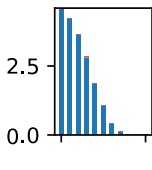
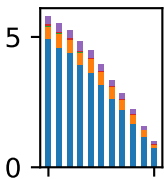
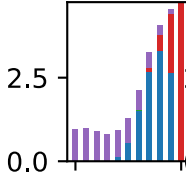
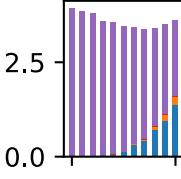
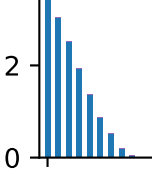
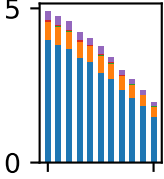
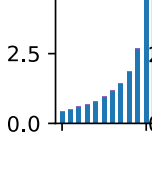
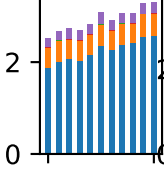
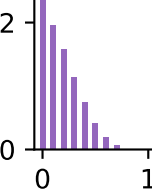
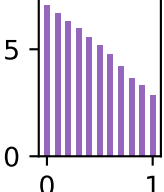
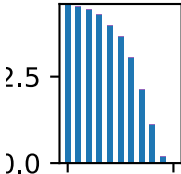
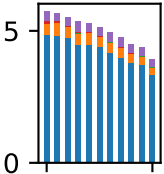
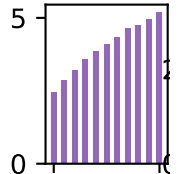
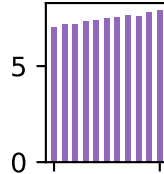
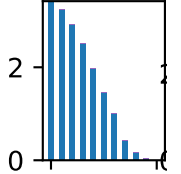
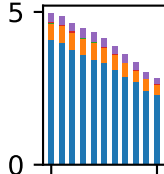
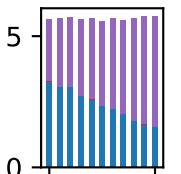
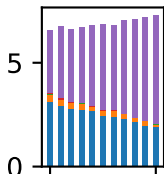
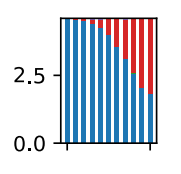
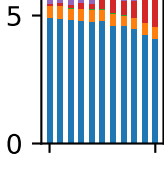
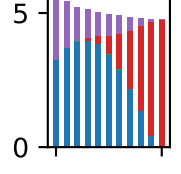
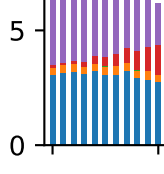
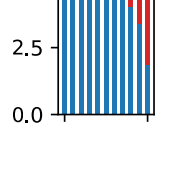
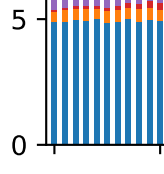
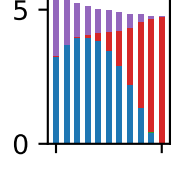
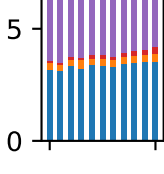
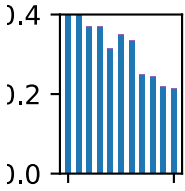
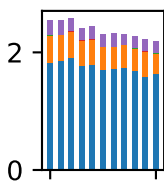
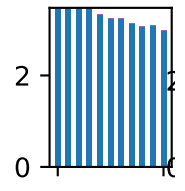
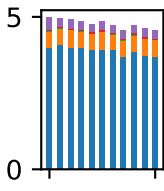
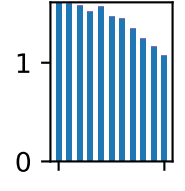
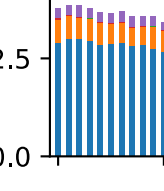
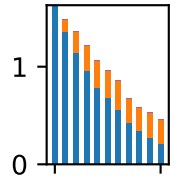
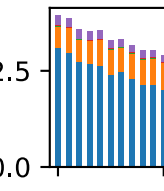
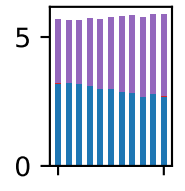
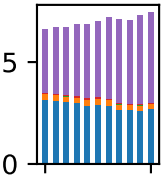
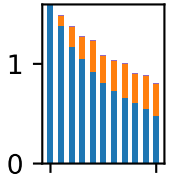
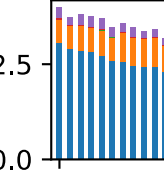
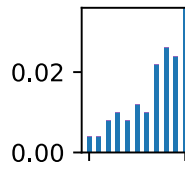
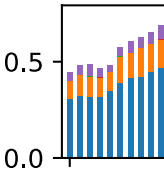
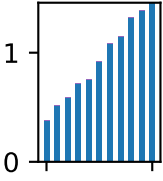
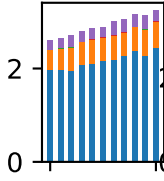
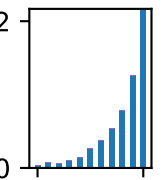
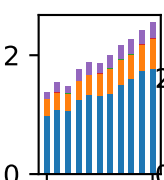
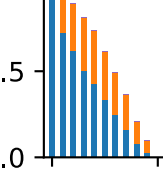
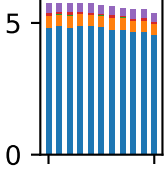
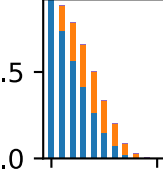
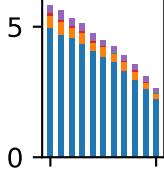
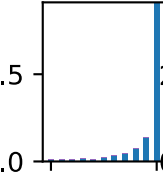
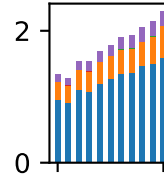


