Molecular Mimicry by an F-Box Effector of *Legionella pneumophila* Hijacks a Conserved Polyubiquitination Machinery within Macrophages and Protozoa

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Abstract

The ability of *Legionella pneumophila* to proliferate within various protozoa in the aquatic environment and in macrophages indicates a remarkable evolution and microbial exploitation of evolutionarily conserved eukaryotic processes. Ankyrin B (AnkB) of *L. pneumophila* is a non-canonical F-box-containing protein, and is the only known Dot/Icm-translocated effector of *L. pneumophila* essential for intra-vacuolar proliferation within both macrophages and protozoan hosts. We show that the F-box domain of AnkB and the L¹⁰P conserved residues are essential for intracellular bacterial proliferation and for rapid acquisition of polyubiquitinated proteins by the *Legionella*-containing vacuole (LCV) within macrophages, *Dictyostelium discoideum*, and *Acanthamoeba*. Interestingly, translocation of AnkB and recruitment of polyubiquitinated proteins in macrophages and *Acanthamoeba* is rapidly triggered by extracellular bacteria within 5 min of bacterial attachment. Ectopically expressed AnkB within mammalian cells is localized to the periphery of the cell where it co-localizes with host SKP1 and recruits polyubiquitinated proteins, which results in restoration of intracellular growth to the *ankB* mutant similar to the parental strain. While an ectopically expressed AnkB-L¹⁰P/AA variant is localized to the cell periphery, it does not recruit polyubiquitinated proteins and fails to trans-rescue the *ankB* mutant intracellular growth defect. Direct *in vivo* interaction of AnkB but not the AnkB-L¹⁰P/AA variant with the host SKP1 is demonstrated. Importantly, RNAi-mediated silencing of expression of SKP1 renders the cells non-permissive for intracellular proliferation of *L. pneumophila*. The role of AnkB in exploitation of the polyubiquitination machinery is essential for intrapulmonary bacterial proliferation in the mouse model of Legionnaires’ disease. Therefore, AnkB exhibits a novel molecular and functional mimicry of eukaryotic F-box proteins that exploits conserved polyubiquitination machinery for intracellular proliferation within evolutionarily distant hosts.


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Introduction

Intracellular pathogens have evolved with remarkable mechanisms to exploit host cell processes to evade degradation within the lysosomes, which is the first line of defense against microbial invasion. The intracellular bacterial pathogen *Legionella pneumophila* is ubiquitous in natural aquatic environments and man-made water systems, where it replicates within various protozoan hosts [1]. Once transmitted to humans by aerosols, *L. pneumophila* replicates within pulmonary macrophages causing pneumonia [1]. Remarkably, intracellular trafficking and intra-vacuolar proliferation of *L. pneumophila* within amoebae and human macrophages is very similar, at the cellular and molecular levels [1]. Within both evolutionarily distant host cells, *L. pneumophila* evades endocytic fusion and intercepts ER-to-Golgi vesicle traffic to remodel its phagosome into a rough endoplasmic reticulum (RER)-derived vacuole [2,3]. The Dot/Icm type IV secretion system [4,5] is required for *L. pneumophila* to modulate various mammalian and protozoan cellular processes through translocation of ~200 effectors into the host cell, but the role of most of these effectors in the intracellular infection is minimal or not known [2,3,6].

The similarity in the intracellular life cycle of *L. pneumophila* within protozoan and mammalian cells suggests that co-evolution of this bacterium with protozoa in the aquatic environment has allowed this bacterium to undergo patho-adaptation and evolution to exploit evolutionarily conserved eukaryotic processes that have enabled this bacterium to proliferate within human macrophages [1]. It is not surprising that bioinformatic genomic analyses of *L. pneumophila* have revealed numerous eukaryotic-like genes, such as ankyrin (ank)-encoding genes [7,8] which have been suggested to be acquired by cross-kingdom horizontal gene transfer. Among ~200 Dot/Icm-exported effectors, AnkB and SidJ are the only effectors essential for proliferation within the two evolutionarily-distant hosts, mammalian macrophages and protozoa [9–11]. In
Author Summary

*Legionella pneumophila* is abundantly found in the aquatic environment within various protozoa and can cause a severe pneumonia called Legionnaires’ disease when it invades human macrophages in the lung. The ability of *L. pneumophila* to invade and proliferate within macrophages and protozoa is dependent on the translocation of specific proteins into the invaded cell via a specialized secretory device, and these proteins modulate various host cell processes. Of these translocated proteins, AnkB is indispensable for intracellular growth of *L. pneumophila* within macrophages and protozoa. Here we show that AnkB is essential for establishing a favorable intracellular replicative niche by promoting the decoration of the *Legionella* containing vacuole (LCV) with polyubiquitinated proteins. The AnkB effector achieves this by mimicking the action of host cell F-box proteins, a highly conserved component of the SCF ubiquitin ligase complex that is found in both unicellular organisms and mammalian cells. Our study provides new insights into the ability of intracellular pathogens to hijack evolutionarily conserved host cell processes through molecular mimicry to establish a favorable replicative niche within various hosts and to cause disease in mammals.

addition to the two Ankyrin domains (ANK), AnkB harbors an F-box domain [9,10], which is a prevalent domain in eukaryotic proteins known to be involved in polyubiquitination within eukaryotic cells [12,13].

