

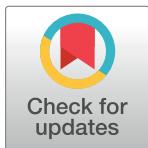
REVIEW

Global prevalence of 4 neglected foodborne trematodes targeted for control by WHO: A scoping review to highlight the gaps

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Abstract

Background

Foodborne trematodiases (FBTs) are a group of trematodes targeted for control as part of the World Health Organization (WHO) road map for neglected tropical diseases from 2021 to 2030. Disease mapping; surveillance; and capacity, awareness, and advocacy building are critical to reach the 2030 targets. This review aims to synthesise available data on FBT prevalence, risk factors, prevention, testing, and treatment.

Methods

We searched the scientific literature and extracted prevalence data as well as qualitative data on the geographical and sociocultural risk factors associated with infection, preventive/protective factors, and methods and challenges of diagnostics and treatment. We also extracted WHO Global Health Observatory data representing the countries that reported FBTs during 2010 to 2019.

Results

One hundred and fifteen studies reporting data on any of the 4 FBTs of focus (*Fasciola* spp., *Paragonimus* spp., *Clonorchis* sp., and *Opisthorchis* spp.) were included in the final selection. Opisthorchiasis was the most commonly reported and researched FBT, with recorded study prevalence ranging from 0.66% to 88.7% in Asia, and this was the highest FBT prevalence overall. The highest recorded study prevalence for clonorchiasis was 59.6%, reported in Asia. Fascioliasis was reported in all regions, with the highest prevalence of 24.77% reported in the Americas. The least data was available on paragonimiasis, with the highest reported study prevalence of 14.9% in Africa. WHO Global Health Observatory data indicated 93/224 (42%) countries reported at least 1 FBT and 26 countries are likely co-endemic to 2 or more FBTs. However, only 3 countries had conducted prevalence estimates for multiple FBTs in the published literature between 2010 to 2020. Despite differing epidemiology, there were overlapping risk factors for all FBTs in all geographical areas, including proximity

Competing interests: None declared.

to rural and agricultural environments; consumption of raw contaminated food; and limited water, hygiene, and sanitation. Mass drug administration and increased awareness and health education were commonly reported preventive factors for all FBTs. FBTs were primarily diagnosed using faecal parasitological testing. Triclabendazole was the most reported treatment for fascioliasis, while praziquantel was the primary treatment for paragonimiasis, clonorchiasis, and opisthorchiasis. Low sensitivity of diagnostic tests as well as reinfection due to continued high-risk food consumption habits were common factors.

Conclusion

This review presents an up-to-date synthesis on the quantitative and qualitative evidence available for the 4 FBTs. The data show a large gap between what is being estimated and what is being reported. Although progress has been made with control programmes in several endemic areas, sustained effort is needed to improve surveillance data on FBTs and identify endemic and high-risk areas for environmental exposures, through a One Health approach, to achieve the 2030 goals of FBT prevention.

Background

The cluster of selected foodborne trematodes (FBTs) listed by the World Health Organization (WHO) consists of 4 genera of trematodes (*Fasciola* spp., *Paragonimus* spp., *Clonorchis* sp., and *Opisthorchis* spp.) [1]. Despite the prominent public health impacts of the FBTs, they remain listed as neglected tropical diseases (NTDs) by WHO. FBTs, like other NTDs, impact the most impoverished populations and lack the surveillance systems and tools to adequately ascertain their true burden [2,3]. FBTs are zoonotic diseases, with a complex lifecycle involving a primary intermediate snail host, and a secondary intermediate host for all except *Fasciola* spp. (crustaceans for *Paragonimus* spp., freshwater fish for *Clonorchis* sp. and *Opisthorchis* spp.), with humans becoming infected via the consumption of contaminated food [2].

FBT infections in humans result in a range of clinical symptoms including abdominal pain and fever, but could escalate to result in damage to internal organs including severe damage to the liver and lungs [1].

Fasciola hepatica is globally distributed, with *F. gigantica* distribution restricted to Africa and Asia. Infection sources are diverse and include contaminated food and water [4–6]. Fascioliasis most commonly results in inflammation of the bile ducts, gallbladder, and liver, resulting in liver fibrosis [5,6], but adult trematodes can also occur in the eyes and central nervous system, resulting in severe neurological and ocular symptoms [7]. Infection with *Paragonimus* spp. is acquired through the consumption of undercooked crab or crayfish and is found in Africa, Asia, and Latin America [4,5]. Adult flukes lodge in the lung tissue of the final host and can result in a chronic cough with bloody sputum, chest pain, and dyspnoea [5,6]. Ectopic infection in the brain is uncommon and may result in headaches, convulsions, and cerebral haemorrhages [5,6]. *Clonorchis sinensis* and *Opisthorchis viverrini* are largely confined to Asia, with infection acquired via the consumption of undercooked fish [5,6]. These parasites are classified as carcinogenic, as adult flukes can lodge in the bile ducts of the liver, causing inflammation of tissues and resulting in cholangiocarcinoma, a fatal bile duct cancer [5,6]. Infection with *Opisthorchis felineus* may result in acute abdominal pain due to gallbladder obstruction, and there is evidence that this is also carcinogenic [8].

FBTs have been reported in more than 70 countries worldwide [1]. The WHO Foodborne Disease Burden Epidemiology Reference Group attributes 200,000 illnesses and more than 7,000 deaths to FBTs annually [9]. In 2015, this was estimated to result in 1,066 thousand years disability-adjusted life years (DALYs) lost globally [10].

FBTs largely affect populations with poor sanitation and poor public health awareness [11,12]. The diversity of FBTs, range of clinical presentation, lack of sensitive diagnostic tools and dedicated surveillance systems, differences in host susceptibility, and different cultural food habits contribute to the underestimation and underreporting of these pathogens [2,6]. The true burden remains unknown as accurate prevalence data are scarce, and many endemic countries do not have national surveillance programmes in place [2].

FBTs are targeted for control as part of the WHO NTD road map 2021–2030, with mapping and surveillance and advocacy, capacity, and awareness building identified as critical actions required to reach the 2030 targets. A core component of this road map is the promotion of integrated One Health approaches in the development and implementation of NTD prevention and control programmes. Such One Health initiatives have already been implemented in several endemic areas, and while these initiatives have highlighted the complexity of FBT epidemiology and control, promising outcomes have been demonstrated which can be used as an example in other areas.

While several publications have introduced estimates of FBTs in 2012, up-to-date global estimates of foodborne trematodes focussing on prevalence data are needed to support the 2030 WHO NTD road map [6].

Considering these FBTs are confined to areas where the intermediate host species inhabit and the specific cultural or food habits of people leading to exposure to the pathogens, identifying the FBTs in relation to the geographical area is important. Furthermore, knowledge on co-endemicity of multiple FBTs in certain countries may inform planning campaigns of preventive chemotherapy against more than 1 FBT simultaneously, which could improve the cost-effectiveness of these campaigns [2,11,13–15]. Therefore, the overarching objective of this review is to bring together available data on reporting, prevalence, risk factors, prevention, testing, and treatment of the FBTs while identifying the potential for co-endemicity and data gaps at the country level.

Methods

Search strategy and selection criteria of published scientific literature

An initial broad-based search of PubMed, IRIS, Web of Science, Science Direct, and Cochrane electronic databases was performed using a combination of search terms, including each FBT and the terms “burden,” “prevalence,” “incidence,” and “cases.” A complete list of the specific search terms used can be found in Box A in [S1 Supplementary material](#). No language restrictions were set, although all search terms were in English. All references published from January 2010 through February 2020 were included in the review. Additional records were identified through a snowballing approach, whereby the bibliographies of full text articles included from the initial literature search were screened, and any reference published after 2010 was reviewed using the same inclusion/exclusion criteria.

Records were initially screened based on title and abstract and were excluded if the abstract focused on an animal population and did not identify a human population of interest, and if the abstract did not mention 1 of the 4 trematode species of interest (*Fasciola* spp., *Paragonimus* spp., *Clonorchis* sp., and *Opisthorchis* spp.). Records were retained for full-text review if they identified a human population of interest and identified 1 or more of the 4 FBTs of interest. Full-text articles were evaluated for inclusion, and a second reviewer was consulted where

there was ambiguity. Records were excluded if there was no prevalence or incidence recorded for a human population, for 1 or more of the 4 FBTs of interest. Records were also excluded if they did not specify the geographical area where data was collected, or if no diagnostic method was identified for determining prevalence. While the focus was on prevalence, 2 publications reported an incidence rate, and these records were included to highlight the presence of *O. felineus* captured in the literature. Extracted data included: parasite species and subspecies, geographical location, reported prevalence, time frame of study, diagnostic methods used, diagnostic challenges, treatments used, treatment challenges, environmental risk factors, sociocultural risk factors, and preventive factors.

Extracting quantitative and presence only data

The prevalence of the 4 FBTs (i.e., fascioliasis, paragonimiasis, clonorchiasis, and opisthorchiasis) were extracted.

Prevalence data at national level were scarce; therefore, prevalence studies of smaller spatial areas were recorded for the purpose of this review. Where multiple records were available for a specific country and parasite, the minimum and maximum prevalence were recorded. The full list of countries with WHO data and the selected studies and prevalence reported are included in Table A in [S1 Supplementary material](#).

Presence only data on FBTs reported between 2010 and 2019 were extracted from the WHO Global Health Observatory [16].

Extracting qualitative data

We extracted qualitative data related to the risk and preventive factors associated with FBTs and grouped them into the following categories: environmental and sociocultural risk factors for infection, preventive measures, diagnostic methods and challenges, and treatment methods and challenges from each record. Qualitative data that were extracted included all risk and preventive factors that were discussed in each record and not only those which were the focus of the study.

Mapping endemicity and co-endemicity of FBTs

Reported “Presence only” data from the WHO Global Health Observatory were mapped using Adobe Illustrator CS5, version 15.1.0 on WHO official template of world map, to represent the combinations of FBTs reported to the WHO Global Health Observatory. Countries that had reported more than 2 FBTs were considered to have potential for geographical co-endemicity. WHO records were compared with the records extracted through the review process to identify countries that have conducted epidemiological studies on the FBTs between 2010 and 2020 even if the records were not submitted to WHO.

Results

Literature review

After screening 8,926 records, 115 eligible full-text records providing prevalence data from 25 countries were included in the final dataset ([Fig 1](#)).

Reported species, geography, and prevalence

Opisthorchiasis was the most recorded FBT globally, with 41 of the final full-text records reporting prevalence from 7 countries ([Table 1](#)). The highest recorded study prevalence for opisthorchiasis was in Lao People’s Democratic Republic, with a prevalence of up to 88.7%

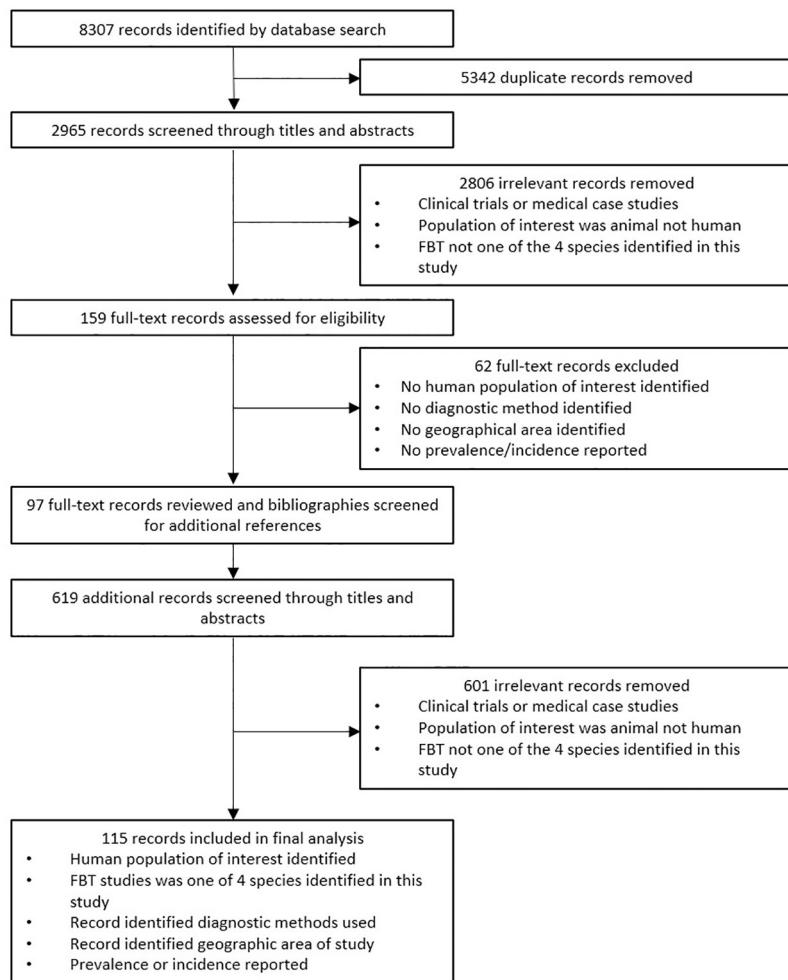


Fig 1. Flow diagram summarising the selection process of the studies in the review process.

