

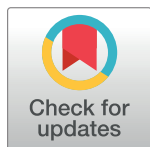
RESEARCH ARTICLE

Maternal *Leishmania infantum* infection status has significant impact on leishmaniasis in offspring

Angela J. Toepp^{1,2}, Carlyne Bennett^{1,2}, Benjamin Scott^{1,2}, Reid Senesac^{1,2}, Jacob J. Oleson³, Christine A. Petersen^{1,2*}

1 Department of Epidemiology, College of Public Health, University of Iowa, Iowa City, Iowa, United States of America, **2** Center for Emerging Infectious Diseases, University of Iowa Research Park, Coralville, Iowa, United States of America, **3** Department of Biostatistics, College of Public Health, University of Iowa, Iowa City, Iowa, United States of America

* christine-petersen@uiowa.edu



OPEN ACCESS

Citation: Toepp AJ, Bennett C, Scott B, Senesac R, Oleson JJ, Petersen CA (2019) Maternal *Leishmania infantum* infection status has significant impact on leishmaniasis in offspring. PLoS Negl Trop Dis 13(2): e0007058. <https://doi.org/10.1371/journal.pntd.0007058>

Editor: Mary Ann McDowell, University of Notre Dame, UNITED STATES

Received: August 16, 2018

Accepted: December 5, 2018

Published: February 13, 2019

Copyright: © 2019 Toepp et al. This is an open access article distributed under the terms of the [Creative Commons Attribution License](https://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Data Availability Statement: All data have been placed into Research Gate at this URL: <https://www.researchgate.net/project/Leishmania-transmission>

Funding: This work was funded by a grant from the National Institutes of Health NIAID R01TW010500 to CAP and JO. The funders had no role in study design, data collection and analysis, decision to publish or preparation of the manuscript.

Competing interests: The authors have declared that no competing interests exist.

Abstract

Visceral Leishmaniasis is a deadly disease caused by *Leishmania infantum*, endemic in more than 98 countries across the globe. Although the most common means of transmission is via a sand fly vector, there is growing evidence that vertical transmission may be critical for maintaining *L. infantum* infection within the reservoir, canine, population. Vertical transmission is also an important cause of infant morbidity and mortality particularly in sub-Saharan Africa. While vertical transmission of visceralizing species of *Leishmania* has been reported around the globe, risk factors associated with this unique means of *Leishmania* transmission have not been identified therefore interventions regarding this means of transmission have been virtually non-existent. Furthermore, the basic reproductive number, (R_0), or number of new *L. infantum* infections that one infected mother or dam can cause has not been established for vertical transmission, also hampering the ability to assess the impact of this means of transmission within reservoir of human hosts. Canine Leishmaniosis (CanL) is enzootic within a U.S. hunting dog population. CanL is transmitted within this population via transplacental transmission with no reported vector transmission, despite many repeated attempts to find infected sand flies associated with these dogs and kennels. This population with predominantly, if not solely, vertical transmission of *L. infantum* was used to evaluate the critical risk factors for vertical transmission of *Leishmania* and establish the R_0 of vertical *L. infantum* infection. Evaluation of 124 animals born to eighteen dams diagnostically positive for infection with *L. infantum* showed that there was a 13.84x greater chance of being positive for *L. infantum* within their lifetime if the mother was also positive within her lifetime (RR: 13.84, 95% CI: 3.54–54.20, p-value: <0.0001). The basic reproductive number for vertically transmitted *L. infantum* within this cohort was 4.12. These results underscore that there is a high risk of *L. infantum* infection to transmit from mother to offspring. Targeted public health interventions and control efforts that address vertical transmission of *L. infantum* are necessary in endemic countries to eliminate visceral leishmaniasis.

Author summary

Canine leishmaniosis (CanL) is a deadly disease caused by *Leishmania infantum* parasite, it is found in animal populations, including people, in more than 98 countries across the globe. CanL was first identified within the US in hunting dogs 1980 and then again in 1999 when a large outbreak in a kennel in New York occurred. As the US is usually not considered a tropical country, there was much debate about how this neglected, vector borne, tropical disease had made its way into these dogs. We found that within the U.S. hunting dog population CanL is transmitted from mom to pup with no reported sand fly transmission in the population, despite multiple attempts to find infected sand flies associated with these dogs. While vertical transmission of this disease has been reported in case reports around the globe, risk factors associated with this unique means of *Leishmania* transmission are not known. Furthermore, the basic reproductive number, (R_0), or number of new infections that one infected animal can cause has not been reported for vertical transmission of *L. infantum*. It is important to know the R_0 as it helps identify how infectious a route of transmission can be and therefore how easy it might be to control this infection. A cohort of 124 dogs from 18 dams was analyzed from 1999 to 2016 for factors related to vertical transmission. Offspring from dams ever diagnostically positive for infection with *L. infantum* were 13.84x more likely to become positive for *L. infantum* themselves within their lifetime (RR: 13.84 95% CI: 3.54–54.20 p-value: <0.0001). The basic reproductive number for vertically transmitted *L. infantum* within this cohort was 4.12. These results underscore that an infected mom is highly likely to infect her offspring if treatment is not started to prevent transmission. There is a need for any public health prevention and control efforts to address vertical as well as vector transmission of canine leishmaniosis in endemic countries.

Introduction

Leishmaniosis is a disease caused by the obligate intracellular protozoan parasite *Leishmania infantum* [1–3]. Visceral Leishmaniasis (VL) can also be caused by *Leishmania donovani* which causes anthroponotic human visceral leishmaniasis in many countries including areas of Asia and Africa [4, 5]. Zoonotic visceral leishmaniasis (ZVL) occurs in countries where the disease is endemic/enzootic in both human and animal populations. Within these countries the parasite is transmitted primarily via the phlebotomine sand fly [6, 7], although the role of other means of transmission, particularly vertical transmission, is not known. Dogs play an important role in the ecology and control of ZVL as they are the predominant domestic reservoir for the disease, with greater than 10% seropositivity often evident in dogs prior to emergent VL observed in people [8]. Dog ownership is a risk factor of human visceral leishmaniasis in multiple endemic countries with ZVL including Iran, Ethiopia, and Brazil [9–11]. As such, control measures in locations where ZVL is prominent include insecticide treatment or culling of dogs. Dogs remain an important model system for understanding the ecology and epidemiology of VL [12–14].

In recent years vertical, and specifically transplacental, transmission of *L. infantum* has been shown to be able to maintain infection within population(s) of dogs [15, 16]. Dogs in Brazil have been shown to have infected *in utero* pups [17–19]. Multiple case reports and case series have identified vertical transmission of VL as an important cause of infant morbidity and mortality [20–22]. Compared to sand fly transmitted infection [23–25], there is very little

known about the risk of vertical transmission in dogs or people [16, 26–29]. Therefore, understanding the impact and risk factors associated with parasite transmission *in utero* is important for education and treatment of infected mothers and for control of *Leishmania* infection within reservoir hosts.

In the United States leishmaniasis is enzootic in hunting dogs. CanL was first identified in a dog with no travel outside of the United States in 1980, but it was not until a large outbreak in a kennel in New York in 1999 that a larger scale study was performed to understand the broad burden of disease in the U.S. hunting dog population [30, 31]. Further examination found that the primary route of transmission in this population was vertical, from dam to pup [15, 32] and not via sand fly transmission despite many studies looking for infected sand flies associated with these infected dogs [33, 34]. Despite experimental studies that indicate that vector transmission of the *Leishmania infantum* found in US hunting dogs is possible, there is no evidence that vector transmission occurs naturally from the U.S. hunting dog population [34–36]. A decade of surveillance of this hunting hound population found that the prevalence of CanL from vertical transmission was higher than expected and similar to the rates seen in countries where VL is endemic [37, 38].

The basic reproductive number, R_0 , or the number of secondary infections one infected individual can cause within a susceptible population is an important epidemiological value for public health officials interested in control and elimination of this disease in endemic countries [39]. Previous calculations of the R_0 for leishmaniasis have been restricted as these studies did not include vertical transmission as a potential route of transmission or lacked data to assess the true rate of transmission in a population [40–42].

