fang F, Fischer K, Lespine A, Aho LS, Mullins AJ, et al. Efficacy and Pharmacokinetics Evaluation of a Single Oral Dose of Afoxolaner against Sarcoptes scabiei in the Porcine Scabies Model for Human Infestation. Antimicrob Agents Chemother. 2018; AAC.02334-17. https://doi.org/10.1128/ AAC.02334-17 PMID: 29914951