**S4 Table. Knockout and overexpression model behaviors vs. experimental data**



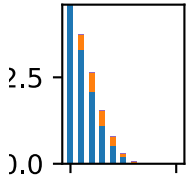
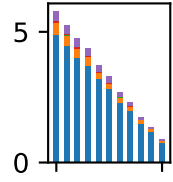
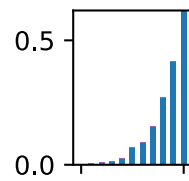
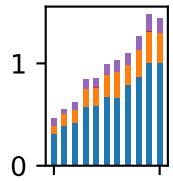




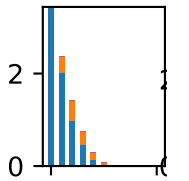
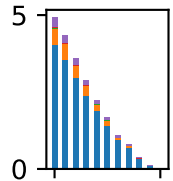
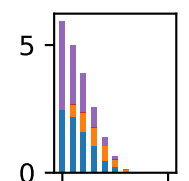
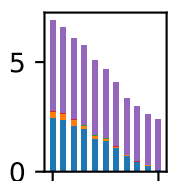
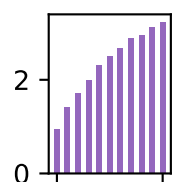
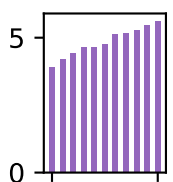
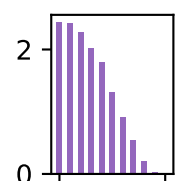
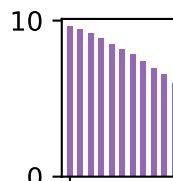

