

Opinions

HIV-1 Group M Conserved Elements Vaccine

Morgane Rolland, David C. Nickle, James I. Mullins*

Recent HIV vaccine designs have sought to block viral escape pathways by compressing antigenic diversity. In light of HIV's propensity to mutate and thereby to ever ramify viral populations, could it be that providing sufficient protection against global diversity is an insurmountable problem? We propose an alternative HIV-1 vaccine design that deliberately includes viral segments conserved across the entire main group (or M group) of HIV-1 strains and excludes variable segments. We describe a prototype conserved elements (CE) vaccine constituted of 45 viral segments at least eight amino acids long that fulfill stringent conservation criteria.

Our paradigm contends that the best way to cope with HIV-1 diversity may be to avoid it altogether. We argue that a successful vaccine must elicit responses against conserved regions of the viral proteome in which mutations would severely compromise the viability of the virus. Simultaneously, it must not elicit responses against variable, "decoy" elements of the virus, i.e., features that can mutate while retaining function, and that can absorb much of the adaptive host immune response.

Coping with HIV-1's extensive diversity is a major challenge for vaccine design strategies. Centralized (consensus and ancestral) immunogens [1–3] have in some cases improved the breadth of responses, and recent designs seek to compress the more common variant features among circulating strains into immunogens [4–6]. However, there is a practical limit to antigenic complexity that may prevent inclusion of all escape pathways in realistically sized immunogens. Besides, HIV's propensity to mutate has been shown to provide means for HIV to escape from antiretrovirals and antibody and cytotoxic T lymphocyte (CTL) pressures.

The foregoing considerations led us to propose a vaccine exclusively composed of viral segments strictly conserved in all HIV-1 M group proteins and specifically devoid of mutable segments. The presence of segments that are nearly invariant in all HIV-1 M group proteomes strongly suggests that those CE are both obligatory for viral viability and are the Achilles' heels of the virus. Additionally, considering that variable segments can readily escape CTL pressures and can be highly immunogenic epitopes, we propose that mutable segments may act as immunologic decoys, subverting responses away from conserved elements.

Rationale

Despite HIV-1's extreme diversity, certain segments are nearly invariant (Figure 1). Near-total conservation of some sites implies that tight functional constraints obviate certain mutations. Indeed, escape from CTL responses, as with antiretroviral resistance, sometimes results in fitness decrease [7–14]. Specifically, escape mutations in Gag impaired viral fitness significantly more than mutations in Env ($p = 0.0033$, Mann-Whitney test) (unpublished data). These findings

corroborate the amount of polymorphisms that can typically be accommodated in each protein, as also hinted by a study of *env* V3 arguing for site-specific amino acid (AA) conservation as a predictor of viral fitness [15].

As HIV-1 establishes infection, it relentlessly mutates away from the founding strain [16]. However, some mutations recover consensus-like AAs [14,17,18], and these transitions to ancestral states may reverse CTL escape mutations back to susceptible and possibly increasingly fit forms upon transmission to human leukocyte antigen (HLA)-mismatched individuals [10,11,18,19,20].

HIV-1's predilection to mutate away from CTL-susceptible sequences highlights the critical importance of CTL responses. Gag, the second-most conserved HIV-1 protein, is immunodominant during infection [21] in different ethnic groups [22]. Numerous studies have suggested a superior role for Gag-specific CTL in viral containment [23–33], whereas CTL targeting of variable proteins may not contribute or even negatively impact immune control in HIV infection in humans [23,30] and in mouse studies [34]. Likewise, the emergence of escapes from Env-specific antibodies against variable regions hinders efforts to generate neutralizing antibodies [35,36].

Many studies underscored the challenge of broadening CTL recognition through vaccination as they reflected typical HIV immunodominance profiles: the immune system focuses on relatively few immunodominant epitopes, leaving many epitopes subdominant or cryptic. Since subdominant responses may be critical to effective suppression [37], mitigating immunodominance patterns could prove critical for successful vaccines.