Ubiquitination of proteins is an ancient and highly conserved eukaryotic post-translational modification process that covalently links a 76-residue ubiquitin polypeptide to a specific protein to target it for proteasomal degradation or to modulate its function in a wide range of important cellular processes [13]. A major group of ubiquitin ligases is the SCF1 (RBX1-CUL1-SKPI) tri-molecular complex that binds the F-box domain [12,14], which is highly conserved throughout the eukaryotic kingdom, including yeast and the social amoeba *Dictyostelium discoideum* [15,16]. Substitution of the two conserved LP residues of the F-box domain in unicellular or multi-cellular eukaryotes abolishes polyubiquitination by the SCF1 complex [15,16]. Polyubiquitination by the SCF1 complex in the social amoeba *D. discoideum*, *Tetrahymena*, *Acanthamoeba*, and yeast appears to function in an identical manner to those of mammalian cells [15–17,18,19,20,21,22–24]. Although members of the F-box family of human proteins typically have a substrate-binding domain such as leucine-rich repeat (LRR) or WD40 [25,26], the ANK domain has never been described in eukaryotic F-box proteins [12,13], indicating a novel structure of the microbial F-box AnkB protein.

Many intracellular bacterial pathogens have been shown to exploit or intercept the host polyubiquitination machinery [27]. The *Legionella*-containing vacuole (LCV) is decorated with polyubiquitinated proteins, and the Dot/Icm transport system is essential for this process [28]. We show that AnkB is a non-canonical F-box protein that exhibits molecular and functional mimicry of evolutionarily conserved eukaryotic F-box proteins that enables *L. pneumophila* to hijack a conserved polyubiquitination machinery within mammalian and protozoan cells. Interestingly, AnkB is rapidly translocated into the host cell upon bacterial attachment to the host cell plasma membrane and polyubiquitinated proteins are rapidly recruited to the plasma membrane beneath the sites of bacterial attachment. This molecular and functional mimicry by the F-box domain of AnkB in exploitation of the polyubiquitination machinery within evolutionarily distant host cells is also essential for intrapulmonary bacterial proliferation in the animal model of Legionnaires’ disease. The host factor SKP1 interacts with AnkB, and knockdown expression of SKP1 renders the cells non-permissive for intracellular proliferation.

**Results**

The F-box-containing AnkB is essential for acquisition of polyubiquitinated proteins by the LCV within macrophages

We determined whether the F-box protein AnkB (Fig. S1) mimicked the action of host F-box proteins and recruited polyubiquitinated proteins to the LCV [28]. The wild type strain *L. pneumophila* AA100/130b, the *ankB* mutant, and a dotA type IV translocation-defective mutant control were used to infect the U937 human monocytic cell line for 5–120 min and 14 h. At different time intervals post-infection, the cells were labeled using an anti-polyubiquitin antibody and examined by confocal microscopy. The data showed that as early as 5 min post-infection, >70% of the LCVs harboring the WT strain were decorated with polyubiquitinated proteins, while the *ankB* mutant failed to recruit polyubiquitinated proteins, similar to the *dotA* translocation-defective mutant (Fig. 1). Defect of the *ankB* mutant in decorating the LCV with polyubiquitinated proteins was restored by complementation with the WT *ankB* gene on a plasmid (data not shown).

These observations were further confirmed in semi-purified LCVs in post-nuclear supernatant (PNS) of infected U937 cells. Control LCVs in PNS labeled with anti-*Legionella* antibody prior to permeabilization showed that the majority of the bacteria (95%) were within intact LCVs (data not shown). The data showed that 80% of the purified LCVs harboring the WT strain were decorated with polyubiquitin, while less than 10% of LCVs containing the *ankB* mutant were decorated with polyubiquitin, similar to the *dotA* mutant (Fig. 2). To validate our observations in primary cells, we utilized infection of human monocyte- derived macrophages (hMDMs). Essentially, similar results of polyubiquitination of the LCV were obtained in hMDMs at all time points examined up to 14 h (Fig. 2 and data not shown). Defect of the *ankB* mutant in decorating the LCV with polyubiquitinated proteins was restored by complementation with the WT *ankB* gene on a plasmid (Fig. 2). We conclude that the *L. pneumophila* Dot/Icm-translocated F-box-containing AnkB [9,10] is essential for acquisition of polyubiquitinated proteins by the LCV within human macrophages.

AnkB-mediated acquisition of polyubiquitinated proteins by the LCV in *A. polyphaga* and *D. discoideum*

Whether the LCV hijacks the polyubiquitination machinery of protozoan hosts is not known. Therefore, we determined whether the LCV within *Acanthamoeba polyphaga* and the social amoeba *Dictyostelium discoideum* was decorated with polyubiquitinated proteins and whether the Dot/Icm system type IV secretion system and the AnkB effector were involved in this process. The data showed that similar to human macrophages, the LCVs harboring the wild type strain with *A. polyphaga* (Fig. 3) and *D. discoideum* (Fig. S2) acquired polyubiquitinated proteins as early as 5 min post-infection, while the *ankB* mutant failed to recruit polyubiquitinated proteins, similar to the *dotA* mutant.

These observations were further confirmed in semi-purified LCVs in post-nuclear supernatant (PNS) of infected *A. polyphaga* cells at 2 h post-infection. Control LCVs in PNS labeled with anti-*Legionella* antibody prior to permeabilization showed that the
majority of the bacteria (95%) were within intact LCVs (data not shown). The data showed that the wild type strain purified LCVs were heavily decorated with polyubiquitinated proteins, while LCVs of the dotA and ankB mutants were both severely defective in acquisition of polyubiquitinated proteins (Fig. 3). We conclude that the AnkB effector is essential for acquisition of polyubiquitinated proteins by the LCV within protozoan cells, similar to human macrophages.

