

CORRECTION

Correction: ADAMTS5 Is a Critical Regulator of Virus-Specific T Cell Immunity

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An influenza-specific CD8+ T cell peptide encoded within the nucleoprotein (NP) was incorrectly referred to as NP366-372 throughout the article. The correct designation should be NP366-374. The peptide sequence ASNENMETM stated throughout the 'Methods' section should also be attributed to NP366-374.

The authors have provided corrected versions of [Fig 3](#), [Fig 4](#), [Fig 5](#), [Fig 8](#), [S10 Fig](#) and [S11 Fig](#).



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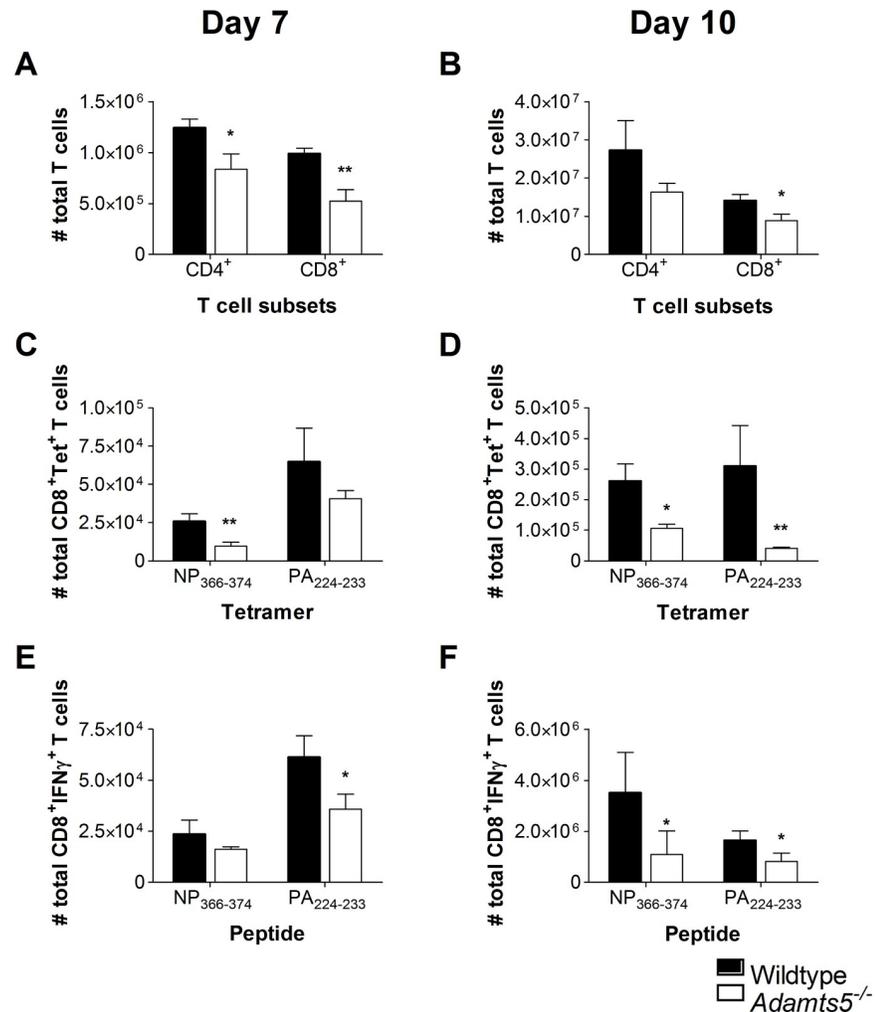


Fig 3. Reduced CD4⁺ and CD8⁺ T cell numbers in the spleens of influenza-infected *Adamts5*^{-/-} mice. C57.BL/6 and *Adamts5*^{-/-} mice were intranasally (i.n.) infected (10⁴ pfu/mouse) with X31 (H3N2) influenza virus. Spleens were removed at day 7 and day 10 p.i., and CD8⁺ T cell responses determined. Total CD4⁺ and CD8⁺ T cells numbers were calculated at days (A) 7 and (B) 10 p.i. Influenza-specific D^bNP₃₆₆₋₃₇₄ CD8⁺ and D^bPA₂₂₄₋₂₃₃ CD8⁺ tetramer-positive T cell numbers were measured at days (C) 7 and (D) 10 p.i. Functional influenza-specific D^bNP₃₆₆₋₃₇₄IFN γ ⁺CD8⁺ and D^bPA₂₂₄₋₂₃₃IFN γ ⁺CD8⁺ T cell activity was determined by ICS, and total IFN γ ⁺ T cells enumerated at days (E) 7 and (F) 10 after infection. WT denotes C57.BL/6. The results are expressed as means \pm SD, and statistical significance (relative to C57.BL/6) was determined by Student's *t* test (**p* \leq 0.05, ***p* \leq 0.01, *n* = 5 representing three experiments). Underlying data are provided in S1 Data.

