

**Supplementary Table 1. Primers used for Cas9/sgRNA-mediated genome engineering in sweet orange**

Primer name	Primer sequence from 5' to 3'	Note
Cas9-5-BamHI	AGGT <u>GGATCC</u> CGTTGGACCGGTGCCACCAGT	
Cas9-3-EcoRI	TGATCAGCGAGCTCTAG <u>GAATTCTTA</u>	
NosT-5-EcoRI	AGGATCCACC <u>GGTGACGAATT</u> CCGAATTCCCCGATCGTTCAA	
NosT-3-XhoI-AscI	<u>AGGCGGCCATTAAAT</u> TCGAGCCGATCTAGAACATAGATGA	
sgRNA-5-BamHI	TCGACT <u>GGATCC</u> GGTACCAAG	
sgRNA-3-SacI	AGGT <u>GAGCTTC</u> GCCCTTAATGCCAACTT	
CaMV35S-5-XhoI	<u>ACTCGAG</u> ACTAGTACCATGGTGGACTCCTCTAA	
sgRNA-CsPDS2	TTGTGCACAACCTCTCAAATGAAATGAACCTTC	5'-phosphorylated
sgRNA-CsPDS1	GCAATTGTACGTTTAGAGCTAGAAATAGCAA	5'-phosphorylated
NosT-3-AscI	ACCTGGGCC <u>GGCGCGCC</u> GATCTAGAACATAGATGA	
CsPDS-5-P1	AGGATTCTTCATTTCAATGA	
CsPDS-3-P2	TGAACATTCAATTACTTAGG	
7290-5-P1	TCCACTGGA <u>ACTGGTGG</u> AATCACT	
7290-3-P2	TGATCATGGATATAAGAAGACT	
2370-5-P1	TACCCACAATCACAGGATAAAGT	
2370-3-P2	TGACTAAGGATTGAGCCAGGTTCG	
4890-5-P1	AGACATCATCCAAGGGTACTCT	
4890-3-P2	TGACTGA <u>ACTTCCTACTTC</u> CATG	
3808-5-P1	ACGATCTATTGGCAACATGCCAAA	
3808-3-P2	AGCTGGTAGGTGTCCTTGTTC	
3020-5-P1	TGGGATCGATTATTGTTGGATTCA	
3020-3-P2	ACACCAAAACCC <u>TTGTTAACAAATC</u>	
5862-5-P1	TCATCCAACCGGATTGATATGCTC	
5862-3-P2	ACAAACACGGTAGCAA <u>ATGCTCCC</u>	
9370-5-P1	ACCTTCTATCC <u>CATCTTATC</u> CTCAC	
9370-3-P1	ACACAACATACACAGGAGCAAGCA	
5100-5-P1	TGTGGTGCTTGT <u>CTTAATTAGCAC</u>	
5100-3-P2	ATACTCC <u>CTGATTGCTGCAAGTCA</u>	
6180-5-P1	TGTCCA <u>AAATTGAGCCC</u> GAATTAAG	
6180-3-P2	TCATCTAAT <u>CTCACCTACCTCTTC</u>	
2600-5-P1	AGTGA <u>ATGTCGTT</u> CATTCACTG	
2600-3-P2	TCATACATT <u>TCACATGACCC</u> ACTC	
8390-5-P1	TGAGCTTAT <u>CACAGAGCCC</u> AAATTC	
8390-3-P2	TCCATGTAT <u>GAAGACTCC</u> ACCCAA	
5080-5-P1	ACTGGCTGA <u>ATGATGGG</u> AAACCT	
5080-3-P2	TGCTCGTGT <u>CTATGTCG</u> TACAA	
0160-5-P1	TGTTGAA <u>ACATCTAGCAAC</u> CTGTAG	
0160-3-P2	ACGTATCGTAC <u>ATAATGGGT</u> GA	
2870-5-P1	TGCTTG <u>GTGCTTCACCTT</u> CGTGTGA	
2870-3-P2	TGCTTCT <u>CCGTGACCACACAC</u> CGC	

