Finding Missing Heritability in Less Significant Loci and Allelic Heterogeneity: Genetic Variation in Human Height

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Abstract

Genome-wide association studies (GWAS) have identified many common variants associated with complex traits in human populations. Thus far, most reported variants have relatively small effects and explain only a small proportion of phenotypic variance, leading to the issues of ‘missing’ heritability and its explanation. Using height as an example, we examined two possible sources of missing heritability: first, variants with smaller effects whose associations with height failed to reach genome-wide significance and second, allelic heterogeneity due to the effects of multiple variants at a single locus. Using a novel analytical approach we examined allelic heterogeneity of height-associated loci selected from SNPs of different significance levels based on the summary data of the GIANT (stage 1) studies. In a sample of 1,304 individuals collected from an island population of the Adriatic coast of Croatia, we assessed the extent of height variance explained by incorporating the effects of less significant height loci and multiple effective SNPs at the same loci. Our results indicate that approximately half of the 118 loci that achieved stringent genome-wide significance (p-value < 5 × 10^{-8}) showed evidence of allelic heterogeneity. Additionally, including less significant loci (i.e., p-value < 5 × 10^{-6}) and accounting for effects of allelic heterogeneity substantially improved the variance explained in height.

Introduction

Genome-wide association studies (GWAS) have identified more than one thousand common variants associated with complex traits in human populations [1]. However, most identified variants confer relatively small effects, and in combination explain only a small fraction of phenotypic variance [2]. Using human height as an example, a classic quantitative trait where an estimated 80–90% of the normal variation is attributed to additive genetic factors [3–6], but the recently identified 180 SNPs by the GIANT (The Genetic Investigation of ANthropometric Traits) study account for only ~10% of the overall height variance [7]. The same study also revealed two possible sources of missing heritability. First, many common variants with small effects contribute to phenotypic variation, though the strengths of association of these variants do not achieve genome-wide significance (p-value < 5 × 10^{-8}). Second, multiple variants at a single locus may jointly influence a trait (i.e. allelic heterogeneity) and explain additional phenotypic variation.

The dissection of allelic heterogeneity is complicated by the correlation between SNPs due to linkage disequilibrium (LD), which often results in multiple SNP signals in any significant locus. Usually, only the lead SNP (the SNP with smallest p-value) of a locus is reported to represent the significant association. Some GWAS have conducted conditional association analyses to identify secondary signals associated with complex traits by accounting for the effects of lead SNPs [7–9]. As an alternative, we propose an analytical approach that estimates allelic effects and dissects allelic heterogeneity from GWA summary data (p-values, allele frequencies and sample size) rather than requiring individual-level data.

Using this method, we have examined allelic heterogeneity in height loci that include SNPs of different significance levels based on the GIANT summary data. To assess the extent to which additional variance may be explained by incorporating multiple effect SNPs at the same loci, we studied the variance explained in a sample of 1,304 individuals collected from an island population of the Adriatic Coast of Croatia. Our results indicated that a substantial fraction of height loci showed evidence of allelic heterogeneity and by including loci of lower significance and accounting for allelic heterogeneity, we were able to explain a considerably higher proportion of trait variance.


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Methods

Clustering significant height loci based on GIANT summary data

The height summary data from the Stage I meta-analysis of GIANT studies were downloaded from the GIANT consortium website (http://www.broadinstitute.org/collaboration/giant). The data file contains p-values, direction of effect, and number of observations at nearly 2.5 M (2,469,635) genotyped or imputed SNPs. Including SNPs with lower levels of significance may increase the variance explained [7], therefore we examined SNPs at six different levels of significance (a = 5 \times 10^{-6} to 5 \times 10^{-8}). SNPs meeting our significance criteria were clustered into distinct loci if they were physically adjacent to one another; different lengths (500 kb–50 kb) were used to define “physical adjacency” since lowering the significance level substantially increased the number of significant SNPs (Table 1).

Estimation of effect sizes and allelic heterogeneous effects

The GIANT summary data do not include effect size estimates (beta coefficients), therefore we first estimated the allelic effect (b) of each SNP using the following approximation function:

\[ b \approx \frac{z}{\sqrt{2p(1-p)n}} \]

where z is the z-score computed from reported p-values, and is either positive or negative according to the reported direction of effect. The reference allele frequency is denoted by p and n is the number of observations (sample size).

