PEARLS

Mechanisms of Host Behavioral Change in *Toxoplasma gondii* Rodent Association

Ajai Vyas*

School of Biological Sciences, Nanyang Technological University, Singapore

* avyas@ntu.edu.sg

Introduction

The behavioral manipulation hypothesis predicts that parasites can change host behavior in a way that benefits the parasites and not the host (extensively reviewed in [1-9]). In other words, the hypothesis predicts that genes of a parasite can produce an "extended" phenotype that manifests beyond a parasite's soma [10]. Protozoan parasite *Toxoplasma gondii* (henceforth toxoplasma) is an often-cited example. Chronic toxoplasma infection reduces aversion of rodents to cat odors, plausibly increasing predation by its definitive felid host [11]. Here, I enumerate main narratives that have emerged in the past decade about biological mechanisms of behavioral change in rodents after toxoplasma infection.

Cats are infected by toxoplasma when they eat infected prey. The parasite undergoes gametogenesis in cat intestines, resulting in eventual shedding of fecal oocysts that are ingested by intermediate hosts. Entry in the cat is important for the parasite because it permits a) sexual recombination; b) infection of herbivore hosts who otherwise cannot be infected through carnivory between intermediate hosts; and c) the discharge of highly infectious and resilient oocysts into the environment. Yet, entry of the parasite in the cat is constrained by predation rates. Preys of cats avoid cats and cat odors [12]. Apropos, toxoplasma infection leads to reduced aversion of rodents to cat odors [11]. A subset of animals also develops an atypical and "fatal" attraction [11,13]. These behavioral observations suggest, but do not prove, that the parasite creates an extended phenotype in the host behavior. The caution in the preceding sentence is necessary because it is yet unknown if infected rodents are indeed predated more frequently by cats.

Toxoplasma is also sexually transmitted through the male ejaculate in rats [14]. Apropos, male rats infected with toxoplasma become more attractive to females [15]. Uninfected females spend greater time near infected males and allow them greater reproductive access [14]. These observations suggest a second parasitic manipulation of the host behavior, whereby being infected creates greater avenues for sexual transmission of the parasite itself [9].

Biological pathways underlying mate choice and innate aversion to predator odor are relatively well-studied in rodents. This has allowed researchers to study proximate mechanisms of parasitic behavioral manipulation in greater detail in this association compared to other host– parasite relationships. This mechanistic research has focused on three main narratives.

Narrative #1: Tropism to Specific Regions of the Brain

This narrative posits that toxoplasma preferentially concentrates in certain brain regions; and this tropism can explain host behavioral changes through local manipulation of neuronal signaling and/or damage. Toxoplasma exhibits a decided tropism to brain, testes, and eyes. These



OPEN ACCESS

Citation: Vyas A (2015) Mechanisms of Host Behavioral Change in *Toxoplasma gondii* Rodent Association. PLoS Pathog 11(7): e1004935. doi:10.1371/journal.ppat.1004935

Editor: Laura J Knoll, University of Wisconsin Medical School, UNITED STATES

Published: July 23, 2015

Copyright: © 2015 Ajai Vyas. This is an open access article distributed under the terms of the <u>Creative</u> <u>Commons Attribution License</u>, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Funding: This work was supported by grants from Ministry of Education, Singapore. The funder had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing Interests: The author has declared that no competing interests exist.

organs are immune-privileged, in the sense that immune cells have limited access to these sites. Some experimental evidence suggests that toxoplasma gains entry into these sites through brain endothelial cells [16] or by using dendritic immune cells as Trojan horses [17]. The parasite then forms bradyzoite-containing tissue cysts that undergo periodic cycles of rupture and encystment (i.e., recrudescence). Several studies have mapped sites of encystment within the brain, with the hope that location of these cysts has some bearing on the host behavior. Two earliest studies in this regard reported a rather widespread occurrence of tissue cysts in a variety of brain regions [13,18]. Both of these reports suggested a mild tropism to nucleus accumbens, ventromedial hypothalamus, or amygdala. These brain structures are involved in decision making and generation of fear [19,20]. Thus, the suggestion was that presence of cysts somehow compromised normal functioning of these brain regions and led to deficits in processing of fear or decision-making capacities. Yet, subsequent detailed analysis of cyst distribution failed to reveal substantial tropism in any of these three brain structures in mice and rats, instead reporting a rather "probabilistic" spread of the parasites [21]. Even more surprisingly, further experiments demonstrated that change in host behavior could be observed even after extensive clearance of parasite cysts within brain [22]. This suggests that toxoplasma tropism or lack of tropism does not have a causal relationship with the behavioral change. A nontropic model must thus be sought. Opinions still remain divided on this issue. For example, it has been suggested that within infected animals, those with presence of parasitic cysts within certain brain regions experience greater magnitude of behavioral change [23]. Another possibility in this regard is that toxoplasma could "coopt" brain cells without invading them [24]. The parasite is known to inject effector proteins inside host cells during early phases of invasion [24]. In many of the brain cells, the parasite injects these effector proteins but then does not, or fails to, gain residence. Such covert tropism will be difficult to detect by mere enumeration of parasite presence in various brain regions. Excitingly, new transgenic methods now allow visualization of "toxoplasma-kissed" neurons [25], although selective tropism of such events is presently unstudied.