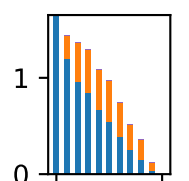
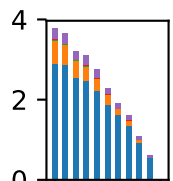
Model behavior						Experimental support
KO/ OE	Node	Env. sign	Description	Synchronous result	Biased asynch. result	
				<b>x-axis:</b> % inhibition/over-expression <b>y-axis:</b> occurrence in 100 time-steps <div><div></div> Normal cell cycle <div></div> G2 → G1 → Genome dupl. <div></div> Aneuploidy + Genome dupl. <div></div> No cytokinesis → Genome dupl. <div></div> Apoptosis</div>		
Growth Signaling Module						
↘	Ras	95% GF <sub>H</sub>	Ras knockdown limits proliferation under strong mitogenic stimulation.			Ras is a known proto-oncogene; constitutive activation of Ras is found in a variety of tumors. It activates MAPK and PI3K signaling, and is a potent inducer of proliferation and survival signaling [1].
↗		20% Trail (no GF <sub>H</sub> )	Ras hyper activation enhances proliferation and blocks Trail-mediated apoptosis.			
↘	ERK	80% GF <sub>H</sub>	ERK knockdown limits proliferation under strong mitogenic stimulation.			As the often oncogenic pro-proliferative MAPK signaling cascade goes through ERK, its inhibition is under investigation as a potentially potent cancer drug against a range of tumors with MAPK pathway mutations [2].
↗		50% GF <sub>H</sub>	ERK hyper activation enhances proliferation under weak mitogenic stimulation.			
↗		50% GF (no GF <sub>H</sub> )	ERK hyper activation blocks apoptosis in the absence of survival signals.			


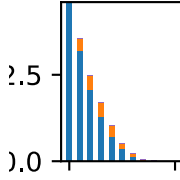
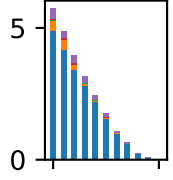

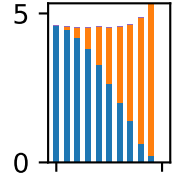
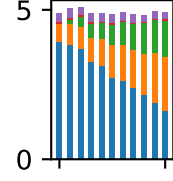

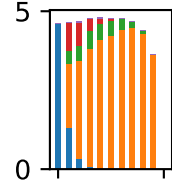
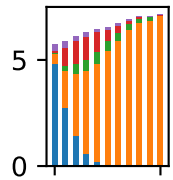

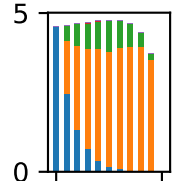
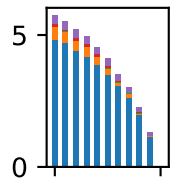

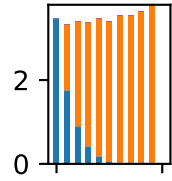
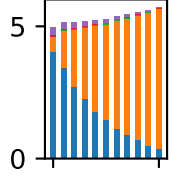
↓	<b>mTORC2</b>	95% $GF_H$	<b>mTORC2</b> knockdown limits proliferation under strong mitogenic stimulation.			Inhibition of the <i>mTORC2</i> subunit <i>Rictor</i> blocks proliferation and promotes apoptosis in gastric cancer [3] and glioblastoma [4].
		50% $GF$ (no $GF_H$ )	<b>mTORC2</b> knockdown enhances apoptosis in response to a loss of survival signals.			
↓	<b>PDK1</b>	80% $GF_H$	<b>PDK1</b> knockdown limits proliferation under strong mitogenic stimulation.			PDK1 inhibition by RNAi in melanoma and colon cancer cells resulted in significant cell growth inhibition and apoptosis [5].
		50% <i>Trail</i>	<b>PDK1</b> knockdown potentiates <i>Trail</i> -induced apoptosis in cycling cells.			
↓	<b>PLCγ</b>	95% $GF_H$	<b>PLCγ</b> knockdown enriches for cells that fail to complete cytokinesis.			<i>prediction</i>
		50% <i>Trail</i>	<b>PLCγ</b> knockdown protects against <i>Trail</i> -mediated apoptosis in high growth factor environments.			<i>prediction</i>
↓	<b>NeddL4</b>	95% $GF_H$	<b>NeddL4</b> knockdown enriches for cells that fail to complete cytokinesis.			<i>prediction</i>
		50% <i>Trail</i>	<b>NeddL4</b> knockdown protects against apoptosis in high growth factor environments.			<i>prediction</i>