Part of the observed effect at a SNP j may be due to the impact of an adjacent lead SNP i with effect (b_i). Assuming an additive effect model, this part of apparent effect of SNP j (hereafter referred to as projected effect) can be written as:

\[ b_j^\ast = b_i \frac{D_{ij}}{p_i(1-p_i)} \]

where \( D_{ij} \) is the linkage disequilibrium coefficient between the two SNPs and \( p_i \) is the frequency of the reference allele SNP j. Thus, the allelic effect of SNP j conditioned on the primary SNP i can be calculated as the difference between the observed effect and the projected effect: \( b_j = b_j^\ast - b_j^\ast \) (hereafter referred to as conditional effect). Accordingly, the “conditional p-value” of SNP j can be approximated following equation:

\[ P_j \approx 2 \times \left[ 1 - \Phi\left( b_j \sqrt{2p_i(1-p_i)n} \right) \right] \]

where \( \Phi(.) \) is the standard normal cumulative distribution function. For each significant locus, we selected the most significant SNP as the primary SNP and estimated the conditional effects and p-values of all adjacent SNPs using the above functions. The SNP with the smallest conditional p-value was chosen as the secondary SNP if the conditional p-value was less than a Bonferroni-corrected significance level (0.05 divided by the number of SNPs at the locus). By applying this procedure iteratively (similar to stepwise conditioning), we also obtained conditional effect estimates of tertiary and quaternary SNPs, until conditional p-values were no longer significant. Since most GIANT study samples are European in origin, we used allele frequencies and linkage disequilibrium parameters calculated from haplotype data of HapMap Phase 2 CEU samples (release 24) [10].

Analysis of variance explained

To assess height variance explained, we used a study sample derived from a genetic study of metabolic syndrome in a relatively isolated population [11–14]. Briefly, study participants were recruited from Hvar, a middle Dalmatian island on the eastern Adriatic coast of Croatia. Blood samples and anthropometric data were collected in two field seasons of May 2007 and May 2008. For this study, we used the data on 1,304 individuals for whom genome-wide SNP data and height measurements were available. The study was approved by the Ethics Committee of the Institute for Anthropological Research in Zagreb, Croatia and the Institutional Review Board of the University of Cincinnati. Written informed consent was obtained from all participants.

Genome-wide SNP genotype data were obtained using the Affymetrix Human SNP Array 5.0 according to the manufacturer’s protocol. Genotype calls were determined using the CRIMM algorithm [15,16]. After QC filtering (MAF>0.02, HWE p-value>0.0001, call rate>95%), we performed genotype imputation using MACH [17] and the reference haplotype data from the Phase 2 CEU HapMap, yielding a final genotype data set of 2.5 million SNPs in 1,304 individuals (565 males and 739 females).

We estimated the fraction of variance explained using genetic scores that combine information from primary SNPs and conditional SNPs selected from significant loci. The weighted genetic score was constructed as \( S = \sum b_i G_i \) where \( G_i = 0, 1 \) or 2

<table>
<thead>
<tr>
<th>Table 1. Numbers of significant loci and conditional signals.</th>
</tr>
</thead>
<tbody>
<tr>
<td>Significance level</td>
</tr>
<tr>
<td>---------------------</td>
</tr>
<tr>
<td>5.E-08</td>
</tr>
<tr>
<td>5.E-07</td>
</tr>
<tr>
<td>5.E-06</td>
</tr>
<tr>
<td>5.E-05</td>
</tr>
<tr>
<td>5.E-04</td>
</tr>
<tr>
<td>5.E-03</td>
</tr>
</tbody>
</table>

*Lowering the significance level substantially increased the number (or the density) of significant SNPs used in clustering height loci. Therefore, shorter context lengths were arbitrarily selected in defining “physical adjacency” when relaxed significance levels were used, which might artificially reduce the length of significant loci and hence the chance of allelic heterogeneity in these loci clustered at lower significance level.

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indicates the number of reference alleles for a specific SNP and $b_i$ is the estimated allelic effect (for primary SNPs) or conditional effect (for secondary and tertiary SNPs). The variance explained ($r^2$) was then calculated based on a linear regression model using the constructed scores as predictor and age-, gender-, and their interaction term adjusted height residuals as outcome.