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recorded in some areas. This study prevalence was overall the highest recorded of all the FBTs in this review.

Fascioliasis had 36 records included in the final review, with the widest geographical range, representing 16 countries and all regions. The highest fascioliasis prevalence reported was 24.77% in Bolivia.

Clonorchiasis had 32 records included in the final review, reporting prevalence from 3 Asian countries. The highest recorded study prevalence for clonorchiasis was in China, with a prevalence of up to 59.6% recorded in some areas.

The least number of records were available on paragonimiasis, with only 11 papers reporting study prevalence from 7 countries, and the highest study prevalence of 14.9% was reported in Cameroon.

The following countries had the most available records on FBTs: 15 records for clonorchiasis in the Republic of Korea and 13 records in China, 12 records for fascioliasis in the Islamic Republic of Iran and 7 records in Peru, 19 records for opisthorchiasis in Thailand and 11 records in Lao People's Democratic Republic.

Table 1. Study prevalence range of FBTs recorded in the literature from 2010–2020, presented by country and parasite.

Geographical region (and WHO region)	Country	Parasite	Study prevalence (range provided when more than 1 record)	Time frame of study	Study prevalence identified in most recent record	Range of sample sizes	No. of records included	Diagnostic methods used	References
Africa (AFR)	Ethiopia	<i>Fasciola</i> spp.	3.30%	2007–2008	3.3%	520	1	Parasitological (faecal)	[17]
	United Republic of Tanzania	<i>Fasciola</i> spp.	21.00%	2012–2013	21.0%	1,460	1	Parasitological (faecal)	[18]
	Cameroon	<i>Paragonimus africanus</i>	14.90%	2004–2006	14.9%	168	1	Immunological (serological) Parasitological (faecal and sputum)	[19]
Americas (AMR)	Argentina	<i>Fasciola hepatica</i>	11.90%	Not stated	11.9%	42	1	Immunological (serological) Parasitological (faecal)	[20]
	Brazil	<i>Fasciola hepatica</i>	1.80%	2013	1.8%	434	1	Immunological (serological) Parasitological (faecal)	[21]
	Bolivia	<i>Fasciola hepatica</i>	21.7–24.77%	2008	24.77%	436–437	2	Immunological (ELISA for coproantigen detection) Parasitological (faecal)	[22,23]
	Cuba	<i>Fasciola hepatica</i>	0.33%	2011–2012	0.33%	300	1	Parasitological (faecal)	[24]
	Haiti	<i>Fasciola hepatica</i>	6.50%	Not stated	6.5%	216	1	Immunological (serological)	[25]
	Mexico	<i>Fasciola hepatica</i>	5.78%	Not stated	5.78%	865	1	Parasitological (faecal)	[26]
	Peru	<i>Fasciola hepatica</i>	2.3–24.4%	2007–2017	10.10%	223–2,515	7	Immunological (serological) Parasitological (faecal)	[22, 27–32]
	Kyrgyzstan	<i>Fasciola hepatica</i>	1.90%	2009	1.9%	1,262	1	Parasitological (faecal)	[33]
European (EUR)	Turkey	<i>Fasciola hepatica</i>	0.04–5.6%	1977–2010	5.60%	1,600–69,633	2	Immunological (serological) Parasitological (faecal)	[34,35]
	Kazakhstan	<i>Opisthorchis felineus</i>	*incidence reported 7.4/100,000	1997–2011			1	Parasitological (faecal)	[36]
	Russian Federation	<i>Opisthorchis felineus</i>	*incidence reported 24.7/100,000	2011–2013			1	Parasitological (faecal)	[37]
	Iran (Islamic Republic of)	<i>Fasciola</i> spp.	0.13–24.8%	2003–2016	2.60%	206–1984	12	Immunological (serological) Parasitological (faecal)	[38–49]
Eastern Mediterranean (EMR)	Pakistan	<i>Fasciola</i> spp.	0.74–1.18%	2003–2005	0.74%	540–7,200	2	Parasitological (faecal)	[50,51]

(Continued)

Table 1. (Continued)

Geographical region (and WHO region)	Country	Parasite	Study prevalence (range provided when more than 1 record)	Time frame of study	Study prevalence identified in most recent record	Range of sample sizes	No. of records included	Diagnostic methods used	References
Southeast Asia (SEAR)	India	<i>Paragonimus</i> spp.	6.16–11.0%	2008–2011	11.00%	624–4,371	2	Immunological (serological) Parasitological (faecal and sputum)	[52,53]
	Myanmar	<i>Opisthorchis viverrini</i>	0.7–9.3%	2015–2016	9.30%	364–2,057	2	Parasitological (faecal)	[54,55]
	Thailand	<i>Opisthorchis viverrini</i>	2.48–45.69%	1990–2017	17.00%	245–18,900	19	Immunological (urine ELISA) Parasitological (faecal)	[56–74]
Western Pacific (WPR)	Cambodia	<i>Opisthorchis viverrini</i>	4.6–47.5%	2006–2012	7.66%	228–32,201	6	Parasitological (faecal)	[75–80]
	China	<i>Fasciola gigantica</i>	8.8%	2011	8.8%	3,177	1	Immunological (serological) Parasitological (faecal)	[81]
		<i>Paragonimus</i> spp.	0.02–7.46%	2006–2013	0.02%	724–8,396	2	Immunological (serological) Parasitological (faecal)	[82,83]
		<i>Clonorchis sinensis</i>	0.03–59.6%	1989–2018	38.72%	718–356,629	13	Immunological (serological) Parasitological (faecal)	[82,84–95]
	Japan	<i>Paragonimus</i> spp.	8.5%	2001–2012	8.5%	5,200	1	Immunological (serological)	[96]
	Philippines	<i>Paragonimus westermani</i>	6.70%	2011–2013	6.7%	836	1	Parasitological (sputum)	[97]
	Lao People's Democratic Republic	<i>Opisthorchis viverrini</i>	0.66–88.7%	2006–2015	87.90%	207–6,178	11	Parasitological (faecal)	[98–108]
	Republic of Korea	<i>Paragonimus westermani</i>	1.6–2.8%	1993–2011	1.60%	720–74,448	3	Immunological (serological)	[109–111]
		<i>Clonorchis sinensis</i>	0.2–28.2%	1993–2017	0.20%	231–99,451	15	Immunological (serological) Parasitological (faecal)	[109–123]
Viet Nam		<i>Fasciola</i> spp.	5.90–7.8%	2012–2013	7.8%	1,612–10,084	2	Immunological (serological)	[124,125]
		<i>Paragonimus westermani</i>	5.40%	Not stated	5.4%	590	1	Immunological (serological) Parasitological (sputum)	[126]
		<i>Clonorchis sinensis</i>	16.47–22.72%	2009–2017	19.50%	400–1,857	4	Parasitological (faecal)	[127–130]
		<i>Opisthorchis viverrini</i>	11.40%	2015	11.4%	254	1	Parasitological (faecal)	[131]

* Incidence rates were reported, but no prevalence rate. These records were included to highlight that *O. felineus* presence was captured in the literature, in the specified time frame.

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Global health observatory

Among the 224 countries and territories reporting data to the WHO Global Health Observatory from 2010 to 2019, 93/224 (42%) countries reported at least 1 of the 4 FBTs (i.e., 131/224 (58%) did not report any), and 2019 was the last available year of data as the process of reporting and validation of NTD surveillance data to WHO entails approximately 1-year lag in data

availability. Fascioliasis was reported in 75/224 (33%), paragonimiasis in 44/224 (20%), clonorchiasis in 10/224 (4%), and opisthorchiasis in 7/224 (3%) countries. Among the reporting countries, 26 were co-endemic to 2 or more FBTs.

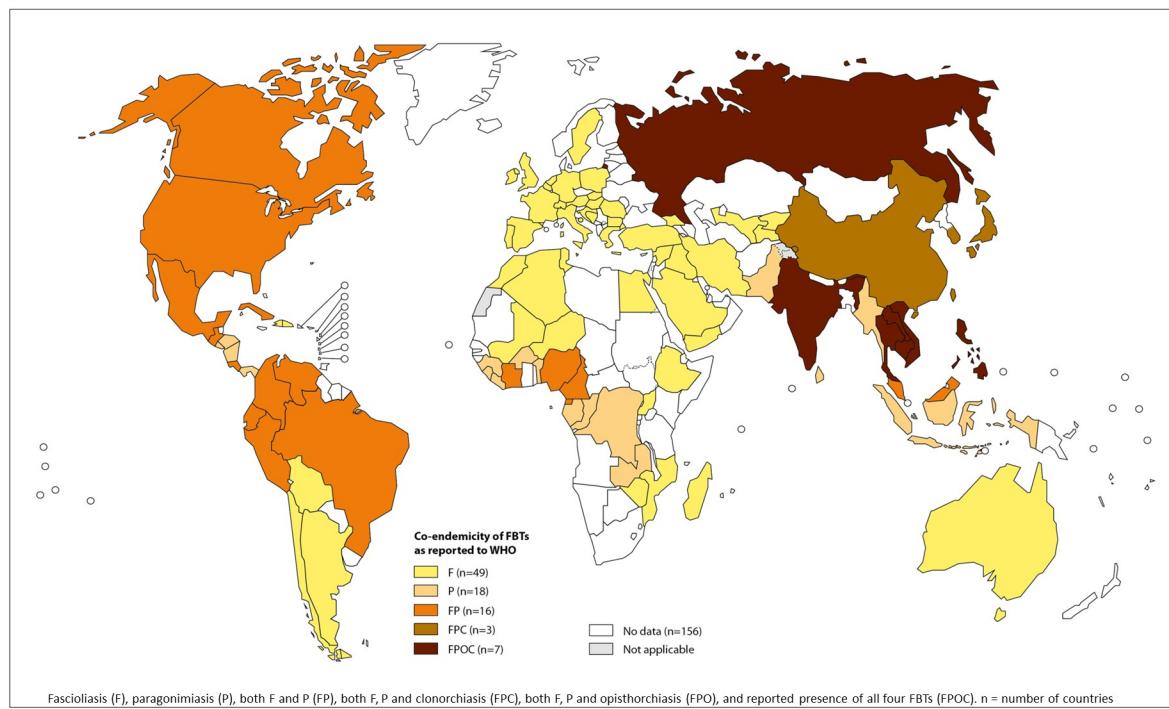
Co-endemicity (2 or more FBTs identified)

Countries reporting co-endemicity of FBTs to WHO included: (i) 16 countries reporting fascioliasis and paragonimiasis; (ii) 3 countries reporting fascioliasis, paragonimiasis, and clonorchiasis (China, Japan, and the Republic of Korea); and (iii) 7 countries reported all 4 FBTs (Cambodia, Lao People's Democratic Republic, India, the Philippines, Russian Federation, Thailand, and Viet Nam) (Fig 2). Except for the United Republic of Tanzania and Kazakhstan, all countries that were included in the scoping review process also reported to WHO for at least 1 of the 4 FBTs during the 2010 to 2019 period. However, among these 26 countries with co-endemicity, only 3 countries (China, Republic of Korea, and Viet Nam) had conducted prevalence estimates for multiple FBTs in the published literature between 2010 and 2020.