This study examines both *L. infantum* positive and negative dams their offspring over the course of their lifetime to determine risk factors associated with vertical transmission and the corresponding crude basic reproductive number of vertical transmission. We hypothesize that the crude R_0 of vertical transmission will be greater than one: *Leishmania* will maintain infection by infecting at least one pup from a diagnostically positive dam. Understanding the risk factors associated with vertical transmission remains an important public health concern as elimination and control programs focusing on vector control does not show 100% reduction of VL in endemic countries with zoonotic disease [43–45], and vertical transmission appears to be a major risk for maintaining disease within an area or population.

Materials and methods

Study design

A retrospective cohort study based on data collected regarding *Leishmania infantum* infection and exposure in U.S. hunting dogs since the 1999 outbreak [33, 34, 46] was completed. A subset of dams that were diagnostically positive and never diagnostically positive were identified. All pups from these two respective groups, ever positive or never positive, were tracked to determine their *Leishmania* diagnostic status. All historical data was collected from studies performed by Centers for Disease Control and Prevention [33, 34] and the our laboratory at Iowa State University and the University of Iowa [15, 32, 35, 47–49].

Ethics statement

All dogs were enrolled in this retrospective study with informed consent from their caretakers and all protocols followed were approved by the University of Iowa Institution Animal Care and Use Committee (IACUC) an AAALAC accredited institution following the requirements for the US National Institutes of Health Office of Laboratory Animal Welfare Assurances

which operates under the 2015 reprint of the Public Health service Policy on Humane Care and Use of Laboratory Animals, under protocol #6041721.

Animals

An active surveillance cohort of 4 large (>50 dogs each) kennels was established and observed over a 9-year period. Our laboratory visited each of these kennels biannually for at least three years, at which point two of the kennels elected to control visceral leishmaniasis in their kennel via euthanasia. Licensed veterinarians collected 1–5 cc whole blood and serum from all dogs present at these kennels. Demographic information regarding time of pregnancy, sex and age were collected. The active surveillance cohort testing period extended from 2007 to 2017. This surveillance effort started eight years, or at least one hunting-dog life-span, after the major *L. infantum* outbreak in 1999 with CanL surveillance performed on these same dogs passively by the CDC as reported in [33, 34].

Leishmania diagnostic status PCR

DNA was isolated from canine peripheral whole blood samples collected in heparinized or ethylenediaminetetraacetic acid (EDTA) via the QIAmp DNA Blood Mini Kit (Qiagen, Valencia, CA) per manufacturer protocol. The quality and quantity of DNA was assessed using a Nano-Drop 2000 (Thermo, Scientific, Waltham, MA). Real time quantitative polymerase chain reaction (RT-qPCR) was performed as previously described with all samples run in duplicate with positive samples determined as samples with 1 or more positive wells and negative samples with no amplification in any wells [37, 49–51]. All RT-qPCR included both positive, negative control blood spiked with 10^6 *Leishmania infantum* parasites, and negative controls. Between 2007 and 2011 kinetoplastid primer and probe targets were used. The primer and probe sequences were as follows: F 5'-CCGCCCCGCTCAAGAC, R 5'-TGCTGAATATTGGTGGT TTTGG, (Integrated DNA Technologies, Coralville, IA) and TaqMan probe, 5'-6FAM-AGCC GCGAGGACC-MGBNFQ, were used (Applied Biosystems, Foster City, CA). From 2012 to present ribosomal primer and probe targets were utilized. The sequences were as follows: F 5'-AAGTGCTTTCCCATCGCAACT, R 5' CGCACTAAACCCCTCCAA (Invitrogen, Life Technologies, Grand Island, NY), probe: 5' 6FAM-CGGTTCGGTGTGTGGCGCC-MGBNFQ (Applied Biosystems, Life Technologies, Grand Island, NY). Assays were performed on ABI 7000 systems until 2016 when they were run on ABI 7900 systems (Applied Biosystems). Analysis was performed using ABI 7000 System SDS Software and ABI 7900 HT Sequence Detection Systems Version 2.4.1. (Applied Biosystems).

Serological status

Serological status was determined via the Dual Path Platform Canine Visceral Leishmaniasis (DPP CVL) assay (Chembio Diagnostic Systems Inc., Medford, NY) or via immunofluorescent antibody test (IFAT). The DPP CVL assay detects *Leishmania*-specific antibodies via rK28 antigen, a *Leishmania* recombinant antigen. The assay was utilized as previously described with positives determined as dogs with a test and control line appearing at 4 minutes or less [51]. All positives or questionable samples were confirmed using the Chembio microreader system. The system detects the intensity of the control and test lines. Immunofluorescent antibody test (IFAT) was utilized on canine samples before 2015. This test was performed by the Division of Parasitic Diseases at the Centers for Disease Control and Prevention as previously described [33, 52]. Positive tests were determined as tests where immunofluorescence was reported in 50% of organisms at serum dilutions equal to or above 1/64. These tests were

performed without identifying each dog (blindly) and were repeated four times at each dilution to determine positivity.

Statistical methods

Univariate analyses were performed to determine unadjusted relative risk values for dam's age at the time of birth, diagnostic status during the year of birth, and other variables. Pearson chi-squared test and Fisher's exact test were used to assess categorical variables against disease status. Mann-Whitney test was used to compare dam's age between disease states as age not normally distributed. An unpaired t-test with the Welch's correction was utilized to compare litter size between infected and uninfected groups. For assessment purposes the dam's diagnostic status via qPCR or serology during the same year she gave birth was utilized. Feasibility restrictions, the fact the gestational period for a dog is two months, prevented the researchers from obtaining information on the dam's diagnostic status during pregnancy.

Multivariable logistic regressions were performed to determine adjusted relative risk. Due to the fact that the dam's diagnostic status can be determined via qPCR and serology, diagnostic status was assessed different ways through three models. One model included the overall diagnostic status of the dam (ever diagnostic positive vs never diagnostic positive), the dam's age at the time she gave birth (older than six years of age vs younger than or equal to six years of age), and the sex of the puppy (male vs female). To further assess the dam's diagnostic status impact a second model was created with qPCR and serology as separate variables. A third model was created separating the dam's serology and dam's PCR status in the year she gave birth into two explanatory variables. P-values of less than 0.05 were determined as statistically significant. Each model was fit assuming a binomial distribution with a log link function.

Kaplan-Meier time to event analysis was performed to assess whether dam's diagnostic status altered time to pup diagnostic positive.

Basic reproductive number was calculated using dams who were ever diagnostically positive for *Leishmania*, from which their average proportion of puppies per litter that became *Leishmania* diagnostic positive was determined. Hunting dogs are a medium size dog with average litter size in the study was between 6–7 [53]. Using the average litter size, the proportion of puppies in a litter that would become positive for *Leishmania* was determined as the basic reproductive number of vertical transmission in US hunting dogs.

For all analyses, as observation of transmission of *L. infantum* infection was the goal, *L. infantum* exposure/diagnostic result status for each dog was identified as “ever diagnostically positive” for *Leishmania* or “never diagnostically positive” for *Leishmania*. Positivity was determined as qPCR positive and/or serologically positive at any point during the dog's lifetime.

All statistical analyses were performed using SAS 9.4 (SAS Institute, Cary, NC) and Graph Pad Prism 6 (GraphPad Software, Inc, La Jolla, CA).

Results

Study demographics and univariate analysis

Compared to sand fly transmission, little is known about the risk factors of vertical transmission of *Leishmania infantum*. Understanding these risk factors and the corresponding likelihood of transmission as measured by the basic reproductive number, R_0 , provides valuable information for assessing control and elimination programs for zoonotic leishmaniasis. We hypothesized that a dam's positive *Leishmania* diagnostic status during pregnancy would be a risk factor of *L. infantum* transmission. A retrospective cohort study examined the health records from 130 dogs born to eighteen dams for risk factors associated with vertical transmission and the corresponding individual level basic reproductive number calculation. Six dogs were

Table 1. Dam and litter demographics based on dam's *Leishmania* diagnostic status.