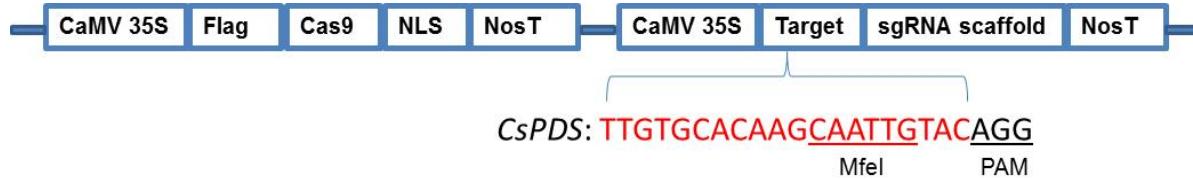
**Supplementary Table 2. Forty-six potential off-target sequences of the *CsPDS* gene in the sweet orange genome**

Off-target site	Off-target sequence	Test by MfeI-suppressed PCR
Cs9g07290	GAGAGCACAAG <b>CAATTGTAC</b>	Yes
Cs9g12370	<b>CATGACACAAGCAATTGTAG</b>	Yes
Cs7g14890	TGACACACAAG <b>CAATTGTAA</b>	Yes
Cs5g10470	<b>CCATGCACAAGCAATTGCAC</b>	No
Cs5g05850	CTGTGCACAAGCAAT <b>CGTGC</b>	No
orange1.1t03808	<b>GATGACACAAGCAATTGTT</b>	Yes
orange1.1t03020	TTTT <b>CAACAAGCAATTGTAT</b>	Yes
orange1.1t05862	<b>CACTTCGCAAGCAATTGTAC</b>	Yes
orange1.1t01105	<b>ATCTGCACAAGCAATTTACA</b>	No
orange1.1t00848	TTGTGCACAAG <b>CACTTGGA</b>	No
orange1.1t00315	<b>TCGAGCACAAGCAATTGCCA</b>	No
Cs9g09370	<b>AACCCTGCAAGCAATTGTAC</b>	Yes
Cs9g05100	<b>GAAGACCCAAGCAATTGTAC</b>	Yes
Cs8g17760	<b>AAATGCACAAGCAATTACA</b>	No
Cs8g16180	TAACATCCAAG <b>CAATTGTAC</b>	Yes
Cs8g12600	<b>TGTGGCACAAGCAATTGAAT</b>	Yes
Cs7g18390	<b>AAGGATCCAAGCAATTGTAC</b>	Yes
Cs7g09470	TTTTGCACAAGCAATT <b>ACAT</b>	No
Cs7g04320	TTTT <b>CCACAAGCAATTGTGT</b>	No
Cs7g01860	<b>TAGTGCACAAGCAATATGAC</b>	No
Cs6g15500	<b>ATGTGCACAAGCAAACCAAG</b>	No
Cs6g05090	<b>TAGCACACAAGCAATTGTCA</b>	No
Cs6g02660	<b>CCCTGCGCAAGCAATTGTAC</b>	No
Cs5g31970	<b>ACTTGCACAAGCAATTTACA</b>	No
Cs5g20160	TTGTGCACAAG <b>CAGTTTCTT</b>	No
Cs5g18400	<b>CTGTGCACAAGCAAACACTA</b>	No
Cs5g11660	<b>ACAATCACAAGCAATTGTT</b>	No
Cs5g10520	TGGTGCACAAG <b>CAATCACCA</b>	No
Cs4g12540	TTGTGCACAAG <b>GCACCCATTG</b>	No
Cs4g12530	TTGTGCACAAG <b>GCACCCATTG</b>	No
Cs3g25770	<b>AGGTGCACAAGCAATATCAA</b>	No
Cs3g25080	<b>AACTTCACAAGCAATTGTGA</b>	Yes
Cs3g21130	<b>TGAATCACAAGCAATTGTGC</b>	No
Cs3g20160	<b>CAGTAGACAAGCAATTGTAA</b>	Yes
Cs3g19160	<b>GATTGCACAAGCAATTAGCA</b>	No
Cs3g12870	<b>TCTACACAAGCAATTGTGA</b>	Yes

**Supplementary Table 2. Forty-six potential off-target sequences of the *CsPDS* gene in the sweet orange genome (continued)**