Fascioliasis

Environmental risk factors. Rural/agricultural environments were commonly identified risk factors for all geographical areas where fascioliasis was recorded. Rural and agricultural

Presence of the four neglected foodborne trematodes targeted for control by WHO (As reported to WHO between 2010 – 2020)



The boundaries and names shown and the designations used on this map do not imply the expression of any opinion whatsoever on the part of the World Health Organization concerning the legal status of any country, territory, city or area or of its authorities, or concerning the delimitation of its frontiers or boundaries. Dotted and dashed lines on maps represent approximate border lines for which there may not yet be full agreement. © WHO 2014. All rights reserved.

Data source: World Health Organization
Map production: Control of Neglected Tropical Diseases (NTD)
World Health Organization



Fig 2. Map depicting the endemicity or co-endemicity of the 4 neglected FBTs as reported to the WHO between 2010 and 2019. The categories include fascioliasis (F); paragonimiasis (P); both F and P (FP); both F, P, and clonorchiasis (FPC); both F, P, and opisthorchiasis (FPO); and reported presence of all 4 FBTs (FPOC). The number of countries under each category is listed.

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environments were often remote, with poor health care access, limited access to safe water, poor personal and environmental sanitation, and suitable intermediate and reservoir hosts to maintain transmission [17,18,24,26,27,31–33,35,39,43,47,46].

Suitable aquatic environments were identified as necessary for sustaining populations of snail intermediate hosts and maintaining the life cycle of *Fasciola* spp., making high rainfall areas, floodplains, and irrigated areas high-risk environments for *Fasciola* transmission [17,21,26,31,32,35,40,43,44,48,50,51].

High altitude, mountainous areas were identified as a risk factor for fascioliasis transmission in many records from the Region of the Americas, with hyperendemic areas identified in populations in higher altitude communities [24–29,31,32,132].

Areas with warm temperatures, irrigation, or environmental conditions resulting in high soil moisture favoured fascioliasis transmission in the Eastern Mediterranean Region, with the emergence of new disease foci observed in central and southwestern parts of the Islamic Republic of Iran [40,43,44,48,50,51].

Sociocultural risk factors. The consumption of raw vegetables and freshwater plants was a key risk factor in every geographical area where fascioliasis was reported.

Poor sanitation and hygiene was identified as a risk factor for all regions [18,19,21,24,26,28,29,31–33,35,37,42,47–51,54]. Poor water and sanitation included drinking of contaminated water, poor sanitation infrastructure, open defaecation, and poor food hygiene. Inadequate water, sanitation, and hygiene (WASH) contributed to ongoing transmission cycles, with poor sanitation resulting in contamination of water supplies that were used for drinking, cooking, and irrigation of crops [17,18,21,24,25,29,31,32,35,42,47,51,124].

Close contact with livestock was an important risk factor identified in the African, Americas, Eastern Mediterranean, and Western-Pacific regions, with free-roaming livestock and the use of animal manure on crops resulting in contamination of food and water sources [17,18,21,25,26,32,38,43,46,47,49–51,124].

Poverty and low socioeconomic settings were identified as risk factors in the Americas, Eastern Mediterranean, European, and Western Pacific regions [28,29,31,35,124].

Diagnostics. Faecal coprological tests were the commonly reported diagnostic method in all regions, except for the Eastern Mediterranean Region (represented primarily by the Islamic Republic of Iran), where serological methods were used for diagnosis in most of the included records [38–49].

Faecal parasitological tests were considered simple, cheap, and rapid for diagnosis and mass screening of populations. However, frequently identified challenges to faecal coprological tests included the low sensitivity of these tests due to low egg burdens, intermittent egg shedding, or acute infections, resulting in possible false negatives and underestimation of prevalence [19–21,26,28–30,32–34,47,77,81]. Increasing the number of faecal samples collected and increasing the number of slides examined from each sample were both identified as options to improve the sensitivity of these methods, but required further resources, had a lower compliance from patients, and faced logistical challenges with collection in remote areas [40,41,51].

The Eastern Mediterranean Region was the only region where serological diagnostic methods were more frequently used than faecal coprological tests and were considered more sensitive [40,48,49]. However, serological testing was unable to determine between past and current infections, and the possibility of cross-reactivity with other parasites could result in false positives [20,21,31,48,124]. These challenges meant that serological testing did not necessarily equate to true active cases and could overestimate *Fasciola* prevalence [31,45,124].

Using both parasitological and serological testing methods improved sensitivity and specificity of diagnosis; however, this was logistically challenging in many settings [21,25,35,41,124]. Coproantigen techniques were reported to be an easy and fast way in which

to mass screen communities, allowing for rapid selective treatment, and reducing the probability of drug resistance [22].

Treatment and preventive measures. Treatment with triclabendazole was the most reported treatment for fascioliasis, although challenges to treatment included drug resistance, adverse drug reactions, and difficulty in obtaining the medication [18,20,29,33,34,47,81]. Four records reported the dosing regimen used for treatment, with 3 records using a single dose of 10 mg/kg [23,34,47] and the third record using 10 mg/kg per day for 2 days [81]. One record from Tanzania reported the use of nitazoxanide in areas with limited supply of triclabendazole, noting that not all patients were cleared with this treatment [18]. Another record reported the use of nitazoxanide in Mexico, noting its potential as an alternative to triclabendazole in countries where triclabendazole is not registered or where triclabendazole resistance is found [26].

Improved health education and awareness around disease transmission and risk factors was a commonly reported preventive measure, although it was noted that this needed to be implemented alongside improved WASH, disease surveillance, and improved access to diagnosis and treatment [17,33].

Management of reservoir species to interrupt transmission cycles was identified as an important preventive measure, with recommendations including surveillance and treatment of reservoir species, management of intermediate snail host populations, and grazing management of livestock, highlighting the need for an integrated approach to fascioliasis surveillance and management [20,21,25,27,31,38–40,43,45–49].

Paragonimiasis

Environmental risk factors. The Three Gorges Reservoir area of China was reported to be an area of high paragonimiasis endemicity, with hilly and forested areas, and natural bodies of water that sustain populations of both the intermediate snail host and intermediate freshwater crab host [93]. Mountainous areas of Vietnam were reported endemic and provided suitable habitats for intermediate mountainous crab hosts [128], while hyperendemic foci of paragonimiasis were identified in rural, remote hilly, and forested areas of northeastern India where populations had poor access to health care [53].

Sociocultural risk factors. The consumption of raw or undercooked crab was identified as a key risk factor in the Southeast Asia and Western Pacific regions [53,83,97,109–111,126], with the consumption of raw boar meat also identified as a risk factor in Japan [96]. Poverty and lack of education were identified as factors that contributed to high paragonimiasis prevalence in endemic areas of India [53].

Diagnostics. Reported diagnostic methods included examination of sputum and faeces for eggs, serological diagnosis, and intradermal testing. Serological methods were reported as a more sensitive diagnostic tool in both the African and the Southeast Asia regions, as ova were expectorated or shed intermittently in sputum and faeces, respectively [19,52]. Although positive serological results could include past infections and cross-reactions, it was considered a reliable diagnostic method [111]. Collection of multiple sputum samples increased diagnostic sensitivity of microscopy methods, with a record from the Philippines recommending 2 sputum samples to allow same day diagnosis, thereby improving patient compliance and reducing costs of diagnostics compared to samples collected over multiple days [97]. The same record recommended the collection of an early morning sputum sample followed by a spot sample, as the early morning sample improved sensitivity [97].

Co-endemicity and similarity of clinical symptoms with pulmonary tuberculosis, most notably a cough and hemoptysis, was a challenge for diagnosis in India and the Philippines [52,53,97]. Assumptions that these symptoms were indicative of tuberculosis contributed to

underdiagnosis of paragonimiasis. These records recommended the integration of pulmonary tuberculosis and paragonimiasis surveillance and control programmes, noting that serological testing of symptomatic but tuberculosis negative patients, could improve paragonimiasis detection [52,53,97].

Treatment and preventive measures. Praziquantel was the only treatment reported, with 1 record reporting a treatment regime of 20 mg/kg per day for 6 days [83] and another reporting 75 mg/kg per day for 3 days [96].

Targeted active case detection and treatment of infected cases together with community education was found to reduce the prevalence of paragonimiasis in highly endemic areas of northeastern India [53].

Challenges to treatment included cases of underdosing reported in Japan, and a need for increased awareness among clinicians [96], and reinfection due to continued high-risk cultural food habits in Viet Nam despite more than 15 years of mass screening, treatment, and education [126].

Clonorchiasis

Environmental risk factors. Rural/agricultural areas with poor sanitary conditions and river basin areas were identified as risk factors for clonorchiasis, as these environments supported both the intermediate freshwater snail host and the intermediate freshwater fish hosts [87,113,115,117].

Sociocultural risk factors. Consumption of raw freshwater fish was identified as the key risk factor for clonorchiasis, with reinfection in communities common due to deeply entrenched cultural food habits [87,117]. Poor sanitary conditions and food hygiene was also identified as a risk factor that could result in the contamination of food and utensils, leading to ingestion of metacercariae [87,88,90,117].

Diagnostics. Diagnosis by faecal parasitological methods were commonly reported, using either formalin-ether concentration technique (FECT) or the Kato–Katz thick smear method [122]. Such methods were widely used as they were simple, non-invasive, rapid, inexpensive, and were able to determine both the diagnosis and the intensity of infection [84,90,91]. However, microscopy methods could result in false negative results, particularly in light infections, and there were challenges in differentiating *C. sinensis* from other minute intestinal flukes [85,87,92,117]. Increasing the size of the faecal sample, repeating egg counts, and increasing the number of slides improved the sensitivity but also increased time and labour costs and was not always acceptable to communities [87].

Serological methods such as ELISA improved the sensitivity, but had poor specificity and were unable to detect early phases of infection [87]. Cross-reactivity with other parasites could also result in additional false-positive results [87,111]. However, while the ELISA method was recommended to be an auxiliary method to faecal microscopy for the diagnosis of individuals, ELISA methods were noted as being a potential option for large-scale screening to monitor community prevalence [87].

Molecular methods such as PCR were able to improve specificity and sensitivity but the high cost, need for trained personnel, and laboratory facilities, made this impractical for point-of-care diagnosis [87,90].

Several records noted that there may be value in using reports of raw freshwater fish consumption as a screening tool to identify high-risk communities, as this was simple, low cost, and had improved compliance from communities compared to faecal collection [92,117].

Treatment and preventive measures. Repeated mass drug administration (MDA) using praziquantel, combined with health education were commonly recommended prevention/

treatment policies; however, reinfection was common due to the difficulty in changing deeply rooted cultural food habits [84,88,114,116]. Health education programmes that targeted community leaders were possibly more effective at promoting and maintaining changed behaviour in communities [116].

25 mg/kg every 5 h for 3 doses was the reported praziquantel dose regime by 4 records [92,116,117,129], with another record reporting a single treatment of 40 mg/kg [118]. Poor compliance with taking the second and third dose of praziquantel was identified as a possible factor for low cure rates in some communities [84]. Repeated mass or selective treatment every 6 to 12 months was recommended for reducing prevalence and reinfection in heavily endemic areas [8,115,119].

Reservoir hosts were an additional challenge to control, with diagnosed cases reported in dogs, cats, and pigs, and these reservoir hosts contributed to environmental contamination and sustained parasite lifecycle [84,89]. Treatment of reservoir hosts may contribute to reducing the source of infection in communities [84].

An integrated approach was recommended, with improved access to anthelmintics, improved health education, avoidance of raw fish consumption, active screening for early disease detection, treatment of reservoir hosts, and elimination of intermediate host snails [84,114,115].

Opisthorchiasis

Environmental risk factors. River basins were reported risk areas for the transmission of *Opisthorchis* species, as these environments supported both the snail and fish intermediate hosts [37,60]. Similarly, areas of higher rainfall and rural, lowland villages with surrounding land that was dominated by high water content (wetlands, paddies, streams, ponds, and lakes) were reported to have high prevalence of *O. viverrini* due to the suitable environments for intermediate hosts [60,99,103]. The development of water resources for aquaculture and irrigation may have contributed to the high prevalence found in northeast Thailand [60]. Higher altitude areas were reported to have a lower risk of opisthorchiasis, although this was likely related to accessibility to freshwater fish and the difference of cultural food preferences [99].