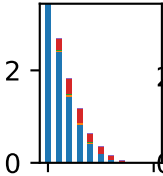
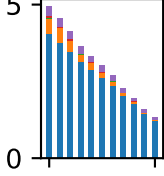
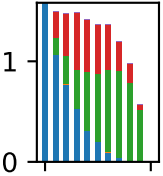
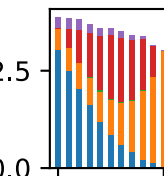
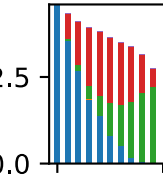
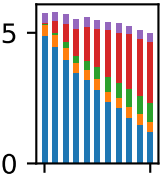
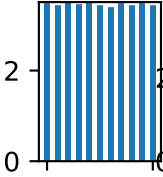
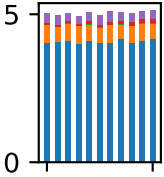
↗	<b>FoxO1</b>	50% GF <sub>H</sub>	<b>FoxO1</b> over expression slows down proliferation.			Overexpression of the active form of FOXO family members promotes cell cycle arrest at the G1/S boundary in a variety of cell lines [6].
↗	<b>TSC2</b>	80% GF <sub>H</sub>	<b>mTORC1</b> over expression slows down proliferation.			Over-expression of TSC2 slows proliferation in oral cancer cell lines [7]. and increased the fraction of human osteosarcoma cells arrested in G1 [8].
↘	<b>Rheb</b>	65% GF <sub>H</sub>	<b>Rheb</b> inhibition slows proliferation under moderate mitogenic stimulation.			Fruitfly Rheb promotes cell cycle progression and cell growth in vivo and in vitro [9].
↘	<b>mTORC1</b>	65% GF <sub>H</sub>	<b>mTORC1</b> inhibition severely limits proliferation under moderate mitogenic stimulation.			Inhibition of <i>mTORC1</i> blocks both cell growth (in volume) and cell cycle progression [10].
↘	<b>S6K</b>	50% Trail	<b>S6K</b> inhibition increases Trail-mediated apoptosis in cycling cells.			Silencing S6K lead to a substantial increase in TNF-α as well as TRAIL-mediated apoptosis [11].
↘	<b>eIF4E</b>	65% GF <sub>H</sub>	<b>eIF4E</b> inhibition decreases cell proliferation at non-saturating mitogenic exposure, and leads to a small fraction of endo-reduplicating cells.			4E-BPs inhibit cell proliferation via inhibitory binding to <i>eIF4E</i> , and selectively blocking translation of proteins involved in cell cycle progression [10].
↘	<b>GSK3</b>	20% GF <sub>H</sub>	<b>GSK3</b> inhibition increases proliferation and cell cycle entry under low mitogenic stimulation			GSK-3 inhibitors maintain proliferative capacity in mouse embryonic stem cells (ESCs) as well as mammalian cardiomyocytes [12].