Variable	Dam <i>Leishmania</i> dx Positive (ever) (N = 8)	Dam <i>Leishmania</i> dx Negative (N = 10)
Average age of dam at pregnancy, ±SD (Min-Max)	5.14 ± 1.83 (2–9)	4.00 ± 1.70 (2–7)
Proportion of male puppies*, %, N	48.61, 35	38.64, 17
Average litter size ±SD (Min-Max)	7.02 ± 3.24 (1–15)	6.3 ± 2.58 (2–11)
Average number of previous litters (Min-Max)	0.57 (0–2)	0 (0–0)
Proportion of puppies <i>Leishmania</i> diagnostic positive ever, %, N	62.16, 46	4.00, 2

*Data incomplete due to missing information

Leishmania diagnostic status determined as dam ever positive via IFAT serology, DPP CVL assay, or PCR.

<https://doi.org/10.1371/journal.pntd.0007058.t001>

removed from analysis due to incomplete data to use in statistical models. There were eight dams identified as *Leishmania* diagnostic positive at some point in their lifetime and ten dams were diagnostically negative throughout their lives. Most dogs were not multiparous. The average litter size was 6–7 pups (Table 1).

Dogs that ever became diagnostically positive were born to dam's that were slightly older in age, 5.10 years compared to 4.04 years (p-value = 0.0004) and were more likely to be born to dams who had previously had at least one litter (p-value <0.0001, RR = 3.351 95% CI:2.32–4.83). There was no significant difference between *Leishmania* diagnostic outcome in male vs. female dogs. Dogs ever diagnostically positive were more likely to be from large(r) litters. This difference have been skewed by on particularly large litter of fifteen puppies from a dam that was diagnostically positive during her year of pregnancy at six years of age. When this litter is removed the significance of dam age and litter size is reduced.

Additional analysis shows that dogs born to a dam that was qPCR positive for *Leishmania infantum* at the time of pregnancy had a relative risk of being diagnostically positive during their lifetime 10.46x greater than the risk than when the dam was PCR negative at the time of pregnancy (Unadjusted RR: 10.46, 95% CI: 3.57–31.82, p-value <0.0001). The dam's serological status during the year she gave birth was also found to increase the risk of offspring testing diagnostically positive within their lifetime. Pups born to dams seropositive during the year they gave birth were 2.69x more likely to test positive for *Leishmania* within their life (Unadjusted RR: 2.69 95% CI: 1.32–5.52, p-value 0.0054, Table 2).

Table 2. Univariate analysis of risk factors for vertical transmission from dam to pup.

Variable	Pup ever <i>Leishmania</i> dx positive (N = 48)	Pup never <i>Leishmania</i> dx positive (N = 76)	P-value	Unadjusted Relative Risk
Average age of dam at pregnancy ±SD (Min-Max)	5.10 ± 1.61 (3.00–9.00)	4.04 ± 1.45 (2.00–7.00)	0.0004	N/A
Proportion of male puppies*, %, N	40.43, 19	47.83, 33	0.6731	0.91
Average litter size, ±SD (Min-Max)	9.5 ± 4.02 (1–15)	7.62 ± 2.51 (2–15)	0.0049	N/A
Proportion dam <i>Leishmania</i> PCR positive**	46.15	4.41	<0.0001	10.46
Proportion dam <i>Leishmania</i> Seropositive**	34.04	12.68	0.0054	2.69
Proportion dam ever <i>Leishmania</i> dx positive	95.83	36.84	<0.0001	2.601

*Data incomplete due to missing information.

**Diagnostic status of dam during year gave birth

Outcome defined as pups diagnostic positivity for *Leishmania* via qPCR or serology throughout lifetime.

<https://doi.org/10.1371/journal.pntd.0007058.t002>

Controlling for all variables, transmission of *Leishmania* is dramatically higher from dams diagnostically positive for *Leishmania*

A series of three logistic regression models were created to determine the risk factors associated with vertical transmission of *L. infantum*. The models were labeled as A, B, and C. Whether the puppy became diagnostically positive within their lifetime or not was used as the outcome for these models. Model A assessed a dam’s diagnostic status as ever positive for *Leishmania* during their lifetime as an explanatory variable along with age at the time of pregnancy, and sex of the dog. When adjusting for all other explanatory variables it is found that dogs born to a dam that was ever positive for *Leishmania* have a relative risk **13.84x** greater than dogs born to a dam that was never diagnostically positive (Adjusted RR: 13.84, 95% CI: 3.54–54.20, p-value 0.0002).

Transmission of *Leishmania* is higher in dams qPCR positive for *Leishmania*

In order to assess the impact of seropositivity/ *Leishmania* exposure vs detectable parasite infection via qPCR from the blood in transmission two additional models were created; models B and C. Model B utilized a dam’s diagnostic status during the year she gave birth (positive vs negative), age of dam during pregnancy (older than six vs younger), and sex of the puppy as explanatory variables. In this model puppies born to dams diagnostically positive via qPCR or serology during the year of pregnancy were 2.27x more likely to become positive for *Leishmania* compared to dogs born to a dam that was diagnostically negative at the time of pregnancy. Model C used the dam’s qPCR status, serostatus and age during the year of pregnancy and progenys’ sex as explanatory variables. This model allows for the assessment of how parasite infection via qPCR from the blood vs seropositivity/ *Leishmania* exposure could affect *Leishmania* transmission. Pups born to a dam that was qPCR positive for *Leishmania* during pregnancy were 3.14x more likely to become positive for *Leishmania* in their lifetime (Adjusted RR: **3.14**, 95% CI: 1.37–7.18, p-value: 0.0067, **Table 3**). Two dogs born to a dam that was never qPCR or serologically positive for *Leishmania* were found to be positive during their lifetime. One dog was identified as ever qPCR positive and one as ever serologically positive.

A dam’s serological status during the year of pregnancy was not statistically significantly associated with her offspring becoming diagnostically positive. This was an interesting finding

Table 3. Dam ever dx positive and qPCR positive during year of pregnancy significantly associated with *Leishmania* transmission to pups.

Model	Variable	Sample Size N = number of dams n = number of puppies	Adjusted RR of pup lifetime exposure	95% CI	p-value
A	Dam ever <i>Leishmania</i> dx positive	N = 18 n = 116	13.84	3.54–54.20	0.0002
B	Dam dx positive during yr. of pregnancy	N = 14 n = 100	2.27	1.40–3.68	0.0009
C	Dam qPCR positive during pregnancy	N = 14 n = 100	3.14	1.37–7.18	0.0067
C	Dam seropositive during pregnancy	N = 14 n = 100	1.07	0.46–2.55	0.8639

Multivariate logistic regression analysis for risk factors associated with *Leishmania* vertical transmission. Model A: Explanatory variables are dam’s *Leishmania* diagnostic status (ever positive vs. never positive), age of dam during pregnancy (older the 6 years old vs. 6 or younger), and sex of offspring. Model B: Explanatory variables are dam’s *Leishmania* diagnostic status during pregnancy, age of dam during pregnancy, sex of offspring. Model C: Explanatory variables are dam’s *Leishmania* qPCR serostatus and age during pregnancy, sex of offspring.

<https://doi.org/10.1371/journal.pntd.0007058.t003>

as qPCR is a measure of parasite DNA within the peripheral blood. As the transplacental blood supplied each *in utero* puppy with nutrients, and apparently parasites, this may have increased the risk of the puppy becoming infected with *Leishmania* parasites.

Offspring of dams diagnostically positive for *Leishmania* are more likely to become positive for *Leishmania*

Based on our findings via univariate and logistic regression, we were interested in evaluating the risk of becoming *Leishmania* diagnostic positive over years of a pup's life based on its mother's diagnostic status. To better assess when dogs became diagnostically positive for *Leishmania*, time to event Kaplan-Meier curves were created. To visualize the overall relationship between age at which offspring became *Leishmania* diagnostically positive this was compared between the groups of dam *Leishmania* positive vs negative ever. Dogs born to positive dams (red) were statistically significantly more likely to become positive at a younger age than dogs born to negative dams (blue) (chi-square: 40.33, p-value <0.0001, Fig 1).