High infection rates were reported for *O. felineus* in Western Siberia [37] and northern Kazakhstan [36], while high prevalence of *O. viverrini* was reported throughout the Mekong Basin in Southeast Asia, including endemic areas in north and northeast Thailand, central and southern Lao PDR, southern Cambodia, and southern Viet Nam [54,56,57,59,60,62,63,79].

Sociocultural risk factors. The consumption of raw or undercooked freshwater fish was the most commonly reported risk factor for infection with *Opisthorchis* spp., and this food preference had a strong cultural basis contributing to frequent reinfection [37,54,58,60,64,65,67,80,99].

Cultural habits that may contribute to transmission include the eating of food with fingers, potentially resulting in hand contamination [58], and traditional food sharing habits that was reported to result in clusters of opisthorchiasis infection in villages [65].

Poor food hygiene and sanitation was reported to result in contamination of other food, surfaces, and utensils [54,106]. Several records reported poor access to toilets and defaecation in fields, resulting in contamination of water bodies and facilitating ongoing transmission in communities [59,60,80,99,100].

Free-roaming domestic cats and dogs were reported to act as reservoir hosts, with dogs often accompanying owners to rice fields, contributing to maintenance of transmission cycles in communities [105].

Lower levels of education was associated with higher risk of infection [64,66].

Diagnostics. Similar to the other FBTs reported, diagnosis of opisthorchiasis was primarily based on the detection of eggs from faecal samples with microscopy, as this was a low cost and non-invasive method [54,56,59,67,99]. However, this had limited diagnostic sensitivity and specificity, required skill parasitologists for diagnosis, and it was difficult to differentiate opisthorchiid eggs from other small intestinal flukes [54,63,67,80,99,103,105,106]. Additional smears from multiple stool samples were reported to improve sensitivity [99,100,103,105,106].

Diagnosis using a monoclonal antibody-based enzyme-linked immunosorbent assay for measuring OV excretory-secretory antigens in urine was discussed as a potential diagnostic method, as this was non-invasive and could be useful for mobile screening [67].

One record reported the use of an *O. viverrini* verbal screening test, which included questions around the consumption of raw fish [56]. This method was a simple, low-cost, and rapid screening test that may be of use for identifying at risk populations and screening for large-scale prevention and control [56].

Treatment and preventive measures. Treatment consisted of praziquantel, using a single dose of 40 mg/kg [62,64,65,67,75,80,99,102,117,127], with 1 record reporting 25 mg/kg for 3 doses over a single day [70]. Limited awareness about the severity of opisthorchiasis, community complacency driven by the belief that symptoms were mild and an effective treatment was available, and deeply rooted cultural food habits contributed to frequent reinfection in endemic communities [59,62,67,71,73,74,100,106]. It was also noted that repeated cycles of reinfection and treatment with praziquantel could result in drug resistance and potentially increase the risk of cholangiocarcinoma [77].

Mass drug administration combined with culturally appropriate health education around food consumption attitudes, personal hygiene, and sanitation was noted to be important in reducing transmission and reinfection [58,59,62,99].

Several records noted that there were high levels of infection in reservoir species, including free-roaming cats and dogs, which were contributing to community transmission [64]. Recommendations for opisthorchiasis control included the treatment of reservoir species in an integrated approach to break the epidemiological cycle and improve the effectiveness of interventions [62,64,102,105].

Access to sanitation was reported to be protective, as this reduced environmental contamination and infection intensity; however, further awareness building around sanitation was required as often latrines were not used even if these were available [100].

Discussion

There was a mismatch between the data that are reported to WHO and that which are reported in the literature reviewed, with the data captured in this review representing 25 countries despite 93 countries reporting FBT “presence only” data to WHO between 2010 and 2019. Similarly, although 26 countries reported ≥ 2 FBTs to WHO, only 3 countries in this review conducted studies for multiple FBTs (*Fasciola*, *Paragonimus*, *Clonorchis* in China; *Paragonimus* and *Clonorchis* in Republic of Korea; all FBTs in Viet Nam). This mismatch of data highlights the limited availability of reliable data in some regions and challenges in reporting FBTs and the need for more research in many geographical areas. Additional countries were identified during the screening process of records that reported clinical cases of FBTs, without reporting prevalence, further enforcing the need to instil surveillance and mapping where cases are reported.

Although the geographical distribution of each FBT discussed in this review reflects what has previously been reported, there were differences in reported prevalences. The highest FBT study prevalence was reported for *O. viverrini* in Lao People’s Democratic Republic. This is

consistent with the reported high prevalence of 37.02% by Furst and colleagues; however, the records in our review identified study prevalences as high as 88.7% in some areas [6,100]. While this could be due to increasing cases or improved surveillance resulting in increased case detection, the prevalences depicted in this review are based on very limited data and are unlikely to reflect true national prevalence. The highest fascioliasis study prevalence identified in this review was in the Region of the Americas, consistent with previous literature [132]. A high study prevalence was also identified in the United Republic of Tanzania, despite no reporting of FBTs to WHO in this time period [18]. The highest recorded study prevalence of paragonimiasis was also reported in the African Region, in Cameroon [18]. The available data on FBTs in Africa is limited, and the reports of high prevalences for both fascioliasis and paragonimiasis support an urgent need for research to better understand the epidemiology and burden of FBTs in this region.

Several limitations exist with our method of reporting prevalence. Although no language restrictions were set, all search terms were in English, potentially resulting in the exclusion of records from areas that have a high burden of FBTs, including Latin America and the Russian Federation. This bias is most notable with the absence of records for *F. Hepatica* in the Americas, despite the well-documented hyperendemic areas in this region [132]. Due to the limited availability of national prevalence data, we instead extracted the study prevalence from each record, using the mean study prevalence when multiple prevalences were recorded. However, as study size and methodology varied greatly between records, and there was a wide range of study prevalences reported even between geographically close areas with similar risk factors, this data cannot be used to reliably estimate national prevalence.

Nevertheless, the data gaps recognised here demonstrate the need for continued improvement of mapping and surveillance to confirm focal points of disease that should be targeted for public health interventions. While disease burden can be estimated within assumptions and available data [6], the limited knowledge available on FBTs in many areas highlights the need for more research and the promotion of successful frameworks, guidelines, and control programmes for surveillance and reporting of FBTs. For example, initiatives in highly endemic fascioliasis areas of Bolivia [133–139], Peru [22,140], Argentina [141], Egypt [142], Pakistan [143] and Vietnam [144], and highly endemic opisthorchiasis areas of Thailand [145–147] have included experimental studies and field surveys to assess transmission and infection sources, field evaluation of diagnostic tools, passive and active surveillance, mass chemotherapy, and the promotion of health awareness among a wide variety of stakeholders. Comprehensive education programmes aimed at community leaders and schoolchildren have been implemented in China [148] and the Republic of Korea [116] to complement mass screening and MDA for clonorchiasis, and reduce transmission and infection risk, and studies in the Philippines have explored the integration of paragonimiasis surveillance and control with tuberculosis control to improve finding and treatment of cases [97,149,150]. These efforts have helped to improve surveillance and case detection [22,135], map FBT endemic areas [140], understand and reduce transmission and infection risk [138,139,147], lower FBT prevalence in humans and intermediate hosts [116,147], and helped to prioritise where resources should be most effectively used in control programmes [135,136,139]. Such initiatives could help inform future studies in other endemic areas to identify within country variation of the endemic areas and delineate treatment strategies according to the level of geographical co-endemicity.

Several themes were highlighted from the qualitative data extracted from the literature; however; as these data reflect what each record discussed and were not necessarily supported by strong epidemiological data, the trends identified in this review may not align with the true epidemiological situation in communities. In addition to this, as our search strategy focused on human prevalence studies, prevalence studies of other species were not captured, and the

extracted qualitative data does not take into account the evidence provided in many valuable epidemiological or transmission risk studies available. This is a significant limitation in our review of qualitative data, and a more comprehensive review focusing on such qualitative data, and prevalence in intermediate and reservoir hosts should be considered to capture these aspects. The inclusion of robust epidemiological data in future prevalence studies would also be valuable to support ongoing surveillance and targeted prevention and treatment programmes.

Despite the epidemiological differences between FBTs, there was significant overlap in the geographical and sociocultural factors that promoted infection and sustained transmission for each FBT. Rural and agricultural communities with appropriate aquatic environments provided a suitable ecological niche for intermediate snail hosts, and secondary fish and crustacean hosts, and also promoted activities that increased the risk of exposure to FBTs (poor sanitation, high-risk food consumption, contact with livestock). Identifying communities that meet these risk criteria can help predict distribution at non-surveyed locations, inform disease surveillance, and help target control programmes against multiple FBTs [99]. However, the interplay of these factors is complex, and endemic and non-endemic villages often coexist within close proximity despite meeting the same risk criteria [80]. The wide range of study prevalences identified in this review, even between geographically close areas with similar risk factors, highlights the need for local health systems to be strengthened and engaged in order for FBTs to be prioritised and dealt with by local health authorities [97,104]. Improved screening is needed to confirm these focal areas of endemicity and co-endemicity. As recognised by previous studies, this review also highlighted the logistical and diagnostic challenges of available methods and the need for an efficient and accurate diagnostic methods to improve surveillance [18]. Although MDA and improved community awareness were frequently discussed as preventive factors for FBT infection, this review highlighted that even in communities that received regular treatment with anthelmintics and were aware of the risks of infection, there continued to be ongoing community transmission and reinfection [80,128,131]. Communities with strong cultural dietary habits and poor sanitation will continue to have a high risk of exposure to infection [17,58]. High infection rates in untreated domestic reservoir species will contribute to ongoing transmission, as well as resulting in veterinary and economic impacts in communities that are dependent on agriculture for their livelihoods [2,17,25,64,105].

One Health approaches are needed to reduce environmental contamination, improve access to clean water and adequate sanitation, and address the role of reservoir hosts in FBT transmission [81,104,105]. One Health is defined as an “integrated, unifying approach that aims to sustainably balance and optimise the health of people, animals, and ecosystems,” and is core to the NTD road map [151,152]. However, while the concept of One Health is becoming more familiar, there is still significant work to be done to promote a consistent understanding across sectors and finding practical ways to operationalise One Health in disease prevention and control programmes. One Health approaches need to address more than the zoonotic pathway of disease transmission, but should also promote coordinated resource allocation and planning, and explore opportunities for treatment implementation to be integrated with that of other diseases and protocols. Implementing such approaches, at both local and national levels, are integral to effectively and sustainably reducing the burden of FBTs. These approaches should further be integrated into Universal Health Care programmes to ensure equitable implementation of the road map for NTD control by 2030 [153]. Examples of successfully implemented One Health approaches have been demonstrated in several areas, including *Fascioliasis* endemic areas of Northern Bolivian Altiplano and *Opisthorchiasis* endemic areas of Khon Kaen Province in Thailand, and such examples can provide a framework that can help build similar One Health programmes in other endemic areas.

Conclusions

FBTs are targeted for control as part of the WHO NTD road map 2021–2030, with mapping and surveillance and capacity, awareness, and advocacy building identified as critical actions required to reach the 2030 targets. FBTs, like other NTDs, impact the most impoverished populations and lack the surveillance systems and tools to adequately ascertain their true burden. The continued high burden of FBTs identified in this review, and the mismatch between the data that are reported to WHO and that which are reported in the literature, demonstrates the need for programmes to improve mapping and surveillance in order to target public health interventions. Integrated One Health approaches across environmental, animal, and human health sectors are needed to meet the 2030 goals.