Translocation of AnkB and recruitment of polyubiquitinated proteins upon attachment of L. pneumophila to macrophages and A. polyphaga

Due to the rapid recruitment of polyubiquitinated proteins to the LCV within macrophages and amoebae, we hypothesized that the bacteria might initiate the process upon attachment to the host cell membrane. To test this hypothesis, macrophages were pre-treated with cytochalasin D to block bacterial entry, and the extracellular bacteria were labeled with an antibody prior to permeabilization to differentiate them from intracellular bacteria. Interestingly, within 5 minutes of bacterial attachment, >50% of the WT attached bacteria recruited polyubiquitinated proteins to their attachment sites at the macrophage plasma membrane but the ankB mutant was defective, similar to the dotA mutant (Fig. 1). Similar results were also obtained in A. polyphaga (Fig. S3). We conclude that upon attachment of L. pneumophila to human macrophages and A. polyphaga, AnkB rapidly recruits polyubiquitinated proteins to the host plasma membrane beneath the sites of bacterial attachment.

Our data above led us to hypothesize that AnkB was rapidly translocated into the host cell by attached extracellular bacteria.

Figure 1. AnkB is essential for recruitment of polyubiquitinated proteins by the LCV and bacterial attachment to the macrophage plasma membrane is sufficient to trigger the recruitment process. U937 cells were infected with the wild type (WT) L. pneumophila (Lpn) and the isogenic dotA or ankB mutants. A) Representative confocal microscopy images of infected U937 cells at 2 h post-infection for co-localization of the LCVs with polyubiquitinated (PolyUb) proteins. The arrow indicates heavy co-localization of polyubiquitin with the WT strain. B) Quantitation of the co-localization during infection at different time points. In panels C and D, the U937 cells were treated with 1 μM cytochalasin D for 30 min prior to infection, to inhibit bacterial entry. The cells were infected with GFP-expressing bacteria for 5, 15, 30 and 60 min. Prior to permeabilization, extracellular L. pneumophila were labeled with an anti-Lpn antibody (blue) resulting in a dual labeling of the extracellular bacteria (green and blue). Host polyubiquitin was labeled after permeabilization (red). C) Representative confocal microscopy images that show an extracellular WT L. pneumophila bacterium co-localized with high levels of polyubiquitinated proteins (arrow) at the site of bacterial attachment after 5 min of infection. D) Quantitation of the %co-localization of attached bacteria with polyubiquitinated proteins was determined by analysis of 100 attached extracellular bacteria. The data represent analyses of 100 infected cells and are representative of three independent experiments, and the error bars represent standard deviation.

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To test this hypothesis, we examined whether AnkB was translocated into macrophages prior to bacterial internalization. We utilized the calmodulin-dependent adenylate cyclase-AnkB (CyaA-AnkB) reporter system [10], which becomes active only after translocation into the host cell, to quantify translocation of the AnkB protein to the host cell. For controls of Dot/Icm-translocated effectors that are only translocated by intracellular and not extracellular bacteria, we used RalF-CyaA and AnkD-CyaA fusions [7]. The results showed that in contrast to the RalF effector and AnkD controls, AnkB was efficiently translocated by attached extracellular bacteria (Fig. 4A). These results are consistent with our data above that showed extracellular bacteria recruited polyubiquitinated proteins to the cytosolic face of the plasma membrane beneath the sites of bacterial attachment.

Role of the F-box domain of AnkB in acquisition of polyubiquitinated proteins within human macrophages and *A. polyphaga*

The F-box domain is highly conserved through evolution and the two characterized *D. discoideum* F-box proteins harbor a highly conserved F-box [19,22]. In addition, the LP conserved residues of
the F-box domain, which are essential for binding of eukaryotic F-box proteins to the cellular factor SKP1, are also conserved from *D. discoideum* to mammals [15,16]. To analyze the role of the F-box domain of AnkB in decorating the LCV with polyubiquitinated proteins, we engineered a mutant allele of *ankB* with an in-frame deletion of the F-box domain (*ankBΔF-box*) (Fig. S1). In addition, the conserved 9L10P residues of the F-box domain were substituted by alanine residues (*ankB-9L10P/AA*) (Fig. S1). Macrophages and *A. polyphaga* were infected as described above by the wild type strain, the *ankB* mutant, or the *ankB* mutant harboring plasmids containing the WT *ankB* or the *ankBΔF-box* or the *ankBΔF-box* allele. The data showed that within U937 human macrophages, hMDMs, and *A. polyphaga* the LCVs harboring the *ankB* mutant complemented with the *ankBΔF-box* or the *ankBΔF-box* allele failed to become decorated with polyubiquitinated proteins, similar to the *ankB* null mutant or the *dotA* mutant control (Fig. 2 and 3). The LCVs harboring the *ankB* mutant complemented with the WT *ankB* allele were decorated with polyubiquitinated proteins, similar to the WT strain (Fig. 2 and 3). These results were further confirmed in semi-purified LCVs harboring the