Based on the previous finding that dam qPCR status during the year she was pregnant was also highly correlated with the pup becoming *Leishmania* diagnostic positive, dam's qPCR status (negative during year of birth vs. positive) was utilized. Offspring born to dams who were qPCR positive (red) during the year they gave birth were significantly more likely to become positive for *Leishmania* via qPCR at younger ages than offspring from dams that were qPCR negative (blue) (Fig 2, chi-squared: 49.54 p-value <0.0001).

This was similar to the relationship between dams who were seropositive during the year they gave birth and the age at which their puppies became seropositive for *Leishmania* (Fig 3, chi-squared 18.43, p-value <0.0001).

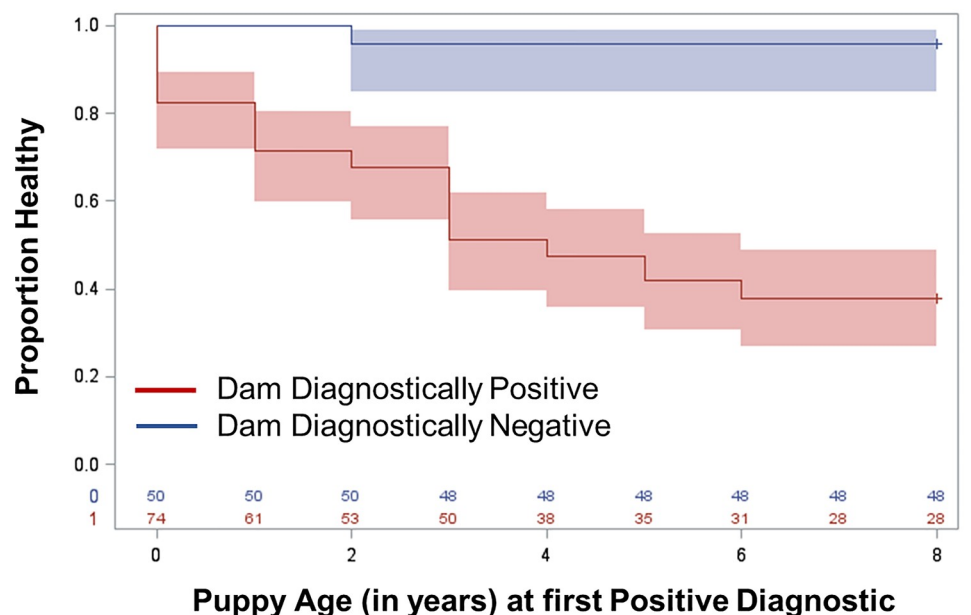


Fig 1. Kaplan-Meier time to offspring diagnostic positive based on dam's diagnostic status ever. Proportion healthy refers to the proportion of dogs that were diagnostically negative via qPCR and ELISA and DPP CVL assay. Blue represents the diagnostic status of pups from dams who were diagnostically negative during their lifetime. Red represents the diagnostic status of pups from dams who were diagnostically positive at any point during their lifetime via qPCR or serology. Shaded area represents variance around the mean. (chi-squared: 26.28 p-value <0.0001).

<https://doi.org/10.1371/journal.pntd.0007058.g001>

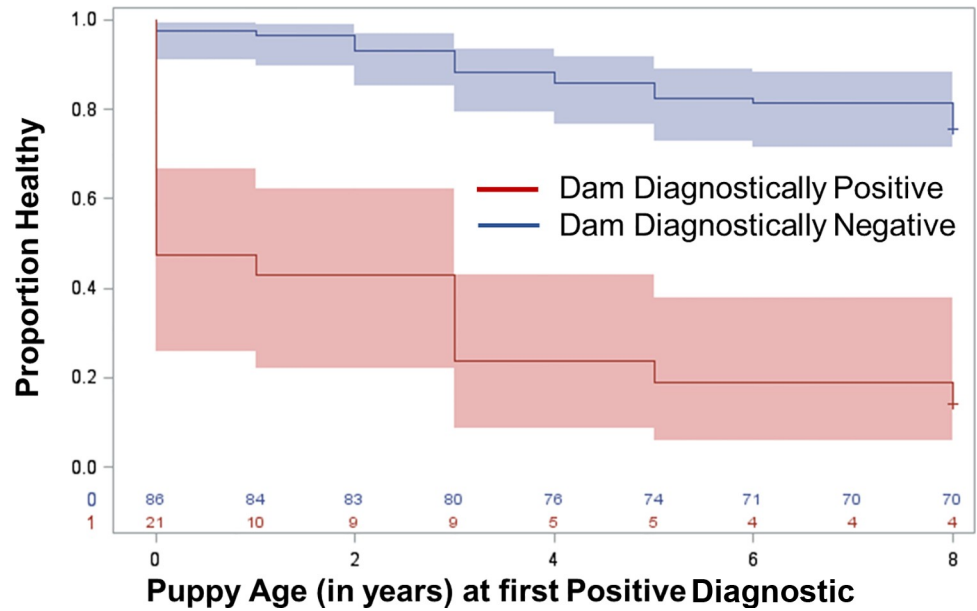


Fig 2. Kaplan-Meier time to dog diagnostic positive based on dam’s qPCR diagnostic status during the year of pregnancy. Proportion healthy refers to the proportion of dogs that are diagnostically negative via qPCR. Blue represents the diagnostic status of pups from dams who were diagnostically negative during year of pregnancy via qPCR. Red represents the diagnostic status of pups from dams who were diagnostically positive via qPCR during year of pregnancy. (chi-squared: 55.70 p-value <0.0001).

<https://doi.org/10.1371/journal.pntd.0007058.g002>

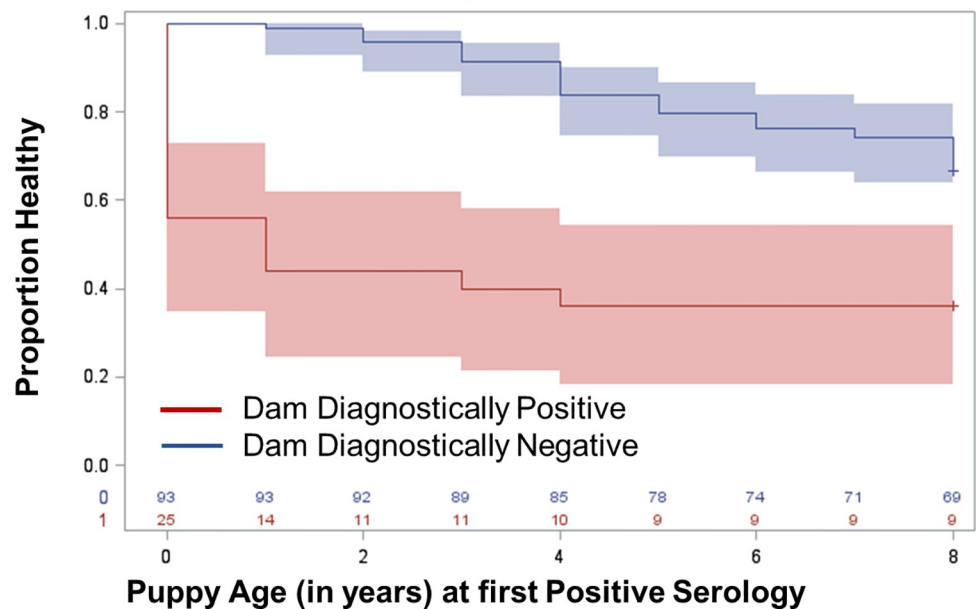


Fig 3. Kaplan-Meier time to dog diagnostic positive based on mother’s serological diagnostic status during pregnancy. Proportion healthy refers to the proportion of dogs that are diagnostically negative via ELISA and DPP CVL assay. Blue represents the diagnostic status of pups from dams who were diagnostically negative during year of pregnancy via ELISA and/or DPP CVL assay. Red represents the diagnostic status of pups from dams who were diagnostically positive via ELISA and/or DPP CVL during year of pregnancy.

<https://doi.org/10.1371/journal.pntd.0007058.g003>

Transmission of *L. infantum* persists across three generations of dogs

Within this study cohort we found two instances and three litters in which three generations of infected dogs were identified. In these specific families, on average the second generation had evidence of infection in 79.2% of dogs (seropositive or PCR positive at some point of their lives). To date, dogs in the third generation were 60.4% sero- or PCR positive for *L. infantum*. It should be noted that one of these litters are dogs currently 3 years old. These younger dogs may become qPCR or seropositive as they age and experience immunosuppressive conditions.

