RESEARCH ARTICLE

Safety and pharmacokinetics of single, dual, and triple antiretroviral drug formulations delivered by pod-intravaginal rings designed for HIV-1 prevention: A Phase I trial

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Abstract

Background

Intravaginal rings (IVRs) for HIV pre-exposure prophylaxis (PrEP) theoretically overcome some adherence concerns associated with frequent dosing that can occur with oral or vaginal film/gel regimens. An innovative pod-IVR, composed of an elastomer scaffold that can hold up to 10 polymer-coated drug cores (or “pods”), is distinct from other IVR designs as drug release from each pod can be controlled independently. A pod-IVR has been developed for the delivery of tenofovir (TFV) disoproxil fumarate (TDF) in combination with emtricitabine (FTC), as daily oral TDF-FTC is the only Food and Drug Administration (FDA)-approved regimen for HIV PrEP. A triple combination IVR building on this platform and delivering TDF-FTC along with the antiretroviral (ARV) agent maraviroc (MVC) also is under development.

Methodology and findings

This pilot Phase I trial conducted between June 23, 2015, and July 15, 2016, evaluated the safety, pharmacokinetics (PKs), and acceptability of pod-IVRs delivering 3 different ARV regimens: 1) TDF only, 2) TDF-FTC, and 3) TDF-FTC-MVC over 7 d. The crossover, open-
Policy and the authors of this manuscript have the following competing interests: During part of the time of the study, IB, SAC, and JMC played a role in study design but had no role in data collection or analysis (with the possible exception pertaining to MG in the competing interests disclosure) or manuscript preparation.

MG was a 50% shared employee of Auritec and participated in the residual data collection or analysis (with the possible exception pertaining to MG in the competing interest disclosure) or manuscript preparation.

Auritec Pharmaceuticals, Inc. was the sponsor of the study and prime recipient of the NIH grant (Eunice Kennedy Shriver National Institute of Child Health & Human Development of the National Institutes of Health under Award Number R44HD075636) that funded part of the study. Study materials (e.g., IVRs) were provided to UTMB (clinical site) by Oak Crest in collaboration with Auritec under that Award; however, there was no direct funding provided by Auritec. Auritec employed the only FDA-approved PrEP combination regimen can be advanced to safety and efficacy testing. An alternate, more adherence-independent, longer-acting prevention device based on oral dosing, suggesting that efficacy for vaginal HIV PrEP is achievable. These results show that an alternate, more adherence-independent, longer-acting prevention device based on the only FDA-approved PrEP combination regimen can be advanced to safety and efficacy testing.

**Conclusions**

An innovative pod-IVR delivery device with 3 different formulations delivering different regimens of ARV drugs vaginally appeared to be safe and acceptable and provided drug concentrations in CVFs and tissues exceeding concentrations achieved by highly protective oral dosing, suggesting that efficacy for vaginal HIV PrEP is achievable. These results show that an alternate, more adherence-independent, longer-acting prevention device based on the only FDA-approved PrEP combination regimen can be advanced to safety and efficacy testing.

**Trial registration**

ClinicalTrials.gov NCT02431273

**Author summary**

**Why was this study done?**

- Pre-exposure Prophylaxis (PrEP) for HIV prevention has been shown to be effective in highly adherent users.
Preclinical studies of an innovative pod-intravaginal ring (IVR) design delivering antiretroviral (ARV) regimens have shown protection against Simian Human Immunodeficiency Virus (SHIV) in nonhuman primates.

The goal of this clinical study was to assess safety, pharmacokinetics (PKs), and acceptability of pod-IVRs delivering 3 different ARV regimens: 1) tenofovir disoproxil fumarate (TDF) only, 2) TDF and emtricitabine (FTC), and 3) TDF, FTC, and maraviroc (MVC) via the vaginal route.

What did the researchers do and find?

A 7-d vaginal ring study was performed, sequentially delivering TDF, TDF-FTC, and TDF-FTC-MVC in 6 women to determine safety and PKs.

There were no concerning safety findings that would inhibit further development of the vaginal rings.

Vaginal secretion and tissue levels were detected in ranges that have been associated with protection against HIV/SHIV infection in preclinical and clinical studies.

What do these findings mean?

Vaginal delivery of combinations of ARV drugs, including the only Centers for Disease Control (CDC)-recommended regimen for HIV PrEP, was achieved in this early Phase I clinical study.

The interventions appeared safe and acceptable and therefore merit further study.

Introduction

Scale-up of prevention and treatment efforts to curb the HIV epidemic have resulted in decreasing the number of new infections by half per year in 2012 since the peak in 1996 [1]. In Fast-Track, the Joint United Nations Programme on HIV/AIDS (UNAIDS) has set aggressive goals, including 500,000 (or fewer) new annual infections by 2020, a 75% reduction from 2010 numbers, and an end to the AIDS epidemic by 2030 [2]. Unfortunately, the number of annual, new HIV infections has stalled around 1.9 million since 2010, suggesting that a prevention gap has been reached [3]. To meet these ambitious UNAIDS Fast-Track goals, further work is needed to develop new highly effective strategies for HIV prevention.

Pre-exposure prophylaxis (PrEP) using Food and Drug Administration (FDA)-approved antiretroviral (ARV) drugs holds significant promise as a strategy for preventing HIV infection. Multiple HIV PrEP clinical trials have demonstrated that vaginal and oral ARV regimens based on the nucleoside reverse transcriptase inhibitor (NRTI) tenofovir (TFV), alone or in combination with the NRTI emtricitabine (FTC), can be effective in susceptible men, women, and partners of HIV-infected individuals [4–11]. Oral tenofovir disoproxil fumarate (TDF)-FTC (Truvada, Gilead Sciences, Foster City, CA) is the only FDA-approved regimen for HIV...
PrEP [12]. Centre for the AIDS Programme of Research in South Africa (CAPRISA)-004 demonstrated that topical dosing—in this case pericoitally with a 1% TFV gel—can be effective in preventing vaginal HIV transmission [4]. In post hoc analyses of both Vaginal and Oral Interventions to Control the Epidemic (VOICE) gel and Follow-on African Consortium for Tenofovir Studies (FACTS) 001 trials, TFV gel was effective in highly adherent women [13]. A critical factor driving success in these trials appears to involve sustaining high adherence to frequent dosing [14].

Treatment of any medical condition that requires long-term use of medications must address the challenge to maintain adherence to therapy; adherence has been shown to be inversely related to dosing frequency regardless of dosing formulation [15–18]. Use of extended-release or long-acting formulations, including intravaginal rings (IVRs) [19], implantable devices, and injectable formulations [20], can decrease the burden of frequent dosing and potentially improve adherence. Two Phase 3, randomized, double-blind, placebo-controlled trials (MTN-020–A Study to Prevent Infection with a Ring for Extended Use (ASPIRE) and IPM 027–The Ring Study) recently evaluated a monthly IVR delivering the non-nucleoside HIV reverse-transcriptase inhibitor dapivirine (DPV) [21,22]. The trials enrolled 2,629 and 1,959 women, respectively, between the ages of 18 and 45 years in Malawi, South Africa, Uganda, and Zimbabwe and demonstrated that an IVR delivering an ARV agent can reduce the risk of HIV acquisition by as much as 56% in highly adherent users, as determined through quarterly plasma and residual IVR DPV concentrations.