**Figure 3. AnkB is essential for acquisition of polyubiquitinated proteins by the LCV in *A. polyphaga***. Representative images of co-localization of the LCVs within *A. polyphaga* cells (A) or in semi-purified LCVs (B) with polyubiquitinated proteins at 2 h post-infection. The cells or LCVs were labeled with anti-Lpn antibody (green) and anti-polyubiquitin (red) and then analyzed by confocal microscopy. The arrow indicates heavy co-localization of polyubiquitin with the WT strain. Quantification of co-localization of the LCVs with polyubiquitinated proteins at 2 h is shown in the merged images in the third column in A and B. C) Quantitation of co-localization of the LCVs with polyubiquitinated proteins at various time points post-infection. The data represent analyses of 100 infected cells or LCVs and are representative of three independent experiments.
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Role of the F-box domain of AnkB in intracellular proliferation within human macrophages

To determine the role of the F-box domain in intracellular proliferation in macrophages, the ankB mutant was complemented by the ankBAF-box or the ankB-3L10P/AA construct on a plasmid. The data showed that complementation of the ankB mutant with the respective wild type gene on a plasmid fully restored intracellular growth within hMDMs to levels similar to the wild type strain (Fig. S4). In contrast, complementation of the ankB mutant with the ankBAF-box, or the ankB-3L10P/AA allele failed to restore the intracellular growth defect of the ankB mutant in hMDMs. Therefore, the F-box domain of AnkB and its two LP conserved residues are essential for decoration of the LCV with polyubiquinated proteins and for intracellular proliferation within macrophages.

Translocation of AnkB into the host cell is essential for its crucial role in intracellular proliferation [9,10]. Therefore, it was essential to confirm that the failure of the ankBAF-box allele to restore to the ankB null mutant intracellular proliferation and recruitment of polyubiquitinated proteins was not due to a defect in translocation. We utilized the Calmodulin-dependent adenylate cyclase assay to determine translocation of the truncated AnkBAFbox variant. Immunoblot analyses of bacterial cell lysates showed equivalent expression of the Cya-AnkB fusions among all the tested strains (Fig. 4). The data showed that the AnkBAFbox was efficiently translocated into the host cell, albeit at slightly reduced levels compared to the WT AnkB (Fig. 4). Thus, the defect in polyubiquitination and intracellular proliferation of the AnkBAFbox variant was not likely to be due to reduced expression or stability of the AnkBAFbox variant protein.

Ectopic expression of AnkB and its distribution in mammalian cells

Although AnkB is clearly translocated into the host cell by the Dot/Icm system, we have been unable to detect AnkB during infections using AnkB-specific antibodies (data not shown), indicating that the protein is either extremely unstable or translocated by the bacterium at undetectable concentrations. Several other Dot/Icm effectors such as LpxA and LpxX [29,30] have also been shown to be undetectable, which may ensure limited interference with host cell processes without compromising viability of the host cell. To overcome this limitation and to elucidate the distribution of AnkB in the host cell, we ectopically expressed a 3X-flag AnkB fusion construct in the HEK-293 human cell line. Ectopic expression was not toxic to the cells, which was further confirmed by our ability to generate stable HEK293 cell transfectants (Fig. 5A, data not shown). Transient transfection of HEK-293 cells with a plasmid encoding 3X-Flag AnkB revealed a striking localization of this protein to the periphery of the cell. In contrast, the control protein 3X-Flag-bacterial alkaline phosphatase (BAP) was distributed throughout the cytosol (Fig. 5A). Localization of the 3X-Flag AnkB to the periphery of the cell, beneath or at the inner layer of the plasma membrane, was further supported by labeling the plasma membrane with wheat germ agglutinin (WGA) prior to permeabilization and fixation, which revealed close association of AnkB to the periphery of the cell, or the inner layer of the plasma membrane (Fig. 5A). In addition, AnkB was also observed as a few punctate spots within the cytosol (Fig. 5). Substitution of the conserved 3L10P residues of the F-box domain to AA did not of conserved cellular F-box proteins to hijack the ubiquitination machinery of evolutionarily distant hosts.

Figure 4. Translocation of AnkB by attached extracellular bacteria and efficient translocation of the AnkBAF-box reporter by intracellular bacteria. A) Translocation of AnkB into U937 cells by attached WT or dotA mutant extracellular bacteria was determined at 30 min post-infection. Strains harbored either empty vector (pCya) or adenylate cyclase RafF, AnkB or AnkD fusions (pCya-RafF, pCya-AnkB and pCya-AnkD). B) Translocation of the reporter constructs by intracellular bacteria at 2 h post-infection. C) Equivalent expression of the AnkB-Cya fusion alleles in L. pneumophila was determined by immunoblots of cell lysates and total bacterial cultures probed with anti-CAT antibodies.
impair localization of AnkB to the cell periphery (Fig. 5C). Similar results were obtained using AnkB-GFP fusions (data not shown).

To determine whether AnkB recruits the host SKP1 protein to its peripheral localization sites in the cells, we co-transfected HEK293 cells with plasmids encoding 3X-flag AnkB and SKP1-GST (Fig. 5B). There was a clear and distinct co-localization of SKP1 at the cell periphery where AnkB was concentrated and preferentially localized (Fig. 5B).