Those considerations converge onto the approach of exclusively using conserved regions as components of HIV-1 vaccines [38–40]. Focusing on CTL induction, we articulate the conserved elements approach to vaccine design and describe here our candidate antigen. We consider that: (1) an HIV vaccine must be composed of rigorously conserved elements of the virus that cannot mutate without greatly

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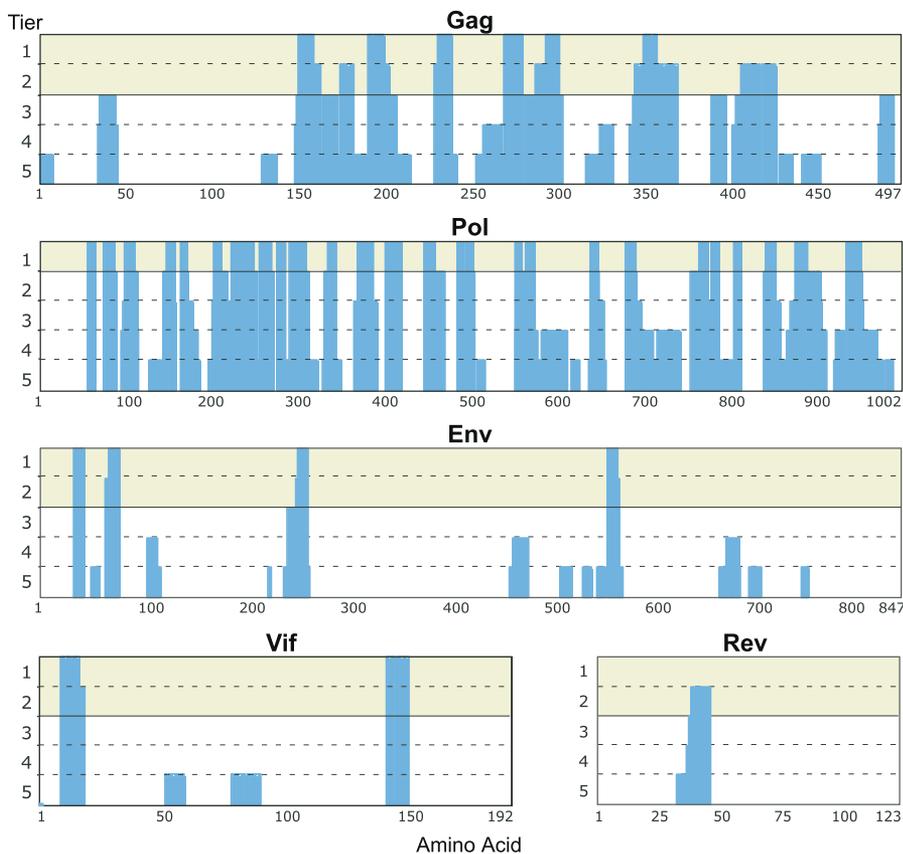
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Abbreviations: AA, amino acid; CE, conserved elements; CTL, cytotoxic T lymphocyte; HIVDB, HIV sequence database; HLA, human leukocyte antigen; M, main

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Figure 1. Conservation of M Group Sequences across HIV-1 Proteins

The full lengths of each of the viral proteins containing 1st and/or 2nd tier CE are shown. The histogram indicates the tier of conservation for each segment of at least eight AAs in length, with tier number shown on the y-axis. 1st tier segments correspond to segments at least eight AAs long in which the AA at each site is found in more than 98% of HIVDB database sequences. 2nd tier include sites at which the most common AA is found in less than 98% of sequences in the database, but at which two AAs together make up more than 99% of the AAs found at that site in the database. For comparison, additional tiers of potential use for vaccine design are shown. 3rd tier expands the set to include two variable sites, and 4th tier includes n variable sites, each of which satisfy the criteria of having two AAs that together make up more than 99% of the AAs in the database. The 5th tier corresponds to peptides that have n variable sites that satisfy the criteria for 4th tier, but the requirement for conservation encompassed by the two AAs at each site is relaxed to more than 98%.

detracting or eliminating viability; (2) mutable epitopes can act as immunodominant decoys, thereby sapping responses against protective, invariant epitopes; and (3) most HIV targets that can mutate without drastically impairing virus functionality do not contribute to the durable efficacy of the immune response.

CE Vaccine Design

To define CE, we evaluated the conservation in a database of all HIV-1 M group nucleotide sequences available (one sequence/person; sequence accession numbers and alignments available at <http://mullinslab.microbiol.washington.edu/HIV/Rolland2007/>). We then identified protein stretches at least eight AAs long, the minimal size of a CTL epitope, in which every AA is nearly conserved across the entire HIV-1 M group. Given the preponderance of defective HIV sequences in vivo, we did not require absolute conservation but included segments where each AA is found in more than 98% of all HIV-1 M group sequences (1st tier CE). This level of conservation is consistent with our recent study of fitness loss associated with mutations in Gag (unpublished observations). We have also included CE in

which two variant forms at a given position together account for more than 99% of the amino acids found in the database (2nd tier CE) (Figure 1). Although the Los Alamos HIV sequence database (HIVDB) is overwhelmingly constituted of clades B and C, other M group subtypes were sufficiently represented in sequence alignments such that a subtype-specific AA would drag that site below the 1st tier criterion of conservation.