User adherence to IVRs depends on a number of factors including user sensory perceptions and experiences of product characteristics and impact such as dimensionality and perceptions of increased discharge [23]. Users ascribe meaning to product characteristics, and those meanings can influence perceptions of product efficacy and the user’s willingness to engage in consistent and correct product use [23,24].

The primary purpose of this exploratory, open label clinical trial was to evaluate the safety, pharmacokinetics (PKs), and user perceptibility of an innovative IVR design (termed “pod-IVR”) [25–27] delivering three formulations: first, TDF alone over 7 d; second, TDF-FTC over 7 d in a crossover design; and third, TDF-FTC-maraviroc (MVC) over 7 d. The pod-IVR consists of a silicone scaffold that holds up to 10 individual "pods" of polymer-coated drug cores, allowing independently controlled drug release from each pod through delivery channels.

**Materials and methods**

**Ethics statement**

All human research was approved by the University of Texas Medical Branch Institutional Review Board (IRB #14–0479), conducted according to the Declaration of Helsinki, and registered in clinicaltrials.gov (NCT02431273). All participants provided written informed consent.

**Clinical trial design**

Women were recruited through announcements to use IVRs releasing TDF, TDF-FTC, and TDF-FTC-MVC for 7 d each between June 23, 2015, and July 15, 2016. Women were prescreened to confirm basic eligibility and then scheduled for a screening visit where inclusion/exclusion criteria were reviewed, medical and sexual history were obtained, and baseline labs with blood counts, liver and kidney function, and sexually transmitted infection (STI) and HIV screening were collected. Inclusion criteria were women aged 18–45 with regular or suppressed menses, on contraception, and willing to abstain from sexual intercourse during IVR use. Women with STIs, HIV, Hepatitis B, abnormal screening labs, allergy to study product, currently using an IVR, or at high risk for HIV were excluded.
The trial consisted of an open-label, crossover design of a pod-IVR with 3 different ARV regimens. There were 3 separate 7-d treatment periods (Treatment Period 1, TDF-only pod-IVR; Treatment Period 2, TDF-FTC pod-IVR; and Treatment Period 3, TDF-FTC-MVC pod-IVR) with 6 participants per treatment period. Each participant used each pod-IVR for 7 d with a washout period of at least 14 d between each treatment period. Progression to the next treatment period was contingent upon the absence of grade 3 or 4 genitourinary adverse events (AEs) considered to be drug related by the investigator or other investigator-assessed drug-related serious adverse event (SAE). There was a 3-mo pause and re-consent between the second and third treatment periods to allow for determination of safety and PKs of the combination IVR before proceeding to the use of the TDF-FTC-MVC pod-IVR in the third treatment period. The study design was chosen with the ultimate goal of developing a TDF-FTC and TDF-FTC-MVC pod-IVR; however, the FDA required assessment of a TDF-only pod-IVR prior to use of the TDF-FTC and TDF-FTC-MVC formulations.

Following the screening visit, women who were clinically deemed eligible returned to the clinic for IVR insertion (Visit 1, TDF IVR; Visit 5, TDF-FTC IVR) after cessation of menses. Women were instructed on vaginal ring placement, which was performed during this visit, and were asked to be abstinent during IVR use. They returned for Visit 2 (TDF IVR) or Visit 6 (TDF-FTC IVR) on Day 2 (± 1 d) after IVR insertion and for Visit 3 (TDF IVR) or Visit 7 (TDF-FTC IVR) on Day 7 (± 1 d) when the IVRs were removed. Women returned for follow-up 1–2 wk after IVR removal for Visit 4 (TDF IVR) or Visit 8 (TDF-FTC IVR).

The TDF-FTC-MVC pod-IVR arm was carried out separately from the TDF and TDF-FTC IVR portion of the study. Women who were deemed clinically eligible returned to the clinic for IVR insertion (Visit 1) after cessation of menses, instructed on vaginal ring placement (which was performed during this visit), and asked to be abstinent during IVR use. Women returned for Visit 2 on Day 2 (± 1 d) after IVR insertion and for Visit 3 on Day 7 (± 1 d) when the IVRs were removed. Women returned for follow-up 1–2 wk after IVR removal for Visit 4.

During all visits, AEs were reviewed and colposcopy was performed to evaluate the vagina and cervix for safety assessments. For PK assessments, blood (plasma) and cervicovaginal samples (cervicovaginal fluid [CVF], Dacron swabs; cervicovaginal lavage [CVL], 2.5 mL sterile phosphate-buffered saline solution containing 1 mM LiCl) were collected. In summary, for PK assessments, two samples were collected during pod-IVR use (Visit 2/6/2 and Visit 3/7/3) and one sample after pod-IVR removal (Visit 4/8/4). Additionally, at Visit 3/7/3, the IVRs were collected and vaginal biopsies were obtained. One biopsy was flash-frozen in liquid nitrogen for the analysis of ARV drug concentrations. A second vaginal biopsy was fixed in formalin and hematoxylin–eosin stained. The slides were reviewed by a trained pathologist with expertise in reproductive tract pathology.

User perception and product acceptability

Behavioral assessments of perceptibility, acceptability, and adherence were obtained through daily diaries during IVR use, surveys completed using computer-assisted self-interview (CASI) format at the time of IVR removal, and qualitative interviews conducted at Visit 4/Visit 8/Visit 4. Baseline demographics and sexual history surveys were completed prior to product initiation. A user sensory perception and experience (USPE) survey, acceptability and adherence measures, and an in-depth interview were completed following IVR use.

Chemicals and reagents

TDF and FTC labeled for human use were purchased from commercial vendors with a Drug Master File (DMF) registered with the FDA. MVC was isolated from the commercial...
formulation (Pfizer, Inc., New York, NY), which consists of film-coated tablets for oral administration containing 300 mg of MVC and inactive ingredients, as described previously [26]. All other reagents were obtained from Sigma-Aldrich, unless otherwise noted.

Fabrication of pod-intravaginal rings

Polydimethylsiloxane ([PDMS], silicone) pod-IVRs were fabricated in a multistep process that has been described in detail elsewhere [25,26,28,29], and only a brief description is provided here. Each pod contained a single drug. The drug powder was compacted into cores of 3.2-mm outer diameter in a manual tablet press (MTCM-I; Globe Pharma, New Brunswick, NJ). Drug cores were coated with poly(vinyl alcohol) to yield pods, which were placed in the corresponding IVR cavities and sealed in place by back-filling with room-temperature-cure silicone. Each pod was matched with the appropriate configuration of mechanically punched delivery channels. The IVR drug loadings were as follows: TDF, 180 mg; FTC, 125 mg; MVC, 90 mg. Residual drug content of used IVRs was measured by high-performance liquid chromatography (HPLC) according to published methods [26,28] and used to calculate in vivo release rates.

Safety measures

AEs were reviewed at all study visits along with colposcopy. Histology was performed on VT samples collected on Day 7 (Visit 3/7/3). Vaginal pH and Nugent scores [30] were measured at each study visit. Vaginal microbial community profiles also were measured at each study visit by a custom quantitative polymerase chain reaction (qPCR) array described previously [31]. The array targets 46 distinct key vaginal microbiota to the species level along with several housekeeping genes.