Since AnkB is essential for recruitment of polyubiquitinated substrates to the LCV, we determined whether ectopic expression of 3X-Flag AnkB and its recruitment of host SKP1 to the periphery of the cell also altered distribution of polyubiquitinated proteins in the host cell. Strikingly, strong co-localization of 3X-flag AnkB and polyubiquitinated proteins was observed at the periphery of transfected HEK-293 cells (Fig. 5C). In contrast, 3X-flag AnkB-1LP/AA failed to co-localize strongly with polyubiquitinated proteins, similar to the BAP control (Fig. 5E), despite its similar distribution in the cell to that of the native AnkB protein. These data indicate that the two LP conserved residues of the F-box domain do not affect localization of the AnkB protein in HEK-293 cells, but are crucial for the ability of AnkB to hijack the host ubiquitination machinery. Thus, ectopically expressed AnkB recruits host SKP1 protein and acts as a platform for acquisition of polyubiquitinated proteins at the periphery of the cell.

**Figure 5.** Ectopically expressed 3X-Flag AnkB is distributed at the periphery of HEK-293 cells, co-localizes with the three SCF components and poly-ubiquitinated proteins, and restores intracellular growth to the ankB mutant. A) Representative confocal images of HEK-293 cells transfected with plasmids encoding 3X-Flag BAP or 3X-flag AnkB. At 24 h following transfection, cells were labeled using a mouse anti-Flag (green) and WGA conjugated to Alexa Fluor-647 (red). Nuclei were stained with DAPI (purple). The white arrow indicates peripheral co-localization of 3X-Flag AnkB. B) Representative confocal images of HEK-293 cells that were co-transfected with plasmids encoding 3X-flag AnkB or BAP and SKP1-GST. Cells were stained with anti-Flag (green) and anti-GST (red) and nuclei were visualized with DAPI (blue). Arrows indicate peripheral co-localization of AnkB and SKP1. C) 3X-Flag AnkB co-localizes with polyubiquitinated proteins at the cell periphery. At 24 h following transfection cells were labeled using a rabbit anti-flag antibody (green) and a mouse anti-polyubiquitin antibody (red). Nuclei were labeled with DAPI (blue). The white arrow indicates heavy co-localization of 3X-Flag AnkB with polyubiquitin. D) Representative confocal images at 12 h post-infection of HEK-293 cells transfected for 24 h with a plasmid encoding 3X-Flag AnkB. The cells were labeled with an anti-Lpn antibody (green) and an anti-Flag antibody (red). The data are representative of three independent experiments. doi:10.1371/journal.ppat.1000704.g005
Ectopic expression of AnkB in mammalian cells rescues growth of the ankB mutant

Since the ankB mutant is severely defective in intracellular proliferation, we determined whether ectopically expressed 3X-Flag AnkB and its recruitment of polyubiquitinated proteins to the periphery of the cell would be sufficient to restore intracellular growth to the ankB mutant. The 3X-flag AnkB-transfected HEK-293 cells were infected for 1 h and the number of intracellular bacteria was determined at 12 h using single cell analysis by microscopy. At 2 h post-infection most cells harbored a single organism (data not shown). The data revealed that indeed ectopically expressed 3X-Flag AnkB restored intracellular growth to the ankB mutant at 12 h post-infection, similar to wild type bacteria in transfected or non-transfected cells (Fig. S5). Importantly, the ankB mutant failed to replicate in un-transfected cells or cells transfected with the 3X-Flag BAP control (Fig. S5). Despite the trans-rescue of the ankB mutant for its defect in intracellular proliferation in cells expressing AnkB at high levels, the AnkB protein was not detected on the LCV. These data indicate that AnkB may function at a distance from the LCV to hijack the polyubiquitination machinery to enable intracellular proliferation. However, it can not be excluded that possible rapid turnover of AnkB on the LCV may preclude its detection.

Host SKP1 interacts with AnkB in vivo and is essential for intracellular proliferation of L. pneumophila

To determine whether AnkB mimicked a host F-box protein and interacted directly with the host factor SKP1, we performed in vivo co-immunoprecipitation. We co-transfected HEK-293 cells with plasmids encoding 3X-Flag tagged AnkB, AnkB\textsuperscript{L}\textsuperscript{108/PA}, BAP or SKP2 and SKP1-GST. The transfected cells were lysed and reciprocal co-immunoprecipitation with anti-flag agarose and anti-GST agarose was performed (Fig. 6A, data not shown). As expected, our data showed that SKP1 and SKP2 co-precipitated whilst SKP1 and BAP did not co-precipitate. Importantly, our data showed that AnkB and SKP1 specifically co-precipitated (Fig. 6A). To test the specificity of this interaction and the direct role of the F-box domain in binding to SKP1, we analyzed if 3X-Flag AnkB\textsuperscript{L}\textsuperscript{108/PA} would co-precipitate with SKP1-GST. The data were very clear that the 3X-Flag AnkB\textsuperscript{L}\textsuperscript{108/PA} did not interact with SKP1-GST (Fig. 6A). This was consistent with data above using scanning confocal microscopy of cells co-transfected with plasmids encoding 3X-Flag AnkB and SKP-GST, where distinct co-localization of SKP1 with AnkB at the cell periphery was detected (Fig. 5B). No specific co-localization with SKP1 and BAP control was observed around the cell periphery (Fig. 5B). Taken together, these results show that AnkB specifically interacts with the host SKP1 in vivo, and this interaction absolutely requires the functional F-box domain of AnkB.