Dogs diagnostically positive for *L. infantum* highly likely to die from clinical visceral leishmaniasis

A small subset of 20 dogs within the study were more closely followed through their entire lives and cause of death was established. Of the 20 dogs from infected dams for which a cause of death was identified 95%, or 19 of these dogs, died from clinical visceral leishmaniasis. The one dog identified as being diagnostically positive for *Leishmania infantum* but not dying from clinical visceral leishmaniasis died from a secondary infection with *Ehrlichia spp.* as identified at necropsy. Neither of the two dogs born to uninfected mothers found to be infected with *L. infantum* have died from VL, but this is a very small sample size.

Basic reproductive number for vertical transmission of *Leishmania* above 4 new infections

The basic reproductive number for vertical transmission of *Leishmania* remains of interest in order to determine effectiveness of control efforts that are in many cases focused on vector transmission. R_0 was calculated based on information regarding each litter from this population. On average, 64% of dogs born to a dam who were ever diagnostically positive for *Leishmania* will become positive in their lifetime. Using the average litter size of our population, between 6 and 7, we calculate an average R_0 of 4.16.

Discussion

A retrospective cohort study was performed to assess risk factors associated with vertical transmission of *Leishmania infantum* and a crude basic reproductive number was calculated for the population. The mother's *L. infantum* diagnostic status during the year she was pregnant was a statistically significant risk factor for her offspring to be *L. infantum* positive during their lifetime, with a significant 13 times greater risk of infection than dogs without maternal exposure to *Leishmania*. Despite these dramatic findings in this retrospective cohort study, there is an overall paucity of reported cases of congenital VL. This is likely for several reasons; first the diagnostic difficulties of confirming that a case is due to congenital transmission vs. exposure to sand fly transmitted disease in endemic areas. To date there is no way to distinguish *L. infantum* infection by route of transmission, so in endemic areas the presumption is that cases are vector borne, although this may not be true. The second reason is availability of treatment of mothers for ZVL during pregnancy reducing the maternal parasite load and therefore decreasing transmission to the child/offspring [27, 54]. This study is the first study to calculate the basic reproductive number and determine risk factors associated with vertical transmission of *Leishmania infantum* in a population where vertical transmission is the main route of transmission and there is no known vectorial transmission [55, 56].

Vertical transmission occurs not only in leishmaniosis but other infections as well, such as human immunodeficiency virus (HIV) and malaria [57, 58]. In HIV infection, anti-retroviral treatment during pregnancy and caesarean births have been associated with decreased risk of

transmission likely due to a reduced exposure to the mother's blood and virus [59]. In malaria, mothers with malaria during pregnancy are at risk of vertical transmission [60]. This is similar to CanL where dogs born to mothers that were qPCR positive during pregnancy had a much higher risk of becoming positive for *Leishmania*. This is likely due to the fact that a positive qPCR test identifies that there was parasite DNA in the blood which is shared between mother and pup across the placenta. The mother's combined diagnostic status of seropositive or qPCR positive was a significant risk factor in predicting whether a puppy would become positive during their life. This was also reasonable as dogs become immunocompromised there can be increased disease progression and parasite replication with higher serological diagnostic values in dogs with more severe clinical disease [47, 50].

Within this study there were two sets of three generations of dogs that were followed and data indicating that transmission occurred across these generations. These results provide additional evidence that vertical transmission is capable of maintaining visceral leishmaniasis in a population over multiple generations.

Within this study two *Leishmania*-positive dogs were born to dams that were never qPCR or serologically positive for *Leishmania*. In the hunting dog community, dogs are commonly drafted or traded between groups and across international borders from endemic to non-endemic areas. Such movement of dogs greatly increases the difficulty of consistent testing across different locations and disease risk levels. This testing limitation may have led to a false negative status for the mother [61]. The two puppies that were identified as serologically/qPCR positive without maternal exposure could also have been exposed to *Leishmania* via fighting or wound cleaning of infected pen mates as blood to blood contact is possible due to the fact the dogs are housed in communal areas.

A small subset of 20 dogs (15% of the study population) were followed until death and a cause of death was identified. 100% of the dogs with an established cause of death were diagnostically positive for *Leishmania infantum* at some time throughout their life. Of those dogs with an established cause of death in this cohort, 95% died from clinical visceral leishmaniasis. These results highlight that without treatment many of these animals will progress with clinical disease. Therefore, it remains an important public health goal to identify ways to prevent *L. infantum* transmission from mother to child in both animals and people.

The basic reproductive number was calculated via an individual level model system, thus the number refers to the number of dogs in each litter that one mother could infect. This calculation provides a direct assessment of the R_0 within this cohort. An R_0 of approximately 4 (rounded to the nearest whole number to refer to number of puppies in the litter) shows that this disease is capable of maintaining at high levels within a population without vector transmission. The R_0 of other diseases, such as influenza, which remain important public health concerns across the globe are as low as 2 [62]. Astonishingly, in comparison the R_0 identified for an average canine litter coming from an infected dam was greater than 4, similar to the estimated basic reproductive number of smallpox [63]. As these studies all occur in an area where there is not holoendemic pressure of sand fly transmission, establishing the R_0 and effect of vertical transmission in dogs from endemic areas would be valuable. These studies would all be limited by the inability to distinguish sand fly transmitted and vertical transmission once pups are born and it is hard to know the outcome of maternal infection on in utero pups.

Current control programs for leishmaniasis in countries where the disease remains endemic in both humans and animals include vector control, vaccination, and dog culling, which has been shown to be ineffective. Based on the data evaluated here, there is a significant need to also address vertical transmission through canine sterilization programs [64–66]. Recent studies have identified vaccination of infected/exposed asymptomatic dogs as safe, so vaccination to boost a better immunity prior to pregnancy may be of value to reduce

transmission to the next generation [51]. Larger scale xenodiagnosis studies need to be performed to determine what skin burden of parasites is required to transmit CanL and the effectiveness of vaccination [67], allopurinol or additional (immuno)therapies to reduce parasite load immediately before or during pregnancy. Further analysis using Bayesian compartmental model techniques combining both vector and vertical transmission should be used to better understand the basic reproductive number for the full ecology of *Leishmania* infection in endemic areas and subsequently model how this number can be altered by public health control and prevention measures to assess elimination potential.

The findings of this study underscore the need for risk management through spaying and neutering animals by dog owners to reduce vertical transmission of *L. infantum* from their dogs. This action would decrease propagation of CanL within the canine reservoir for reduced transmission to people.

Acknowledgments

The authors would like to thank the dog owners who helped us gather this data as well as previous members of the Petersen laboratory who ran diagnostics, performed data entry and ran analyses over the years.

Author Contributions

Conceptualization: Angela J. Toepp, Reid Senesac, Jacob J. Oleson, Christine A. Petersen.

Data curation: Angela J. Toepp, Carolyne Bennett, Benjamin Scott, Reid Senesac.

Formal analysis: Angela J. Toepp, Carolyne Bennett, Benjamin Scott, Reid Senesac, Jacob J. Oleson.

Funding acquisition: Christine A. Petersen.

Investigation: Angela J. Toepp, Christine A. Petersen.

Methodology: Angela J. Toepp, Christine A. Petersen.

Project administration: Christine A. Petersen.

Supervision: Christine A. Petersen.

Visualization: Angela J. Toepp, Christine A. Petersen.

Writing – original draft: Angela J. Toepp, Christine A. Petersen.

Writing – review & editing: Jacob J. Oleson, Christine A. Petersen.