Bioanalysis of in vivo samples

Concentrations of TDF, TFV, tenofovir diphosphate (TFV-DP), FTC, and MVC were determined via previously described liquid chromatographic-tandem mass spectrometric (LC-MS/MS) assays [32–35]. All assays were developed and validated following the FDA Guidance for Industry, Bioanalytical Method Validation recommendations and met all acceptability criteria [36]. Isotopically labeled internal standards were used for all compounds and the determination of drug concentrations in all specimen sources.

PK analysis

PK parameter values for CVF were determined by noncompartmental analysis (NCA) using Phoenix WinNonlin 6.4 (Pharsight Corporation, Sunnyvale, CA). The NCA was run using the
linear trapezoidal rule for increasing concentration data and the logarithmic trapezoidal rule for decreasing concentration data (linear up and log down) as the calculation method. Post-dose concentrations below the corresponding lower limit of quantification (LLOQ) \( (C_{\text{LLOQ}}) \) were imputed as follows:

\[
C_{\text{LLOQ}} = \frac{\text{LLOQ of assay}}{2 \times \text{median sample mass}}
\]  

(1)

Data analysis

Qualitative data were analyzed by applying study-specific thematic identification and summarizing verbatim transcript data per theme, with illustrative quotes retained as conventional [38]. Quantitative data were analyzed using GraphPad Prism (version 7.00, GraphPad Software, Inc., La Jolla, CA). Statistical significance is defined as two-sided \( P < 0.05 \). The unpaired, nonparametric Mann–Whitney test was used to compare two groups, including vaginal pH, Nugent scores, in vivo TDF release rates, CVF FTC concentrations in the TDF-FTC and TDF-FTC-MVC pod-IVR treatment periods and terminal half-lives of ARV drug elimination from CVF within one IVR treatment period and across IVR treatment periods. The nonparametric (i.e., do not assume a Gaussian distribution) Kruskal–Wallis tests with no matching/pairing of the data was used to compare three IVRs, including CVF TDF and TFV concentrations and CVF:VT concentration ratios and terminal half-lives of ARV drug elimination from CVF within one IVR arm.

Results

Study participants

Six participants were enrolled and completed the first 2 treatment periods using the TDF-only and TDF-FTC pod-IVRs. Due to the delay between treatment periods 2 and 3, 4 participants dropped out and were replaced for treatment period 3. Demographics for the initial 6 study participants who completed the first 2 treatment periods (i.e., TDF-only pod-IVR and TDF-FTC pod-IVR) can be found in Table 1, and demographics for the final treatment period (i.e., TDF-FTC-MVC pod-IVR) can be found in Table 2. There were no missed study visits, and one participant (ID 479–12) had an additional visit (Visit 2A, TDF pod-IVR arm) with visits on Day 1 (Visit 2) and Day 2 (Visit 2A) for evaluation of AEs (see Safety measures below).

| Participants enrolled, \( n \) (percent) | 6 (100) |
| Age (years), median (range) | 26 (24–35) |
| Race and Ethnicity, \( n \) (percent) | |
| Black or African American | 2 (33) |
| White | 4 (67) |
| Hispanic | 2 (33) |
| Non-Hispanic | 2 (33) |
| Asian | 0 (0) |
| Other | 0 (0) |
| Tobacco use, \( n \) (percent) | 1 (17) |
| Alcohol use, \( n \) (percent) | 4 (67) |
| BMI (kg/m\(^2\)), median (range) | 32.4 (27.2–44.2) |

Abbreviation: BMI, body mass index.

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Safety measures

The pod-IVRs were safe and generally well tolerated in all three treatment periods. AEs were recorded with daily diaries and during study visits. There were no concerning safety findings by participant report, examination with colposcopy, evaluation of vaginal microbiome (VMB), or histology from vaginal biopsy. There were no SAEs or grade 3 or 4 genitourinary AEs. Fifty-eight AEs occurred during the study period (54 Grade 1, 4 Grade 2; Tables 3–5). All AEs were Grade 1 except for four Grade 2 findings. Two of the Grade 2 events were pelvic pain/cramping, for which the subjects took over-the-counter medication and were determined to be “possibly related.” One woman had a Grade 2 candida vaginitis determined to be “possibly related,” which did not recur after treatment. One woman reported having Grade 2 diarrhea, which was determined to be “not related” and attributed to “food poisoning.”

The following is a description of findings during IVR use. Thirty-eight AEs occurred during use of the IVRs, 17 during TDF IVR use (Table 3), 9 during TDF-FTC use (Table 4), and 12 during TDF-FTC-MVC use (Table 5). On colposcopy, cervicovaginal erythema was found in 3 women (6 findings) during IVR use, 2 of whom had erythema at baseline. During IVR use, 6 women had pelvic pain/cramping (13 findings). Four women reported vaginal discharge (7

### Table 2. Demographics of study participants completing the 3rd treatment period.

<table>
<thead>
<tr>
<th>Participants enrolled, n (percent)</th>
<th>6 (100)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age (years), median (range)</td>
<td>24.5 (21–41)</td>
</tr>
<tr>
<td>Race and Ethnicity, n (percent)</td>
<td></td>
</tr>
<tr>
<td>Black or African American</td>
<td>2 (33)</td>
</tr>
<tr>
<td>White</td>
<td>4 (67)</td>
</tr>
<tr>
<td>Hispanic</td>
<td>1 (17)</td>
</tr>
<tr>
<td>Non-Hispanic</td>
<td>3 (50)</td>
</tr>
<tr>
<td>Asian</td>
<td>0 (0)</td>
</tr>
<tr>
<td>Other</td>
<td>0 (0)</td>
</tr>
<tr>
<td>Tobacco use, n (percent)</td>
<td>1 (17)</td>
</tr>
<tr>
<td>Alcohol use, n (percent)</td>
<td>5 (83)</td>
</tr>
<tr>
<td>BMI (kg/m²), median (range)</td>
<td>28.4 (23.8–44.6)</td>
</tr>
</tbody>
</table>

**Abbreviation:** BMI, body mass index.

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### Table 3. AEs during TDF pod-IVR use.

<table>
<thead>
<tr>
<th>AE</th>
<th>Number of Participants experiencing AEs</th>
<th>Number of AEs reported</th>
<th>Grade of AE</th>
<th>Relatedness to IVR use</th>
</tr>
</thead>
<tbody>
<tr>
<td>Pelvic Pain</td>
<td>3</td>
<td>5</td>
<td>Grade 1 (n = 4)*</td>
<td>Possibly Related</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>Grade 2 (n = 1)**</td>
<td></td>
</tr>
<tr>
<td>Vaginal Discharge</td>
<td>3</td>
<td>3</td>
<td>Grade 1 (all)</td>
<td>Possibly Related</td>
</tr>
<tr>
<td>Metrorrhagia</td>
<td>1</td>
<td>1</td>
<td>Grade 1</td>
<td>Possibly Related</td>
</tr>
<tr>
<td>Vulvovaginal Itching</td>
<td>3</td>
<td>3</td>
<td>Grade 1 (all)</td>
<td>Possibly Related</td>
</tr>
<tr>
<td>Cervicovaginal Erythema</td>
<td>3</td>
<td>3</td>
<td>Grade 1 (all)</td>
<td>Possibly Related</td>
</tr>
<tr>
<td>Diarrhea</td>
<td>1</td>
<td>1</td>
<td>Grade 1</td>
<td>Possibly Related</td>
</tr>
<tr>
<td>Malaise</td>
<td>1</td>
<td>1</td>
<td>Grade 1</td>
<td>Not Related</td>
</tr>
</tbody>
</table>

*Grade 1 AEs are mild and cause no or minimal interference with activity.
**Grade 2 AEs are controlled with medication or cause greater than minimal interference with activity. The AE was considered Grade 2 because the subject used over-the-counter medication for treatment.