To determine whether the host SKP1 function was required for intracellular proliferation of L. pneumophila, 3 different SKP1-specific RNAi were transfected separately into HEK293 cells. There was knock-down in the expression of SKP1 by RNAi-1, -2 or a combination of both but viability of the cells was not affected by any of the RNAi used (Fig. 6D and data not shown). However, RNAi-3 or control RNAi had no detectable effect on SKP1 expression by 48 h (Fig. 6D). Two days after transfection, cells were infected with the WT strain L. pneumophila or the ankB mutant. Single cell analysis by microscopy was performed at 12 hours post-infection to determine the effect on bacterial proliferation at the single cell level (Fig. 6B, C). Silencing of SKP1 by RNAi-1, RNAi-2, or a combination of both completely blocked intracellular replication of L. pneumophila, which was reminiscent of the defect of the ankB null mutant in non-transfected cells (Fig. 6B, C and data not shown). In contrast, in the non-transfected cells, or cells transfected with control RNAi or RNAi-3, there was no effect on intracellular replication (Fig. 6B, C and data not shown). We conclude that host SKP1 is essential for intracellular replication of L. pneumophila. This supports the evidence that the L. pneumophila AnkB F-box protein mimics the function of eukaryotic F-box proteins to hijack the host polyubiquitination machinery, which is essential for intracellular proliferation of L. pneumophila within mammalian and protozoan cells.

AnkB is essential for intrapulmonary proliferation in the mouse model of Legionnaires’ disease

The role of Dot/Icm-translocated effectors of L. pneumophila in intrapulmonary proliferation in animal models has never been examined for any effector. When groups of 10 A/J mice were infected with 10\textsuperscript{5} CFUs of the WT strain or the ankB mutant to determine lethality by high dose infection, 100% of the WT strain-infected mice succumbed to infection within 24–48 h after inoculation, while 100% survival for the ankB mutant-infected mice was observed up to 7 days. When mice were infected by 10\textsuperscript{6} CFUs, 70% mortality was exhibited by 24–72 h after infection with the WT strain, but 100% survival was observed for the ankB mutant-infected mice.

To determine whether AnkB was required for intrapulmonary proliferation of L. pneumophila, we infected A/J mice with 10\textsuperscript{8} CFUs of L. pneumophila. Proliferation of L. pneumophila in the lungs of infected mice was assessed by enumeration of the CFUs in lung homogenates after 24 h, 48 h, 72 h and 7 days post-infection. While there was robust intrapulmonary proliferation by the WT strain by 24–48 h, the data were very clear that there was no detectable intrapulmonary proliferation for the ankB mutant (student \textit{t}-test, \textit{p}<0.001) (Fig. 7). By 48 h post-infection, at least 2000-fold less bacteria were recovered from lungs infected with the ankB mutant compared to the WT strain (Fig. 7). Due to our repeated observations of the rapid loss of plasmids by the wild type strain AA100 or its derivative mutants in mouse lungs, complementation in mouse lungs was not achieved. We conclude that the function of AnkB in recruitment of polyubiquitinated proteins to the LCV is essential \textit{in vivo} for intrapulmonary proliferation of L. pneumophila in the mouse model of Legionnaires’ disease.

Discussion

L. pneumophila is an environmental bacterium that is believed to have evolved through interaction with various protozoa to acquire mechanisms to allow them to proliferate within mammalian pulmonary macrophages, causing pneumonia. This microbial
adaptation to the intracellular life within protozoa and mammalian cells indicates microbial manipulation of eukaryotic cellular processes that are conserved through evolution. Among Dot/Icm-exported effectors of *L. pneumophila*, AnkB is the only effector that plays a major role in intracellular proliferation within both protozoan and mammalian cells [10]. This indicates that the host target of AnkB is an evolutionarily conserved process in mammalian and protozoan cells. Comparison of the amino acid sequences for the two characterized *D. discoideum* F-box proteins FbxA and MEKKa to mammalian F-box proteins reveals identical sequences in the conserved F-box motif, including the conserved LP residues (data not shown). The F-box domain of the *L. pneumophila* AnkB protein mimics host F-box proteins by interacting with host SKP1 in vivo and SKP1 is essential for intracellular bacterial proliferation. A) In vivo co-immunoprecipitation of AnkB and SKP1 in co-transfected cells. HEK-293 cells were co-transfected with plasmids encoding 3X-Flag AnkB, AnkB<sup>Δ10P/AA</sup>, BAP or Skp2 and Skp1-GST. Total protein input in total cell lysate was equivalent in all the lanes (not shown). Cell lysates were purified using anti-flag resin and the resulting supernatants were analyzed by immunoblotting with antibodies against Flag and GST. B) Representative confocal images of HEK-293 cells that were untreated or transfected with RNAi followed by infection and labeling of the bacteria at 16 h to score replicative vacuoles. C) Quantitative analyses of the formation of replicative vacuoles were based on examination of 100 infected cells. The different color bars represent the number of bacteria/vacuole. The data represent analyses of the mean and standard deviations of 100 infected cells, and are representative of two independent experiments. D) Knockdown expression of SKP1 protein by siRNA silencing in HEK-293 cells was confirmed by Western blot of HEK-293 cell lysates probed with an anti-SKP1 antibody. The blots were re-probed with anti-actin antibodies, which showed equivalent quantities of proteins were loaded onto the gel. The results are representative of three independent experiments.