Most 1st and 2nd tier CE were identified in Pol ($n = 27$; only 1st tier peptides were included in our CE, due to the extreme conservation of Pol) and Gag ($n = 10$), the two most conserved HIV-1 proteins; however, other CE were found in Env (five), Vif (two) and Rev (one) (Table 1). The idiosyncrasy of the CE design means that more than half of CE peptides correspond to Pol segments. However, it should be noted that Pol epitopes are rarely targeted in natural infection. Recent results suggest that the infrequent recognition of Pol epitopes is likely related to the considerably lower ratio of Pol compared to Gag proteins produced by the virus [41] rather than to a lack of immunogenicity. Both observations raise concerns about Pol segments as sagacious choices for vaccine candidates.

Table 1. CE Peptide Coverage

Protein	L	Coverage			Coverage			All Peptides		CTL Epitopes		9-mer Coverage		Epitope Concentration		Supertype Restriction
		1st Tier Peptides			1st + 2nd Tier Peptides			n	L	CE	Non-CE	CE	Non-CE	CE	Non-CE	
		n	L	% Total	n	L	% Total									
Gag	497	6	53	10.7	10	111	22.3	19	211	8	215	5.83	94.17	1.37	2.28	All 9
Pol	1,002	27	312	31.1	(27.0)	312.0	(31.1)	27	312	11	150	9.29	90.71	1.18	1.65	All 9
Vif	192	2	17	8.9	2	19	9.9	3	29	0	26	1.59	98.41	0	0.26	A3, A24
Vpr	96	0	0		0	0										
Tat	92	0	0		0	0										
Rev	123	0	0		1	8	6.9	2	16	0	18	0.2	99.8	0	0.18	B27
Vpu	82	0	0		0	0										
Env	847	4	41	4.8	5	56	6.6	8	92	3	136	1.91	98.09	1.57	1	A2, A24, A1, B62, B58
Nef	206	0	0		0	0								0	1.39	
Total	3,137		423	13.5		506	16.1	59	660	22	545					

CE peptides are described for each protein by the number of fragments (*n*), the corresponding number of AAs (*L*), and the fraction of the whole protein they represent. Coverage is shown separately for 1st tier peptides, 1st and 2nd tier peptides, and for the total combinations of 1st and 2nd tier variant peptides. Due to the extreme conservation of the protein, only 1st tier peptides are considered for Pol (the 1st tier numbers are enclosed in parentheses in the 1st and 2nd tier columns). HIV-1 M group sequences were scanned for CTL epitopes of eight to 11 AAs in length previously reported in the HIVDB (these epitopes are heavily biased for viruses and laboratory reference strains belonging to subtype B, and increasingly, subtype C). Due to the requirement of including multiple peptides to encompass 2nd tier CE, the number of peptides to cover all of the overlapping 9-mer peptides were generated from the sequence dataset used to identify the CE (available at <http://mullinslab.microbiol.washington.edu/HIV/Rolland2007/>); 9-mer coverage corresponds to the percentage of all possible 9-mer peptides found in CE and non-CE segments. The epitope concentration is the number of CTL epitopes divided by the 9-mer coverage. The nine supertypes correspond to B7, A3, A2, A24, B44, A1, B27, B62, and B58 [47].

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Indeed, early vaccine trial results show a preponderance of CTL responses in Pol when Gag and Pol are administered at the same ratio: a study of immune responses elicited by an Ad5 vaccine candidate in 30 HIV-uninfected participants showed that 39 distinct epitopes were recognized in Pol and eight in Gag (Helen Horton, personal communication). As another consequence of not defining our candidate vaccine based on the frequency of recognition in natural infection, our CE vaccine design does not contain Nef epitopes, a traditional component of HIV-1 vaccine candidates, including the recently failed Merck/STEP clinical trial. We reason that while both Gag and Nef are under important CTL pressure, the degree of polymorphisms that can be accommodated in Nef leads to a cycle of CTL escape mutations/de novo responses to variant epitopes, and those readily escapable shifting CTL responses do not afford long-term control of HIV-1 replication. In contrast, a CTL escape mutation in Gag has a strong impact on viral viability and those strong constraints on HIV-1 evolution might coerce the virus toward minimal replicative fitness capacities. Likewise, we consider that a CTL-based vaccine would not benefit from focusing largely on Env antigens.