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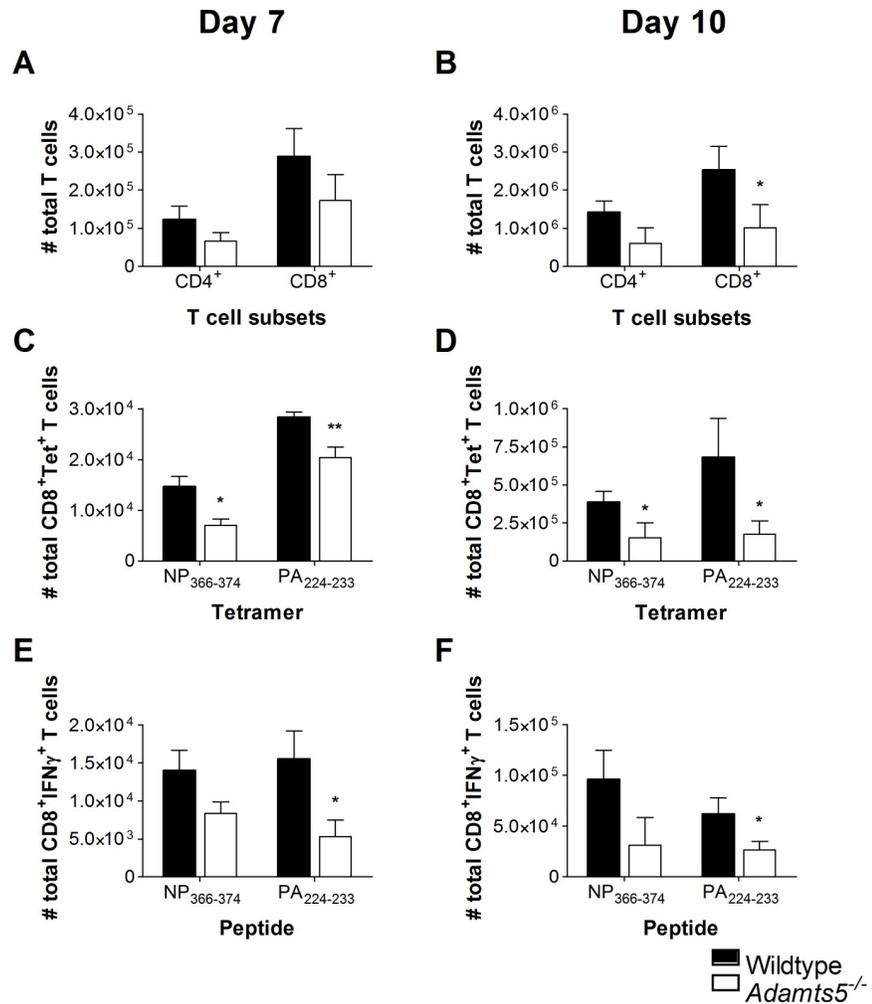


Fig 4. CD4⁺ and CD8⁺ T cell responses in the lungs of influenza-infected *Adamts5*^{-/-} mice. C57.BL/6 and *Adamts5*^{-/-} mice were infected i.n. with 10⁴pfu X31 (H3N2) influenza virus. Lungs were removed at days 7 and 10 p.i., and CD8⁺ T cell immunity characterised. Total CD4⁺ and CD8⁺ T cell numbers are shown at days (A) 7 and (B) 10 p.i. Influenza-specific tetramer⁺ D^bNP₃₆₆₋₃₇₄⁺ CD8⁺ and D^bPA₂₂₄₋₂₃₃⁺ CD8⁺ T cell responses were enumerated at days (C) 7 and (D) 10 p.i. CD8⁺ T cell functionality was assessed by ICS and D^bNP₃₆₆₋₃₇₄⁺IFN γ ⁺CD8⁺ and D^bPA₂₂₄₋₂₃₃⁺IFN γ ⁺CD8⁺ T cell responses enumerated at days (E) 7 and (F) 10 p.i. Wildtype denotes C57.BL/6 mice. The results are expressed as means \pm SD, and statistical significance (relative to C57.BL/6) was determined by Student's *t* test (* = $p \leq 0.05$, ** = $p \leq 0.01$, $n = 5$ representing three experiments). Underlying data are provided in S1 Data.