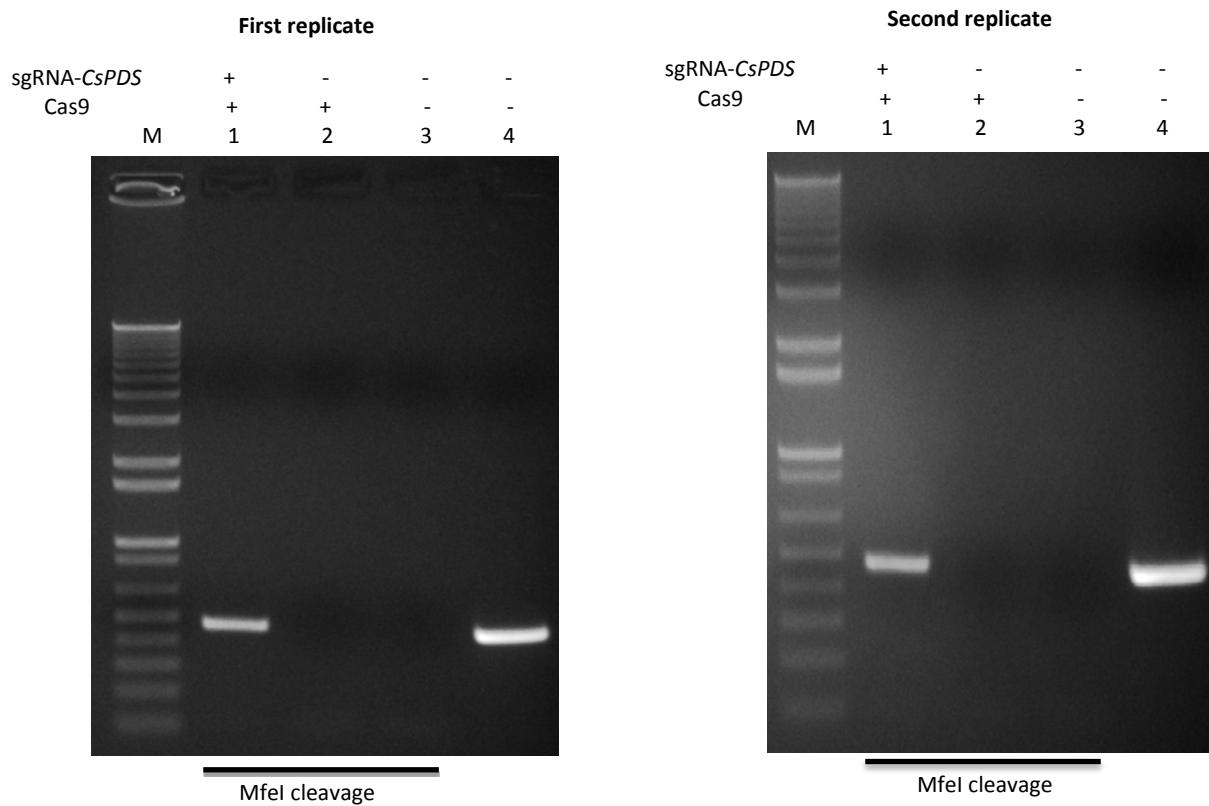
Off-target site	Off-target sequence	Test by MfeI-suppressed PCR
Cs3g07250	TTGTGCACAAGCATTCCAAT	No
Cs2g29170	TTGTGCACAAGCATCTTAAA	No
Cs2g11120	TTGTGCACAAGCATAAACCA	No
Cs2g06350	TTATGCACAAGCAATTATAG	No
Cs2g06110	TCATGCACAAGCAATTAAAGA	No
Cs2g04990	AAATATACAAGCAATTGTAA	No
Cs2g04980	AAATATACAAGCAATTGTAA	No
Cs1g18550	AGCAGCACAAGCAATTGAGT	No
Cs1g16510	TAGTGCACAAGCAATACGAC	No

The *MfeI* site is shown in green; Mismatching bases are shown in red.