**Abbreviations:** AE, adverse event; IVR, intravaginal ring; TDF, tenofovir disoproxil fumarate.

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findings); one episode was a mucus-like discharge that appeared peri-ovulatory that was deemed “possibly related.” Three women had metrorrhagia/intermenstrual bleeding during IVR use (3 findings), one during TDF IVR use and two during TDF-FTC-MVC use. Three women had vulvovaginal itching (3 findings). Two women were diagnosed with candida vaginitis (one Grade 1 and one Grade 2) and were treated. One woman reported an odor (1 finding) on Day 7 of TDF-FTC-MVC IVR use. One woman reported nausea (1 finding) associated with cramping which lasted only 10 min during TDF-FTC-MVC IVR use. One woman had Grade 1 diarrhea during TDF-only IVR use, which was “possibly related.” One woman had malaise after influenza vaccine during TDF-only IVR use, and this was considered “not related.”

With two exceptions, all biopsies were reported as “consistent with a normal vaginal epithelium.” For one subject in the TDF pod-IVR treatment period, there was one section of the vaginal biopsy with minimal to mild inflammatory infiltrates and necrosis, which may be consistent with an infection (she had asymptomatic Candida infection determined by qPCR); a recut from the same specimen showed normal tissue. In the TDF-FTC-MVC pod-IVR treatment period, there was a finding of minimal inflammatory infiltrate and hemorrhage in the subject who had the Grade 2 symptomatic Candida infection (Table 5).

In all 3 treatment periods, the impact of the pod-IVRs on the vaginal microbial community profiles was analyzed using custom qPCR arrays targeting the most common 46 vaginal bacteria at genus or species levels [31]. A second array was used to quantify the levels of 16 additional pathogen targets, including bacteria and viruses common to the vagina. Collectively, the

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**Table 4. AEs during TDF-FTC pod-IVR use.**

<table>
<thead>
<tr>
<th>AE</th>
<th>Number of Participants experiencing AEs</th>
<th>Number of AEs reported</th>
<th>Grade of AE</th>
<th>Relatedness to IVR use</th>
</tr>
</thead>
<tbody>
<tr>
<td>Pelvic Pain</td>
<td>2</td>
<td>3</td>
<td>Grade 1 (n = 1)*</td>
<td>Possibly Related</td>
</tr>
<tr>
<td>Vaginal Discharge</td>
<td>3</td>
<td>3</td>
<td>Grade 1 (all)</td>
<td>Possibly Related</td>
</tr>
<tr>
<td>Cervicovaginal Erythema</td>
<td>1</td>
<td>3</td>
<td>Grade 1 (all)</td>
<td>Possibly Related</td>
</tr>
</tbody>
</table>

*Grade 1 AEs are mild and cause no or minimal interference with activity.

**Grade 2 AEs are controlled with medication or cause greater than minimal interference with activity. The AE was considered Grade 2 because the subject used over-the-counter medication for treatment.

**Abbreviations:** AE, adverse event; FTC, emtricitabine; IVR, intravaginal ring; TDF, tenofovir disoproxil fumarate.

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**Table 5. AEs during TDF-FTC-MVC pod-IVR use.**

<table>
<thead>
<tr>
<th>AE</th>
<th>Number of Participants experiencing AEs</th>
<th>Number of AEs reported</th>
<th>Grade of AE</th>
<th>Relatedness to IVR use</th>
</tr>
</thead>
<tbody>
<tr>
<td>Pelvic Pain</td>
<td>4</td>
<td>5</td>
<td>Grade 1</td>
<td>Possibly Related</td>
</tr>
<tr>
<td>Vaginal Discharge</td>
<td>3</td>
<td>3</td>
<td>Grade 1</td>
<td>Possibly Related</td>
</tr>
<tr>
<td>Metrorrhagia</td>
<td>2</td>
<td>2</td>
<td>Grade 1</td>
<td>Possibly Related</td>
</tr>
<tr>
<td>Vulvovaginal Itching</td>
<td>1</td>
<td>1</td>
<td>Grade 1</td>
<td>Possibly Related</td>
</tr>
<tr>
<td>Odor</td>
<td>1</td>
<td>1</td>
<td>Grade 1</td>
<td>Possibly Related</td>
</tr>
<tr>
<td>Nausea</td>
<td>1</td>
<td>1</td>
<td>Grade 1</td>
<td>Possibly Related</td>
</tr>
<tr>
<td><em>Candida</em> (monilial vulvovaginitis)</td>
<td>1</td>
<td>1</td>
<td>Grade 2**</td>
<td>Possibly Related</td>
</tr>
</tbody>
</table>

*Grade 1 AEs are mild and cause no or minimal interference with activity.

**Grade 2 AEs are controlled with medication or cause greater than minimal interference with activity. The AE was considered Grade 2 because the subject had moderate symptoms.

**Abbreviations:** AE, adverse event; FTC, emtricitabine; IVR, intravaginal ring; MVC, maraviroc; TDF, tenofovir disoproxil fumarate.

https://doi.org/10.1371/journal.pmed.1002655.t005
results show no clear, systematic impact of the pod-IVRs on the stability of the microbial community profiles for the trial participants (S1–S3 Figs). One subject (ID 479–16) had asymptomatic *Candida albicans* noted only by qPCR (saline microscopy with potassium hydroxide [KOH] was negative) during the study.

The median vaginal pH prior to IVR placement (Visits 0 and 1) was 4.0 (IQR, 4.0–4.5), and with the TDF and TDF-FTC pod-IVRs in place was 4.5 (IQR, 4.1–4.5) and 4.0 (IQR, 4.0–4.5), respectively. There was no statistically significant difference in vaginal pH prior to pod-IVR placement compared to during pod-IVR use: TDF pod-IVR, \( P = 0.3405 \); TDF-FTC pod-IVR, \( P = 0.5882 \). The median Nugent score prior to IVR placement (Visits 0 and 1) was 3.0 (IQR, 3.0–3.3) and with the TDF and TDF-FTC pod-IVRs in place was 2.5 (IQR, 2.0–4.0) and 4.5 (IQR, 2.8–5.3), respectively. There was no statistically significant difference in Nugent score prior to pod-IVR placement compared to pod-IVR: TDF pod-IVR, \( P = 0.6068 \); TDF-FTC pod-IVR, \( P = 0.5611 \).