The CE segments contain known CTL epitopes, including some previously associated with long-term nonprogression or recurrent HIV exposure without seroconversion [42–46] (Note: CE sequences are available at <http://mullinslab.microbiol.washington.edu/HIV/Rolland2007/>). Some epitopes are strictly embedded in CE: eight in Gag, 11 in Pol, and three in Env, representing a variety of HLA restrictions. In addition, CE sequences include supertype motifs from all nine supertypes (B7, A3, A2, A24, B44, A1, B27, B62, B58) in both Gag and Pol, providing a population coverage of at least 80%, regardless of ethnicity [47]. Additional epitopes were found to overlap CE/non-CE junctions (data not shown); thus extending CE immunogens could increase the number of

peptides available to CD8⁺ T lymphocytes. However, their inclusion would relax our stringent conservation criteria. It should be noted that attempts to correlate CTL breadth and viral containment have been inconclusive [25,26,48–51]. The conundrum of extending CE without including potentially pathogenic variable positions thus begs for a better understanding of virus viability with respect to sequence conservation.

One intricate issue with the CE strategy is to ensure that antigen structures are recognized outside of native proteins. Thus, CE constructs have to be engineered optimally to elicit immune responses by capitalizing on the mechanisms governing epitope processing and presentation while preventing the creation of junctional immunogenicity or homology to the HIV or human proteome [52]. Furthermore, certain elements of the HIV proteome may be conserved because there is a dearth of features within and surrounding these elements that are capable of mediating efficient processing for presentation on HLA. Such elements would not be useful for inclusion in a vaccine. Thus, configurations should explicitly capitalize on emerging data on the mechanisms that govern epitope processing and presentation.

In eliciting responses to particular segments, another undefined problem is immunodominance [22,53]. With respect to all 9-mer peptides in datasets, epitope concentrations were lower in CE than in non-CE, except in Env (Table 1). However, epitopes in variable regions like Env are under-represented in databases [18] and HLA class I allelic promiscuity is more pervasive than previously thought [54]. In terms of the utility of a CE vaccine, considering the lower representation of epitopes in CE versus non-CE as problematic may be misleading, especially as our CE design is intended to not reproduce certain features of the antiviral response seen in natural infection. Since CE are apparently

not preferentially targeted in natural infection, CE vaccine designs have to deliberately focus the immune response toward segments that are not traditionally immunodominant, thereby assuring that reactivity against CE is not obscured by the immunodominating responses evident in natural infection. For example, isolating subdominant epitopes in CE vaccines might promote their recognition [55,56].

Concluding Remarks

To overcome the stumbling block posed by HIV-1 diversity, we specified a novel HIV vaccine design that exclusively includes portions of the proteome meeting stringent conservation criteria and thereby explicitly deprived of mutable segments. We consider that variable regions act as decoys that divert the immune system from responding to conserved regions, which are seemingly better for long-term viral control. Crucially, our scheme culminates in a single global CE vaccine suited for evaluation against all circulating HIV-1 M group strains. ■