Narrative #2: Disruption of Dopamine Signaling in the Brain

An important candidate in the nontropic model is disruption of brain dopamine signaling. The genome of toxoplasma contains two genes (AAH1 and AAH2) that bear striking sequence similarity to a mammalian enzyme called tyrosine hydroxylase [26]. This enzyme in mammals catalyzes conversion of L-tyrosine to L-3,4-dihydroxyphenylalanine. This reaction is a ratelimiting step in synthesis of dopamine. While dopamine is often presented as the neurotransmitter that signals reward or pleasure, its more parsimonious role in the brain is in motivation and goal-directed behaviors [27]. The narrative then is that toxoplasma increases dopamine signaling in the host brain by virtue of supplying a rate-limiting enzyme for its synthesis [28,29]. This increase in dopamine signaling then interferes with host behavior, creating atypical motivation to explore predator odors. In support of this, drugs that interfere with the binding of dopamine to its receptors ameliorate effects of the infection on aversion to predator odors [30]. Tissue cysts within the infected brain contain high amounts of dopamine, and cultured dopaminergic cells secrete greater dopamine when infected with toxoplasma, suggesting the possibility of hyperdopaminergic drive in the infected brain [28]. This hypothesis does not necessarily require tropism of the parasite within brain regions that endogenously produce this neurotransmitter. A stochastic distribution of the parasite will provide tyrosine hydroxylase to a wide variety of brain regions. Yet, it is unlikely that nondopaminergic neurons will contain complementary enzymes required for dopamine synthesis. Moreover, any residual dopamine production in nonendogenous circuits is unlikely to have any effect because of lack of

dopamine receptors in downstream efferent. In short, an intersection of generalized increase in tyrosine hydroxylase in brain with specific distribution of complementary proteins in endogenous dopaminergic circuit could create a rather specific behavior alteration. This hypothesis can reconcile lack of strong tropism of the parasite. Yet it obligatorily requires persistent presence of the parasite within brain. The relationship of dopamine alteration and increase in sexual attractiveness of infected males also remains unclear at present. A definitive proof of this hypothesis will require a future experiment with possible demonstration that disruption of parasitic tyrosine hydroxylase genes within brain necessarily results in loss of host behavioral change without affecting parasite survival itself.

Recently, a mutant parasite with ablation of one of the AAH genes (AAH2) has been described [29]. This ablation does not affect parasite viability, invasion, and transmission. It will be interesting to ask if infection with this mutant still causes host behavioral change. Contrary to predictions from dopaminergic mediation, infection with wild-type or Δ AAH2 mutant toxoplasma did not result in greater dopamine in mice brain. Similarly, overexpression of AAH2 genes in cell culture failed to augment dopamine content [29]. A double mutant parasite lacking both AAH genes has still not been successfully created.

Narrative #3: Hormonal Upheavals and Concomitant Epigenetic Changes

An alternative nontropic model invokes parasite-induced tweaking of communication lines between brain and gonadal hormones. Toxoplasma invades rat testes upon infection, leading to a heavy cyst burden in epididymis and the ejaculates [14]. This is not surprising because the testes, like the brain, is an immune-privileged site. The infection results in up-regulation of testosterone synthesis within Leydig cells of the testes [31]. The narrative here posits that greater testosterone synthesis results in two simultaneous effects. One, it increases synthesis of male sexual pheromone by virtue of androgen dependence of these molecules [15]. Two, excess testosterone shifts the host towards sexual behaviors and away from defensive behaviors [32]. In support of this narrative, castrating male rats before infection prevents host behavioral change [31]. An important caveat in this experiment is the possibility that removal of testosterone by castration can potentially increase tonicity of the immune response during the acute phase of the infection, thereby changing the course of the infection. An unequivocal demonstration of the direct role of testosterone would require selective ablation of testosterone "increase" postinfection rather than removal of all testicular steroidogenesis preinfection.

