

A Constant-Force Technique to Measure Corneal Biomechanical Changes after Collagen Cross-Linking



Olivier Richoz¹, Sabine Kling¹, Souska Zandi¹, Arthur Hammer¹, Eberhard Spoerl², Farhad Hafezi^{1,3}*

1 Dept. of Ophthalmology, Geneva University Hospitals, Geneva, Switzerland, 2 Dept. of Ophthalmology, University Hospital Dresden, Dresden, Germany, 3 Dept. of Ophthalmology, Keck School of Medicine, University of Southern California, Los Angeles, California, United States of America

Abstract

Purpose: To introduce a constant-force technique for the analysis of corneal biomechanical changes induced after collagen cross-linking (CXL) that is better adapted to the natural loading in the eye than previous methods.

Methods: For the biomechanical testing, a total of 50 freshly enucleated eyes were obtained and subdivided in groups of 5 eyes each. A Zwicki-Line Testing Machine was used to analyze the strain of 11 mm long and 5 mm wide porcine corneal strips, with and without CXL. Before material testing, the corneal tissues were pre-stressed with 0.02 N until force stabilization. Standard strip extensiometry was performed as control technique. For the constant-force technique, tissue elongation (Δ strain, %) was analyzed for 180 seconds while different constant forces (0.25 N, 0.5 N, 1 N, 5 N) were applied.

Results: Using a constant force of 0.5 N, we observed a significant difference in Δ strain between 0.26 \pm 0.01% in controls and 0.12 \pm 0.03% in the CXL-treated group (p=0.003) over baseline. Similarly, using a constant force of 1 N, Δ strain was 0.31 \pm 0.03% in controls and 0.19 \pm 0.02% after CXL treatment (p=0.008). No significant differences were observed between CXL-treated groups and controls with 0.25 N or 5 N constant forces. Standard stress-strain extensiometry failed to show significant differences between CXL-treated groups and controls at all percentages of strains tested.

Conclusion: We propose a constant-force technique to measure corneal biomechanics in a more physiologic way. When compared to standard stress-strain extensiometry, the constant-force technique provides less variability and thus reaches significant results with a lower sample number.

Citation: Richoz O, Kling S, Zandi S, Hammer A, Spoerl E, et al. (2014) A Constant-Force Technique to Measure Corneal Biomechanical Changes after Collagen Cross-Linking. PLoS ONE 9(8): e105095. doi:10.1371/journal.pone.0105095

Editor: Craig Boote, Cardiff University, United Kingdom

Received March 7, 2014; Accepted July 18, 2014; Published August 27, 2014

Copyright: © 2014 Richoz et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Funding: This work was supported in part by a grant of the Swiss National Fonds and the Castier Foundation, Geneva, Switzerland. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing Interests: The authors have declared that no competing interests exist.

* Email: farhad@hafezi.ch

Introduction

Corneal biomechanics are of increasing importance in understanding the pathophysiology of corneal weakening, thinning, and irregularity in diseases such as keratoconus and postoperative ectasia. They also determine the outcome after refractive laser surgery. Corneal stiffness can be modified by corneal collagen cross-linking (CXL), a procedure that uses UV-A light and riboflavin to increase the mechanical and biochemical stability of corneal stroma [1–5]. CXL is effective in delaying or arresting the progression of keratoconus [1,6–10] and postoperative corneal ectasia [11–13] as demonstrated *in vitro and in vivo* when the limitations of the technique are respected [14,15].

Only recently CXL was transformed from a laboratory technique to a widely used clinical method [1,3,4,16]. As with all new technologies, continuous modifications and improvements of the original protocol are tested both experimentally [17–21] and clinically [13,22,23]. Initially, CXL has relied on *in vitro* biomechanical studies with stress-strain measurements that were adapted from standard material testing, using values in a non-physiological range that do not accurately measure biomechanical

changes that are relevant in the physiological and pathological biological tissue like the cornea [1,4,24].