References

1. Alvar J, Gutierrez-Solar B, Molina R, Lopez-Velez R, Garcia-Camacho A, Martinez P, et al. Prevalence of *Leishmania* infection among AIDS patients. *Lancet*. 1992; 339(8806):1427. Epub 1992/06/06. 0140-6736(92)91255-7 [pii]. PMID: [1350846](#).
2. Moreno J, Alvar J. Canine leishmaniasis: epidemiological risk and the experimental model. *Trends Parasitol*. 2002; 18(9):399–405. PMID: [12377257](#).
3. Miro G, Petersen C, Cardoso L, Bourdeau P, Baneth G, Solano-Gallego L, et al. Novel Areas for Prevention and Control of Canine Leishmaniasis: (Trends in Parasitology 33, 718–730; 2017). *Trends Parasitol*. 2017. Epub 2017/11/28. <https://doi.org/10.1016/j.pt.2017.05.005> PMID: [28601528](#).
4. Ready PD. Epidemiology of visceral leishmaniasis. *Clin Epidemiol*. 2014; 6:147–54. Epub 2014/05/17. <https://doi.org/10.2147/CLEP.S44267> PMID: [24833919](#); PubMed Central PMCID: [PMC4014360](#).
5. Werneck GL. Visceral leishmaniasis in Brazil: rationale and concerns related to reservoir control. *Rev Saude Publica*. 2014; 48(5):851–6. Epub 2014/11/06. <https://doi.org/10.1590/S0034-8910.2014048005615> PMID: [25372177](#); PubMed Central PMCID: [PMC4211574](#).

6. Rogers ME, Hajmova M, Joshi MB, Sadlova J, Dwyer DM, Volf P, et al. Leishmania chitinase facilitates colonization of sand fly vectors and enhances transmission to mice. *Cell Microbiol.* 2008; 10(6):1363–72. Epub 2008/02/21. CMI1132 [pii] <https://doi.org/10.1111/j.1462-5822.2008.01132.x> PMID: 18284631; PubMed Central PMCID: PMC2408650.
7. Scorza BM, Wacker MA, Messingham K, Kim P, Klingelutz A, Fairley J, et al. Differential Activation of Human Keratinocytes by Leishmania Species Causing Localized or Disseminated Disease. *J Invest Dermatol.* 2017; 137(10):2149–56. Epub 2017/06/26. <https://doi.org/10.1016/j.jid.2017.05.028> PMID: 28647347; PubMed Central PMCID: PMC5786447.
8. Lima ID, Lima ALM, Mendes-Aguiar CO, Coutinho JFV, Wilson ME, Pearson RD, et al. Changing demographics of visceral leishmaniasis in northeast Brazil: Lessons for the future. *PLoS Negl Trop Dis.* 2018; 12(3):e0006164. Epub 2018/03/07. <https://doi.org/10.1371/journal.pntd.0006164> PMID: 29509765.
9. Gavgani AS, Mohite H, Edrissian GH, Mohebbi M, Davies CR. Domestic dog ownership in Iran is a risk factor for human infection with *Leishmania infantum*. *Am J Trop Med Hyg.* 2002; 67(5):511–5. Epub 2002/12/14. PMID: 12479553.
10. Bsrat A, Berhe M, Gadissa E, Taddele H, Tekle Y, Hagos Y, et al. Serological investigation of visceral Leishmania infection in human and its associated risk factors in Welkait District, Western Tigray, Ethiopia. *Parasite Epidemiology and Control.* 2018; 3(1):13–20. <https://doi.org/10.1016/j.parepi.2017.10.004> PMID: 29774295
11. Lima ÁLM, de Lima ID, Coutinho JFV, de Sousa ÚPST, Rodrigues MAG, Wilson ME, et al. Changing epidemiology of visceral leishmaniasis in northeastern Brazil: a 25-year follow-up of an urban outbreak. *Transactions of The Royal Society of Tropical Medicine and Hygiene.* 2017; 111(10):440–7. <https://doi.org/10.1093/trstmh/trx080> PMID: 29394411
12. Schaut RG, Lamb IM, Toepp AJ, Scott B, Mendes-Aguiar CO, Coutinho JF, et al. Regulatory IgDhi B Cells Suppress T Cell Function via IL-10 and PD-L1 during Progressive Visceral Leishmaniasis. *J Immunol.* 2016; 196(10):4100–9. Epub 2016/04/15. <https://doi.org/10.4049/jimmunol.1502678> PMID: 27076677; PubMed Central PMCID: PMC4868652.
13. Esch KJ, Juelsgaard R, Martinez PA, Jones DE, Petersen CA. Programmed death 1-mediated T cell exhaustion during visceral leishmaniasis impairs phagocyte function. *J Immunol.* 2013; 191(11):5542–50. <https://doi.org/10.4049/jimmunol.1301810> PMID: 24154626; PubMed Central PMCID: PMC3896087.
14. Esch KJ, Schaut RG, Lamb IM, Clay G, Morais Lima AL, do Nascimento PR, et al. Activation of Autophagy and Nucleotide-Binding Domain Leucine-Rich Repeat-Containing-Like Receptor Family, Pyrin Domain-Containing 3 Inflammasome during *Leishmania infantum*-Associated Glomerulonephritis. *The American journal of pathology.* 2015; 185(8):2105–17. <https://doi.org/10.1016/j.ajpath.2015.04.017> PMID: 26079813; PubMed Central PMCID: PMC4530124.
15. Boggiatto PM, Gibson-Corley KN, Metz K, Gallup JM, Hostetter JM, Mullin K, et al. Transplacental transmission of *Leishmania infantum* as a means for continued disease incidence in North America. *PLoS Negl Trop Dis.* 2011; 5(4):e1019. Epub 2011/05/03. <https://doi.org/10.1371/journal.pntd.0001019> PMID: 21532741; PubMed Central PMCID: PMC3075227.
16. Grinnage-Pulley T, Scott B, Petersen CA. A Mother's Gift: Congenital Transmission of *Trypanosoma* and *Leishmania* Species. *PLoS Pathog.* 2016; 12(1):e1005302. Epub 2016/01/29. <https://doi.org/10.1371/journal.ppat.1005302> PMID: 26821216; PubMed Central PMCID: PMC4731145.
17. da Silva SM, Ribeiro VM, Ribeiro RR, Tafuri WL, Melo MN, Michalick MS. First report of vertical transmission of *Leishmania (Leishmania) infantum* in a naturally infected bitch from Brazil. *Vet Parasitol.* 2009; 166(1–2):159–62. Epub 2009/09/08. S0304-4017(09)00497-X [pii] <https://doi.org/10.1016/j.vetpar.2009.08.011> PMID: 19733439.
18. Mancianti F, Sozzi S. Isolation of *Leishmania* from a newborn puppy. *Trans R Soc Trop Med Hyg.* 1995; 89(4):402. Epub 1995/07/01. PMID: 7570879.
19. Pangrazio KK, Costa EA, Amarilla SP, Cino AG, Silva TM, Paixao TA, et al. Tissue distribution of *Leishmania chagasi* and lesions in transplacentally infected fetuses from symptomatic and asymptomatic naturally infected bitches. *Vet Parasitol.* 2009; 165(3–4):327–31. Epub 2009/08/04. S0304-4017(09)00412-9 [pii] <https://doi.org/10.1016/j.vetpar.2009.07.013> PMID: 19647368.
20. Galindo-Sevilla N, Mancilla-Ramírez J. T-cell tolerance as a potential effect of congenital leishmaniasis on offspring immunity. *Parasite Immunology.* 0(0):e12540. <https://doi.org/10.1111/pim.12540> PMID: 29888463
21. Adam GK, Abdulla MA, Ahmed AA, Adam I. Maternal and perinatal outcomes of visceral leishmaniasis (kala-azar) treated with sodium stibogluconate in eastern Sudan. *Int J Gynaecol Obstet.* 2009; 107(3):208–10. Epub 2009/09/22. <https://doi.org/10.1016/j.ijgo.2009.08.002> PMID: 19766208.
22. Adam GK, Omar SM, Ahmed MA, Abdallah TM, Ali AA. Cross-sectional study of the case-fatality rate among patients with visceral leishmaniasis infections during pregnancy in Sudan. *Int J Gynaecol Obstet.* 2018; 140(1):119–20. Epub 2017/09/30. <https://doi.org/10.1002/ijgo.12332> PMID: 28960291.

