

Table S1: Bacterial strains and plasmids

Strains	Description	Reference
<i>P. aeruginosa</i>		
PA01	Wild-type <i>P. aeruginosa</i>	[1]
$\Delta yfiR$	PA01 with <i>yfiR</i> deleted	this study
$\Delta yfiNR$	PA01 with <i>yfiNR</i> deleted	this study
$\Delta yfiBNR$	PA01 with <i>yfiBNR</i> deleted	this study
<i>yfiBNR</i> - complementation strains	$\Delta yfiBNR$ with <i>yfiBNR</i> complementation operons inserted at the <i>Att::Tn7</i> locus. Ten strains in total	this study
PA01 <i>yfiR</i> -M2	$\Delta yfiR$ with <i>yfiR</i> -M2 inserted at the <i>Att::Tn7</i> locus	this study
$\Delta yfiN$ <i>yfiR</i> -M2	$\Delta yfiNR$ with <i>yfiR</i> -M2 inserted at the <i>Att::Tn7</i> locus	this study
$\Delta yfiBN$ <i>yfiR</i> -M2	$\Delta yfiBNR$ with <i>yfiR</i> -M2 inserted at the <i>Att::Tn7</i> locus	this study
PA01-Gm ^R	Gm ^R , PA01 with the gentamycin resistance cassette inserted at the <i>Att::Tn7</i> locus	this study
PA01 <i>cupA::Tn</i>	Tet ^R , PA01 with the <i>lacZ</i> -Tn5 transposon inserted into <i>cupA4</i>	[2]
PA01 <i>cupB::Tn</i>	Tet ^R , PA01 with the <i>phoA</i> -Tn5 transposon inserted into <i>cupB4</i>	[2]
PA01 <i>cupC::Tn</i>	Tet ^R , PA01 with the <i>lacZ</i> -Tn5 transposon inserted into <i>cupC2</i>	[2]
PA01 <i>pelG::Tn</i>	Tet ^R , PA01 with the <i>phoA</i> -Tn5 transposon inserted into <i>pelG</i>	[2]
PA01 $\Delta pslAB$	Gm ^R , PA01 containing the WFP60 $\Delta pslAB$ cassette	[3]
PA01 <i>pelG::Tn</i> $\Delta pslAB$	Tet ^R , Gm ^R , PA01 containing the WFP60 $\Delta pslAB$ cassette, with the <i>phoA</i> -Tn5 transposon inserted into <i>pelG</i>	this study
$\Delta yfiR$ <i>cupA::Tn</i>	Tet ^R , PA01 with the <i>lacZ</i> -Tn5 transposon inserted into <i>cupA4</i>	this study
$\Delta yfiR$ <i>cupB::Tn</i>	Tet ^R , PA01 with the <i>phoA</i> -Tn5 transposon inserted into <i>cupB4</i>	this study
$\Delta yfiR$ <i>cupC::Tn</i>	Tet ^R , PA01 with the <i>lacZ</i> -Tn5 transposon inserted into <i>cupC2</i>	this study
$\Delta yfiR$ <i>pelG::Tn</i>	Tet ^R , PA01 with the <i>phoA</i> -Tn5 transposon inserted into <i>pelG</i>	this study
$\Delta yfiR$ $\Delta pslAB$	Gm ^R , PA01 containing the WFP60 $\Delta pslAB$ cassette	this study
$\Delta yfiR$ <i>pelG::Tn</i> $\Delta pslAB$	Tet ^R , Gm ^R , PA01 containing the WFP60 $\Delta pslAB$ cassette, with the <i>phoA</i> -Tn5 transposon inserted into <i>pelG</i>	this study
ClinSCV-110	SCV strain isolated from CF sputum	this study
<i>E. coli</i>		
BL21-(DE3)	Sm ^R , K12 <i>recF143 lacI^d lacZΔM15, xylA</i>	Novagen
DH5 α	<i>endA1, hsdR17(r_K-m_K+), supE44, recA1, gyrA (Nal^r), relA1, $\Delta(lacIZYA-argF)U169, deoR, \Phi 80dlac\Delta(lacZ)M15$</i>	[4]
MM337	K-12 <i>araD139 flbB5301 ptsF25 rbsR relA1 rpsL150 -(argF-lac)U169 -cya</i>	M. Manson
DY330	W3110 $\Delta lacU169 gal490 lcl857 \Delta(cro-bioA)$	[5]
Plasmids		
pALMAR3	Insertion vector for Tet ^R Mariner transposon	A. Levi
pBV-MCS4	Gm ^R , vanillate inducible high-copy number plasmid	[6]
pBV-PA5295	pBV-MCS4 with PA5295 under the control of vanillate promoter	[7]
pBV-PA5295 _{E328A}	pBV-MCS4 with PA5292 _{E328A} under the control of vanillate promoter	[7]
pET42b	Km ^R , purification vector, His ₆ -tag	Novagen
pET42b- <i>yfiN</i>	pET42b with N-terminal truncated <i>yfiN</i> as <i>XhoI</i> - <i>NdeI</i> fragment	this study
pET42b- <i>yfiB</i>	pET42b with <i>yfiB</i> as <i>XhoI</i> - <i>NdeI</i> fragment	this study
pUC18T-mini-Tn7T-Gm	Amp ^R , Gm ^R , Tn7 insertion vector	[8]
<i>yfiBNR</i> -complementation	pUC18T-mini-Tn7T-Gm with <i>yfiBNR</i> operon alleles as <i>HindIII</i> - <i>BamHI</i>	this study