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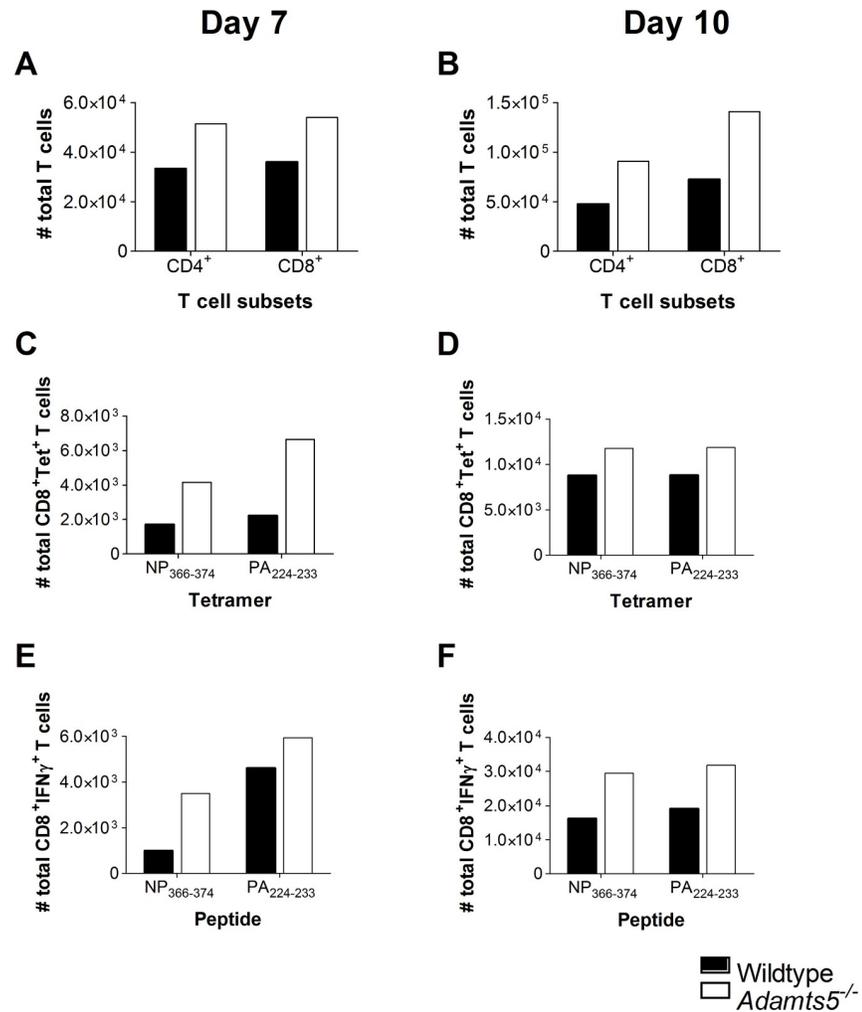


Fig 5. T cell immunity in the pooled MLN. C57.BL/6 and *Adamts5*^{-/-} mice were infected i.n. with 10⁴ pfu/mouse X31 (H3N2) influenza virus. MLNs were removed, pooled, and processed at days 7 and 10 p.i., and single-cell suspensions analysed for influenza-specific immunity. Total CD4⁺ and CD8⁺ T cell numbers were determined at days (A) 7 and (B) 10 p.i. Influenza-specific D^bNP₃₆₆₋₃₇₄⁺ CD8⁺ and D^bPA₂₂₄₋₂₃₃⁺ CD8⁺ tetramer positive T cells were enumerated at days (C) 7 and (D) 10 p.i. CD8⁺ T cell functionality was measured using ICS. Influenza-specific D^bNP₃₆₆₋₃₇₄⁺IFN γ ⁺CD8⁺ and D^bPA₂₂₄₋₂₃₃⁺IFN γ ⁺CD8⁺ T cell responses were characterised at days (E) 7 and (F) 10 p.i. Results are expressed as total pooled means from five mice repeated three times. Wildtype denotes C57.BL/6 mice. Underlying data are provided in S1 Data.