The median vaginal pH prior to IVR placement (Visits 0 and 1) was 4.1 (IQR, 4.1–4.5) and with the TDF-FTC-MVC pod-IVRs in place was 4.4 (IQR, 4.1–4.5). There was no statistically significant difference (0.3775) in vaginal pH prior to pod-IVR placement compared to during pod-IVR use. The median Nugent score prior to IVR placement (Visits 0 and 1) was 4.0 (IQR, 2.8–5.3) and with pod-IVRs in place was 5.0 (IQR, 4.0–5.0). These findings could be attributed to the prevalence of *Gardnerella vaginalis* in some of the participants’ VMBs (S3 Fig). There was no statistically significant difference (\( P = 0.5858 \)) in Nugent score prior to pod-IVR placement compared to during pod-IVR.

**User perception and product acceptability**

All participants in the TDF and TDF-FTC treatment periods were willing to recommend the ring to others, and 4 of 6 would “probably” or “definitely” use the pod-IVR for HIV prevention. The remaining 2 expressed no perceived risk of HIV acquisition. Participants’ confidence in their ability to insert and remove the IVR either held constant between using the first and second study rings or improved. Participants’ willingness to use the pod-IVR for 28 d and access the ring for “real world” use was high. For example, after using the first ring, participants averaged 3.5 (on a 1–5 Likert scale from “not at all confident” to “completely confident”) that she could use the ring for 28 d, while the average increased to 4.0 after using the second ring. Qualitative interview data revealed that, overall, the pod-IVRs were well tolerated by participants. Prior to insertion of the pod-IVR, clinicians reviewed verbal instructions with the participants, which reportedly allowed women to feel more confident in their ability to insert the pod-IVR on their own: clinician’s checking pod-IVR placement on first insertion also led to greater confidence in some. While 2 of 6 women felt the TDF-FTC pod-IVR was more difficult to insert than the TDF pod-IVR, the majority of participants felt both pod-IVRs were flexible and easy to insert. The majority of women noted that they did not feel the pod-IVR in their vaginas at all during the use period. Of note, psychological awareness of the pod-IVR was reported by one woman, which subsided after a few days. Lack of physical awareness of the pod-IVRs allowed most participants to continue their typical physical activity routines during the weeks of use, with one exception (i.e., an elite runner). All participants felt that the pod-IVRs had no impact on their typical hygiene practices. All participants were able to successfully remove the pod-IVRs, with 3 of 6 women reporting difficulty initially locating the TDF pod-IVRs in their vaginas.

All 6 women from the TDF-FTC-MVC pod-IVR study group reported that they would use the device if it were a 7-d HIV prevention ring; 5 of 6 said they would use it if it were a 28-d device. All 6 were willing to recommend the IVR to others. With respect to use behaviors, all
reported being highly confident that they could insert the IVR correctly. The two participants who previously used the TDF and TDF-FTC pod-IVRs noted that inserting the TDF-FTC-MVC pod-IVR was much easier than the first two pod-IVRs they had used, because they were familiar with the insertion process and confident they could insert the device correctly. Of the 6 women who used the TDF-FTC-MVC pod-IVR, none reported concerns with being able to remove the device; ultimately, all thought removing it was easy and were able to do so successfully. When asked about preferred access should the IVR become available, 3 of the women would use the pod-IVR if accessed by prescription, and 4 would use the pod-IVR if accessed over the counter.

Qualitative interview data revealed that, overall, the TDF-FTC-MVC pod-IVR was well tolerated by participants. Prior to insertion of the pod-IVR, clinicians provided verbal instructions. Participants reported that their confidence to insert the pod-IVR was increased by the clinician’s instructions, as well as when the clinician checked for proper pod-IVR placement on first insertion. The majority (5 of 6) of the women noted that they did not feel the pod-IVR in their vaginas at all during the use period. None of the participants felt that the pod-IVRs had an impact on their typical hygiene practices. All 6 expressed the desire for the pod-IVR to have multiple purposes, such as for HIV prevention and contraception. Detailed data of user experiences are reported elsewhere [39].

In vivo ARV drug release rates

In vivo release rate measurements are based on the residual drug mass remaining in the used IVRs and the assumption, supported by in vitro data [26,28,29], that drug release is linear over the period of IVR use. The mean daily in vivo IVR drug release rates in used IVRs were as follows: median (IQR); TDF pod-IVR; TDF, 0.81 (0.66–1.07) mg/d; TDF-FTC pod-IVR; TDF, 0.98 (0.88–1.93) mg/d; FTC, 1.99 (1.70–2.31) mg/d (S1 and S2 Tables). Importantly, >95% of the residual TDF in the used IVR pods was present as the prodrug by HPLC, i.e., no significant prodrug hydrolysis was observed following 1 wk of use in vivo. The in vivo release rates of TDF from the TDF and TDF-FTC pod-IVR were found to be not significantly different ($P = 0.4286$).

For in vivo release rates under 1 mg/d TDF, less than 4% of the IVR drug content would have been released over 1 wk of IVR use, making differential residual drug measurements challenging. The amount of TDF released in the TDF-FTC-MVC pod-IVR treatment period could not be quantified accurately, although it was higher than in the TDF and TDF-FTC pod-IVR treatment periods, based on median TDF CVF concentrations (S3 Table). The mean daily in vivo IVR drug release rates were as follows: median (IQR); FTC, 2.37 (1.94–2.57) mg/d; MVC, 2.07 (1.77–2.09) mg/d.

Summary of drug concentration measurements

Drug and drug metabolite concentrations in key anatomic compartments with the IVRs in place are summarized in S1–S3 Tables. These data show that the IVRs maintained high ARV drug exposure in CVFs and vaginal tissues (VTs) relative to lower concomitant plasma concentrations. FTC and MVC drug concentrations in all matrices were higher than corresponding TDF/TFV concentrations as expected based on the residual drug levels in used IVRs.

CVF drug levels

CVF ARV drug concentrations all exhibited low variability during IVR use in all treatment periods (Figs 1–3), with the exception of one low data point at Visit 3 for MVC (Fig 3D). TDF CVF concentrations in the three groups (i.e., TDF, TDF-FTC, and TDF-FTC-MVC) were not
statistically significantly different ($P = 0.4233$); however, TFV concentrations were different ($P = 0.0118$) with this test. Total TFV CVF concentrations, defined as the molar sum of paired TFV and TDF concentrations reported as TFV, across all three groups also were different ($P = 0.0155$), as detailed below, with higher median concentrations in the TDF-FTC-MVC pod-IVR group. FTC CVF concentrations in the TDF-FTC and TDF-FTC-MVC groups were not significantly different ($P = 0.9323$).

A comparison of the medians and IQRs shows that the corresponding TDF (S1 Table) and TDF-FTC (S2 Table) pod-IVR datasets are similar, but the TDF-FTC-MVC (S3 Table) pod-IVR leads a total (i.e., TDF + TFV, on a molar basis) TFV exposure that is ca. two times higher: median (IQR); TDF pod-IVR, 58.1 (43.9–97.4) ng/mg; TDF-FTC pod-IVR, 43.1 (31.0–65.0) ng/mg; TDF-FTC-MVC pod-IVR, 96.9 (14.5–137.1) ng/mg; TFV; TDF pod-IVR, 13.9 (6.2–19.3) ng/mg; TDF-FTC pod-IVR, 15.9 (7.1–20.0) ng/mg; TDF-FTC-MVC pod-IVR, 28.0 (24.3–31.9) ng/mg; total TFV; TDF pod-IVR, 36.2 (31.3–60.6) ng/mg; TDF-FTC pod-IVR, 34.4 (26.9–48.7) ng/mg; TDF-FTC-MVC pod-IVR, 70.0 (59.2–80.6) ng/mg.