Figure 6. The F-box AnkB protein mimics host F-box proteins by interacting with host SKP1 in vivo and SKP1 is essential for intracellular bacterial proliferation. doi:10.1371/journal.ppat.1000704.g006
L. pneumophila AnkB protein also harbors an identical sequence to the F-box motif and has the conserved LP residues. We show that the F-box domain of AnkB exhibits molecular and functional mimicry of eukaryotic F-box proteins in exploitation of a conserved polyubiquitination machinery [31] to allow microbial parasitism of evolutionarily distant hosts. Our data show that the F-box of AnkB and its two highly conserved L18P19 residues are essential for hijacking the polyubiquitination machinery for intracellular proliferation. This indicates that AnkB exhibits molecular and functional mimicry of an evolutionarily conserved eukaryotic F-box protein and that AnkB has conserved eukaryotic targets in mammalian and protozoan cells, which allows L. pneumophila to hijack the ubiquitination machinery of the evolutionarily distant host cells. The L. pneumophila published genomes indicate that there are at least two other F-box proteins encoding genes but the roles of these in infection are not known [32–35]. In addition, a U-box-containing E3 ligase of L. pneumophila has been reported but does not play a role in polyubiquitination of the LCV [30]. However, the profound effect of AnkB on polyubiquitination of the LCV clearly indicates a major role for AnkB in this process. We have recently identified tremendous heterogeneity in acquisition of polyubiquitinated proteins by the LCV harboring various clinical and environmental isolates of L. pneumophila (Price et al, unpublished data), which indicates potential redundancy for the function of AnkB in various isolates, such as the Lp02 strain [10,32]. Functional redundancy among Dot/Icm effectors is very common and is thought to allow parasitism of various unicellular hosts in the aquatic environment [2].

Interestingly, the F-box family of human proteins typically have a leucine-rich repeat (LRR) or WD40 protein-protein interaction domains, which bind substrates and link them to the SCF1 ubiquitin ligase tri-molecular complex that is bound to the F-box domain [25,26]. However, the ANK domain has never been found in eukaryotic F-box proteins [12,13]. This would render the bacterial F-box-containing AnkB to have a non-canonical or novel structure among the large family of F-box proteins [12,13]. Interestingly, co-expression of AnkB and SKP1 in cells results in co-localization of the two proteins at the periphery of the cell where polyubiquitinated proteins are recruited, and this is sufficient to trans-rescue the defect of intracellular proliferation of the ankB mutant. Importantly, the host SKP1 interacts directly and specifically with AnkB in vivo, and the specificity of this interaction has been demonstrated by failure of the AnkB-L18P/AA variant to interact with SKP1. Importantly, this interaction is essential for bacterial proliferation, since knockdown expression of SKP1 renders the cells non-permissive for intracellular proliferation of the wild type strain. We conclude that the AnkB-SKP1 interaction and its role in exploitation of the host polyubiquitination machinery is essential for intracellular bacterial proliferation.

Recruitment of polyubiquitinated proteins to the phagosome of intracellular pathogens has been described only for L. pneumophila. The quantity, the kind of linkages involved in polyubiquitination, or the identity of the proteins that are polyubiquitinated on the LCV remain to be elucidated. In addition, other components of the SCF1 complex that interact with AnkB need to be identified. Future proteomic studies on these aspects of the LCV to identify these processes would be intriguing. Taken together, we propose a model in which the F-box domain of AnkB interacts with the host SKP1, while protein substrates to be polyubiquitinated are recruited by the two ANK domains of AnkB. The identity of these substrates is currently under investigation.

In summary, we have characterized an essential role for a structurally novel non-canonical F-box translocated effector in a mammalian intracellular bacterial pathogen. This L. pneumophila non-canonical F-box effector exhibits molecular and functional mimicry of an evolutionarily-conserved F-box protein in recruitment of the host polyubiquitination machinery to the bacterial vacuole, which is essential for intra-vacuolar proliferation within mammalian and protozoan cells and is essential for manifestation of pulmonary disease in the animal model. The host SKP1 factor interacts directly and specifically with AnkB, and knockdown of its expression renders the cells non-permissive for intracellular proliferation. Our data provide the first description of a bacterial non-canonical F-box protein and its molecular and functional mimicry of evolutionarily conserved F-box proteins to exploit an evolutionarily conserved polyubiquitination machinery to enable intra-vacuolar proliferation within evolutionarily distant hosts and to cause pulmonary disease in mammals.

Materials and Methods

Ethics statement

All animal work in this manuscript has been performed after approval by the institutional IACUC committee who follows the US federal guidelines for the use and humane handling of animals in research.

Bacterial strains, cell cultures and infections

L. pneumophila serogroup I parental strain AA100/130b and the mutants dotA, ankB, and complemented ankB mutants were grown as described previously [10]. Escherichia coli strain DH5α was used for cloning purposes. Isolation and preparation of human monocyte-derived macrophages (hMDMs) and maintenance of the macrophage-like U937 cells were performed as previously described [9]. Cultures of A. polyphaga and D. discoideum were performed as described previously [9,10].

For intracellular proliferation studies, infections were performed as we described previously [9]. Briefly, macrophages were infected at a multiplicity of infection (MOI) of 10 for 1 h followed by treatment with 50 μg/ml gentamicin for 1 h to kill extracellular bacteria. At each time point, the macrophages were lysed and
dilutions were plated on agar plates. For single cell analysis studies, infections were performed as we described above and at 12 h cells were fixed and processed for confocal microscopy. Phagosomes were isolated from post nuclear supernatants (PNS) of infected cells as described previously [29]. Samples were then fixed and probed as described below. The cya constructs were generated by PCR using specific primers (Table 1). Measurement of cAMP in cell lysates for adenylyl cyclase fusion assays was performed using the Direct Cyclic AMP Enzyme Immunoassay kit (Assay Designs), as we described previously [10].