This narrative nonetheless provides a plausible chain of events. In male rats, testosterone sustains synthesis of major urinary proteins in the liver. These proteins are necessary and sufficient to signal sexual attractiveness in a dose-dependent manner to females when eventually excreted in the urine [33]. Testosterone and/or its metabolic derivatives bind to their receptor found in the medial amygdala, a brain region involved in signaling presence of sexual opportunities [34,35]. This brain region contains population of neurons expressing arginine vasopressin, a neurotransmitter that mediates reproductive behaviors in many species (e.g., [36,37]). Male rats infected with toxoplasma exhibit reduced DNA methylation in promoter sites upstream of the arginine vasopressin gene in the medial amygdala, resulting in its greater production [32]. This is akin to observations noted during testosterone supplementation in uninfected rats [38]. These arginine vasopressin neurons are typically recruited during copulation or sensory stimulation by female presence in uninfected rats [37]. Atypically, the same population of neurons becomes activated during exposure to cat odors in infected male rats. Furthermore, pharmacological mimicry of this molecular change institutes decreased predator aversion akin to the effects of toxoplasma infection [32]. Thus, the proposal here is that the

increase in testosterone synthesis mediates both aspects of behavioral manipulation of sexual attractiveness and reduction in aversion to cats. The presence of the parasite within the brain or its tropism to certain brain regions becomes merely incidental and non-necessary in this narrative.

The narrative detailed above encompasses substrates that are highly dimorphic between genders. Congruent to males, toxoplasma also reduces aversion to cat odors in female mice and rats [13,39]. Thus, proximate mechanisms involving gender-dimorphic biology needs to be yet reconciled with gender-nondimorphic behavioral effects of the parasitism. Mechanisms of the behavioral change in females are currently understudied, though preliminary evidence suggests changes in progesterone levels postinfection [39].

Conclusions

Rats and mice infected with toxoplasma exhibit behavioral change in their aversion to cat odor and their sexual attractiveness to females. Previous work has resulted in three main classes of hypotheses pertaining to proximate mechanism of this phenomenon. Current work continues to test these hypotheses. More clarity about the mechanisms will plausibly inform ultimate causation of host behavioral change.

References

- Adamo SA. Parasites: evolution's neurobiologists. J Exp Biol. 2013 Jan 1; 216(Pt 1):3–10. doi: <u>10.</u> <u>1242/jeb.073601</u> PMID: <u>23225861</u>
- 2. Flegr J. Influence of latent Toxoplasma infection on human personality, physiology and morphology: pros and cons of the Toxoplasma-human model in studying the manipulation hypothesis. J Exp Biol. 2013 Jan 1; 216(Pt 1):127–33. doi: 10.1242/jeb.073635 PMID: 23225875
- 3. Hughes D. Pathways to understanding the extended phenotype of parasites in their hosts. J Exp Biol. 2013 Jan 1; 216(Pt 1):142–7. doi: 10.1242/jeb.077461 PMID: 23225877
- Knight K. How pernicious parasites turn victims into zombies. J Exp Biol. 2013 Jan 1; 216(Pt 1):i–iv. PMID: 23355981
- Lafferty KD, Shaw JC. Comparing mechanisms of host manipulation across host and parasite taxa. J Exp Biol. 2013 Jan 1; 216(Pt 1):56–66. doi: <u>10.1242/jeb.073668</u> PMID: <u>23225868</u>
- Libersat F, Gal R. What can parasitoid wasps teach us about decision-making in insects? J Exp Biol. 2013 Jan 1; 216(Pt 1):47–55. doi: 10.1242/jeb.073999 PMID: 23225867
- McConkey GA, Martin HL, Bristow GC, Webster JP. Toxoplasma gondii infection and behaviour location, location, location? J Exp Biol. 2013 Jan 1; 216(Pt 1):113–9. doi: <u>10.1242/jeb.074153</u> PMID: <u>23225873</u>
- Poulin R. Parasite manipulation of host personality and behavioural syndromes. J Exp Biol. 2013 Jan 1; 216(Pt 1):18–26. doi: <u>10.1242/jeb.073353</u> PMID: <u>23225863</u>
- 9. Vyas A. Parasite-augmented mate choice and reduction in innate fear in rats infected by Toxoplasma gondii. J Exp Biol. 2013 Jan 1; 216(Pt 1):120–6. doi: <u>10.1242/jeb.072983</u> PMID: <u>23225874</u>
- 10. Dawkins R. The extended phenotype: The long reach of the gene: Oxford University Press; 1999.
- Berdoy M, Webster JP, Macdonald D. Fatal attraction in rats infected with Toxoplasma gondii. Proceedings of the Royal Society of London Series B: Biological Sciences. 2000; 267(1452):1591–4. PMID: <u>11007336</u>
- Dielenberg RA, McGregor IS. Defensive behavior in rats towards predatory odors: a review. Neuroscience & Biobehavioral Reviews. 2001; 25(7):597–609.
- Vyas A, Kim SK, Giacomini N, Boothroyd JC, Sapolsky RM. Behavioral changes induced by Toxoplasma infection of rodents are highly specific to aversion of cat odors. Proceedings of the National Academy of Sciences of the United States of America. 2007 Apr 10; 104(15):6442–7. PMID: <u>17404235</u>
- Dass SAH, Vasudevan A, Dutta D, Soh LJT, Sapolsky RM, Vyas A. Protozoan parasite Toxoplasma gondii manipulates mate choice in rats by enhancing attractiveness of males. PLoS One. 2011; 6(11): e27229. doi: <u>10.1371/journal.pone.0027229</u> PMID: <u>22073295</u>
- Vasudevan A, Kumar V, Chiang YN, Yew JY, Cheemadan S, Vyas A. α2u-globulins mediate manipulation of host attractiveness in Toxoplasma gondii–Rattus novergicus association. The ISME Journal. 2015.