The 2-wk window for the follow-up visit after IVR removal (Visit 4/8/4) allowed ARV drug washout kinetics to be measured and the corresponding terminal half-lives of elimination to be calculated (Figs 1–3): median (IQR); TDF pod-IVR; TDF, 11.8 (10.6–14.4) h; TFV, 39.5 (30.0–58.1) h; TDF-FTC pod-IVR; TDF, 14.2 (11.7–15.4) h; TFV, 31.4 (25.9–36.2) h; FTC, 19.1 (13.5–20.1) h; TDF-FTC-MVC pod-IVR; TDF, 18.3 (11.8–24.2) h; TFV, 24.8 (24.4–31.9) h; FTC, 17.0 (15.5–17.8) h; MVC, 16.0 (15.8–18.3) h. Comparison of the TDF and TFV half-lives in the TDF pod-IVR group showed that the datasets were different ($P = 0.0043$). The TDF and TFV half-lives in the TDF-FTC pod-IVR group also were different ($P = 0.0022$), but the TDF...
and FTC half-lives were not different ($P = 0.2403$). The ARV drug half-lives across IVR groups were not significantly different (TDF, $P = 0.4848$; TFV, $P = 0.4286$). Comparison of the TDF, FTC, and MVC groups (Fig 3) showed that they were not significantly different ($P = 0.9320$). However, comparison of just the TDF and TFV groups showed that they were significantly different ($P = 0.0909$).

The measurement of Li$^+$, an exogenous tracer added to the naive CVF fluid, was used to correct the CVF drug concentration analyses for dilution in the TDF-FTC-MVC pod-IVR group and, hence, affords the corresponding drug concentrations in undiluted CVF [37]. The corrected CVF drug concentrations exhibited a moderate (total TFV and MVC) to weak (FTC) correlation with paired CVF (Dacron swab) drug concentrations (S4 Fig): TFV (total TFV, reported as the molar sum of TDF and TFV concentrations); slope, 7.53 ± 0.84; $R^2$, 0.889; FTC; slope, 2.40 ± 1.43; $R^2$, 0.220; MVC; slope, 4.37 ± 1.11; $R^2$, 0.609. The CVF volume collected during the lavage procedure (S4 Fig) with the TDF-FTC-MVC pod-IVRs in place was: median (IQR), 85.6 (28.0–116) μL.

VT drug concentrations

Molar antiviral drug concentrations in VT biopsy homogenate are described in Fig 4A and Fig 5A to allow direct comparison with the pharmacologically active metabolite of TFV against HIV, TFV-DP. The CVF (ng/mg) to VT (ng/mg) drug concentration ratio (Fig 4B and Fig 5B) provides a simple measure of xenobiotic partitioning between the two anatomic compartments: the lower the ratio, the more the antiviral agent distributes into the vaginal mucosa and the higher the vaginal bioavailability. Drug CVF:VT median (IQR) ratios are as follows: TDF...
and TDF-FTC pod-IVRs (data combined); TFV (molar sum of TDF and TFV, as TFV), 6.6 (4.1–28.6); FTC, 17.2 (8.5–66.5). The concentration ratio for TDF is 2.6 times lower than for FTC; TDF-FTC-MVC pod-IVR; TFV, 14.2 (6.0–16.5); FTC, 11.5 (5.8–17.0); MVC 3.5 (0.9–6.6). There was no statistically significant difference between the three groups \( (P = 0.1591) \).

Fig 3. CVF drug levels and washout following TDF-FTC-MVC pod-IVR use. ARV drug levels in CVFs fall off rapidly following IVR removal (grey arrows) but are quantifiable for a further 2 wk. Every circular datum represents an individual sample from one of the participants \( (n = 6) \), while triangles depict samples that were BLQ of the analytical method, and values were calculated as follows: \([\text{assay BLQ}/2]/(\text{median swab mass})\). (A) TDF. (B) TFV. (C) FTC. (D) MVC. (E) Box plots of ARV drug terminal half-lives of elimination from CVF. The box extends from the 25th to 75th percentiles, with the horizontal line in the box representing the median; whiskers represent the lowest and highest datum. Comparison of the TDF, FTC, and MVC groups using an ordinary 1-way ANOVA analysis (no matching or pairing of the data) showed that they were not significantly different \( (P = 0.8973) \). However, comparison of the TDF and TFV groups using an unpaired \( t \) test with Welch’s correction, showed that the groups were significantly different \( (P = 0.0327) \). ANOVA, analysis of variance; ARV, antiretroviral; BLQ, below the lower limit of quantitation; CVF, cervicovaginal fluid; FTC, emtricitabine; IVR, intravaginal ring; MVC, maraviroc; TDF, tenofovir disoproxil fumarate; TFV, tenofovir.

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Fig 4. Drug and drug metabolite exposure in VT at TDF and TDF-FTC pod-IVR removal (Day 7). (A) ARV drug and TFV-DP levels in vaginal biopsies collected on Day 7 within 30 min of pod-IVR removal. Analyte concentrations are all expressed as fmol/mg of tissue homogenate to allow meaningful cross-comparison. Every circular datum represents an individual sample from one of the participants ($n = 6$); horizontal lines represent group medians; blue, TFV; green, TFV-DP; red, FTC. (B) Paired CVF:VT concentration ratios of TFV (molar sum of TDF and TFV concentrations) and FTC at Day 7; horizontal lines represent group means. The ratios provide a measure of the extent of tissue penetration for each analyte following vaginal delivery and, hence, vaginal bioavailability. The CVF TFV concentrations used in this calculation were the molar sum of the measured TFV and TDF concentrations as the prodrug hydrolyzes to TFV in the vaginal mucosa. ARV, antiretroviral; CVF, cervicovaginal fluid; FTC, emtricitabine; IVR, intravaginal ring; TDF, tenofovir disoproxil fumarate; TFV, tenofovir; TFV-DP, tenofovir diphosphate; VT, vaginal tissue.

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Discussion

To our knowledge, the clinical trial described here is the first to evaluate long-acting vaginal delivery of TDF-FTC, the only FDA-approved drug regimen (in the oral formulation Truvada) for HIV PrEP [12]. The innovative open-label, crossover trial format allowed two IVR formulations—TDF alone and in combination with FTC—to be evaluated sequentially with each participant acting as her own control. To our knowledge, the trial also involved the first triple...

Fig 5. Drug and drug metabolite exposure in VTs at TDF-FTC-MVC pod-IVR removal (Day 7). (A) ARV drug and TFV-DP levels in vaginal biopsies collected on Day 7 within 30 min of pod-IVR removal. Analyte concentrations are all expressed as fmol/mg of tissue homogenate to allow meaningful cross-comparison. Every circular datum represents an individual sample from one of the participants ($n = 6$), while inverted triangles depict samples that were ALQ of the analytical method, and values were calculated as follows: (assay ALQ)/(median biopsy mass); horizontal lines represent group medians. (B) Paired CVF:VT concentration ratios of total TFV (molar sum of TDF and TFV concentrations), FTC, and MVC at Day 7; horizontal lines represent group means. The ratios describe the extent of tissue penetration for each analyte following vaginal delivery and, hence, vaginal bioavailability. The CVF TFV concentrations used in this calculation were the molar sum of the measured TFV and TDF concentrations, as the prodrug hydrolyzes to TFV in the vaginal mucosa. The two MVC VT samples that were ALQ were omitted from the analysis. There was no statistically significant difference between the three groups according to an unpaired, nonparametric Kruskal–Wallis test ($P = 0.1591$). ALQ, above the upper limit of quantitation; ARV, antiretroviral; CVF, cervicovaginal fluid; FTC, emtricitabine; IVR, intravaginal ring; MVC, maraviroc; TDF, tenofovir disoproxil fumarate; TFV, tenofovir; TFV-DP, tenofovir diphosphate; VT, vaginal tissue.