vectors	fragments. Twelve plasmids in total	
pTNS2	Amp ^R , helper plasmid for Tn7 integration events	[8]
pME6032	Tet ^R , P _K , 9.8 kb pVS1 derived shuttle vector	[9]
pME6032- <i>yfiB</i>	pME6032 with <i>yfiB</i> as <i>EcoRI-BamHI</i> fragment	this study
pBBR-MCS4	Amp ^R , broad-host range cloning vector	[10]
pBBR-MCS5	Gm ^R , broad-host range cloning vector	[10]
p- <i>yfiR</i>	pBBR-MCS5 with <i>yfiR</i> as <i>BamHI-HindIII</i> fragment	this study
pBBR4- <i>wspR19</i>	pBBR-MCS4 with <i>wspR19</i> as <i>EcoRI-HindIII</i> fragment	this study
pME- <i>araC</i>	pME6032 with <i>araC/pBAD</i> -promoter as <i>EcoRI-BamHI</i> fragment	this study
p- <i>ara-yfiN</i>	pME- <i>araC</i> with <i>yfiN</i> as <i>SacI -BglII</i> fragment	this study
pME- <i>araC-yfiR-phoA</i>	pME- <i>araC</i> with <i>yfiR-phoA</i> fusion as <i>EcoRI-XhoI</i> fragment	this study
pME- <i>araC-tr-yfiR-phoA</i>	pME- <i>araC</i> with trunc <i>yfiR-phoA</i> fusion as <i>EcoRI-XhoI</i> fragment	this study
pME- <i>yfiR-cherry</i>	pME6032 with <i>yfiR-Mcherry</i> fusion as <i>EcoRI-BglII</i> fragment	this study
pMR20	Tet ^R , low copy number and broad host-range vector	[11]
pMR20- <i>yfiBNR</i>	pMR20 with <i>yfiBNR</i> as <i>HindIII-BamHI</i> fragment	this study
pMR20- <i>yfiR-M2</i>	pMR20 with <i>yfiR-M2</i> as <i>HindIII-BamHI</i> fragment	this study
pTn7- <i>yfiR-M2</i>	pUC18T-mini-Tn7T with <i>yfiR-M2</i> as <i>HindIII-BamHI</i> fragment	this study
pAD6Ω	Sm ^R , Tet ^R , pMR20 derivative containing EGFP under the <i>lac</i> promoter	this study
pKT25	pSU40 derivative with T25 fragment of CyaA	[12]
pKT25- <i>zip</i>	pKT25 derivative with leucine zipper of GCN4	[12]
pUT18C- <i>zip</i>	pUT18C derivative with leucine zipper of GCN4	[12]
pUT18C	pUC19 derivative with T18 fragment of CyaA. C-terminal fusions	[12]
pUT18	pUC19 derivative with T18 fragment of CyaA. N-terminal fusions	[12]
pUT18C- <i>yfi</i> vectors	Bacterial two-hybrid vectors with <i>yfi</i> genes as <i>XbaI-BamHI</i> fragments	this study
pKT25- <i>yfi</i> vectors	Bacterial two-hybrid vectors with <i>yfi</i> genes as <i>XbaI-BamHI</i> fragments	this study
pUT18- <i>yfi</i> vectors	Bacterial two-hybrid vectors with <i>yfi</i> genes as <i>HindIII-BamHI</i> fragments	this study
pME6032- <i>luxCDABE</i>	pME6032 with <i>luxCDABE</i> as <i>EcoRI-BamHI</i> fragment	this study
pME6032-promoter- <i>lux</i> vectors	pME6032- <i>luxCDABE</i> with PA01 promoters as <i>XhoI-EcoRI</i> fragments. Eight plasmids in total	this study
pEX18Ap	Amp ^R , <i>oriT</i> ⁺ , <i>sacB</i> ⁺ , gene replacement vector	[13]
pEX18Ap-Δ <i>yfiBNR</i>	pEX18Ap with <i>yfiBNR</i> Gm ^R -FRT deletion cassette as <i>HindIII-KpnI</i> fragment	this study
pEX18Ap-Δ <i>yfiNR</i>	pEX18Ap with <i>yfiNR</i> Gm ^R -FRT deletion cassette as <i>HindIII-KpnI</i> fragment	this study
pEX18Ap-Δ <i>yfiR</i>	pEX18Ap with <i>yfiR</i> deletion cassette as <i>HindIII-KpnI</i> fragment	this study
pFLP2	Amp ^R , FRT cassette excision vector	[13]

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