Transfection of HEK-293 cells
The ankB gene was cloned into the mammalian expression vector, p3XFlag-CMV-10 (Sigma). To generate the ankB−/−/AKI allele, the wild type p3XFlag-CMV-10 AnkB plasmid was used as a template for PCR based site directed mutagenesis. HEK-293 AA allele, the wild type p3XFlag-CMV-10 AnkB plasmid was used.

Silencing of host SKP1 in HEK 293 cells
HEK 293 cells were maintained in DMEM (Invitrogen, Carlsbad, CA) supplemented with 10% heat inactivated fetal bovine serum. For each transfection reaction, oligonucleotide RNAi-1 (GCAGCAGAAUGAGAAAGGUAUA) and (UAAUUUGUCAGGAUAAGUAUA) (Roche).

Infection of A/J mice with L. pneumophila
Female pathogen-free, 6–8 week old A/J mice were used for infection by intratracheal inoculation with 50 μl containing the bacterial dose as we described previously [36]. Mice were humanely euthanized at various times, the lungs were removed, homogenized and dilutions were cultured on BCYE agar for 72 h as described previously [36].

Supporting Information
Figure S1 The F-box domain of AnkB shares similar alpha helix topology to mammalian F-box proteins. A: Amino acid alignment of the F-box domains of AnkB and the mammalian F-box proteins B-TrCP, CDC4 and SKP2. Alignments were performed using ClustalW software. Red residues represent conserved identical residues, while blue residues indicate conserved residues with similar biochemical properties. Boxed areas represent the known alpha helical regions in the mammalian F-box proteins [37]. Alpha helices for the AnkB F-box domain were predicted using PROF software. B: Cartoon representation of the predicted F-box domain deletion mutant harboring the WT 

Table 1. Primers used in this study.

<table>
<thead>
<tr>
<th>Primer Name</th>
<th>Primer Sequence (5’-3’)</th>
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<tbody>
<tr>
<td>AnkB A1 F*</td>
<td>ACCAAATGCTACCTCCTAATAA</td>
</tr>
<tr>
<td>AnkB A1 R*</td>
<td>TCAGCTTTATATGCCTGTC</td>
</tr>
<tr>
<td>AnkB A2 F*</td>
<td>AAAGATACATGATGGATT</td>
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</tr>
<tr>
<td>AnkB fbox F*</td>
<td>CAGAAGCTATAAAGCGA</td>
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<tr>
<td>AnkB fbox R*</td>
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</tr>
<tr>
<td>AnkB cya F</td>
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<tr>
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</tr>
<tr>
<td>AnkB cya R</td>
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<tr>
<td>AnkB fbox cya A1A2 R*</td>
<td>CTGGAGCTAAAGCAGGGCAGTGT</td>
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*Primers modified with 5’-phosphorylation. doi:10.1371/journal.ppat.1000704.s001 (0.17 MB PDF)
ankB alleles. Representative images of co-localization of the LCVs within infected *D. discoideum* (A) or in semi-purified LCVs (B) with polyubiquitinated proteins at 2h post-infection. The cells or LCVs were labeled with anti-Lpn antibody (green) and anti-polyubiquitin (red) and then analyzed by confocal microscopy. The arrow indicates heavy co-localization of polyubiquitin with the WT strain. Quantification of % co-localization of the LCVs with polyubiquitinated proteins at 2h is shown. The data represent analyses of 100 infected cells or LCVs and are representative of three independent experiments.

Figure S4 The F-box domain of AnkB is essential for intracellular proliferation of *L. pneumophila* within hMDMs. Infection of hMDMs with WT and the isogenic mutants *dotA* and *ankB* and the *ankB* mutant harboring the *ankB*ΔF-box allele was carried out in triplicate with an MOI of 10 for 1 h followed by 1 h gentamicin treatment to kill extracellular bacteria. The infected monolayers were lysed at different time points and plated onto agar plates for colony enumeration. The experiment was done 3 times and the data represent a sample of one independent experiment. Error bars represent standard deviation.

Found at: doi:10.1371/journal.ppat.1000704.s004 (0.11 MB PDF)

Figure S5 Rescue of the intracellular growth defect of the *ankB* mutant in HEK293 cells expressing AnkB. After 12 h post-infection of HEK-293 cells, 100 infected cells were analyzed by laser scanning confocal microscopy for formation of replicative phagosomes. Single cell analysis of replicative phagosomes in untransfected (A) or 3X-Flag AnkB transfected (B) HEK-293 cells at 12h post-infection. Quantitative analyses in A and B were based on examination of 100 infected cells and the error bars represent standard deviation. The data are representative of three independent experiments.

Found at: doi:10.1371/journal.ppat.1000704.s005 (0.11 MB PDF)

Author Contributions
Conceived and designed the experiments: CTP SAK TAQ MS AK YAK. Performed the experiments: CTP SAK TAQ MS FH AK YAK. Analyzed the data: CTP SAK TAQ MS FH AK YAK. Contributed reagents/materials/ analysis tools: FH. Wrote the paper: CTP SAK TAQ AK YAK.

References