23. Cortes S, Vaz Y, Neves R, Maia C, Cardoso L, Campino L. Risk factors for canine leishmaniasis in an endemic Mediterranean region. *Vet Parasitol.* 2012; 189(2–4):189–96. Epub 2012/05/12. <https://doi.org/10.1016/j.vetpar.2012.04.028> PMID: 22575278.
24. Moreira ED Jr., de Souza VM, Sreenivasan M, Lopes NL, Barreto RB, de Carvalho LP. Peridomestic risk factors for canine leishmaniasis in urban dwellings: new findings from a prospective study in Brazil. *Am J Trop Med Hyg.* 2003; 69(4):393–7. Epub 2003/12/03. PMID: 14640499.
25. Oliveira LC, Araujo RR, Alves CR, Mouta-Confort E, Lopez JA, Mendonca-Lima FW. Seroprevalence and risk factors for canine visceral leishmaniasis in the endemic area of Dias D'Avila, State of Bahia, Brazil. *Rev Soc Bras Med Trop.* 2010; 43(4):400–4. Epub 2010/08/31. PMID: 20802939.
26. Figueiro-Filho EA, El Beitune P, Queiroz GT, Somensi RS, Morais NO, Dorval ME, et al. Visceral leishmaniasis and pregnancy: analysis of cases reported in a central-western region of Brazil. *Arch Gynecol Obstet.* 2008; 278(1):13–6. Epub 2007/12/19. <https://doi.org/10.1007/s00404-007-0532-0> PMID: 18087708.
27. Pagliano P, Carannante N, Rossi M, Gramiccia M, Gradoni L, Faella FS, et al. Visceral leishmaniasis in pregnancy: a case series and a systematic review of the literature. *J Antimicrob Chemother.* 2005; 55(2):229–33. <https://doi.org/10.1093/jac/dkh538> PMID: 15649998.
28. Maciel DB, Silva TA, Gomes LI, de Oliveira E, Tiburcio MG, de Oliveira RF, et al. Infection with *Leishmania* (*Leishmania*) *infantum* of 0 to 18-Month-old children living in a visceral leishmaniasis-endemic area in Brazil. *Am J Trop Med Hyg.* 2014; 91(2):329–35. Epub 2014/06/18. <https://doi.org/10.4269/ajtmh.13-0418> PMID: 24935952; PubMed Central PMCID: PMC4125257.
29. Mescouto-Borges MR, Maués E, Costa DL, Pranchevicius MC, Romero GA. Congenitally transmitted visceral leishmaniasis: report of two Brazilian human cases. *Braz J Infect Dis.* 2013; 17(2):263–6. Epub 2013/03/05. <https://doi.org/10.1016/j.bjid.2012.10.017> PMID: 23453409.
30. Anderson DC, Buckner RG, Glenn BL, MacVean DW. Endemic canine leishmaniasis. *Vet Pathol.* 1980; 17(1):94–6. Epub 1980/01/01. <https://doi.org/10.1177/030098588001700110> PMID: 7352367.
31. Gaskin AA, Schantz P, Jackson J, Birkenheuer A, Tomlinson L, Gramiccia M, et al. Visceral leishmaniasis in a New York foxhound kennel. *J Vet Intern Med.* 2002; 16(1):34–44. Epub 2002/02/02. PMID: 11822802.
32. Gibson-Corley KN, Hostetter JM, Hostetter SJ, Mullin K, Ramer-Tait AE, Boggiatto PM, et al. Disseminated *Leishmania infantum* infection in two sibling foxhounds due to possible vertical transmission. *The Canadian veterinary journal La revue veterinaire canadienne.* 2008; 49(10):1005–8. PMID: 19119370; PubMed Central PMCID: PMC2553493.
33. Duprey ZH, Steurer FJ, Rooney JA, Kirchhoff LV, Jackson JE, Rowton ED, et al. Canine visceral leishmaniasis, United States and Canada, 2000–2003. *Emerg Infect Dis.* 2006; 12(3):440–6. Epub 2006/05/18. <https://doi.org/10.3201/eid1203.050811> PMID: 16704782.
34. Schantz PM, Steurer FJ, Duprey ZH, Kurpel KP, Barr SC, Jackson JE, et al. Autochthonous visceral leishmaniasis in dogs in North America. *J Am Vet Med Assoc.* 2005; 226(8):1316–22. Epub 2005/04/23. PMID: 15844420.
35. Schaut RG, Robles-Murguía M, Juelsgaard R, Esch KJ, Bartholomay LC, Ramalho-Ortigao M, et al. Vectorborne Transmission of *Leishmania infantum* from Hounds, United States. *Emerg Infect Dis.* 2015; 21(12):2209–12. <https://doi.org/10.3201/eid2112.141167> PMID: 26583260; PubMed Central PMCID: PMC4672406.
36. Weng JL, Young SL, Gordon DM, Claborn D, Petersen C, Ramalho-Ortigao M. First report of phlebotomine sand flies (Diptera: Psychodidae) in Kansas and Missouri, and a PCR method to distinguish *Lutzomyia shannoni* from *Lutzomyia vexator*. *J Med Entomol.* 2012; 49(6):1460–5. Epub 2012/12/29. PMID: 23270176; PubMed Central PMCID: PMC4353249.
37. Toepp AJ, Schaut RG, Scott BD, Mathur D, Berens AJ, Petersen CA. *Leishmania* incidence and prevalence in U.S. hunting hounds maintained via vertical transmission. *Veterinary Parasitology: Regional Studies and Reports.* 2017; 10:75–81. <https://doi.org/10.1016/j.vprsr.2017.08.011>.
38. Coura-Vital W, Marques MJ, Veloso VM, Roatt BM, Aguiar-Soares RD, Reis LE, et al. Prevalence and factors associated with *Leishmania infantum* infection of dogs from an urban area of Brazil as identified by molecular methods. *PLoS Negl Trop Dis.* 2011; 5(8):e1291. <https://doi.org/10.1371/journal.pntd.0001291> PMID: 21858243.
39. Dietz K. The estimation of the basic reproduction number for infectious diseases. *Stat Methods Med Res.* 1993; 2(1):23–41. Epub 1993/01/01. <https://doi.org/10.1177/096228029300200103> PMID: 8261248.
40. Hasibeder G, Dye C, Carpenter J. Mathematical modelling and theory for estimating the basic reproduction number of canine leishmaniasis. *Parasitology.* 1992; 105 (Pt 1):43–53. Epub 1992/08/01. PMID: 1437275.