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References

- Gao F, Weaver EA, Lu Z, Li Y, Liao HX, et al. (2005) Antigenicity and immunogenicity of a synthetic human immunodeficiency virus type 1 group m consensus envelope glycoprotein. *J Virol* 79: 1154–1163.
- Doria-Rose N, Learn GH, Rodrigo AG, Nickle DC, Li F, et al. (2005) Human immunodeficiency virus type 1 subtype B ancestral envelope protein is functional and elicits neutralizing antibodies in rabbits similar to those elicited by a circulating subtype B envelope. *J Virol* 79: 11214–11224.
- Kothe DL, Li Y, Decker JM, Bibollet-Ruche F, Zammit KP, et al. (2006) Ancestral and consensus envelope immunogens for HIV-1 subtype C. *Virology* 352: 438–449.
- Fischer W, Perkins S, Theiler J, Bhattacharya T, Yusim K, et al. (2007) Polyvalent vaccines for optimal coverage of potential T-cell epitopes in global HIV-1 variants. *Nat Med* 13: 100–106.
- Nickle DC, Rolland M, Jensen MA, Pond SLK, Deng W, et al. (2007) Coping with viral diversity in HIV vaccine design. *PLoS Comput Biol* 3: e75. doi:10.1371/journal.pcbi.0030075
- Li F, Horton H, Gilbert PB, McElrath JM, Corey L, et al. (2007) HIV-1 CTL-based vaccine immunogen selection: antigen diversity and cellular response features. *Curr HIV Res* 5: 97–107.
- Fernandez CS, Stratov I, De Rose R, Walsh K, Dale CJ, et al. (2005) Rapid viral escape at an immunodominant simian-human immunodeficiency virus cytotoxic T-lymphocyte epitope exacts a dramatic fitness cost. *J Virol* 79: 5721–5731.
- Martinez-Picado J, Prado JG, Fry EE, Pfafferoth K, Leslie A, et al. (2006) Fitness cost of escape mutations in p24 Gag in association with control of human immunodeficiency virus type 1. *J Virol* 80: 3617–3623.
- Gandhi RT, Wurcel A, Rosenberg ES, Johnston MN, Hellmann N, et al. (2003) Progressive reversion of human immunodeficiency virus type 1 resistance mutations in vivo after transmission of a multiply drug-resistant virus. *Clin Infect Dis* 37: 1693–1698.
- Leslie AJ, Pfafferoth KJ, Chetty P, Draenert R, Addo MM, et al. (2004) HIV evolution: CTL escape mutation and reversion after transmission. *Nat Med* 10: 282–289.
- Allen TM, Altfeld M, Yu XG, O'Sullivan KM, Lichterfeld M, et al. (2004) Selection, transmission, and reversion of an antigen-processing cytotoxic T-lymphocyte escape mutation in human immunodeficiency virus type 1 infection. *J Virol* 78: 7069–7078.