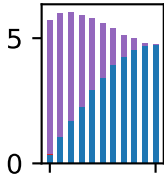
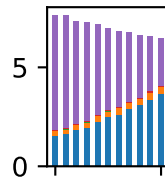
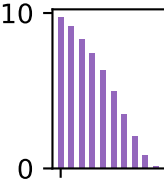
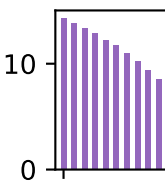
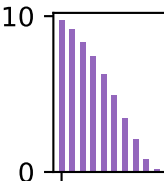
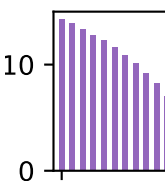
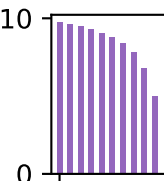
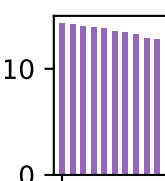
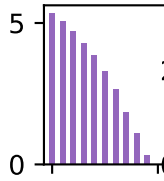
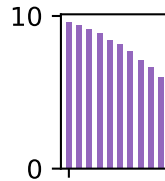
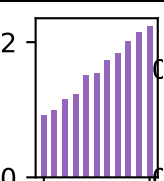
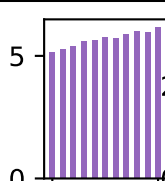
Restriction Switch & Origin of Replication Licensing Switch						
↘	<i>p21</i>	50% $GF_H$	<i>p21</i> inhibition increases proliferation rate in the presence of moderate growth stimulation.			<i>p21</i> -null cells were more likely than their wild-type counterparts to commit to another division before finding the previous one [13]. Generally, <i>p21</i> is known to promote cell cycle arrest in response to several stimuli [14].
↘	<i>pRB</i>	35% $GF_H$	<i>pRB</i> inhibition is a potent inducer of proliferation under weak mitogenic stimulation.			The <i>RB1</i> gene and its protein product, retinoblastoma protein ( <i>pRB</i> ) is the first known tumor suppressor; its loss leads to aberrant cell cycle entry and a lack of an effective G1/S checkpoint [15].
↗	<i>p27<sup>Kip1</sup></i>	95% $GF_H$	<i>p27<sup>Kip1</sup></i> up regulation slows down proliferation.			Overexpression of <i>p27KIP1</i> induced cell cycle arrest in G1 phase in a hepatoma cell line [16].
↘	<i>Myc</i>	95% $GF_H$	<i>Myc</i> inhibition severely limits proliferation even under strong mitogenic stimulation.			<i>Myc</i> is the most frequently amplified human oncogene. orchestrates the transcriptional regulation of cell growth, cell-cycle progression, metabolism, and cell survival. As a result, it is a key therapeutic cancer target [17].
↗		35% $GF_H$	<i>Myc</i> over-expression promotes proliferation even under weak mitogenic stimulation.			

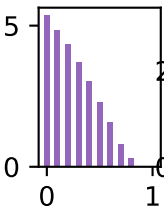
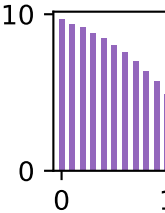
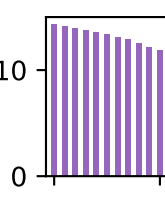
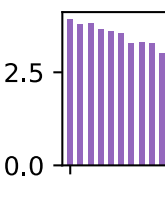
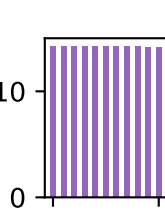
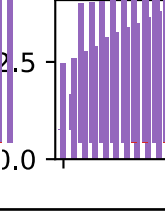
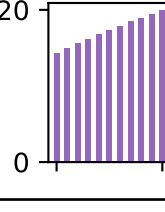


 	<b>Cyclin D1</b>	95% $GF_H$	<b>Cyclin D1</b> inhibition severely limits proliferation even under strong mitogenic stimulation.			<i>Cyclin D1</i> knockdown inhibits proliferation in a variety of cancer cells [18-19].
		20% $GF_H$	<b>Cyclin D1</b> promotes cell cycle entry and proliferation even under weak mitogenic stimulation.			Overexpression of <i>cyclin D1</i> is linked to tumorigenesis of many types of cancer [19].
   	<b>E2F1</b>	80% $GF_H$	<b>E2F1</b> inhibition drastically limits proliferation even in the presence of strong mitogenic stimulation.			The absence of all activating E2F proteins causes cell cycle arrest in flies or mammalian fibroblasts [20]. Reduced <i>E2F1</i> expression impaired rat glioma cell proliferation [21]. E2F-1 expression can sensitize cells to apoptosis by a number of agents [22].
		65% <i>Trail</i>	<b>E2F1</b> inhibition drastically limits Trail-mediated apoptosis in cyclin cells.			
		20% <i>Trail</i>	<b>E2F1</b> over expression strongly increases Trail-mediated apoptosis in quiescent cells.			
		20% $GF$ (no $GF_H$ )	<b>E2F1</b> inhibition blocks apoptosis due to the withdrawal of survival signals.			<b>prediction</b>
	<b>CyclinE</b>	65% $GF_H$	<b>Cyclin E</b> inhibition drastically limits proliferation, and mild inhibition leads to an enrichment of endoreduplicating (multiple S-phases with no intervening M-phase) cells.			<b>prediction</b> Circumstantial evidence: <i>forced Cyclin E</i> expression in fruit fly embryos that normally undergo endoreduplication resulted in strong inhibition of endoreduplication [23].