Supporting information

S1 Supplementary material. Table A in S1 Supplementary material. Box A in S1 Supplementary material: Search terms.
(DOCX)

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References

1. Foodborne trematodiases (Chapter 4.7). In: Investing to overcome the global impact of neglected tropical diseases: third WHO report on neglected tropical diseases. Geneva: World Health Organization; 2015. p. 105–9.
2. Keiser J, Utziner J. Food-borne trematodiases. *Clin Microbiol Rev*. 2009; 22(3):466–483. <https://doi.org/10.1128/CMR.00012-09> PMID: 19597009
3. Bangert M, Molyneux D, Lindsay S, Fitzpatrick C, Engels D. The cross-cutting contribution of the end of neglected tropical diseases to the sustainable development goals. *Infect Dis Poverty*. 2017; 6(73). <https://doi.org/10.1186/s40249-017-0288-0> PMID: 28372566
4. Mas-Coma S, Bargues M, Valero M. Human fascioliasis infection sources, their diversity, incidence factors, analytical methods and prevention measures. *Parasitology*. 2018; 145(13):1665–1699. <https://doi.org/10.1017/S0031182018000914> PMID: 29991363

5. Multicriteria-based ranking for risk management of food-borne parasites: report of a joint FAO/WHO expert meeting, 3–7 September 2012, FAO headquarters, Rome, Italy. WHO and FAO. 2014. <https://apps.who.int/iris/handle/10665/112672>. Accessed 1 May 2021.
6. Furst T, Keiser J, Utzinger J. Global burden of human food-borne trematodiasis: a systematic review and meta-analysis. *Lancet Infect Dis.* 2012; 12(3):210–221. [https://doi.org/10.1016/S1473-3099\(11\)70294-8](https://doi.org/10.1016/S1473-3099(11)70294-8) PMID: 22108757
7. Mas-Coma S, Agramunt V, Valero M. Chapter 2: Neurological and Ocular Fascioliasis in Humans. *Adv Parasitol.* 2014. <https://doi.org/10.1016/B978-0-12-800099-1.00002-8>
8. Bychkov V, Kalyanova L, Khadieva E, Lazarev S, Lukmanov I, Shidin V, et al. Dynamics of the *O. felineus* infestation intensity and egg production in carcinogenesis and partial hepatectomy in the setting of superinvasive opisthorchiasis. *Anal Cell Pathol.* 2019. <https://doi.org/10.1155/2019/8079368> PMID: 31428553
9. WHO estimates of the global burden of foodborne diseases: foodborne diseases burden epidemiology reference group 2007–2015. Geneva: World Health Organization; 2015. <https://apps.who.int/iris/handle/10665/199350>. Accessed 1 May 2021.
10. Mitra A, Mawson A. Neglected tropical diseases: epidemiology and global burden. *Trop Med Infect Dis.* 2017; 2:36. <https://doi.org/10.3390/tropicalmed2030036> PMID: 30270893
11. Keiser J, Utzinger J. Emerging foodborne trematodiasis. *Emerg Infect Dis.* 2005; 11:1507–1514. <https://doi.org/10.3201/eid1110.050614> PMID: 16318688
12. King C, Bertino A. Asymmetries of poverty: why global burden of disease valuations underestimate the burden of neglected tropical diseases. *PLoS Negl Trop Dis.* 2008; 2:e209. <https://doi.org/10.1371/journal.pntd.0000209> PMID: 18365036
13. Hotez P, Molyneux D, Fenwick A, Ottesen E, Sachs S, Sachs J. Incorporating a rapid-impact package for neglected tropical diseases with programs for HIV/AIDS, tuberculosis, and malaria. *PLoS Med.* 2007; 4(9):1553. <https://doi.org/10.1371/journal.pmed.0040277>
14. Knopp S, Mgeni AF, Khamis I, Steinmann P, Stothard JR, Rollinson D, et al. Diagnosis of soil-transmitted helminths in the era of preventive chemotherapy: effect of multiple stool sampling and use of different diagnostic techniques. *PLoS Negl Trop Dis.* 2008; 2(11):e331. <https://doi.org/10.1371/journal.pntd.0000331> PMID: 18982057
15. Fitzpatrick C, Fleming F, Madin-Warburton M, Schneider T, Meheus F, Asiedu K, et al. Benchmarking the cost per person of mass treatment for selected neglected tropical diseases: an approach based on literature review and meta-regression with Web-based software application. *PLoS Negl Trop Dis.* 2016; 10(12):e0005037. <https://doi.org/10.1371/journal.pntd.0005037> PMID: 27918573
16. WHO Global Health Observatory data repository. <https://www.who.int/data/gho/data/themes/neglected-tropical-diseases>. Accessed 28 June 2021.
17. Fentie T, Erqou S, Gedefaw M, Desta A. Epidemiology of human fascioliasis and intestinal parasites among schoolchildren in Lake Tana Basin, northwest Ethiopia. *Trans R Soc Trop Med Hyg.* 2013; 107:480–486. <https://doi.org/10.1093/trstmh/trt056> PMID: 23843557
18. Lukambagire A, Mchaile D, Nyindo M. Diagnosis of human fascioliasis in Arusha region, northern Tanzania by microscopy and clinical manifestations in patients. *BMC Infect Dis.* 2015; 15:578. <https://doi.org/10.1186/s12879-015-1326-9> PMID: 26695775
19. Nkouawa A, Sako Y, Itoh S, Kouojip-Mabou A, Nganou C, Sajio Y, et al. Serological studies of neurologic helminthic infections in rural areas of southwest Cameroon: toxocariasis, cysticercosis and paragonimiasis. *PLoS Negl Trop Dis.* 2010; 4(7):e372. <https://doi.org/10.1371/journal.pntd.0000732> PMID: 20625553
20. Carnevale S, Cabrera M, Cucher M, di Risio C, Malandrini J, Kamenetzky L, et al. Direct, immunological and molecular techniques for a fasciolosis survey in a rural area of San Luis, Argentina. *J Parasit Dis.* 2013; 37(2):251–259. <https://doi.org/10.1007/s12639-012-0175-3> PMID: 24431579
21. Maciel M, Lima W, Moreira de Almeida F, Coelho L, Araujo G, Lima M, et al. Cross-sectional serological survey of human fascioliasis in Canutama municipality in western Amazon, Brazil. *J Parasit Research Dis.* 2018. <https://doi.org/10.1155/2018/6823638> PMID: 29593895
22. Valero M, Periago M, Perez-Crespo I, Angles R, Villegas F, Aguirre C, et al. Field Evaluation of Coproantigen Detection Test for Fascioliasis Diagnosis and Surveillance in Human Hyperendemic Areas of Andean Countries. *PLoS Negl Trop Dis.* 2012; 6(9):e1812. <https://doi.org/10.1371/journal.pntd.0001812> PMID: 23029575
23. Villegas F, Angles R, Barrientos R, Barrios G, Valero MA, Hamed K, et al. Administration of Triclabendazole Is Safe and Effective in Controlling Fascioliasis in an Endemic Community of the Bolivian Altiplano. *PLoS Negl Trop Dis.* 2012; 6(8):e1720. <https://doi.org/10.1371/journal.pntd.0001720> PMID: 22880138

24. Pino Santos A, Nunez Fernandez F, Martinez Sanchez R, Domenech Cañete I, Rodriguez M, Jerez Puebla L, et al. Prevalence and risk factors for intestinal parasitic infections in a rural community in “Consolación del Sur” municipality, Cuba. *West Indian Med J.* 2014; 63(4):333. <https://doi.org/10.7727/wimj.2013.103> PMID: 25429477
25. Agnamey P, Fortes-Lopes E, Racourt C, Boncy J, Totet A. Cross-sectional serological survey of human fascioliasis in Haiti. *J Parasit Research.* 2012. <https://doi.org/10.1155/2012/751951> PMID: 21876782
26. Zumaquero-Rios J, Sarracent-Perez J, Rojas-Garcia R, Rojas-Rivero L, Martinez-Tovilla Y, Valero M, et al. Fascioliasis and intestinal parasitoses affecting schoolchildren in Atlixco, Puebla State, Mexico: epidemiology and treatment with nitazoxanide. *PLoS Negl Trop Dis.* 2013; 7(11):e2553. <https://doi.org/10.1371/journal.pntd.0002553> PMID: 24278492
27. González L, Guillermo Esteban J, Dolores Bargues M, Valero M, Ortiz P, Náquira C, et al. Hyperendemic human fascioliasis in Andean valleys: an altitudinal transect analysis in children of Cajamarca province. *Peru Acta Trop.* 2011; 120:119–129.
28. Lopez M, White A, Cabada M. Burden of *Fasciola hepatica* infection among children from Paucarambo in Cusco, Peru. *Am J Trop Med Hyg.* 2012; 86(3):481–485.
29. Cabada M, Goodrich M, Graham B. Fascioliasis and eosinophilia in the highlands of Cuzco, Peru and their association with water and socioeconomic factors. *Am J Trop Med Hyg.* 2014; 91(5):989–993. <https://doi.org/10.4269/ajtmh.14-0169> PMID: 25200257
30. Cabada M, Lopez M, Arque E, White A. Prevalence of soil-transmitted helminths after mass albendazole administration in an indigenous community of the Manu jungle in Peru. *Pathog Glob Health.* 2014; 108(4):200–205. <https://doi.org/10.1179/204773214Y.0000000142> PMID: 24934795
31. Cabada M, Morales M, Webb C, Yang L, Bravenec C, Lopez M, et al. Socioeconomic factors associated with *Fasciola hepatica* infection among children from 26 communities of the Cusco Region of Peru. *Am J Trop Med Hyg.* 2018; 99(5):1180–1185. <https://doi.org/10.4269/ajtmh.18-0372> PMID: 30226136
32. Rodriguez-Ulloa C, Rivera-Jacinto M, Valle-Mendoza J, Cerna C, Hoban C, Chilón S, et al. Risk factors for human fascioliasis in schoolchildren in Baños del Inca, Cajamarca, Peru. *Trans R Soc Trop Med Hyg.* 2018; 112:216–222. <https://doi.org/10.1093/trstmh/try049> PMID: 29860359
33. Steinmann P, Usupaliyeva J, Imanalieva C, Minbaeva G, Stefiuk K, Jeandron A, et al. Rapid appraisal of human intestinal helminth infections among schoolchildren in Osh oblast, Kyrgyzstan. *Acta Trop.* 2010; 116:178–174. <https://doi.org/10.1016/j.actatropica.2010.06.008> PMID: 20615381
34. Cengiz Z, Yilmaz H, Dülger A, Akdeniz H, Karahocagil M, Çiçek M. Seroprevalence of human fascioliasis in Van province, Turkey. *Turk J Gastroenterol.* 2015; 26:259–262. <https://doi.org/10.5152/tjg.2015.8001> PMID: 26006203
35. Cengiz Z, Yilmaz H, Beyhan Y, Çiçek M. A comprehensive retrospective study: intestinal parasites in human in Van Province. *Turkiye Parazitol Derg.* 2019; 43(2):70–73. <https://doi.org/10.4274/tpd.galenos.2019.5997> PMID: 31204458
36. Sultanov A, Abdybekova A, Abdibaeva A, Shapiyeva Z, Yeshmuratov T, Torgerson PR. Epidemiology of fishborne trematodiasis in Kazakhstan. *Acta Trop.* 2014; 138:60–66. <https://doi.org/10.1016/j.actatropica.2014.04.030> PMID: 24943190
37. Fedorova O, Kovshirina Y, Kovshirina A, Fedotova M, Deev I, Petrovskiy F, et al. *Opisthorchis felineus* infection and cholangiocarcinoma in the Russian Federation: a review of medical statistics. *Parasitol Int.* 2017; 66:365–371.
38. Sarkari B, Ghobakhloo N, Moshfea A, Eilami O. Seroprevalence of human fasciolosis in a new-emerging focus of fasciolosis in Yasuj District, southwest of Iran. *Iran J Parasitol.* 2012; 7(2):15–20. PMID: 23109941
39. Abdi J, Naserifar R, Nejad M, Mansouri V. New features of fascioliasis in human and animal infections in Ilam province, western Iran. *Gastroenterol Hepatol Bed Bench.* 2013; 6(3):152–155. PMID: 24834263
40. Asadian S, Mohebali M, Mahmoudi M, Kia E, Heidari Z, Asgari M, et al. Seroprevalence of human fascioliasis in Meshkin-Shahr District, Ardabil Province, northwestern Iran in 2012. *Iran J Parasitol.* 2013; 8(4):516–521. PMID: 25516731
41. Saberinasab M, Mohebali M, Molawi G, Kia E, Aryaeipour M, Rokni M. Seroprevalence of human fascioliasis using indirect ELISA in Isfahan District, central Iran in 2013. *Iran J Parasitol.* 2014; 9(4):461–465. PMID: 25759726
42. Aryaeipour M, Kia E, Heidari Z, Sayyad Talaie Z, Rokni M. Serological study of human fasciolosis in patients referring to the School of Public Health, Tehran University of Medical Sciences, Tehran, Iran during 2008–2014. *Iran J Parasitol.* 2015; 10(4):517–522. PMID: 26811716