The standard biomechanical measurement method (stress-strain extensiometry) analyzes the force necessary to induce a progressive strain in the corneal tissue [3,4] and serves to characterize the elastic material properties. It was adapted from methods for the mechanical analysis of metals and polymers with rather homogeneous chemical or molecular bonds. In contrast, the mechanical properties of a biological tissue depend on chemical bonds and molecular interactions that may be distributed inhomogeneously and lead to time-dependent, i.e. viscoelastic material properties. Also, the process of cross-linking introduces a variety of new chemical bonds [25], having an effect on the microstructural interactions. These modifications are difficult to measure accurately using conventional stress-strain methods.

Another limitation of standard biomechanical testing is that it is not tension-constant. In the natural environment corneas are hardly subjected to a steady increasing force (such as it is applied in stress-strain tests), it rather has to withstand the normal and constant intraocular pressure (IOP). In this study we introduce a method using a constant-force, which will allow us to analyze the

temporal and hence viscoelastic biomechanical properties of the cornea and their changes with CXL.

Methods

Corneal collagen cross-linking (CXL)

50 fresh enucleated porcine eves were obtained from a local abattoir (Abattoirs d'Orbe SA), stored at 5°C and prepared for the experiments within less than 6 hours after harvest. Only corneas with an intact epithelium, lack of focal stromal edema and a pachymetric thickness of 800±100 µm, as measured by ultrasound pachymetry (SP-2000, Tomey Corporation, Nagoya, Japan), were used. Debridement of the corneal epithelium was performed with a hockey blade. A solution containing 0.9% NaCl (no preservative agent) and 0.1% riboflavin (vitamin B2) was instilled onto the cornea every 2 minutes for 25 minutes. Corneas were then exposed to UV-A irradiation (CXL-365, Peschke Meditrade, Cham, Switzerland) with a large diameter aperture (11 mm), at 9 mW/cm² for 10 minutes at a working distance of 5 cm. After 5 minutes of irradiation, riboflavin drops were instilled once to minimize changes in corneal hydration. Controls were deepithelialized and received riboflavin instillation, but were not exposed to UV-A irradiation.

Biomechanical measurements

After CXL or control preparation, one corneo-scleral strip (5 mm width, full thickness) was obtained centrally in the horizontal axis from each eye. 4 mm of the end of each strip were dedicated to fixation, leaving no more than 11 mm of central corneal strip length, so that the entire central strip had been irradiated and cross-linked. For tensile strength measurement, we used a Zwicki-Line Testing Machine (Zwick, Ulm, Germany), calibrated with a distance accuracy of 2 µm and a tensile sensor with no more than 0.21% of measurement uncertainty between 0.25 N and 50 N. The Zwick Z 0.5 is a classical extensiometer composed of a linear holder extension arm whose speed can be controlled and a Newton meter, which measure the real time Force in Newton exerted by the arm on the held specimen. The conversion from force to stress is calculated from the thickness and width of the specimen. Corneal strips were fixed using pneumatics grips with 164 N. Data analysis was performed using the Xpert II-Testing Software for Static Testing Systems (Zwick, Ulm, Germany).

Before starting either conventional strip-extensiometry tests or constant-force measurements, the corneal tissue was pre-stressed with 0.02 N until force stabilization was achieved. This pre-stressing is equivalent to an IOP of about 6 mmHg.

For the new constant-force measurements (n control = 20 eyes, n CXL = 20 eyes), force was measured every 39 milliseconds. Strain was applied at a speed of 1 mm/min up to a constant force of 0.25 N, 0.5 N, 1 N, and 5 N (n = 5 eyes per force and group). These forces are equivalent to 58, 115, 231 and 1154 mmHg, respectively. Time started (T_0) when the pre-set force was reached. Then the amount of absolute strain was measured at 120 s and 300 s, corresponding to X_{120} and X_{300} , respectively (Figure 1) and Δ strain was determined within this time period (being 0% at 120 s).

Conventional strip-extensiometry (n(control) = 5 eyes, n(cxl) = 5 eyes) was then performed at a speed of 1 mm/min where force was recorded as a function of strain. The testXpert II software was used to calculate the Young's moduli (elasticity moduli) at 2, 4, 6, 8, and 10% of strain and to analyze the variance.