- Peyerl FW, Barouch DH, Letvin NL (2004) Structural constraints on viral escape from HIV- and SIV-specific cytotoxic T-lymphocytes. *Viral Immunol* 17: 144–151.
- Peyerl FW, Bazick HS, Newberg MH, Barouch DH, Sodroski J, et al. (2004) Fitness costs limit viral escape from cytotoxic T lymphocytes at a structurally constrained epitope. *J Virol* 78: 13901–13910.
- Li B, Gladden AD, Altfeld M, Kaldor JM, Cooper DA, et al. (2007) Rapid reversion of sequence polymorphisms dominates early human immunodeficiency virus type 1 evolution. *J Virol* 81: 193–201.
- da Silva J (2006) Site-specific amino acid frequency, fitness, and the mutational landscape model of adaptation in HIV-1. *Genetics* 174: 1689–1694.
- Shankarappa R, Margolick JB, Gange SJ, Rodrigo AG, Upchurch D, et al. (1999) Consistent viral evolutionary changes associated with the progression of human immunodeficiency virus type 1 infection. *J Virol* 73: 10489–10502.
- Herbeck JT, Nickle DC, Learn GH, Gottlieb GS, Curlin ME, et al. (2006) Human immunodeficiency virus type 1 env evolves toward ancestral states upon transmission to a new host. *J Virol* 80: 1637–1644.
- Liu Y, McNeven J, Cao J, Zhao H, Genowati I, et al. (2006) Selection on the human immunodeficiency virus type 1 proteome following primary infection. *J Virol* 80: 9519–9529.
- Jones NA, Wei X, Flower DR, Wong M, Michor F, et al. (2004) Determinants of human immunodeficiency virus type 1 escape from the primary CD8+ cytotoxic T lymphocyte response. *J Exp Med* 200: 1243–1256.
- Allen TM, Altfeld M, Geer SC, Kalife ET, Moore C, et al. (2005) Selective escape from CD8+ T-cell responses represents a major driving force of human immunodeficiency virus type 1 (HIV-1) sequence diversity and reveals constraints on HIV-1 evolution. *J Virol* 79: 13239–13249.
- Coplan PM, Gupta SB, Dubey SA, Pitisuttithum P, Nikas A, et al. (2005) Cross-reactivity of anti-HIV-1 T cell immune responses among the major HIV-1 clades in HIV-1-positive individuals from 4 continents. *J Infect Dis* 191: 1427–1434.
- Frahm N, Korber BT, Adams CM, Szinger JJ, Draenert R, et al. (2004) Consistent cytotoxic-T-lymphocyte targeting of immunodominant regions in human immunodeficiency virus across multiple ethnicities. *J Virol* 78: 2187–2200.
- Zuniga R, Lucchetti A, Galvan P, Sanchez S, Sanchez C, et al. (2006) Relative dominance of Gag p24-specific cytotoxic T lymphocytes is associated with human immunodeficiency virus control. *J Virol* 80: 3122–3125.
- Buseyne F, Le Chenadec J, Corre B, Porrot F, Burgard M, et al. (2002) Inverse correlation between memory gag-specific cytotoxic T lymphocytes and viral replication in human immunodeficiency virus-infected children. *J Infect Dis* 186: 1589–1596.
- Edwards BH, Bansal A, Sabbaj S, Bakari J, Mulligan MJ, et al. (2002) Magnitude of functional CD8+ T-cell responses to the gag protein of human immunodeficiency virus type 1 correlates inversely with viral load in plasma. *J Virol* 76: 2298–2305.
- Masemola A, Mashishi T, Khoury G, Mohube P, Mokgotho P, et al. (2004) Hierarchical targeting of subtype C human immunodeficiency virus type 1 proteins by CD8+ T cells: correlation with viral load. *J Virol* 78: 3233–3243.
- Novitsky V, Gilbert P, Peter T, McLane MF, Gaolekwe S, et al. (2003) Association between virus-specific T-cell responses and plasma viral load in human immunodeficiency virus type 1 subtype C infection. *J Virol* 77: 882–890.
- Ogg GS, Jin X, Bonhoeffer S, Dunbar PR, Nowak MA, et al. (1998) Quantitation of HIV-1-specific T lymphocytes and plasma viral load of viral RNA. *Science* 279: 2103–2106.
- Ramduth D, Chetty P, Mngquandano NC, Nene N, Harlow JD, et al. (2005) Differential immunogenicity of HIV-1 clade C proteins in eliciting CD8+ and CD4+ cell responses. *J Infect Dis* 192: 1588–1596.
- Kiepiela P, Ngumbela K, Thobakgale C, Ramduth D, Honeyborne I, et al. (2007) CD8+ T-cell responses to different HIV proteins have discordant associations with viral load. *Nat Med* 13: 46–53.
- Riviere Y, McChesney MB, Porrot F, Tanneau-Salvadori F, Sansonetti P, et al. (1995) Gag-specific cytotoxic responses to HIV type 1 are associated with a decreased risk of progression to AIDS-related complex or AIDS. *AIDS Res Hum Retroviruses* 11: 903–907.
- Sacha JB, Chung C, Rakasz EG, Spencer SP, Jonas AK, et al. (2007) Gag-specific CD8+ T lymphocytes recognize infected cells before AIDS-virus integration and viral protein expression. *J Immunol* 178: 2746–2754.
- Geldmacher C, Currier JR, Herrmann E, Haule A, Kuta E, et al. (2007) CD8 T-cell recognition of multiple epitopes within specific Gag regions is associated with maintenance of a low steady-state viremia in human immunodeficiency virus type 1-seropositive patients. *J Virol* 81: 2440–2448.
- Toapanta FR, Craigio JK, Montelaro RC, Ross TM (2007) Reduction of anti-HIV-1 Gag immune responses during co-immunization: immune interference by the HIV-1 envelope. *Curr HIV Res* 5: 199–209.
- Wei X, Decker JM, Wang S, Hui H, Kappes JC, et al. (2003) Antibody neutralization and escape by HIV-1. *Nature* 422: 307–312.
- Richman DD, Wrinn T, Little SJ, Petropoulos CJ (2003) Rapid evolution of