	<b>Cdc6</b>	95% GF <sub>H</sub>	<b>Cdc6</b> inhibition abolishes proliferation even under near-saturating mitogenic stimulation.			Cdc6 is required for initiation of DNA replication in mammalian cells [24].
<b>Phase Switch &amp; Cell cycle process nodes</b>						
	<b>Emi1</b>	95% GF <sub>H</sub>	<b>Emi1</b> inhibition limits normal proliferation and induces endo-reduplication (multiple S-phases with no intervening M-phase). <i>In the asynchronous model, Emi1 loss leads to aneuploidy; potentially and artifact of this update.</i>			<b>Emi1</b> knockdown with RNAi caused re-replication in HeLa cells, due to premature activation of APC/C that results in destabilization of cyclin A [25].
	<b>FoxM1</b>	95% GF <sub>H</sub>	<b>FoxM1</b> inhibition drastically limits normal proliferation and potentially induces endo-reduplication, as well as failure to undergo cytokinesis.			FoxM1-deficient cells arrest in G2, but also show mitotic abnormalities such as including mitotic spindle aberrations, chromosome missegregation and overt polyploidization [26]. DNA content of fetal cells from <i>FoxM1</i> knockout mice show 6 to 50-fold polyploidy [27].
	<b>Cdc25A</b>	95% GF <sub>H</sub>	According to our <i>asynchronous</i> model, <b>Cdc25A</b> inhibition limits S-phase entry and proliferation. <i>The synchronous results are likely an artifact.</i>			Inhibition of Cdc25A potentially blocks cell cycle progression in G1 or G2 [28].
	<b>CyclinA</b>	80% GF <sub>H</sub>	<b>Cyclin A</b> inhibition drastically limits normal proliferation and potentially induces endo-reduplication (multiple S-phases with no intervening M-phase).			Cyclin A is required for mitotic entry in primary human fibroblasts [29], and its depletion causes re-replication in HeLa cells [25].

↗	<b>Cdh1</b>	80% $GF_H$	<b>Cdh1</b> inhibition drastically limits the rate of proliferation.			Inhibition of <i>Cdh1</i> binding to APC/C led to significantly prolonged mitosis maintained by a functioning SAC, with no without any spindle defects [30].
↘	<b>CyclinB</b>	65% $GF_H$	Interestingly, our two modeling frameworks do not agree about the effects of <i>CyclinB</i> depletion. <b>Synchronous model:</b> <i>CyclinB</i> knockdown results in severe mitotic defects, where cells start anaphase without passing the SAC — a results supported by evidence. <b>Asynchronous model:</b> <i>CyclinB</i> knockdown predominantly leads to DNA re-duplication from G2 — also supported.			Depletion of cyclin B1/B2 induces a strong increase of G2 cells and increase in a cycling population with double DNA content. In addition, strong mitotic chromosomal aberration such as mal-oriented chromosomes were observed at metaphase, and cells entered anaphase without reaching the SAC [31].
↘	<b>Cdc25C</b>	95% $GF_H$	<b>Cdc25C</b> inhibition limits normal proliferation and (potently) induces aberrant mitotic progression, leading to aneuploidy. In addition, it enriches for cells that do not complete cytokinesis.			<b>chromosome mis-segregation:</b> Excess Cdc25C M phase degradation in a mouse model that impacts Cdk1 activity lead accelerated mitotic progression, failure to arrest at SAC and improper chromosome segregation [32]. <b>failure of cytokinesis: prediction</b>
↘	<b>Mad2</b>	80% $GF_H$	<b>Mad2</b> inhibition in cells that are not exposed to microtubule damage has no effect on cell cycle progression. This result is and artifact of the fact that we do not model the “trial-and-error” nature of chromosome attachments to the spindle, and thus the mechanism that monitors this has little effect on the model.			Mad2 or BubR1 depletion leads to acceleration of mitosis in all cells whereas the depletion of Mad1, Bub1, or Bub3 leaves timing unaffected. The same is also true if the recruitment of Mad2 or BubR1 to kinetochores is blocked by RNAi [33].