For both methods, the total testing time was less than 6 minutes per strip. During and after each test, all junctions between the

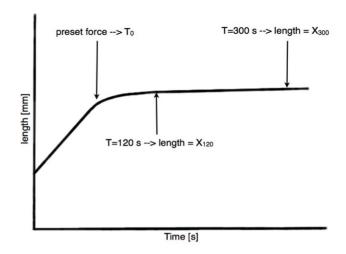


Figure 1. Schematic illustration of the constant-force method. T_0 was determined when the preset force was reached. At t=120 s and t=300 s, respectively, the distances X_{120} and X_{300} were measured. Comparisons were made between the induced strain during the time period from 120 s to 300 s. doi:10.1371/journal.pone.0105095.g001

pneumatic grips and the corneal strips were checked and if any movement or rupture between the strips and grips was suspected, the test was not used for the subsequent analyses.

Statistical Analysis

Statistical analysis was performed using testXpert II software (Zwick Roell Group, Ulm, Germany). Values were expressed as average \pm standard deviation (SD). Differences between the experimental groups were evaluated by the students t-test and considered statistically significant (*) or highly significant (**), when the probability value (P) was <0.05 or <0.01, respectively.

Results

Figure 2 depicts the elongation of cross-linked and control corneal strips over time under various applied forces (0.25 N, 0.5 N, 1 N, 5 N) using the constant-force technique. The corneal strip elongated (slope of the curve) until the point of stabilization (horizontal portion of the curve). After the pre-determined force was reached, elongation stabilized within less than 150 seconds. The time to reach stabilization as well as the amount of strain correlated positively with the applied force: stabilization ($R^2 = 0.79$) occurred more rapidly and strains ($R^2 = 0.87$) were lower in tests using a lesser force.

Using the constant-force technique, significant differences between CXL and control groups were observed (Figure 3), when a constant force of 0.5 N or 1 N was applied. Under a constant force of 0.5 N, untreated corneas elongated by 0.26 \pm 0.01% while CXL-treated corneas increased in length by 0.12 \pm 0.03% (p = 0.003). When 1 N of constant force was applied, CXL-treated corneas elongated by 0.19 \pm 0.02% and non-irradiated controls by 0.31 \pm 0.03% (p = 0.008). No significant differences were observed between CXL-treated groups and controls with 0.25 N or 5 N of applied force.

Figure 4, shows the different elasticity modulus of corneal strips with and without CXL treatment using the conventional stress-strain method. The absolute values for all elasticity moduli in the CXL-treated group were higher than in non-irradiated controls, but did not reach significance: The elasticity modulus at 6% strain

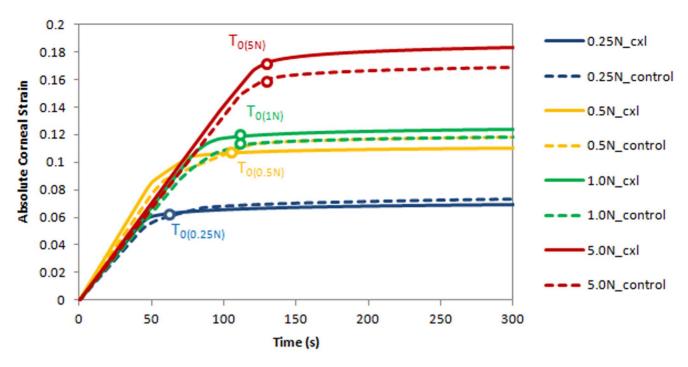


Figure 2. Absolute corneal strain as a function of time under different constant forces. Dashed lines represent control corneas, continuous lines cross-linked corneas. The higher the applied force, the later it was reached (T₀) and the larger strains were observed. doi:10.1371/journal.pone.0105095.g002

was 1.5 ± 0.21 MPa in the CXL group and 1.01 ± 0.06 MPa in non-irradiated controls (p=0.06). At 2% of strain, the elasticity modulus was nearly indistinguishable between the two groups, with 0.29 ± 0.09 MPa for the CXL group and 0.26 ± 0.05 MPa for the non-irradiated controls (p=0.78).

The measurement repeatability was similar in both techniques -0.0017 in the constant-force technique and 0.103 MPa in standard stress-strain extensiometry – which corresponds to a relative standard deviation of about 10%.