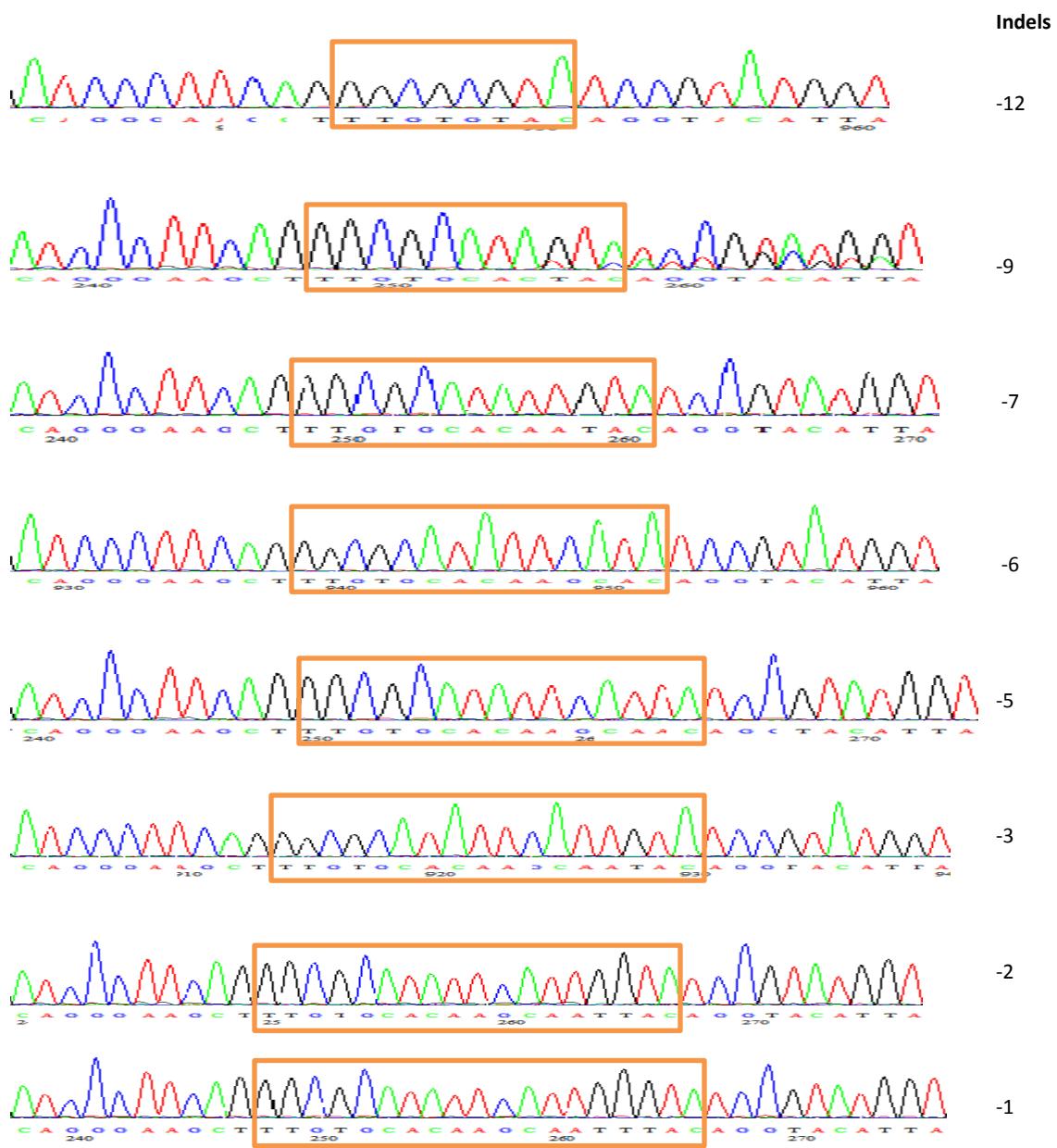


ctcgagACTAGTACCATGGTGGACTCCTCTAAAGCTTGCATGCCGCAGGTCCCCAGATTAGCCTTTCAATTTCAGAAAG  
AATGCTAACCCACAGATGGTAGAGAGGGCTACGCAGCAGGTCTCATCAAGACGATCTACCGAGCAATAATCTCCAGG  
AAATCAAATACCTTCCAAGAAGGTTAAAGATGCAGTAAAAGATTAGGACTAACTGCATCAAGAACACAGAGAAAGA  
TATATTCTCAAGATCAGAAGTACTATTCCAGTATGGACGATTCAAGGCTTGCTCACAAACCAAGGCAAGTAATAGAGA  
TTGGAGTCTCTAAAAGGTAGTCCCCTGAATCAAAGGCCATGGAGTCAAAGATTCAAATAGAGGACCTAACAGAACT  
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AAGGGTAATATCCGAAACCTCCCGATTCCATTGCCAGCTATGTCACTTATTGTGAAGATAGTGGAAAAGGAAG  
GTGGCTCTACAAATGCCATATTGCGATAAAGGAAAGGCCATCGTTGAAGATGCCCTGCCGACAGTGGTCCAAAGA  
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ATCTCCACTGACGTAAGGGATGACGCACAATCCACTATCCTCGAAGACCCCTCCTCTATATAAGGAAGTTCATTCAT  
TTGGAGAGGG**TTGTGCACAAGCAATTGTAC**GTAAAGCTAGAAATAGCAAGTTAAAATAAGGCTAGTCCGTTATCAAC  
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TCGAATTCCCCGATCGTCAAACATTGGCAATAAAGTTCTTAAGATTGAATCCTGTTGCCGGTCTGCGATGATTATC  
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GGTGTATCTATGTTACTAGATCggcgcc