43. Ashrafi K, Saadat F, O'Neill S, Rahmati B, Tahmasbi H, Dalton J, et al. The Endemicity of human fascioliasis in Guilan Province, northern Iran: the baseline for implementation of control strategies. *Iran J Public Health*. 2015; 44(4):501–511. PMID: [26056669](#)
44. Hosseini G, Sarkari B, Moshfe A, Motazedian M, Abdolahi KS. Epidemiology of human fascioliasis and intestinal helminthes in rural areas of Boyer-Ahmad township, southwest Iran: a population based study. *Iran J Public Health*. 2015; 44(11):1520–1525. PMID: [26744710](#)
45. Kheirandish F, Kayedi M, Ezatpour B, Anbari K, Karimi Rouzbahani H, Chegeni Sharafi A, et al. Sero-prevalence of human fascioliasis in Pirabad, Lorestan Province, western Iran. *Iran J Parasitol*. 2016; 11(1):24–29. PMID: [27095965](#)
46. Manouchehri Naeini K, Mohammad Nasiri F, Rokni M, Kheiri S. Seroprevalence of human fascioliasis in Chaharmahal and Bakhtiari Province, southwestern Iran. *Iran J Public Health*. 2016; 45(6):774–780.
47. Heydarian P, Ashrafi K, Mohebali M, Kia E, Aryaeipour M, Chegeni Sharafi A, et al. Seroprevalence of human fascioliasis in Lorestan Province, western Iran, in 2015–16. *Iran J Parasitol*. 2017; 12(3):389–397. PMID: [28979349](#)
48. Bozorgomid A, Nazari N, Kia E, Mohebali M, Hajaran H, Hydarian P, et al. Epidemiology of fascioliasis in Kermanshah Province, western Iran. *Iran J Public Health*. 2018; 47(7):967–972. PMID: [30181994](#)
49. Zoghi S, Emami M, Shahriarirad S, Vahedi R, Cheraghi M, Zamiri B, et al. Human fascioliasis in nomads: a population-based serosurvey in southwest Iran. *Infez Med*. 2019; 1:68–72. PMID: [30882381](#)
50. Qureshi A, Tanveer A, Mas-Coma S. Epidemiological analysis of human fascioliasis in northeastern Punjab. *Pakistan Acta Trop*. 2016; 156:157–164.
51. Quresh A, Zeb A, Mansoor A, Hayat A, Mas-Coma S. *Fasciola hepatica* infection in children actively detected in a survey in rural areas of Mardan district, Khyber Pakhtunkhawa province, northern Pakistan. *Parasitol Int*. 2019; 69:39–46.
52. Devi K, Narain K, Mahanta J, Deori R, Lego K, Goswami D, et al. Active detection of tuberculosis and paragonimiasis in the remote areas in North-Eastern India using cough as a simple indicator. *Pathog Glob Health*. 2013; 107(3):153–156. <https://doi.org/10.1179/204773213Y.0000000086> PMID: [23683370](#)
53. Narain K, Devia K, Bhattacharya S, Negmu K, Rajguru S, Mahanta J. Declining prevalence of pulmonary paragonimiasis following treatment and community education in a remote tribal population of Arunachal Pradesh, India. *Indian J Med Res*. 2015; 141(5):648–652. PMID: [26139784](#)
54. Aung W, Htoo T, Tin H, Thinn K, Sanpool O, Jongthawin J, et al. First report and molecular identification of *Opisthorchis viverrine* infection in human communities from Lower Myanmar. *PLoS ONE*. 2017; 12(5):e0177130. <https://doi.org/10.1371/journal.pone.0177130> PMID: [28472153](#)
55. Sohn W, Jung B, Hong S, Lee K, Park J, Kim H, et al. Low-grade endemicity of opisthorchiasis, Yangon, Myanmar. *Emerg Infect Dis*. 2019; 25(7):1435–1437. <https://doi.org/10.3201/eid2507.190495> PMID: [31211941](#)
56. Kaewpitoon N, Kaewpitoon S, Meererkson T, Chan-Aran S, Sangwalee W, Norkaew J, et al. Detection of *Opisthorchis viverrine* infection among the ASEAN population in Thailand using a verbal screening test and fecal concentrator kit. *Iran J Parasitol*. 2018; 13(2):258–266.
57. Mairiang E, Laha T, Bethony J, Thinkhamrop B, Kaewkes S, Sithithaworn P, et al. Ultrasonography assessment of hepatobiliary abnormalities in 3,359 subjects with *Opisthorchis viverrini* infection in endemic areas of Thailand. *Parasitol Int*. 2012; 61(1):208–211. <https://doi.org/10.1016/j.parint.2011.07.009> PMID: [21771664](#)
58. Boonjaraspinyo S, Boonmars T, Kaewsamut B, Ekobol N, Laummaunwai P, Aukkanimart R, et al. A cross-sectional study on intestinal parasitic infections in rural communities, northeast Thailand. *Korean J Parasitol*. 2013; 51(6):727–34. <https://doi.org/10.3347/kjp.2013.51.6.727> PMID: [24516280](#)
59. Suwannahitatorn P, Klomjitt S, Naaglor T, Taamasri P, Rangsin R, Leelayoova S, et al. A follow-up study of *Opisthorchis viverrini* infection after the implementation of control program in a rural community, central Thailand. *Parasit Vectors*. 2013; 6:188.
60. Wang Y, Feng C, Sithithaworn P. Environmental determinants of *Opisthorchis viverrini* prevalence in northeast Thailand. *Geospat Health*. 2013; 8(1):111–123.
61. Thaewnongiew K, Singthong S, Kutchamart S, Tangsawad S, Promthet S, Sailugkum S, et al. Prevalence and risk factors for *Opisthorchis viverrini* infections in Upper Northeast Thailand. *Asian Pac J Cancer Prev*. 2014; 15(16):6609–6612. <https://doi.org/10.7314/APJCP.2014.15.16.6609> PMID: [25169496](#)
62. Wongsaroj T, Nithikathkul C, Rojikitkul W, Nakai W, Royal L, Rammasut P. National survey of helminthiasis in Thailand. *Asian Biomed*. 2014; 8(6):779–783.

63. Buathong S, Leelayoova S, Mungthin M. Molecular discrimination of *Opisthorchis*-like eggs from residents in a rural community of central Thailand. PLoS Negl Trop Dis. 2017; 11(11):e0006030. <https://doi.org/10.1371/journal.pntd.0006030> PMID: 29095828
64. Prakobwong S, Gunnula W, Chaipibool S, Nimala B, Sangthopo J, Siriveththumrong N, et al. Epidemiology of *Opisthorchis viverrini* in an endemic area of Thailand, an integrative approach. Helminthologia. 2017; 54(4):298–306. <https://doi.org/10.1515/helm-2017-0036>
65. Saenna P, Hurst C, Echaubard P, Wilcox B, Sripa B. Fish sharing as a risk factor for *Opisthorchis viverrini* infection: evidence from two villages in north-eastern Thailand. Infect Dis Poverty. 2017; 6:66. <https://doi.org/10.1186/s40249-017-0281-7> PMID: 28372560
66. Laoraksawong P, Sanpool O, Rodpai R, Thanchomnang T, Kanarkard W, Maleewong W, et al. Current high prevalences of *Strongyloides stercoralis* and *Opisthorchis viverrini* infections in rural communities in northeast Thailand and associated risk factors. BMC Public Health. 2018; 18:940. <https://doi.org/10.1186/s12889-018-5871-1> PMID: 30064407
67. Thinkhamrop K, Khuntikeo N, Sithihaworn P, Thinkhamrop W, Wangdi K, Kelly M, et al. Repeated praziquantel treatment and *Opisthorchis viverrini* infection: a population-based cross-sectional study in northeast Thailand. Infect Dis Poverty. 2019; 8:18. <https://doi.org/10.1186/s40249-019-0529-5> PMID: 30890188
68. Kaewpitoon S, Rujirakul R, Kaewpitoon N. Prevalence of *Opisthorchis viverrini* Infection in Nakhon Ratchasima Province, Northeast Thailand. Asian Pac J Cancer Prev. 2012; 13:5245–5249. <https://doi.org/10.7314/apjcp.2012.13.10.5245> PMID: 23244144
69. Saengsawang PS, Bradshaw P. Prevalence of OV Infection in Yasothon Province, Northeast Thailand. Asian Pac J Cancer Prev. 2012; 13:3399–3402. <https://doi.org/10.7314/apjcp.2012.13.7.3399> PMID: 22994767
70. Songserm N, Promthet S, Wiangnon S, Sithithaworn P. Prevalence and Co-infection of Intestinal Parasites among Thai Rural Residents at High-risk of Developing Cholangiocarcinoma: A Cross-sectional Study in a Prospective Cohort Study. Asian Pac J Cancer Prev. 2012; 13:6175–6179. <https://doi.org/10.7314/apjcp.2012.13.12.6175> PMID: 23464426
71. Saengsawang P, Promthet S, Bradshaw P. Infection with *Opisthorchis viverrini* and Use of Praziquantel among a Working-age Population in Northeast Thailand. Asian Pac J Cancer Prev. 2013; 14:2963–2966. <https://doi.org/10.7314/apjcp.2013.14.5.2963> PMID: 23803062
72. Chaiputcha K, Promthet S, Bradshaw P. Prevalence and Risk Factors for Infection by *Opisthorchis viverrini* in an Urban Area of Mahasarakham Province, Northeast Thailand. Asian Pac J Cancer Prev. 2015; 16:4173–4176. <https://doi.org/10.7314/apjcp.2015.16.10.4173> PMID: 26028068
73. Chudthaisong N, Promthet S, Bradshaw P. Risk factors for *Opisthorchis viverrini* Infection in Nong Khai Province, Thailand. Asian Pac J Cancer Prev. 2015; 16:4593–4596. <https://doi.org/10.7314/apjcp.2015.16.11.4593> PMID: 26107209
74. Yeoh K, Promthet S, Sithithaworn P, Kamsa-Ard S, Parkin DM. Re-examination of *Opisthorchis viverrini* Infection in Northeast Thailand. Asian Pac J Cancer Prev. 2015; 16:3413–3418. <https://doi.org/10.7314/apjcp.2015.16.8.3413> PMID: 25921154
75. Sohn W, Yong T, Eom K, Pyo KH, Lee MY, Lim H, et al. Prevalence of *Opisthorchis viverrini* infection in humans and fish in Kratie Province, Cambodia. Acta Tropica. 2012; 124:215–220. <https://doi.org/10.1016/j.actatropica.2012.08.011> PMID: 22935318
76. Yong T, Chai J, Sohn W, Eom K, Jeoung H, Hoang E, et al. Prevalence of Intestinal Helminths among Inhabitants of Cambodia (2006–2011). Korean J Parasitol. 2014; 52(6):661–666. <https://doi.org/10.3347/kjp.2014.52.6.661> PMID: 25548418
77. Bless P, Schär F, Khiu V, Kramme S, Muth S, Marti H, et al. High prevalence of large trematode eggs in schoolchildren in Cambodia. Acta Trop. 2015; 141:295–302. <https://doi.org/10.1016/j.actatropica.2014.09.007> PMID: 25250828
78. Sohn W, Shin E, Yong T, Eom K, Jeong H, Sinuon M, et al. Adult *Opisthorchis viverrini* flukes in humans, Takeo, Cambodia. Emerg Infect Dis. 2011; 17(7):1302–1304. <https://doi.org/10.3201/eid1707.102071> PMID: 21762595
79. Yong T, Shin E, Chai J, Sohn W, Eom K, Lee D, et al. High prevalence of *Opisthorchis viverrini* infection in a Riparian population in Takeo Province, Cambodia. Korean J Parasitol. 2012; 50(2):173–176. <https://doi.org/10.3347/kjp.2012.50.2.173> PMID: 22711932
80. Miyamoto K, Kirinoki M, Matsuda H, Hayashi N, Chigusa Y, Sinuon M, et al. Field survey focused on *Opisthorchis viverrini* infection in five provinces of Cambodia. Parasitol Int. 2014; 63:366–373.
81. Chen J, Chen M, Ai L, Xu X, Jiao J, Zhu T, et al. An outbreak of human *Fascioliasis gigantica* in Southwest China. PLoS ONE. 2013; 8(8):e71520. <https://doi.org/10.1371/journal.pone.0071520> PMID: 23951181