Discussion

CXL is currently widely used to strengthen the cornea in keratoconus, pellucid marginal degeneration and ectasia after refractive laser surgery. Systems (Ocular Response Analyzer, Corvis ST) based on analyzing corneal deformation following an air-puff have been used to evaluate the effect of CXL in vivo [10,26–28]. A major disadvantage of these techniques is that the recorded geometrical deformation parameters are strongly dependent on the IOP and corneal thickness and do not represent a real mechanical tissue properties. Further factors that impede an

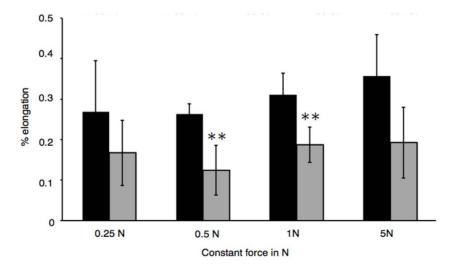


Figure 3. Biomechanical properties of corneal strips and significance from constant-force technique. T_0 was fixed at the beginning of the elongation, corresponding to X_{120} (t=120). Δ Strain under constant forces of 0.25, 0.5, 1 and 5 N (n=5 eyes per force) was determined after 180 seconds, corresponding to X_{300} (t=300). *P<0.05; **P<0.01. doi:10.1371/journal.pone.0105095.g003

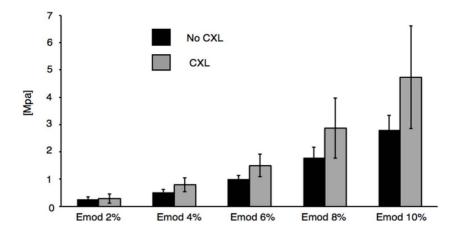


Figure 4. Biomechanical properties of corneal strips and significance from conventional testing. Young's modulus/elasticity modulus (emod %) at 2, 4, 6, 8, and 10% of strain in cross-linked corneas and untreated controls (MPa, n = 5 eyes for both, cxl and control). doi:10.1371/journal.pone.0105095.g004

accurate measurement of in vivo corneal biomechanics is the relatively high inter-individual variability due to age [29], smoking habits [30–32], and the hormonal status of estrogens [15,33,34] and thyroid hormones [35,36]. Ex vivo techniques are therefore still preferred when corneal stiffness needs to be quantified.

Since the surface/volume ratio of a corneal strip is high, rapid dehydration prior to testing was a concern [37]. We therefore performed all biomechanical tests in less than 6 minutes, minimizing tissue dehydration due to evaporation. To transfer the current constant-force technique to human cadaver eyes, forces will have to be reduced by approximately a third in order to account for the difference in thickness between the average porcine (800 μ m) and human cornea (550 μ m) [20,38].

The corneal stroma is mainly composed of collagen fibers and proteoglycans [39]. Recent studies suggest that the corneal stiffening after CXL seems to be due to the creation of additional covalent bonds between collagen fibers and proteoglycans [40,41].

Stress-strain extensiometry in corneas has been derived from elastic material testing, where increasing loads are applied, typically until material break. In the clinical setting however, corneal biomechanics are especially interesting under constant loading conditions. Here, we propose a method that uses constant forces (0.25 N to 5 N) to test biomechanical properties, which might better reflect the physiological loading condition due to the

IOP. The applied forces were chosen in a similar range than in previous stress-strain extensiometry, covering IOPs from 58 to 1154 mmHg. Using the constant force method significant changes after cross-linking were found in two of four forces, while standard stress-strain extensiometry could not find any difference. Therefore our results suggest that the constant force approach is probably more accurate and reliable in detecting differences after CXL than standard stress-strain testing as it addresses the viscoelastic material properties instead of purely elastic properties. Thereby the sensitivity of the constant force technique is force dependent: At low forces, creep behavior is small and depends mainly on the extracellular matrix. With increasing force, the load is more and more carried by the collagen fibers. As significant differences were only found for intermediate forces, we may conclude that cross-linking affects the interaction between collagen fibers and the extracellular matrix and not one of them alone.

Author Contributions

Conceived and designed the experiments: OR ES FH. Performed the experiments: OR SZ. Analyzed the data: OR SK SZ ES FH. Contributed reagents/materials/analysis tools: FH. Wrote the paper: OR SK ES AH FH.