- the neutralizing antibody response to HIV type 1 infection. *Proc Natl Acad Sci U S A* 100: 4144–4149.
37. Frahm N, Kiepiela P, Adams S, Linde CH, Hewitt HS, et al. (2006) Control of human immunodeficiency virus replication by cytotoxic T lymphocytes targeting subdominant epitopes. *Nat Immunol* 7: 173–178.
 38. Altfeld M, Allen TM (2006) Hitting HIV where it hurts: an alternative approach to HIV vaccine design. *Trends Immunol* 27: 504–510.
 39. Kent SJ, Fernandez CS, Dale CJ, Davenport MP (2005) Reversion of immune escape HIV variants upon transmission: insights into effective viral immunity. *Trends Microbiol* 13: 243–246.
 40. Johnston MI, Fauci AS (2007) An HIV vaccine—evolving concepts. *N Engl J Med* 356: 2073–2081.
 41. Wilson W, Braddock M, Adams SE, Rathjen PD, Kingsman SM, et al. (1988 or 89) HIV expression strategies: ribosomal frameshifting is directed by a short sequence in both mammalian and yeast systems. *Cell* 55: 1159–1169.
 42. Kaul R, Dong T, Plummer FA, Kimani J, Rostron T, et al. (2001) CD8+ lymphocytes respond to different HIV epitopes in seronegative and infected subjects. *J Clin Invest* 107: 1303–1310.
 43. Kaul R, Plummer F, Clerici M, Bomsel M, Lopalco L, et al. (2001) Mucosal IgA in exposed, uninfected subjects: evidence for a role in protection against HIV infection. *AIDS* 15: 431–432.
 44. Kaul R, Plummer FA, Kimani J, Dong T, Kiama P, et al. (2000) HIV-1-specific mucosal CD8+ lymphocyte responses in the cervix of HIV-1-resistant prostitutes in Nairobi. *J Immunol* 164: 1602–1611.
 45. Rowland-Jones S, Dong T, Krausa P, Sutton J, Newell H, et al. (1998) The role of cytotoxic T-cells in HIV infection. *Dev Biol Stand* 92: 209–214.
 46. Alimonti JB, Kimani J, Matu L, Wachih C, Kaul R, et al. (2006) Characterization of CD8 T-cell responses in HIV-1-exposed seronegative commercial sex workers from Nairobi, Kenya. *Immunol Cell Biol* 84: 482–485.
 47. Sette A, Sidney J (1999) Nine major HLA class I supertypes account for the vast preponderance of HLA-A and -B polymorphism. *Immunogenetics* 50: 201–212.
 48. Gea-Banacloche JC, Migueles SA, Martino L, Shupert WL, McNeil AC, et al. (2000) Maintenance of large numbers of virus-specific CD8+ T cells in HIV-infected progressors and long-term nonprogressors. *J Immunol* 165: 1082–1092.
 49. Betts MR, Ambrozak DR, Douek DC, Bonhoeffer S, Brenchley JM, et al. (2001) Analysis of total human immunodeficiency virus (HIV)-specific CD4(+) and CD8(+) T-cell responses: relationship to viral load in untreated HIV infection. *J Virol* 75: 11983–11991.
 50. Addo MM, Yu XG, Rathod A, Cohen D, Eldridge RL, et al. (2003) Comprehensive epitope analysis of human immunodeficiency virus type 1 (HIV-1)-specific T-cell responses directed against the entire expressed HIV-1 genome demonstrate broadly directed responses, but no correlation to viral load. *J Virol* 77: 2081–2092.
 51. Musey L, Hughes J, Schacker T, Shea T, Corey L, et al. (1997) Cytotoxic-T-cell responses, viral load, and disease progression in early human immunodeficiency virus type 1 infection. *N Engl J Med* 337: 1267–1274.
 52. Rolland M, Nickle DC, Deng W, Frahm N, Brander C, et al. (2007) Recognition of HIV-1 peptides by host CTL is related to HIV-1 similarity to human proteins. *PLoS ONE* 2: e823. doi:10.1371/journal.pone.0000823
 53. Rolland M, Heckerman D, Deng W, Rousseau C, Coovadia H, et al. (2007) Broad and Gag-biased HIV-1 epitope repertoires are associated with lower viral loads. *PLoS ONE*. In press.
 54. Frahm N, Yusim K, Suscovich TJ, Adams S, Sidney J, et al. (2007) Extensive HLA class I allele promiscuity among viral CTL epitopes. *Eur J Immunol* 37: 2419–2433.
 55. Assarsson E, Sidney J, Oseroff C, Pasquetto V, Bui HH, et al. (2007) A quantitative analysis of the variables affecting the repertoire of T cell specificities recognized after vaccinia virus infection. *J Immunol* 178: 7890–7901.
 56. Schaubert KL, Price DA, Frahm N, Li J, Ng HL, et al. (2007) Availability of a diversely avid CD8+ T cell repertoire specific for the subdominant HLA-A2-restricted HIV-1 Gag p2419–27 epitope. *J Immunol* 178: 7756–7766.

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