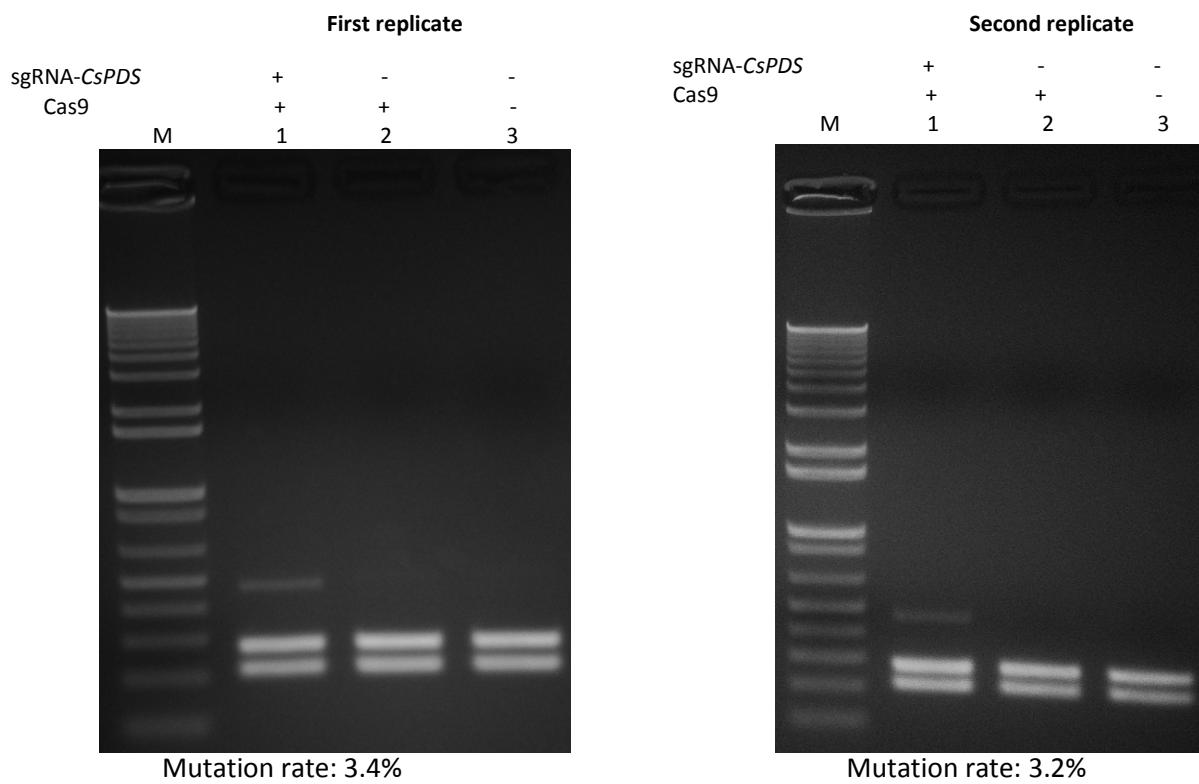
**Supplementary Figure S1. The sequence of the CaMV 35S promoter-CsPDS-targeting sgRNA – NosT of the Cas9/sgRNA construct.** The CaMV 35S promoter is shown in blue. The guide sequence is shown in red. The sgRNA scaffold is shown in purple. The Nos terminator is shown in green. The transcription start site is marked by an arrow.