41. Dye C. The logic of visceral leishmaniasis control. *Am J Trop Med Hyg.* 1996; 55(2):125–30. Epub 1996/08/01. PMID: [8780448](#).
42. Zou L, Chen J, Ruan S. Modeling and analyzing the transmission dynamics of visceral leishmaniasis. *Math Biosci Eng.* 2017; 14(5–6):1585–604. Epub 2017/11/23. <https://doi.org/10.3934/mbe.2017082> PMID: [29161877](#).
43. Courtenay O, Kovacic V, Gomes PA, Garcez LM, Quinnell RJ. A long-lasting topical deltamethrin treatment to protect dogs against visceral leishmaniasis. *Med Vet Entomol.* 2009; 23(3):245–56. <https://doi.org/10.1111/j.1365-2915.2009.00815.x> PMID: [19712155](#).
44. Gavani AS, Hodjati MH, Mohite H, Davies CR. Effect of insecticide-impregnated dog collars on incidence of zoonotic visceral leishmaniasis in Iranian children: a matched-cluster randomised trial. *Lancet.* 2002; 360(9330):374–9. Epub 2002/09/21. S0140673602096095 [pii]. PMID: [12241778](#).
45. Maroli M, Mizzon V, Siragusa C, D'Oorazi A, Gradoni L. Evidence for an impact on the incidence of canine leishmaniasis by the mass use of deltamethrin-impregnated dog collars in southern Italy. *Med Vet Entomol.* 2001; 15(4):358–63. PMID: [11776454](#).
46. Rosypal AC, M. ZA, S. LD. Canine visceral leishmaniasis and its emergence in the United States. *Vet Clin Small Anim.* 2003; 33:921–37.
47. Larson M, Toepp A, Scott B, Epid Kurtz M, Fowler H, et al. Semi-quantitative measurement of asymptomatic *L. infantum* infection and symptomatic visceral leishmaniasis in dogs using Dual-Path Platform (R) CVL. *Appl Microbiol Biotechnol.* 2017; 101(1):381–90. Epub 2016/11/01. <https://doi.org/10.1007/s00253-016-7925-6> PMID: [27796441](#).
48. Toepp AJ, Schaut RG, Scott BD, Mathur D, Berens AJ, Petersen CA. *Leishmania* incidence and prevalence in U.S. hunting hounds maintained via vertical transmission. *Veterinary Parasitology: Regional Studies and Reports.* 2017; 10(Supplement C):75–81. <https://doi.org/10.1016/j.vprsr.2017.08.011>.
49. Vida B, Toepp A, Schaut RG, Esch KJ, Juelsgaard R, Shimak RM, et al. Immunologic progression of canine leishmaniasis following vertical transmission in United States dogs. *Vet Immunol Immunopathol.* 2016; 169:34–8. Epub 2016/02/02. <https://doi.org/10.1016/j.vetimm.2015.11.008> PMID: [26827836](#); PubMed Central PMCID: [PMC4799997](#).
50. Boggiatto PM, Ramer-Tait AE, Metz K, Kramer EE, Gibson-Corley K, Mullin K, et al. Immunologic indicators of clinical progression during canine *Leishmania infantum* infection. *Clin Vaccine Immunol.* 2010; 17(2):267–73. Epub 2009/12/25. CVI.00456-09 [pii] <https://doi.org/10.1128/CVI.00456-09> PMID: [20032217](#); PubMed Central PMCID: [PMC2815526](#).
51. Toepp A, Larson M, Grinnage-Pulley T, Bennett C, Anderson M, Parrish M, et al. Safety Analysis of *Leishmania* Vaccine Used in a Randomized Canine Vaccine/Immunotherapy Trial. *Am J Trop Med Hyg.* 2018. Epub 2018/03/08. <https://doi.org/10.4269/ajtmh.17-0888> PMID: [29512486](#).
52. Badaro R, Reed SG, Carvalho EM. Immunofluorescent antibody test in American visceral leishmaniasis: sensitivity and specificity of different morphological forms of two *Leishmania* species. *Am J Trop Med Hyg.* 1983; 32(3):480–4. PMID: [6407345](#).
53. Lenard ZM, Hopper BJ, Lester NV, Richardson JL, Robertson ID. Accuracy of prediction of canine litter size and gestational age with ultrasound. *Aust Vet J.* 2007; 85(6):222–5. Epub 2007/06/06. <https://doi.org/10.1111/j.1751-0813.2007.00162.x> PMID: [17547634](#).
54. Figueiró-Filho EA, El Beitune P, Queiroz GT, Somensi RS, Morais NO, Dorval ME, et al. Visceral leishmaniasis and pregnancy: analysis of cases reported in a central-western region of Brazil. *Arch Gynecol Obstet.* 2008; 278(1):13–6. <https://doi.org/10.1007/s00404-007-0532-0> PMID: [18087708](#).
55. Petersen CA. Leishmaniasis, an emerging disease found in companion animals in the United States. *Top Companion Anim Med.* 2009; 24(4):182–8. Epub 2009/12/01. S1938-9736(09)00074-9 [pii] <https://doi.org/10.1053/j.tcam.2009.06.006> PMID: [19945086](#); PubMed Central PMCID: [PMC2805016](#).
56. Petersen CA, Barr SC. Canine leishmaniasis in North America: emerging or newly recognized? *Vet Clin North Am Small Anim Pract.* 2009; 39(6):1065–74, vi. Epub 2009/11/26. [https://doi.org/10.1016/j.cvsm.2009.06.008\(09\)00100-4](https://doi.org/10.1016/j.cvsm.2009.06.008(09)00100-4) [pii] PMID: [19932363](#); PubMed Central PMCID: [PMC2824922](#).
57. Ouedraogo A, Tiono AB, Diarra A, Bougouma EC, Nebie I, Konate AT, et al. Transplacental Transmission of *Plasmodium falciparum* in a Highly Malaria Endemic Area of Burkina Faso. *J Trop Med.* 2012; 2012:109705. Epub 2011/12/17. <https://doi.org/10.1155/2012/109705> PMID: [22174725](#); PubMed Central PMCID: [PMC3235890](#).
58. Schuster RC, McMahan DE, Young SL. A comprehensive review of the barriers and promoters health workers experience in delivering prevention of vertical transmission of HIV services in sub-Saharan Africa. *AIDS Care.* 2016; 28(6):778–94. Epub 2016/02/18. <https://doi.org/10.1080/09540121.2016.1139041> PMID: [26883903](#); PubMed Central PMCID: [PMC4978422](#).
59. de Andrade SD, Sabido M, Marcelo Monteiro W, Canellas L, Prazeres V, Schwartz Benzaken A. Mother-to-child Transmission of HIV From 1999 to 2011 in the Amazonas, Brazil: Risk Factors and

- Remaining Gaps in Prevention Strategies. *Pediatr Infect Dis J.* 2016; 35(2):189–95. Epub 2015/10/21. <https://doi.org/10.1097/INF.0000000000000966> PMID: 26484428.
60. Poespoprodjo JR, Fobia W, Kenangalem E, Hasanuddin A, Sugiarto P, Tjitra E, et al. Highly effective therapy for maternal malaria associated with a lower risk of vertical transmission. *J Infect Dis.* 2011; 204(10):1613–9. Epub 2011/09/13. <https://doi.org/10.1093/infdis/jir558> PMID: 21908728; PubMed Central PMCID: PMC3192188.
 61. Sudarshan M, Singh T, Chakravarty J, Sundar S. A Correlative Study of Splenic Parasite Score and Peripheral Blood Parasite Load Estimation by Quantitative PCR in Visceral Leishmaniasis. *J Clin Microbiol.* 2015; 53(12):3905–7. Epub 2015/09/25. <https://doi.org/10.1128/JCM.01465-15> PMID: 26400788; PubMed Central PMCID: PMC4652099.
 62. Biggerstaff M, Cauchemez S, Reed C, Gambhir M, Finelli L. Estimates of the reproduction number for seasonal, pandemic, and zoonotic influenza: a systematic review of the literature. *BMC Infect Dis.* 2014; 14:480. Epub 2014/09/05. <https://doi.org/10.1186/1471-2334-14-480> PMID: 25186370; PubMed Central PMCID: PMC4169819.
 63. Gani R, Leach S. Transmission potential of smallpox in contemporary populations. *Nature.* 2001; 414(6865):748–51. Epub 2001/12/14. <https://doi.org/10.1038/414748a> PMID: 11742399.
 64. Hailu T, Yimer M, Mulu W, Abera B. Challenges in visceral leishmaniasis control and elimination in the developing countries: A review. *J Vector Borne Dis.* 2016; 53(3):193–8. Epub 2016/09/30. PMID: 27681541.
 65. Mondal D, Das ML, Kumar V, Huda MM, Das P, Ghosh D, et al. Efficacy, Safety and Cost of Insecticide Treated Wall Lining, Insecticide Treated Bed Nets and Indoor Wall Wash with Lime for Visceral Leishmaniasis Vector Control in the Indian Sub-continent: A Multi-country Cluster Randomized Controlled Trial. *PLoS Negl Trop Dis.* 2016; 10(8):e0004932. Epub 2016/08/18. <https://doi.org/10.1371/journal.pntd.0004932> PMID: 27533097; PubMed Central PMCID: PMC4988640.
 66. Courtenay O, Quinell RJ, Garcez LM, Shaw JJ, Dye C. Infectiousness in a cohort of Brazilian dogs: why culling fails to control visceral leishmaniasis in areas of high transmission. *J Infect Dis.* 2002; 186(9):1314–20. Epub 2002/10/29. <https://doi.org/10.1086/344312> PMID: 12402201.
 67. Roatt BM, Aguiar-Soares RD, Reis LE, Cardoso JM, Mathias FA, de Brito RC, et al. A Vaccine Therapy for Canine Visceral Leishmaniasis Promoted Significant Improvement of Clinical and Immune Status with Reduction in Parasite Burden. *Front Immunol.* 2017; 8:217. Epub 2017/03/23. <https://doi.org/10.3389/fimmu.2017.00217> PMID: 28321217; PubMed Central PMCID: PMC5338076.