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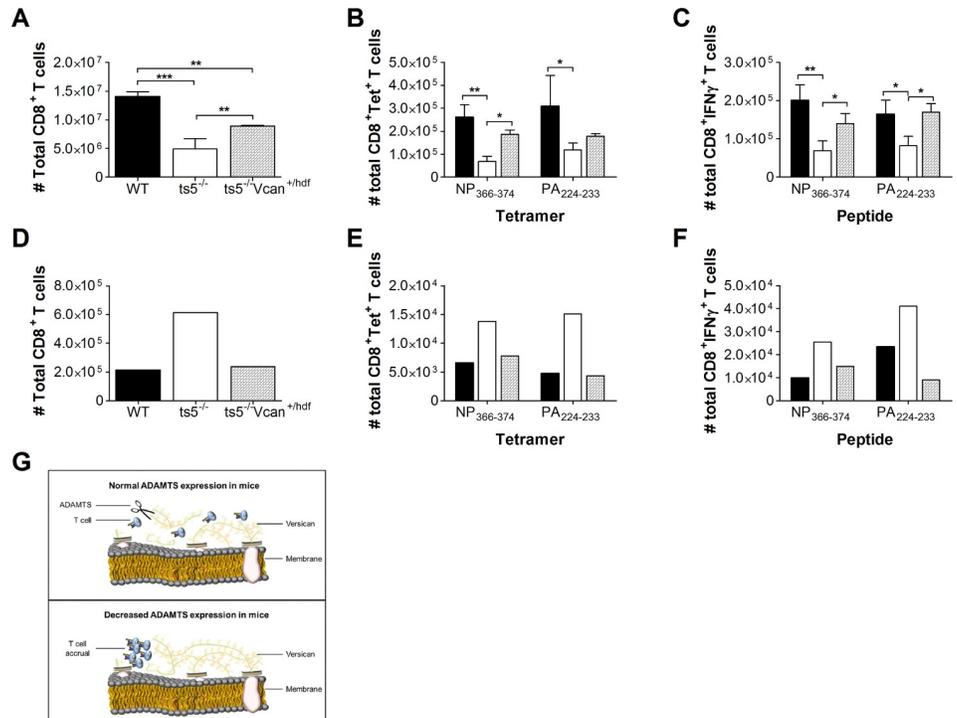


Fig 8. Versican reduction in *Adamts5*^{-/-}*Vcan*^{+/hdf} mice restores normal T cell function. Spleen and MLNs were removed from C57.BL/6, *Adamts5*^{-/-}*Vcan*^{+/+}, and *Adamts5*^{-/-}*Vcan*^{+/hdf} mice and processed at day 10 p.i., and single cell suspensions were analysed for influenza-specific immunity. (A) Total CD8⁺ T cell numbers were determined at day 10 p.i. in the spleen. (B) Influenza-specific D^bNP₃₆₆₋₃₇₄⁺ CD8⁺ and D^bPA₂₂₄₋₂₃₃⁺ CD8⁺ tetramer positive T cells in the spleen were enumerated at day 10 p.i. CD8⁺ T cell functionality was measured using ICS. (C) Influenza specific D^bNP₃₆₆₋₃₇₄⁺IFNγ⁺CD8⁺ and D^bPA₂₂₄₋₂₃₃⁺IFNγ⁺CD8⁺ T cell responses were characterised in the spleen at days 10 p.i. (D) Total CD8⁺ T cell numbers were assessed at day 10 p.i. in the pooled MLN. (E) Influenza-specific D^bNP₃₆₆₋₃₇₄⁺ CD8⁺ and D^bPA₂₂₄₋₂₃₃⁺ CD8⁺ tetramer positive T cells in the pooled MLN were enumerated at day 10 p.i. CD8⁺ T cell functionality was measured using ICS. (F) Influenza-specific D^bNP₃₆₆₋₃₇₄⁺IFNγ⁺CD8⁺ and D^bPA₂₂₄₋₂₃₃⁺IFNγ⁺CD8⁺ T cell responses were characterised in the pooled MLN at day 10 p.i. The results are expressed as means ± SD (spleen data) or as pooled means (MLN data), and statistical significance (relative to C57.BL/6 mice) was determined by one-way ANOVA (**p* ≤ 0.05, ****p* ≤ 0.005 relative to C57.BL/6, *n* = 5 representing three individual experiments). WT denotes C57.BL/6 mice and *ts5*^{-/-} denotes *Adamts5*^{-/-}. (G) Our model for ADAMTS5 enzyme activity and T cell migration proposes that versican can inhibit T cell effector function by acting as a physical block. Cleavage of versican by ADAMTS5 removes the ECM blockade, allowing migration (top panel). Moreover, versican accumulation in the absence of ADAMTS5 enzyme activity results in T cell clustering (bottom panel). Underlying data are provided in S1 Data.