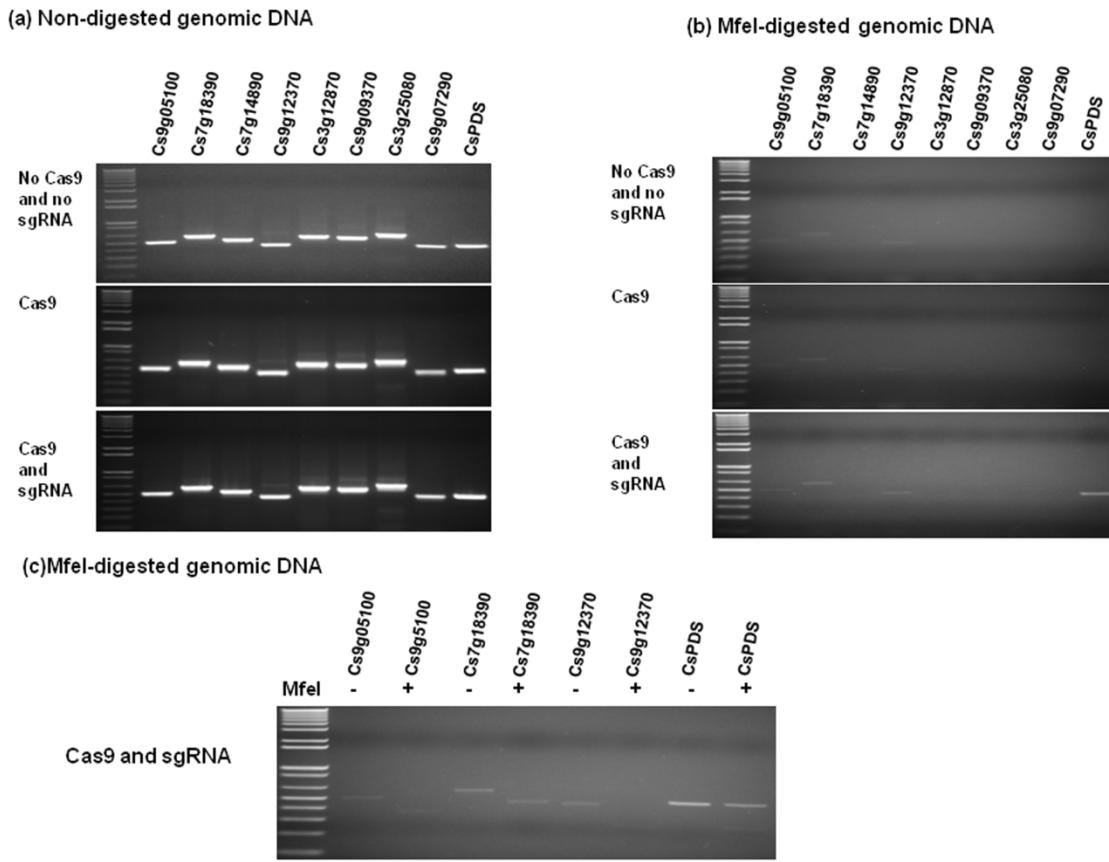
**Supplementary Figure S2. Two replicates of the experiment presented in Figure 2 (b).** Restriction-enzyme-digestion-suppressed PCR was used to detect the Cas9/sgRNA-induced mutation *in planta*. PCR amplification was conducted using the primers CsPDS-5-P1 and CsPDS-3-P2, which flank the target site within the *CsPDS* gene. Lanes 1-3, the template genomic DNA was digested with *Mfe*I. Lane 4, non-digested genomic DNA was used as a template. The PCR product in lane 1 resulted from Cas9/sgRNA-induced disruption of *Mfe*I, which indicates the expected disruption of the *Mfe*I site within the *CsPDS* gene. M, 1 kb DNA ladder.



**Supplementary Figure S3. The representative indel chromatograms of the *CsPDS12* mutations induced by Cas9/sgRNA.** The targete sequence within the *CsPDS* gene is highlighted by an orange rectangle.



**Supplementary Figure S4.** Two replicates of the experiment presented in Figure 2 (d). Measurement of the mutation rate of the *CsPDS* gene induced by Cas9/sgRNA. After PCR amplification of the targeted PDS region, the products were subjected to *Mfe*I digestion. After separation on an agarose gel, the intensities of the bands were quantified using AlphaImager EP. The mutation rate was calculated by dividing the intensity of the uncut band by the intensity of all the bands in the lane. M, 1 kb DNA ladder.



**Supplementary Figure S5. Analysis of potential off-target sequences of the *CsPDS*-targeting Cas9/sgRNA by *MfeI*-suppressed PCR.** (a) Eight potential off-target sequences were amplified by PCR when non-digested genomic DNA was used as the template. (b) When *MfeI*-digested genomic DNA was used, no PCR products or alleviated PCR products were observed. (c) After selective PCR amplification of mutagenized *CsPDS* genes, only the PCR product from the *CsPDS* gene showed resistance to *MfeI* digestion. These results indicated that the *MfeI* restriction sites in the 8 potential off-target sequences were not disrupted by Cas9/sgRNA cleavage and NHEJ repair.