References

- Wollensak G, Spoerl E, Seiler T (2003) Riboflavin/ultraviolet-a-induced collagen crosslinking for the treatment of keratoconus. Am J Ophthalmol 135: 620–627.
- Spoerl E, Wollensak G, Seiler T (2004) Increased resistance of crosslinked cornea against enzymatic digestion. Curr Eye Res 29: 35–40.
- 3. Spoerl E, Seiler T (1999) Techniques for stiffening the cornea. J Refract Surg 15: 711–713.
- Spoerl E, Huhle M, Seiler T (1998) Induction of cross-links in corneal tissue. Exp Eye Res 66: 97–103.
- Spoerl E, Huhle M, Kasper M, Seiler T (1997) [Increased rigidity of the cornea caused by intrastromal cross-linking]. Ophthalmologe 94: 902–906.
- Coskunseven E, Jankov MR 2nd, Hafezi F (2009) Contralateral eye study of corneal collagen cross-linking with riboflavin and UVA irradiation in patients with keratoconus. J Refract Surg 25: 371–376.
- Hashemi H, Seyedian MA, Miraftab M, Fotouhi A, Asgari S (2013) Corneal Collagen Cross-linking with Riboflavin and Ultraviolet A Irradiation for Keratoconus: Long-term Results. Ophthalmology.
- Hersh PS, Greenstein SA, Fry KL (2011) Corneal collagen crosslinking for keratoconus and corneal ectasia: One-year results. J Cataract Refract Surg 37: 149–160.

- Raiskup-Wolf F, Hoyer A, Spoerl E, Pillunat LE (2008) Collagen crosslinking with riboflavin and ultraviolet-A light in keratoconus: long-term results. J Cataract Refract Surg 34: 796–801.
- Vinciguerra P, Albe E, Mahmoud AM, Trazza S, Hafezi F, et al. (2010) Intraand Postoperative Variation in Ocular Response Analyzer Parameters in Keratoconic Eyes After Corneal Cross-Linking. J Refract Surg: 1–8.
- Hafezi F, Kanellopoulos J, Wiltfang R, Seiler T (2007) Corneal collagen crosslinking with riboflavin and ultraviolet A to treat induced keratectasia after laser in situ keratomileusis. J Cataract Refract Surg 33: 2035–2040.
- Kohlhaas M, Spoerl E, Speck A, Schilde T, Sandner D, et al. (2005) [A new treatment of keratectasia after LASIK by using collagen with riboflavin/UVA light cross-linking]. Klin Monatsbl Augenheilkd 222: 430–436.
- Richoz O, Mavrakanas N, Pajic B, Hafezi F (2013) Corneal Collagen Cross-Linking for Ectasia after LASIK and Photorefractive Keratectomy: Long-Term Results. Ophthalmology.
- Hafezi F (2011) Limitation of collagen cross-linking with hypoosmolar riboflavin solution: failure in an extremely thin cornea. Cornea 30: 917–919.
- Hafezi F, Iseli HP (2008) Pregnancy-related exacerbation of iatrogenic keratectasia despite corneal collagen crosslinking. J Cataract Refract Surg 34: 1219–1221.