**Results**

To evaluate the validity of our proposed analytical procedure for estimating effect sizes and allelic heterogeneity from GIANT summary data, we first compared the concordance between our estimated effect sizes with the reported effect sizes listed in the GIANT meta-analysis report [7]. Of the 180 genome-wide significant SNPs, our estimated allelic effects correlated near perfectly ($r^2 = 0.98$) with their reported values (Figure 1A and Table S1). Of the 19 SNPs showing significant secondary signals listed in Table 1 of [7], the conditional allelic effects estimated using our proposed procedure correlated reasonably well ($r^2 = 0.71$) with their reported values (Figure 1B and Table S2). These results demonstrated that our proposed method accurately estimated effect sizes from the GIANT summary data (p-value, allele frequency and sample size). By incorporating LD information from HapMap (CEU), our procedure could also approximately dissect heterogeneous allelic effects (secondary or tertiary signals) of a significant locus.

Using this procedure we evaluated the allelic heterogeneity of significant loci that were associated with height at different significance levels (Table 1), based on the GIANT summary data. Approximately half of the 118 significant loci that achieved stringent genome-wide significance (with p-value at the primary SNP less than $5 \times 10^{-8}$) contained secondary signals (Figure 2A), and approximately one fourth had tertiary or quaternary signals after multiple rounds of conditioning (Figure 2B). All of the 18 regions with secondary signals reported by Lango Allen et al. [7] were recovered by our method (Table S3). A close examination of the significant loci with secondary signals revealed three additional features of the observed allelic heterogeneity. First, half of the identified secondary SNPs occurred within 200 kb from the primary SNPs (Figure S1). Second, we did not observe consistent LD between the primary and secondary SNPs ($\sim 10\%$ of secondary SNPs were in LD with primary SNPs with $r^2 > 0.1$), and third, there was no clear correlation between the estimated effect sizes between the primary and secondary SNPs (Figure S2).

As anticipated, relaxing the significance level substantially increased the number of significant loci. However, the percentage of loci with significant secondary or tertiary signals declined as the significance of the primary SNPs decreased (Table 1). For example, when the significance level was set at $5 \times 10^{-6}$, only 75 ($34.6\%$) of the 217 significant loci included significant secondary signals, which was smaller than 50.8% - the percentage of significant loci with secondary signals identified at more stringent significance level ($5 \times 10^{-8}$). This decline was mainly due to the shorter sizes of the significant loci clustered by SNPs with lesser statistical significance.

In our study samples, we assessed the extent of variance explained in age and gender adjusted height using genetic scores based on the estimated allelic effects of the clustered significant loci. Height was normally distributed in both males ($N = 565$) and females ($N = 739$) with larger variability observed in males (Figure S3). In Table 2, we listed the fractions of variance explained by various significant loci selected at different levels of significance, with or without secondary and tertiary SNPs. As demonstrated by Figure 3, two important patterns in the variance explained could be identified. First, the fraction of variance explained increased with relaxing the significance level, with the maximum around $5 \times 10^{-4}$. Second, additional proportion ($\sim 30\%$) of variance could be explained by including secondary and tertiary SNPs. Adding quaternary SNPs did not explain additional variance (data not shown). In our samples, the highest level of variance explained

![Figure 1. Correlation between reported and estimated effect sizes of the 180 primary height SNPs (A) and the 19 secondary SNPs (B) reported by Lango Allen et al. * The reference study did not report the effect sizes of the secondary signals. Here we used the values converted from the reported p-values based on conditional analyses in a subset of Stage 1 GIANT studies (Table 1 of Lango Allen et al). doi:10.1371/journal.pone.0051211.g001](image)
Figure 2. Two example loci with allelic heterogeneity. (A) The GHSR locus included a secondary signal (rs7652177) after accounting for the primary signal (rs572169). (B) The HMGA1 locus had a more complicated pattern of allelic heterogeneity; with significant secondary, tertiary and quaternary signals after multiple rounds of conditioning (only the first round of conditioning is shown). The secondary p-values (bottom plots) conditioning on the primary SNP were estimated from GIANT summary data using the analytical approach described in main text.

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Figure 3. Additional fraction of variance explained could be obtained by including less significant SNPs and secondary/tertiary SNPs.

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(13.8%) was achieved by the weighted-genetic score that included primary SNPs with p-value<5×10^{-4} as well as significant secondary and tertiary SNPs. The proportions of variance explain were constantly higher in females than in males (Table 2).

**Discussion**

GWAS have successfully established many robust associations between common variants and human complex traits, yet the extent of phenotypic variability explained by these variants remains disappointingly low. In this study, we examined two possible sources of missing heritability: first, variants associated with height at non-GWA significance, and second multiple variants at a single locus jointly influencing a trait.