82. Lee M, Shin H, Chung B, Lee S. Intestinal parasite infections among inhabitants in Yanbian prefecture, Jilin Province, China. *Korean J Parasitol*. 2017; 55(5):579–582. <https://doi.org/10.3347/kjp.2017.55.579> PMID: 29103276
83. Zhang X, Wang Y, Wang G, Chen W, He X, Niu H, et al. Distribution and clinical features of *Paragonimiasis skrjabini* in Three Gorges Reservoir Region. *Parasitol Int*. 2012; 61:645–649. <https://doi.org/10.1016/j.parint.2012.06.007> PMID: 22814214
84. Choi M, Park S, Li Z, Ji Z, Yu G, Feng Z, et al. Effect of control strategies on prevalence, incidence and re-infection of clonorchiasis in endemic areas of China. *PLoS Negl Trop Dis*. 2010; 4(2):e601. <https://doi.org/10.1371/journal.pntd.0000601> PMID: 20169061
85. Han S, Zhang X, Wen J, Li Y, Shu J, Ling H, et al. A combination of the Kato-Katz methods and ELISA to improve the diagnosis of clonorchiasis in an endemic area, China. *PLoS ONE*. 2012; 7(10):e46977. <https://doi.org/10.1371/journal.pone.0046977> PMID: 23056547
86. Jeon H, Lee D, Park H, Min D, Rim H, Zhang H, et al. Human infections with liver and minute intestinal flukes in Guangxi, China: analysis by DNA sequencing, ultrasonography, and immunoaffinity chromatography. *Korean J Parasitol*. 2012; 50(4):391–394. <https://doi.org/10.3347/kjp.2012.50.4.391> PMID: 23230343
87. Han S, Zhang X, Chen R, Wen J, Li Y, Shu J, et al. Trends in prevalence of clonorchiasis among patients in Heilongjiang Province, northeast China (2009–2012): implications for monitoring and control. *PLoS ONE*. 2013; 8(11):e80173. <https://doi.org/10.1371/journal.pone.0080173> PMID: 24260354
88. Qian M, Chen Y, Fan Y, Tan T, Zhu T, Zhou C, et al. Epidemiological profile of *Clonorchis sinensis* infection in one community, Guangdong, People's Republic of China. *Parasit Vectors*. 2013; 6:194.
89. Qian M, Chen Y, Yang Y, Lu M, Jiang Z, Wei K, et al. Increasing prevalence and intensity of foodborne clonorchiasis, Hengxian County, China, 1989–2011. *Emerg Infect Dis*. 2014; 20(11):1872–1875. <https://doi.org/10.3201/eid2011.131309> PMID: 25340976
90. Dai Y, Xu X, Liu J, Jin X, Shen M, Wang X, et al. Prevalence of intestinal helminth infections in Jiangsu Province, eastern China; a cross-sectional survey conducted in 2015. *BMC Infect Dis*. 2019; 19:604. <https://doi.org/10.1186/s12879-019-4264-0> PMID: 31291911
91. Xie W, Deng Y, Chen S, Yang Q. Association between eosinophil count and cholelithiasis among a population with *Clonorchis sinensis* infection in Foshan City, China. *J Helminthol*. 2019; 94:1–8.
92. Qian M, Jiang Z, Ge T, Wang X, Zhou C, Zhu H, et al. Rapid screening of *Clonorchis sinensis* infection: performance of a method based on raw-freshwater fish-eating practice. *Acta Trop*. 2020; 207:105380. <https://doi.org/10.1016/j.actatropica.2020.105380> PMID: 32007446
93. Lo T, Chang J, Lee H, Kuo H. Risk factors for and prevalence of clonorchiasis in Miaoli County, Taiwan. *Southeast Asian J Trop Med Public Health*. 2013; 44(6):950–958. PMID: 24450231
94. Qian M, Zhuang S, Zhu S, Deng X, Li Z, Zhou X. Improving diagnostic performance of the Kato-Katz method for *Clonorchis sinensis* infection through multiple samples. *Parasit Vectors*. 2019; 12:336. <https://doi.org/10.1186/s13071-019-3594-5> PMID: 31287026
95. Xue-Ming L, Ying-Dan C, Yi O, Hong-Man Z, Rui L, Weil M. Overview of human clonorchiasis sinensis in China. *Southeast Asian J Trop Med Public Health*. 2011; 42(2):248–254. PMID: 21710843
96. Nagayasu E, Yoshida A, Hombu A, Horii Y, Maruyama H. Paragonimiasis in Japan: a twelve-year retrospective case review (2001–2012). *Intern Med*. 2015; 54:179–186. <https://doi.org/10.2169/internalmedicine.54.1733> PMID: 25743009
97. Belizario V, Totanes F, Asuncion C, De Leon W, Jorge M, Ang C, et al. Integrated surveillance of pulmonary tuberculosis and paragonimiasis in Zamboanga del Norte, the Philippines. *Pathog Glob Health*. 2014; 108(2):95–102. <https://doi.org/10.1179/2047773214Y.0000000129> PMID: 24601907
98. Sayasone S, Mak T, Vanmany M, Rasphone O, Vounatsou P, Utzinger J, et al. Helminth and intestinal protozoa infections, multiparasitism and risk factors in Champasack Province, Lao People's Democratic Republic. *PLoS Negl Trop Dis*. 2011; 5(4):e1037. <https://doi.org/10.1371/journal.pntd.0001037> PMID: 21532735
99. Forrer A, Sayasone S, Vounatsou P, Vonghachack Y, Bouakhasith D, Vogt S, et al. Spatial distribution of, and risk factors for, *Opisthorchis viverrini* infection in Southern Lao PDR. *PLoS Negl Trop Dis*. 2012; 6(2):e1481. <https://doi.org/10.1371/journal.pntd.0001481> PMID: 22348157
100. Phongluxa K, Xayaseng V, Vonghachack Y, Akkhavong K, van Eeuwijk P, Odermatt P. Helminth infection in southern Laos: high prevalence and low awareness. *Parasit Vectors*. 2013; 6:328. <https://doi.org/10.1186/1756-3305-6-328> PMID: 24499584
101. Eom K, Yong T, Sohn W, Chai J, Min D, Rim H, et al. Prevalence of helminthic infections among inhabitants of Lao PDR. *Korean J Parasitol*. 2014; 52(1):51–56. <https://doi.org/10.3347/kjp.2014.52.1.51> PMID: 24623882

102. Chai J, Sohn W, Jung B, Yong T, Eom K, Min D, et al. Intestinal helminths recovered from humans in Xieng Khouang Province, Lao PDR with a particular note on *Haplchorhis pumilio* infection. Korean J Parasitol. 2015; 53(4):439–445. <https://doi.org/10.3347/kjp.2015.53.4.439> PMID: 26323842
103. Saiyachak K, Tongtsotsang S, Saenrueang T, Moore M, Promthet S. Prevalence and factors associated with *Opisthorchis viverrini* infection in Khammouane Province, Lao PDR. Asian Pac J Cancer Prev. 2016; 17:1589–1593. <https://doi.org/10.7314/APJCP.2016.17.3.1589> PMID: 27039810
104. Ribas A, Jollivet C, Morand S, Thongmalayvong B, Somphavong S, Siew C, et al. Intestinal parasitic infections and environmental water contamination in a rural village of northern Lao PDR. Korean J Parasitol. 2017; 55(5):523–532. <https://doi.org/10.3347/kjp.2017.55.5.523> PMID: 29103267
105. Vonghachack Y, Odermatt P, Taisayyavong K, Phoumsavath S, Akkhavong K, Sayasone S. Transmission of *Opisthorchis viverrini*, *Schistosoma mekongi* and soil-transmitted helminthes on the Mekong Islands, southern Lao PDR. Infect Dis Poverty. 2017; 6:131. <https://doi.org/10.1186/s40249-017-0343-x> PMID: 28866984
106. Araki H, Ong K, Lorphachan L, Soundala P. Mothers' *Opisthorchis viverrini* infection status and raw fish dish consumption in Lao People's Democratic Republic: determinants of child infection status. Trop Med Health. 2018; 46:29. <https://doi.org/10.1186/s41182-018-0112-y> PMID: 30093819
107. Sayasone S, Utzinger J, Akkhavong K, Odermatt P. Repeated stool sampling and use of multiple techniques enhance the sensitivity of helminth diagnosis: A cross-sectional survey in southern Lao People's Democratic Republic. Acta Trop. 2015; 141:315–321. <https://doi.org/10.1016/j.actatropica.2014.09.004> PMID: 25225157
108. Sato M, Pongvongsa SS, Sanguankiat S, Yoonuan T, Kobayashi J, Boupha B, et al. Patterns of trematod infections of *Opisthorchis viverrini* (Opisthorchiidae) and *Haplchorhis taichui* (Heterophyidae) in human populations from two villages in Savannakhet Province, Lao PDR. J Helminthol. 2015; 89(4):439–445.
109. Lee M, Hong S, Kim H. Seroprevalence of tissue invading parasitic infections diagnosed by ELISA in Korea. J Korean Med Sci. 2010; 25:1272–1276. <https://doi.org/10.3346/jkms.2010.25.9.1272> PMID: 20808668
110. Choi S, Lee S, Song H, Ryu J, Ahn M. Parasitic infections based on 320 clinical samples submitted to Hanyang University, Korea (2004–2011). Korean J Parasitol. 2014; 52(2):215–220. <https://doi.org/10.3347/kjp.2014.52.2.215> PMID: 24850969
111. Jin Y, Kim E, Choi M, Oh M, Hong S. Significance of serology by multi-antigen ELISA for tissue helminthiases in Korea. J Korean Med Sci. 2017; 32:1118–1123. <https://doi.org/10.3346/jkms.2017.32.7.1118> PMID: 28581268
112. Cho S, Cho P, Lee D, Kim T, Kim I, Hwang E, et al. Epidemiological survey on the infection of intestinal flukes in residents of Muan-gun, Jeollanam-do, the Republic of Korea. Korean J Parasitol. 2010; 48(2):133–138. <https://doi.org/10.3347/kjp.2010.48.2.133> PMID: 20585529
113. Kim H, Cheun H, Cheun B, Lee K, Kim T, Lee S, et al. Prevalence of *Clonorchis sinensis* infections along the five major rivers in Republic of Korea, 2007. Public Health Res Perspective. 2010; 1(1):43–49. <https://doi.org/10.1016/j.phrp.2010.12.010> PMID: 24159439
114. Shin H, Oh J, Lim M, Shin A, Kong H, Jung K, et al. Descriptive epidemiology of cholangiocarcinoma and clonorchiasis in Korea. J Korean Med Sci. 2010; 25:1011–1016. <https://doi.org/10.3346/jkms.2010.25.7.1011> PMID: 20592891
115. June K, Cho S, Lee W, Kim C, Park K. Prevalence and risk factors of clonorchiasis among the populations served by primary healthcare posts along five major rivers in South Korea. Osong Public Health Res Perspect. 2013; 4(1):21–26. <https://doi.org/10.1016/j.phrp.2012.12.002> PMID: 24159525
116. Oh J, Lim M, Yun E, Cho H, Park E, Choi M, et al. Control of clonorchiasis in Korea: effectiveness of health education for community leaders and individuals in an endemic area. Trop Med Int Health. 2014; 19(9):1096–1104. <https://doi.org/10.1111/tmi.12338> PMID: 24862476
117. Park D, Na S, Cho S, June K, Cho Y, Lee Y. Prevalence and risk factors of clonorchiasis among residents of riverside areas in Muju-gun, Jeollabuk-do, Korea. Korean J Parasitol. 2014; 52(4):391–397. <https://doi.org/10.3347/kjp.2014.52.4.391> PMID: 25246718
118. Jung S, Ahn M, Oh J, Nam H, Hong S, Yun Y, et al. Infection status of endoparasites in foreigner workers living in Cheonan City, Chungnam Province, Korea. Korean J Parasitol. 2015; 53(2):243–246. <https://doi.org/10.3347/kjp.2015.53.2.243> PMID: 25925187
119. Jeong Y, Shin H, Lee S, Cheun H, Ju J, Kim J, et al. Prevalence of *Clonorchis sinensis* infection among residents along 5 major rivers in the Republic of Korea. Korean J Parasitol. 2016; 54(2):215–219. <https://doi.org/10.3347/kjp.2016.54.2.215> PMID: 27180582
120. Choi S, Lee C, Yang J, Kwak M, Chung G, Kang H, et al. Identifying helminth infections via routine fecal parasitological examinations in Korea. Am J Trop Med Hyg. 2017; 97(3):888–895. <https://doi.org/10.4269/ajtmh.17-0084> PMID: 28749758