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ARV combination IVR to be evaluated clinically. The pod-IVRs were safe and generally well tolerated, despite high VT ARV concentrations especially for FTC and MVC (Fig 5A). The implications of our findings are discussed below in the context of developing a viable HIV PrEP candidate targeted at resource-poor regions.

The 3 pod-IVRs maintained high, controlled ARV drug concentrations in CVF over the period of product use, while leading to low systemic exposures (S1–S3 Tables). The low plasma ARV drug concentrations are a benefit of topical dosing as the risks of systemic toxicity and the emergence of drug resistance are reduced. The CVF drug levels decreased sharply post-IVR removal on Day 7 (Figs 1–3). The drug washout profiles allowed the CVF half-lives of the three drugs as well as TFV, the hydrolysis product of TDF, to be measured for the first time (Fig 3E). In all three groups, TFV had a significantly longer half-life (median, 24.8–39.5 h) than the other agents. We previously have described a mechanism in sheep [40] in which TDF, delivered via IVR, distributed efficiently from the CVF into the VT, where it hydrolyzed enzymatically to TFV. The accumulation of TFV in the VT formed a depot that could release TFV back into the lumen. A similar mechanism may operate here in women, accounting for the longer TFV half-life in CVF relative to the other agents.

The systematically higher (2.4–7.5-fold) CVF drug concentrations collected via lavage versus Dacron swab in the TDF-FTC-MVC pod-IVR group was unexpected and remains largely unexplained. It is possible that the lavage is more efficient at extracting drug that is associated with the surface of the vaginal epithelium. While we have used the Li⁺ tracer technique previously to measure CVF dilution in CVL in sheep [29], this is the first example of its application in a clinical setting for PK analysis. During IVR use, the median CVF volumes (0.086 mL; IQR, 0.028–0.12 mL) in the 6 participants were lower than the volumes collected previously in women at different phases of the menstrual cycle, 0.30 ± 0.22 mL (follicular phase) and 0.45 ± 0.21 mL (luteal phase), using a different assay [41]. This was expected based on the collection of multiple samples (one swab for VMB analysis, one pH swab, two Dacron swabs for CVF drug analyses, and one vaginal sidewall scraping for microbial biofilm imaging) prior to the lavage. As a result, a significant fraction of the available CVF was removed before CVL collection.

In HIV PrEP, there is no biomarker of ARV drug effect in susceptible, uninfected individuals to guide product development, unlike treatment of HIV-1/AIDS. The choice of ARV agent and target drug levels in key pharmacological compartments largely is based on theoretical arguments and results from preclinical studies. The strategy employed here was based on using FDA-approved ARV drugs that have shown clinical efficacy in HIV PrEP using oral or topical regimens [4,5,7–11,42]. Combined with PK data from oral dosing randomized clinical trials demonstrating efficacy [7,42], one can bracket the concentration targets associated with vaginal protection. A randomized, PK, cross-over study (MTN-001) compared TFV vaginal gel and oral TDF tablets by measuring drug and drug metabolite levels in VT from 144 HIV-uninfected women [32]. The TFV concentrations in VT homogenate at end-of-period visit were as follows: median (IQR); vaginal TFV gel, 113 (27–265) ng/mg; oral TDF, 0.15 (0.15–0.27) ng/mg. Assuming that the range of drug concentrations obtained from these dosing modalities is a key determinant for efficacy in preventing vaginal acquisition of HIV, the TFV concentrations in vaginal biopsies collected on Day 7 at IVR removal in this study, 8.4 (4.7–11.2) ng/mg (S1 Table), are suggestive of positive pharmacodynamic outcomes. However, this analysis assumes oral dosing is not fundamentally different than topical dosing in terms of tissue concentrations required for high PrEP efficacy.

A previous clinical trial evaluating a different TDF IVR platform—a reservoir IVR, in which a solid TDF formulation is contained in a hollow hydrophilic polyether urethane tube and the drug is delivered through the ring elastomer—reported median CVF TDF and TFV

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Phase I trial of antiretroviral intravaginal rings
concentrations during ring use of 110 ng/mg and 70 ng/mg, respectively [43]. These levels are higher than the corresponding drug concentrations measured here (S1–S3 Tables), although the TDF:TFV ratio was considerably higher in our study (2.7–4.2-fold, depending on the formulation, compared to 1.5) and the TDF CVF concentrations were more stable (Figs 1–2). Together, these results suggest less TDF hydrolysis either in the IVRs during use or sample handling in our trial. Sample bioanalysis in both trials was carried out by the same laboratory using the same methods. The observation may be significant since the VT bioavailability in sheep of the TDF prodrug is nearly 100 times higher than for parent TFV [40].

Immune cells in the vaginal mucosa are believed to be the key pharmacologic compartment determining the efficacy of vaginal HIV PrEP. It is not feasible in early-stage clinical trials to collect sufficient VT to extract immune cells for intracellular drug analysis. VT biopsy homogenate has been shown to correlate well with CD4+ cells extracted from VT [44], and the relevant analytes are TFV, TFV-DP, the pharmacologically active metabolite of TFV against HIV, and FTC (Figs 4A and 5A). Due to the small amount of VT collected, the active, triphosphorylated metabolite of FTC could not be measured here. Median VT homogenate TFV-DP concentrations of 303 (TDF pod-IVR), 289 (TDF-FTC pod-IVR), and 302 (TDF-FTC-MVC pod-IVR) fmol/mg were measured post-IVR removal. These concentrations are 2–3-fold higher than the 120 fmol/mg median ectocervical biopsy levels obtained with a TDF reservoir IVR in a recent clinical trial [43]. Importantly, the median TFV and FTC VT concentrations (TDF pod-IVR; TFV, 8.4 ng/mg; TDF-FTC pod-IVR; TFV, 5.1 ng/mg; FTC, 75 ng/mg) were lower than those obtained in pigtailed macaques (TFV 28–35 and FTC 460–650 ng/mg, depending on the sampling location) with TDF-FTC pod-IVRs [26], which provided complete protection from Simian Human Immunodeficiency Virus (SHIV) infection in pigtailed macaques using the rigorous, repeat low-dose challenge model [45]. Smith and colleagues used the same model to evaluate a reservoir TDF IVR and also obtained complete protection from SHIV infection [46]. The median VT TFV concentration around 10 ng/mg observed with these reservoir TDF IVRs is comparable to the VT TFV values obtained in our study. When complemented by ca. 10 times higher tissue FTC levels, as observed here, these results suggest that the devices may be effective in HIV PrEP.