- Lachenmaier SM, Deli MA, Meissner M, Liesenfeld O. Intracellular transport of Toxoplasma gondii through the blood-brain barrier. Journal of neuroimmunology. 2011 Mar; 232(1–2):119–30. doi: <u>10.</u> <u>1016/j.jneuroim.2010.10.029</u> PMID: <u>21106256</u>
- Bierly AL, Shufesky WJ, Sukhumavasi W, Morelli AE, Denkers EY. Dendritic cells expressing plasmacytoid marker PDCA-1 are Trojan horses during Toxoplasma gondii infection. Journal of immunology (Baltimore, Md: 1950). 2008 Dec 15; 181(12):8485–91.
- Gonzalez LE, Rojnik B, Urrea F, Urdaneta H, Petrosino P, Colasante C, et al. < i> Toxoplasma gondii infection lower anxiety as measured in the plus-maze and social interaction tests in rats: A behavioral analysis. Behavioural brain research. 2007; 177(1):70–9. PMID: <u>17169442</u>
- Dielenberg RA, Hunt GE, McGregor IS. "When a rat smells a cat": the distribution of Fos immunoreactivity in rat brain following exposure to a predatory odor. Neuroscience. 2001; 104(4):1085–97. PMID: <u>11457592</u>
- McGregor IS, Hargreaves GA, Apfelbach R, Hunt GE. Neural correlates of cat odor-induced anxiety in rats: region-specific effects of the benzodiazepine midazolam. The Journal of neuroscience: the official journal of the Society for Neuroscience. 2004 Apr 28; 24(17):4134–44.
- Berenreiterova M, Flegr J, Kubena AA, Nemec P. The distribution of Toxoplasma gondii cysts in the brain of a mouse with latent toxoplasmosis: implications for the behavioral manipulation hypothesis. PLoS One. 2011; 6(12):e28925. doi: <u>10.1371/journal.pone.0028925</u> PMID: <u>22194951</u>
- Ingram WM, Goodrich LM, Robey EA, Eisen MB. Mice infected with low-virulence strains of Toxoplasma gondii lose their innate aversion to cat urine, even after extensive parasite clearance. PloS one. 2013; 8(9):e75246. doi: 10.1371/journal.pone.0075246 PMID: 24058668
- Evans AK, Strassmann PS, Lee IP, Sapolsky RM. Patterns of Toxoplasma gondii cyst distribution in the forebrain associate with individual variation in predator odor avoidance and anxiety-related behavior in male Long-Evans rats. Brain, behavior, and immunity. 2014 Mar; 37:122–33. doi: <u>10.1016/j.bbi</u>. <u>2013.11.012</u> PMID: <u>24269877</u>
- Koshy AA, Dietrich HK, Christian DA, Melehani JH, Shastri AJ, Hunter CA, et al. Toxoplasma co-opts host cells it does not invade. PLoS pathogens. 2012; 8(7):e1002825. doi: <u>10.1371/journal.ppat.</u> 1002825 PMID: 22910631
- Koshy AA, Fouts AE, Lodoen MB, Alkan O, Blau HM, Boothroyd JC. Toxoplasma secreting Cre recombinase for analysis of host-parasite interactions. Nature methods. 2010; 7(4):307–9. doi: <u>10.1038/</u> <u>nmeth.1438</u> PMID: <u>20208532</u>
- Gaskell EA, Smith JE, Pinney JW, Westhead DR, McConkey GA. A unique dual activity amino acid hydroxylase in Toxoplasma gondii. PLoS One. 2009; 4(3):e4801. doi: <u>10.1371/journal.pone.0004801</u> PMID: <u>19277211</u>