121. Jeong J, Lee J, Chung B, Choi Y, Alley A, Kim H. A new method for estimating the prevalence of clonorchiasis in Korea: a proposal to replace arbitrary riverside sampling. *Medicine*. 2017; 96:13. <https://doi.org/10.1097/MD.0000000000006536> PMID: 28353615
122. Shin H, Lee M, Ju J, Jeong B, Park M, Lee K, et al. Epidemiological and clinical parameters features of patients with clonorchiasis in the Geum River Basin, Republic of Korea. *Interdiscip Perspect Infect Dis*. 2017;1–7. <https://doi.org/10.1155/2017/7415301> PMID: 28529523
123. Bahk Y, Park Y, Na B, Sohn W, Hong S, Chai J, et al. Survey on intestinal helminthic infection status of students in two counties, Hadong-gun and Goseong-gun, Korea. *Korean J Parasitol*. 2018; 56(4):335–339. <https://doi.org/10.3347/kjp.2018.56.4.335> PMID: 30196665
124. Nguyen T, Cheong F, Liew J, Lau Y. Seroprevalence of fascioliasis, toxocariasis, strongyloidiasis and cysticercosis in blood samples diagnosed in Medic Medical Center Laboratory, Ho Chi Minh City, Vietnam in 2012. *Parasit Vectors*. 2016; 9:486. <https://doi.org/10.1186/s13071-016-1780-2> PMID: 27595647
125. Quy T, Yeatman H, Flood V, NC C, BV T. Prevalence and risks of fascioliasis among adult cohorts in Binh Dinh and Quang Ngai provinces—central Viet Nam. *Vietnam J Public Health*. 2015; 3(1):46–61.
126. Doanh P, Dung D, Thach D, Horii Y, Shinohara A, Nawa Y. Human paragonimiasis in Viet Nam: epidemiological survey and identification of the responsible species by DNA sequencing of eggs in patients' sputum. *Parasitol Int*. 2011; 60:534–537. <https://doi.org/10.1016/j.parint.2011.09.001> PMID: 21946337
127. Dao T, Bui T, Abatih E, Gabriël S, Nguyen T, Huynh Q, et al. *Opisthorchis viverrini* infections and associated risk factors in a lowland area of Binh Dinh Province. *Central Vietnam Acta Trop*. 2016; 157:151–157. <https://doi.org/10.1016/j.actatropica.2016.01.029> PMID: 26872984
128. Hung N, Dung D, Anh N, Van P, Thanh B, Ha N, et al. Current status of fish-borne zoonotic trematode infections in Gia Vien district, Ninh Binh province, Vietnam. *Parasit Vectors*. 2015; 8:21. <https://doi.org/10.1186/s13071-015-0643-6> PMID: 25586313
129. Van De N, Hoa LT. Human infections of fish-borne trematodes in Vietnam: Prevalence and molecular specific identification at an endemic commune in Nam Dinh province. *Exp Parasitol*. 2011; 129:355–361. <https://doi.org/10.1016/j.exppara.2011.09.005> PMID: 21959023
130. Vinh H, Phimraphai W, Tangkawattana S, Smith J, Kaewkes S, Dung D, et al. Risk factors for Clonorchis sinensis infection transmission in humans in northern Vietnam: A descriptive and social network analysis study. *Parasitol Int*. 2017; 66(2):74–82. <https://doi.org/10.1016/j.parint.2016.11.018> PMID: 27939296
131. Tran A, Doan H, Do A, Nguyen V, Hoang S, Le H, et al. Prevalence, species distribution, and related factors of fish-borne trematode infection in Ninh Binh Province, Vietnam. *Biomed Res Int*. 2019. <https://doi.org/10.1155/2019/8581379> PMID: 31467915
132. Mas-coma S, Funatsu I, Bargues M. *Fasciola hepatica* and lymnaeid snails occurring at very high altitude in South America. *Parasitology*. 2003; 123(7):115–127. <https://doi.org/10.1017/S0031182001008034> PMID: 11769277
133. Mas-Coma S, Buchon P, Funatsu I, Angles R, Mas-Bargues C, Artigas P, et al. Donkey fascioliasis within a One Health control action: Transmission Capacity, Field Epidemiology, and Reservoir Role in a Human Hyperendemic Area. *Front Vet Sci*. 2020. <https://doi.org/10.3389/fvets.2020.591384> PMID: 33251272
134. Mas-Coma S, Funatsu I, Angles R, Buchon P, Mas-Bargues C, Artigas P, et al. Domestic pig prioritised in One Health action against fascioliasis in human endemic areas: Experimental assessment of transmission capacity and epidemiological evaluation of reservoir role. *One Health*. 2021; 16(13):100249. <https://doi.org/10.1016/j.onehlt.2021.100249> PMID: 33997234
135. Mas-Coma S, Cafrune M, Funatsu I, Mangold A, Angles R, Buchon P, et al. Fascioliasis in Llama, *Lama glama*, in Andean Endemic Areas: Experimental Transmission Capacity by the High Altitude Snail Vector *Galba truncatula* and Epidemiological Analysis of Its Reservoir Role. *Animals*. 2021; 11(9):2693. <https://doi.org/10.3390/ani11092693> PMID: 34573658
136. Mas-Coma S, Buchon P, Funatsu I, Angles R, Artigas P, Valero M, et al. Sheep and Cattle Reservoirs in the Highest Human Fascioliasis Hyperendemic Area: Experimental Transmission Capacity, Field Epidemiology, and Control Within a One Health Initiative in Bolivia. *Front Vet Sci*. 2020; 27(7):583204. <https://doi.org/10.3389/fvets.2020.583204> PMID: 33195605
137. Bargues M, Angles R, Coello J, Artigas P, Funatsu I, Cuervo P, et al. One Health initiative in the Bolivian Altiplano human fascioliasis hyperendemic area: Lymnaeid biology, population dynamics, microecology and climatic factor influences. *Braz J Vet Parasitol*. 2021; 30(2):e025620. <https://doi.org/10.1590/S1984-29612021014> PMID: 34076053
138. Bargues M, Artigas P, Angles R, Osca D, Duran P, Buchon P, et al. Genetic uniformity, geographical spread and anthropogenic habitat modifications of lymnaeid vectors found in a One Health initiative in

the highest human fascioliasis hyperendemic of Bolivian Altiplano. *Parasit Vectors*. 2020; 13:171. <https://doi.org/10.1186/s13071-020-04045-x> PMID: 32252808

139. Angles R, Buchon P, Valero M. One Health Action against Human Fascioliasis in the Bolivian Altiplano: Food, Water, Housing, Behavioural Traditions, Social Aspects, and Livestock Management Linked to Disease Transmission and Infection Sources. *Int J Environ Res Public Health*. 2022; 19(3):1120. <https://doi.org/10.3390/ijerph19031120> PMID: 35162146
140. Bardales-Valdivia J, Bargues M, Hoban-Vergara C, Bardales-Bardales C, Goicochea-Portal C, Bazán-Zurita H, et al. Spread of fascioliasis endemic areas assessed by seasonal follow-up of rDNA ITS-2 sequenced lymnaeid populations in Cajamarca, Peru. *One Health*. 2021; 13. <https://doi.org/10.1016/j.onehlt.2021.100265> PMID: 34041348
141. Bargues M, Malandini J, Artigas P, Soria C, Velásquez J, Carnevale S, et al. Human fascioliasis endemic areas in Argentina: multigene characterization of the lymnaeid vectors and climatic environmental assessment of the transmission pattern. *Parasit Vectors*. 2016; 9(306).
142. Periago M, Valero M, Artigas P, Agramunt V, Bargues M, Curtale F, et al. Very High Fascioliasis Intensities in Schoolchildren from Nile Delta Governorates, Egypt: The Old World Highest Burdens Found in Lowlands. *Pathogens*. 2021; 10. <https://doi.org/10.3390/pathogens10091210> PMID: 34578242
143. Afshan K, Fortes-Lima C, Artigas P, Valero AM, Qayyum M, Mas-Coma S. Impact of climate change and man-made irrigation systems of transmission risk, long-term trend and seasonality of human and animal fascioliasis in Pakistan. *Geospat Health*. 2014; 8(2):317–334.
144. Quy T, Yeatman H. Evaluation of a broadly-based control model of fascioliasis (liver fluke) in Central Vietnam. *Vietnam J Public Health*. 2016; 4(1):40–52.
145. Tangkawattana S, Sripa B. Integrative EcoHealth/One Health Approach for Sustainable Liver Fluke Control: The Lawa Model. *Adv Parasitol*. 2018; 102:115–139. <https://doi.org/10.1016/bs.apar.2018.07.002> PMID: 30442307
146. Sripa B, Tangkawattana S, Sangnukul T. The Lawa Model: A sustainable, integrated opisthorchis control program using the EcoHealth approach in the Lawa Lake region of Thailand. *Parasitol Int*. 2017; 66(4):346–354. <https://doi.org/10.1016/j.parint.2016.11.013> PMID: 27890720
147. Sripa B, Tangkawattana S, Laha T, Kaewkes S, Mallory F, Smith J, et al. Towards integrated opisthorchis control in northeast Thailand: the Lawa project. *Acta Trop*. 2015; 141:361–367. <https://doi.org/10.1016/j.actatropica.2014.07.017> PMID: 25102053
148. Qian M, Gan X, Zhao J, Zheng W, Li W, Jiang Z, et al. Effectiveness of health education in improving knowledge, practice and belief related to clonorchiasis in children. *Acta Trop*. 2020; 2017:105436. <https://doi.org/10.1016/j.actatropica.2020.105436> PMID: 32278640
149. Belizario V, Jorge M, Ng J, Mistica M, Lam H, Rivera L, et al. A Model for Integrating Paragonimiasis Surveillance and Control with Tuberculosis Control Program. *Southeast Asian J Trop Med Public Health*. 2016; 47(4):625–637.
150. Trinos J, Sison O, Armino M, Lacuna J, Jorge M, Belizario V. Identification of suspected paragonimiasis-endemic foci using a questionnaire and detection of *Paragonimus* ova using the Ziehl-Neelsen technique in Zamboanga Region, the Philippines. *Pathog Glob Health*. 2020; 114(3):127–135. <https://doi.org/10.1080/20477724.2020.1741900> PMID: 32191613
151. WHO. 2021. Tripartite and UNEP support OHHEP's definition of "One Health". <https://www.who.int/news-room/01-12-2021-tripartite-and-unep-support-ohhlep-s-definition-of-one-health>. [Accessed 17 March 2022]
152. WHO. 2021. Ending the neglect to attain the Sustainable Development Goals: A road map for neglected tropical diseases 2021–2030. <https://www.who.int/publications/item/9789240010352>. [Accessed 17 March 2022]
153. Fitzpatrick C, Bangert M, Mbabazi P, Mikhailov A, Zouré H, Rebollo M, et al. Monitoring equity in universal health coverage with essential services for neglected tropical diseases: an analysis of data reported for five diseases in 123 countries over 9 years. *Lancet Glob Health*. 2018; 6(9):E980–E988. [https://doi.org/10.1016/S2214-109X\(18\)30307-3](https://doi.org/10.1016/S2214-109X(18)30307-3) PMID: 30054258