Apoptotic Switch						
↓	<b>DR4_5</b>	95% <i>Trail</i> ( $GF_H$ ON)	Partial knockdown of the death receptors <b>DR4</b> and <b>DR5</b> decrease the fraction of cycling and quiescent cells that undergo <i>Trail</i> -mediated apoptosis.			An adenovirus shown to intrude internalization of DR4 and DR5 from the cell surface blocs <i>Trail</i> -induced apoptosis [34].
		95% <i>Trail</i> ( $GF_H$ OFF)				
↓	<b>Casp8</b>	95% <i>Trail</i> ( $GF_H$ OFF)	<b>Caspase 8</b> knockdown decreases the fraction of cycling and quiescent cells that undergo <i>Trail</i> -mediated apoptosis.			Knock down or deletion of Caspase-8 suppressed <i>Trail</i> -induced apoptosis [35]
↗	<b>BCL2</b>	95% <i>Trail</i> ( $GF_H$ OFF)	<b>BCL2</b> over-expression decreases the fraction of quiescent cells that undergo <i>Trail</i> -mediated apoptosis even under near-saturating <i>Trail</i> exposure.			Overexpression of Bcl-2 or Bcl-XL has been shown to inhibit TRAIL-induced apoptosis in multiple studies [36].
↗	<b>BCL-XL</b>	20% <i>GF</i> (no $GF_H$ )	<b>BCL-XL</b> over-expression decreases the fraction of quiescent cells that undergo growth factor withdrawal mediated apoptosis.			
↗	<b>Casp2</b>	65% <i>GF</i> (no $GF_H$ )	<b>Caspase 2</b> hyper-activation potentiates apoptosis in cells exposed to weakened survival signals.			Over-expression of the active, truncated form of Caspase2 triggers apoptosis [37].

↘	<b>BAD</b>	20% GF (no GF <sub>H</sub> )	<b>BAD</b> inhibition blocks apoptosis due to the withdrawal of survival signals.			Reduction of BAD expression by RNAi prevents apoptosis in response to the inhibition of survival pathways [38].
↘	<b>tBID</b>	95% Trail (GF <sub>H</sub> OFF)	<b>BID</b> inhibition blocks <i>Trail</i> -induced apoptosis in quiescent cells.			Depletion / over-expression of Bid in human ovarian carcinoma cells decreased / increased TRAIL-mediated apoptosis [39].
↘	<b>BAK</b>	20% Trail (GF <sub>H</sub> OFF)	<b>BAK</b> inhibition weakens apoptosis in response to <i>Trail</i> in quiescent and cycling cells.			BAK-null cells show deficient Smac/DIABLO release from mitochondria in response to Trail, and show decreased rates of apoptosis [40].
↘	<b>BAX</b>	95% Trail (GF <sub>H</sub> OFF)	<b>BAX</b> inhibition does not alter apoptosis in response to <i>Trail</i> . This model behavior is not supported by experiments, and it is a likely artifact of the fact that in the model BAK alone can trigger MOMP.			BAX-null cells also show deficient Smac/DIABLO release from mitochondria in response to <i>Trail</i> , and show decreased rates of apoptosis (more potent than BAK) [40].
↘	<b>IAPs</b>	95% Trail (GF <sub>H</sub> OFF) 50% GF (GF <sub>H</sub> OFF)	Inhibition of IAP proteins increases the rate of apoptosis in response to Trail (quiescent and cycling cells) as well as in response to the withdrawal of survival signaling.	 	 	Down-regulation of all IAPs simultaneously induces apoptosis in multiple melanoma cell lines, and sensitized them to both FASL and Trail [41].

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