- 27. Salamone JD, Correa M. The mysterious motivational functions of mesolimbic dopamine. Neuron. 2012 Nov 8; 76(3):470–85. doi: 10.1016/j.neuron.2012.10.021 PMID: 23141060
- Prandovszky E, Gaskell E, Martin H, Dubey J, Webster JP, McConkey GA. The neurotropic parasite Toxoplasma gondii increases dopamine metabolism. PLoS One. 2011; 6(9):e23866. doi: <u>10.1371/</u> journal.pone.0023866 PMID: <u>21957440</u>
- Wang ZT, Harmon S, O'Malley KL, Sibley LD. Reassessment of the role of aromatic amino acid hydroxylases and the effect of infection by Toxoplasma gondii on host dopamine levels. Infection and immunity. 2014 Dec 29.
- Webster JP, Kaushik M, Bristow GC, McConkey GA. Toxoplasma gondii infection, from predation to schizophrenia: can animal behaviour help us understand human behaviour? The Journal of experimental biology. 2013; 216(1):99–112.
- Lim A, Kumar V, Hari Dass SA, Vyas A. Toxoplasma gondii infection enhances testicular steroidogenesis in rats. Molecular ecology. 2013; 22(1):102–10. doi: <u>10.1111/mec.12042</u> PMID: <u>23190313</u>
- Hari Dass SA, Vyas A. Toxoplasma gondii infection reduces predator aversion in rats through epigenetic modulation in the host medial amygdala. Molecular ecology. 2014; 23(24):6114–22. doi: <u>10.1111/</u> mec.12888 PMID: 25142402
- Kumar V, Vasudevan A, Soh LJT, Le Min C, Vyas A, Zewail-Foote M, et al. Sexual Attractiveness in Male Rats Is Associated with Greater Concentration of Major Urinary Proteins. Biology of reproduction. 2014:biolreprod. 114.117903.
- 34. Baum MJ, Bakker J. Roles of sex and gonadal steroids in mammalian pheromonal communication. Frontiers in neuroendocrinology. 2013 Oct; 34(4):268–84. doi: <u>10.1016/j.yfrne.2013.07.004</u> PMID: <u>23872334</u>
- Petrulis A. Chemosignals, hormones and mammalian reproduction. Horm Behav. 2013 May; 63(5): 723–41. doi: 10.1016/j.yhbeh.2013.03.011 PMID: 23545474

- Ho JM, Murray JH, Demas GE, Goodson JL. Vasopressin cell groups exhibit strongly divergent responses to copulation and male-male interactions in mice. Hormones and behavior. 2010; 58(3): 368–77. doi: 10.1016/j.yhbeh.2010.03.021 PMID: 20382147
- **37.** Dass SAH, Vyas A. Copulation or sensory cues from the female augment Fos expression in arginine vasopressin neurons of the posterodorsal medial amygdala of male rats. Frontiers in zoology. 2014; 11(1):42.
- Auger CJ, Coss D, Auger AP, Forbes-Lorman RM. Epigenetic control of vasopressin expression is maintained by steroid hormones in the adult male rat brain. Proceedings of the National Academy of Sciences. 2011; 108(10):4242–7.
- Golcu D, Gebre RZ, Sapolsky RM. Toxoplasma gondii influences aversive behaviors of female rats in an estrus cycle dependent manner. Physiology & behavior. 2014 Aug; 135:98–103.