- Wollensak G, Spoerl E, Seiler T (2003) Stress-strain measurements of human and porcine corneas after riboflavin-ultraviolet-A-induced cross-linking. J Cataract Refract Surg 29: 1780–1785.
- Schumacher S, Oeftiger L, Mrochen M (2011) Equivalence of biomechanical changes induced by rapid and standard corneal cross-linking, using riboflavin and ultraviolet radiation. Invest Ophthalmol Vis Sci 52: 9048–9052.
- Diakonis VF, Grentzelos MA, Tzatzarakis MN, Kankaria V, Karavitaki A, et al. (2012) Riboflavin's time-dependent degradation rate induced by ultraviolet A irradiation. Eur J Ophthalmol 22 Suppl 7: S51–56.
- Kamaev P, Friedman MD, Sherr E, Muller D (2012) Photochemical kinetics of corneal cross-linking with riboflavin. Invest Ophthalmol Vis Sci 53: 2360–2367.
- Vetter JM, Brueckner S, Tubic-Grozdanis M, Vossmerbaumer U, Pfeiffer N, et al. (2012) Modulation of central corneal thickness by various riboflavin eyedrop compositions in porcine corneas. J Cataract Refract Surg 38: 525–532.
- Wernli J, Schumacher S, Spoerl E, Mrochen M (2013) The efficacy of corneal cross-linking shows a sudden decrease with very high intensity UV light and short treatment time. Invest Ophthalmol Vis Sci 54: 1176–1180.
- Hafezi F, Mrochen M, Iseli HP, Seiler T (2009) Collagen crosslinking with ultraviolet-A and hypoosmolar riboflavin solution in thin corneas. J Cataract Refract Surg 35: 621–624.
- Chatzis N, Hafezi F (2012) Progression of Keratoconus and Efficacy of Corneal Collagen Cross-linking in Children and Adolescents. J Refract Surg 28: 753– 758
- Kohlhaas M, Spoerl E, Schilde T, Unger G, Wittig C, et al. (2006)
 Biomechanical evidence of the distribution of cross-links in corneas treated with riboflavin and ultraviolet A light. I Cataract Refract Surg 32: 279–283.
- Brummer G, Littlechild S, McCall S, Zhang Y, Conrad GW (2011) The role of nonenzymatic glycation and carbonyls in collagen cross-linking for the treatment of keratoconus. Invest Ophthalmol Vis Sci 52: 6363–6369.
- Luce DA (2005) Determining in vivo biomechanical properties of the cornea with an ocular response analyzer. Journal of cataract and refractive surgery 31: 156–169
- Noguera GE, Castro-Combs J, Taylor D, Behrens A (2007) Ocular Response Analyser Uses to Measure Corneal Biomechanics. Invest Ophthalmol Vis Sci 48: 1860

- Spoerl E, Terai N, Scholz F, Raiskup F, Pillunat LE (2011) Detection of biomechanical changes after corneal cross-linking using Ocular Response Analyzer software. J Refract Surg 27: 452–457.
- Robert L, Robert AM, Fulop T (2008) Rapid increase in human life expectancy: will it soon be limited by the aging of elastin? Biogerontology 9: 119–133.
- Hafezi F (2009) Smoking and corneal biomechanics. Ophthalmology 116: 2259 e2251.
- Hafezi F (2010) Tobacco smoking and its impact on corneal biomechanics. Invest Ophthalmol Vis Sci 51: 6892.
- Spoerl E, Raiskup-Wolf F, Kuhlisch E, Pillunat LE (2008) Cigarette smoking is negatively associated with keratoconus. J Refract Surg 24: S737–740.
- Hafezi F, Koller T, Derhartunian V, Seiler T (2012) Pregnancy may trigger late onset of keratectasia after LASIK. Journal of refractive surgery 28: 242–243.
- Hoogewoud F, Gatzioufas Z, Hafezi F (2013) Transitory topographical variations in keratoconus during pregnancy. J Refract Surg 29: 144–146.
- Gatzioufas Z, Thanos S (2008) Acute keratoconus induced by hypothyroxinemia during pregnancy. J Endocrinol Invest 31: 262–266.
- Kahan IL, Varsanyi-Nagy M, Toth M, Nadrai A (1990) The possible role of tear fluid thyroxine in keratoconus development. Exp Eye Res 50: 339–343.
- Borja D, Manns F, Lamar P, Rosen A, Fernandez V, et al. (2004) Preparation and hydration control of corneal tissue strips for experimental use. Cornea 23: 61–66.
- Doughty MJ, Zaman ML (2000) Human corneal thickness and its impact on intraocular pressure measures: a review and meta-analysis approach. Surv Ophthalmol 44: 367–408.
- Almubrad T, Akhtar S (2011) Structure of corneal layers, collagen fibrils, and proteoglycans of tree shrew cornea. Mol Vis 17: 2283–2291.
- Zhang Y, Mao X, Schwend T, Littlechild S, Conrad GW (2013) Resistance of corneal RFUVA-cross-linked collagens and small leucine-rich proteoglycans to degradation by matrix metalloproteinases. Invest Ophthalmol Vis Sci 54: 1014– 1095
- Zhang Y, Conrad AH, Conrad GW (2011) Effects of ultraviolet-A and riboflavin on the interaction of collagen and proteoglycans during corneal cross-linking. J Biol Chem 286: 13011–13022.