No clinical efficacy data for HIV PrEP using intravaginal MVC currently are available, and the levels in the pharmacologically relevant compartments required to afford protection are, therefore, unknown. The published in vitro antiviral potencies of MVC against HIV-1 primary and laboratory-adapted isolates in peripheral blood mononuclear cells (PBMCs) span a wide range of inhibitory concentrations: IC50, 0.1–4.5 nM; IC90, 0.5–13.4 nM [47]. The observed median MVC concentrations of 142 ng/mg (276 μM) in VTs on Day 7 were more than 20,000 times higher than the highest IC90, suggesting favorable pharmacodynamic outcomes. A randomized clinical trial (MTN-013/IPM 026) in 48 HIV-negative women evaluating matrix-IVRs delivering MVC alone or in combination with DPV measured MVC CVF concentrations of 2.5 and 1.1 ng/mg, respectively, at Day 28 when the IVRs were removed [48]. These concentrations are 170–390 times lower than those obtained in the current study (S3 Table). It should be noted that matrix-IVRs tend to have a drug release burst in the first week. VT MVC levels in MTN013 were below the lower limit of quantification for all subjects using the DPV-MVC IVR and were only quantifiable in 4 of the 12 MVC IVR users, with a 0.13–4.4 ng/mg concentration range, much lower than the median concentration (142 ng/mg) measured following pod-IVR delivery reported here (S3 Table).

To our knowledge, the ASPIRE study and The Ring Study were the first published Phase 3 studies of an ARV (i.e., DPV) delivered via IVR for HIV PrEP [49,50]. In the ASPIRE study, the incidence of HIV infection in the DPV group was 27% lower than in the placebo group; this improved to 56% protection in women over the age of 21 years; the difference was
attributed to better adherence in women over the age of 21 years [49]. Both studies found low rates of adherence, particularly in young women raising concerns about the viability of IVRs for HIV PrEP in resource-limited regions like sub-Saharan Africa, where contraceptive IVRs are not as commonly used as in the developed world. It is believed by some that with increased familiarity with IVRs, adherence to IVR use will improve [51–55] and make IVR-based ARV regimens viable. This belief is based on previous experience with oral HIV PrEP regimens that demonstrated a dramatic increase in adherence when moving from the initial, blinded, placebo-controlled trials to the open-label follow-on trials [11,56–58].

In terms of the user perception and acceptability data, the pod-IVR delivery device was both easy to use and well tolerated. Women were willing to use it within the context of this cross-over study and anticipated being confident in their ability to use it for longer periods of time in “real-world” settings for HIV prevention. While, on average, participant confidence in their ability to access and use the IVR stayed the same or increased between uses, one participant’s data suggest that her confidence in her ability to insert and remove the IVR decreased with her second experience, indicating that further study is necessary to determine the factors related to confidence and what is needed to become skilled in IVR insertion and removal. Women were willing to use the TDF-FTC-MVC pod-IVRs effectively within the context of the study and anticipated feeling confident in their ability to use it for HIV prevention for sustained periods of use. Additionally, given the design of the current study, future work will need to assess user perceptions and experiences of pod-IVRs for longer periods of use, as well as in relation to sexual activity and use during menstruation.

Limitations of this study are as follows. Women at low risk for acquisition of HIV were recruited for this study since it was a first in humans and was primarily focused on PK and safety; these women may have different perceptions about HIV prevention than women at high risk for HIV, who will be the targeted population for these prevention products. The small sample size was also a limitation; however, the primary goal of the study was to show drug release and initial safety, as well as to get initial understanding of women’s perceptions and acceptability of the IVRs. The short duration was also a limitation since the IVRs will be used for 28 d; however, this initial study of 7 d duration was required by the FDA prior to a 28-d study. A 28-d study is planned as a follow-on to this study. Lastly, as described above, due to the small size of the vaginal biopsies, we were unable to measure FTC triphosphate in VT.

In conclusion, a crossover Phase I clinical trial sequentially evaluated TDF, TDF-FTC, and TDF-FTC-MVC pod-IVRs and demonstrated that the devices exhibited favorable safety and PK profiles across all three treatment periods. The results justify longer and larger follow-on clinical trials in the future.

Supporting information
S1 TREND Checklist. TREND Checklist with associated paragraph numbers. (PDF)

S1 Fig. Microbial community profiles for the six trial participants in the TDF pod-IVR trial arm determined by a custom qPCR array. The IVRs were inserted at V1 and removed at V3. IVR, intravaginal ring; qPCR, quantitative polymerase chain reaction; TDF, tenofovir disoproxil fumarate. (TIF)

S2 Fig. Microbial community profiles for the six trial participants in the TDF-FTC pod-IVR trial arm determined by a custom qPCR array. The IVRs were inserted at V5 and removed at V7. FTC, emtricitabine; IVR, intravaginal ring; qPCR, quantitative polymerase
chain reaction; TDF, tenofovir disoproxil fumarate.

S3 Fig. Microbial community profiles for the six trial participants in the TDF-FTC-MVC pod-IVR trial arm determined by a custom qPCR array. The IVRs were inserted at V1 and removed at V3. FTC, emtricitabine; IVR, intravaginal ring; MVC, maraviroc; qPCR, quantitative polymerase chain reaction; TDF, tenofovir disoproxil fumarate.

S4 Fig. Measurement of CVF volume collected in the lavage procedure allows for dilution correction of the CVL analyses. Drug concentrations with the IVR in place (V2–V3) in CVL samples corrected for CVF dilution (y-axis) plotted against paired drug concentrations in neat CVF samples collected using Dacron swabs (x-axis) exhibit a moderate-weak correlation, with systematically higher values in CVL samples. (A) TFV (total TFV, reported as the molar sum of TDF and TFV concentrations in the samples); slope, 7.53 ± 0.84; R2, 0.889; FTC; slope, 2.40 ± 1.43; R2, 0.220; MVC; slope, 4.37 ± 1.11; R2, 0.609. (D) Box plots of CVF volume collected for all participants (n = 6) at each study visit (V1–V4). The box extends from the 25th to 75th percentiles, with the horizontal line in the box representing the median; whiskers represent the lowest and highest datum. CVF, cervicovaginal fluid; CVL, cervicovaginal lavage; FTC, emtricitabine; IVR, intravaginal ring; MVC, maraviroc; TDF, tenofovir disoproxil fumarate; TFV, tenofovir.

S1 Table. Summary of drug and drug metabolite concentrations in key anatomic compartments measured with TDF pod-IVR in place (6 participants), i.e., Visits 2 and 3. IVR, intravaginal ring; TDF, tenofovir disoproxil fumarate.

S2 Table. Summary of drug and drug metabolite concentrations in key anatomic compartments measured with TDF-FTC pod-IVR in place (6 participants), i.e., Visits 6 and 7. FTC, emtricitabine; IVR, intravaginal ring; TDF, tenofovir disoproxil fumarate.

S3 Table. Summary of drug and drug metabolite concentrations in key anatomic compartments measured with TDF-FTC-MVC pod-IVR in place (6 participants), i.e., Visits 2 and 3. FTC, emtricitabine; IVR, intravaginal ring; MVC, maraviroc; TDF, tenofovir disoproxil fumarate.

S1 Protocol. Summary of clinical trial protocol.

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References


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