Based on the summary data of the GIANT (stage 1) studies, we clustered significant SNPs selected at different significance levels into distinct significant loci and dissected the allelic effects of SNPs at each locus using a novel analytical approach that accounts for the LD between pair of SNPs. Compared to the reported results [7], our proposed method accurately estimated the effect sizes of primary SNPs and efficiently detected allelic heterogeneity in many significant loci. We observed evidence of allelic heterogeneity in roughly half of the significant loci (60 out of 118) that reached stringent genome wide significance (p-value<5×10^{-8}). All of the 18 regions with second signals reported by Lango Allen et al. were recovered in our list. The larger fraction of loci with allelic heterogeneity recovered by our method (~50% vs ~10% by Lango Allen et al) may be due to the less stringent significance cut off for secondary signals used in the current study. We controlled multiple testing for each locus independently by a Bonferroni-corrected significance level (0.05 divided by the number of SNPs at the locus). Whereas, Lango Allen et al. used a more stringent significance level (3.3×10^{-8}) for all of the 180 significant loci simultaneously.

A recent paper by Yang et al. [18] reported a conditional and joint association analysis of GWAS summary-level statistics and using the GIANT summary data identified 36 loci with multiple associated variants for height. Although targeting the same analytical problem, our method differs from their approach. Yang et al. used a multivariate approach to model the joint effects of multiple SNPs simultaneously and estimated the conditional effect iteratively over all the SNPs across the whole genome. Our method is much simpler by only considering the LD between pair of SNPs once at a time and detects secondary, tertiary SNPs in a stepwise manner within each significant locus. Second, they employed a stringent genome-wide significance level (5×10^{-8}) aiming to robustly establish the significant association of the identified SNPs. While our objective was to assess the proportion of “missing heritability” that could be explained by allelic heterogeneity and therefore, we controlled multiple testing for each locus separately. Nonetheless, 33 of the 36 loci reported by Yang et al. were included in our top list of 60 loci with multiple associated SNPs (Table S3). The concordance of these results again demonstrated the validity of our analytical method.

As demonstrated by Figure 2B, the allelic heterogeneity for some significant loci might involve more than two effect variants and span several million base pairs in length, covering multiple genes. Even in the simple example (Figure 2A), the secondary SNP (rs7652177) is located in a different gene (FNDC3B) than the primary SNP (a synonymous SNP in GHSR). In addition, rs7652177 is a non-synonymous SNP and might have functional consequence. This complex pattern reflects the complexity of allelic heterogeneity in complex phenotypes, which may go beyond the traditional perception of allelic heterogeneity for Mendelian traits as “different mutations within a single gene locus cause the same disorder”, in which a “gene” is usually interpreted as a functional gene (e.g. protein-coding gene) with relatively clear structural boundaries. This complex pattern of allelic heterogeneity also suggests that the search for causal variants in a significant locus would require comprehensive examination of a broader region that extends beyond individual gene with plausible functional relevance. In addition, the primary and secondary SNPs of a single locus were likely to cluster together, usually within several hundred kb. However, there was no obvious LD or correlation of effect sizes between these SNPs, which suggested that, although physically adjacent, multiple SNPs might confer their effects to a polygenic trait in a relatively independent manner.

The 118 primary SNPs that reached genome-wide significance (p-value<5×10^{-8}) explained 6.6% height variance and the 217 primary SNPs with suggestive genome-wide significance (p-value<5×10^{-6}) explained 7.7% variance. These results agreed closely with the proportion of variance explained in the same study samples [19] using the 180 lead SNPs identified by the joint analysis of Stage I+II GIANT studies. Consistent with previous findings [7], including SNPs with less significant p-values increased the variance explained in age- and gender- adjusted height (Table 2 and Figure 3). The highest level of variance explained was observed when the significance level was set between (5×10^{-4} and 5×10^{-3}). This observation suggests that an appreciable fraction of SNPs far from genome-wide significance might have small but genuine effects, and including these SNPs could substantially increase the variance explained. In addition, incorporating the