<https://doi.org/10.1371/journal.pbio.3000558.g004>

The relevant figure legends have been amended to reflect this change and are presented below.

Supporting information

S10 Fig. Influenza virus infection of *Vcan*^{+/hdf} mice. Lung tissue and MLNs were removed from influenza virus infection C57.BL/6 and *Vcan*^{+/hdf} mice and processed to generate single cell suspensions at day 10 p.i. for analysis of influenza-specific immunity. (A) Total CD8⁺ T cell numbers were determined at day 10 p.i. in the lung. (B) Influenza-specific D^bNP₃₆₆₋₃₇₄⁺ CD8⁺ and D^bPA₂₂₄₋₂₃₃⁺ CD8⁺ tetramer positive T cells in the lung were enumerated at day 10 p.i. CD8⁺ T cell functionality was measured using ICS. (C) Influenza specific D^bNP₃₆₆₋₃₇₄⁺IFNγ⁺CD8⁺ and D^bPA₂₂₄₋₂₃₃⁺IFNγ⁺CD8⁺ T cell responses were characterised in the lung at day 10 p.i. (D) Total CD8⁺ T cell numbers were determined at day 10 p.i. from pooled MLN samples. (E) Influenza-

specific D^bNP₃₆₆₋₃₇₄⁺ CD8⁺ and D^bPA₂₂₄₋₂₃₃⁺ CD8⁺ tetramer positive T cells in pooled MLN were enumerated at day 10 p.i. (F) CD8⁺ T cell functionality was measured using ICS to assess influenza-specific D^bNP₃₆₆₋₃₇₄⁺IFNγ⁺CD8⁺ and D^bPA₂₂₄₋₂₃₃⁺IFNγ⁺CD8⁺ T cell responses at day 10 p.i. The results are expressed as means ± SD or as pooled means (MLN data) and statistical significance (relative to C57.BL/6 mice) determined by a Student's *t* test (**p* ≤ 0.05, ****p* ≤ 0.005 relative to C57.BL/6, *n* = 5 representing three individual experiments). WT denotes C57.BL/6 mice. Underlying data are provided in S2 Data.

(TIF)

S11 Fig. Influenza infection of *Adamts5*^{-/-} and WT littermate controls. *Adamts5*^{-/-} and WT mice were infected i.n with X31 (H3N2) influenza virus and spleens, lungs, and MLNs removed from C57.BL/6 and *Adamts5*^{-/-} mice days 7 p.i. Single cell suspensions were then analysed for influenza-specific immunity. (A) Weight loss was calculated over the time course of infection. Total CD4⁺ and CD8⁺ T cells were enumerated in the (B) spleen, (C) lung, and (D) MLN. Influenza-specific D^bNP₃₆₆₋₃₇₄⁺ CD8⁺ and D^bPA₂₂₄₋₂₃₃⁺ CD8⁺ tetramer positive T cell numbers were also characterised in the (B) spleen, (C) lung, and (D) MLN. Lung and spleen results are expressed as means ± SD or as pooled means (MLN data), and statistical significance (relative to C57.BL/6 mice) was determined by a Student's *t* test (**p* ≤ 0.05, ***p* ≤ 0.01 relative to C57.BL/6 mice, *n* = 5 representing three individual experiments). WT denotes C57.BL/6 mice. Underlying data are provided in S2 Data.

(TIF)

Reference

1. McMahon M, Ye S, Izzard L, Dlugolenski D, Tripp RA, Bean AGD, et al. (2016) ADAMTS5 Is a Critical Regulator of Virus-Specific T Cell Immunity. *PLoS Biol* 14(11): e1002580. <https://doi.org/10.1371/journal.pbio.1002580> PMID: 27855162