**Table 2. Fraction of height variance explained.**

<table>
<thead>
<tr>
<th>Significance level</th>
<th>Number of SNPs</th>
<th>All (N = 1304)</th>
<th>Female (N = 739)</th>
<th>Male (N = 565)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>(1st)</td>
<td>(2nd)</td>
<td>(3rd)</td>
<td>(1st)</td>
</tr>
<tr>
<td>S.E.08</td>
<td>118</td>
<td>178</td>
<td>204</td>
<td>0.066</td>
</tr>
<tr>
<td>S.E.07</td>
<td>151</td>
<td>221</td>
<td>252</td>
<td>0.071</td>
</tr>
<tr>
<td>S.E.06</td>
<td>217</td>
<td>292</td>
<td>323</td>
<td>0.077</td>
</tr>
<tr>
<td>S.E.05</td>
<td>354</td>
<td>441</td>
<td>475</td>
<td>0.072</td>
</tr>
<tr>
<td>S.E.04</td>
<td>781</td>
<td>873</td>
<td>897</td>
<td>0.113</td>
</tr>
<tr>
<td>S.E.03</td>
<td>2668</td>
<td>2716</td>
<td>2726</td>
<td>0.116</td>
</tr>
</tbody>
</table>

The number of SNPs used in constructing the genetic score. (1st) primary SNPs only; (2nd) primary+secondary SNPs; and (3rd) primary+secondary+tertiary SNPs.

#The highest level of variance explained was achieved by including less significant SNPs plus significant secondary and tertiary SNPs.

*The highest level of variance explained was achieved by including less significant SNPs plus significant secondary and tertiary SNPs.

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secondary or tertiary SNPs resulted an average increase of ~30% in height variance explained (Figure 3). This percentage is consistent with a recent report of allelic heterogeneity in cis-expression quantitative traits [20]. Taken together, including less significant SNPs as well as secondary and tertiary SNPs yielded the highest level of variance explained (3.8%) observed in our samples. Another interesting observation was the fraction of variance explained in female samples were consistently higher (~50%) than in male samples (Table 2). This difference might partially be due to the larger variance in age- and gender- adjusted height in male (SD = 6.9 cm) versus the female samples (SD = 6.0 cm), or it might indicate intrinsic gender differences in effect sizes of significant SNPs.

Our study has several limitations. First, we estimated the effect sizes and allelic heterogeneity from summary data. The estimation procedure is approximate and depends largely on the assumption of additivity, including both additive allelic effects and between-SNP additive effects, although this assumption is generally supported by theoretical [21] and empirical data [7]. Second, our estimation method can not explicitly distinguish genuine allelic heterogeneity from multiple SNPs in partial LD with a functional variant – the apparently “independent” effects might be projected from a hidden functional variant. Third, because there is no consensus definition of “significant loci”, arbitrarily selected lengths were used to cluster significant SNPs, which could break down a continuous significant locus into small pieces if the length definition was short (i.e. <100 kb). Given these limitations, our analytical approach can only be regarded as a rough evaluation of the allelic effects of significant height-associated loci. Although we have not tested the accuracy of our method, its validity is supported by the increased variance explained in our independent cohort when integrating the estimated allelic heterogeneity and the close agreement between our results and those reported by Lango Allen et al. [7] and Yang et al [18] (Table S3). The detailed dissection of allelic heterogeneity will require deep sequencing of the significant loci to identify the real functional variants.

In summary, we have investigated allelic heterogeneity of height-associated loci using an analytical approximation approach. Our results demonstrated that a substantial fraction of significant loci showed evidence of allelic heterogeneity and a significant proportion may involve more than two effective SNPs. We also examined the extent of height variance explained by the genetic scores constructed based on the identified significant primary and secondary/tertiary SNPs in a sample collected from an isolated eastern European population. We confirmed that including loci with lower significance levels and accounting for multiple variants at a locus considerably increased the variance explained. We anticipate that further analyses of allelic heterogeneity using sequencing technology and more accurate estimation of allelic effects through an elaborated analytical model will lead identification of additional variants with independent effects, and in turn increase the proportion of variance explained.

### Supporting Information

**Figure S1** Distribution of distance between the primary and secondary SNPs. (TIF)

**Figure S2** Correlation between secondary effect and primary effect. (TIF)

**Figure S3** Distribution of height in females (N = 739) and males (N = 565). (TIF)

**Table S1** Effect sizes of the 180 height-associated SNPs reported by Lango Allen et al. [Nature 467 (7317): 832–8](PDF)

**Table S2** Effect sizes and p-values of the 19 secondary signals reported by Lango Allen et al. [Nature 467 (7317): 832–8](PDF)

**Table S3** The 60 significant loci with secondary signals (ordered by p-value of the secondary signal). (PDF)

### Acknowledgments

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### Author Contributions

Conceived and designed the experiments: GZ RD PR RC. Performed the experiments: GS SRI HC. Analyzed the data: GZ. Contributed reagents/materials/analysis tools: DH NN ZD SM PR. Wrote the paper: GZ RD RK RC.

### References