HSPG-Deficient Zebrafish Uncovers Dental Aspect of Multiple Osteochondromas

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Abstract

Multiple Osteochondromas (MO; previously known as multiple hereditary exostosis) is an autosomal dominant genetic condition that is characterized by the formation of cartilaginous bone tumours (osteochondromas) at multiple sites in the skeleton, secondary bursa formation and impingement of nerves, tendons and vessels, bone curving, and short stature. MO is also known to be associated with arthritis, general pain, scarring and occasional malignant transformation of osteochondroma into secondary peripheral chondrosarcoma. MO patients present additional complaints but the relevance of those in relation to the syndromal background needs validation. Mutations in two enzymes that are required during heparan sulphate synthesis (EXT1 or EXT2) are known to cause MO. Previously, we have used zebrafish which harbour mutations in ext2 as a model for MO and shown that ext2+/− fish have skeletal defects that resemble those seen in osteochondromas. Here we analyse dental defects present in ext2−/− fish. Histological analysis reveals that ext2−/− fish have very severe defects associated with the formation and the morphology of teeth. At 5 days post fertilization 100% of ext2−/− fish have a single tooth at the end of the 5th pharyngeal arch, whereas wild-type fish develop three teeth, located in the middle of the pharyngeal arch. ext2−/− teeth have abnormal morphology (they were shorter and thicker than in the WT) and patchy ossification at the tooth base. Deformities such as split crowns and enamel lesions were found in 20% of ext2−/− adults. The tooth morphology in ext2−/− was partially rescued by FGF8 administered locally (bead implants). Our findings from zebrafish model were validated in a dental survey that was conducted with assistance of the MHE Research Foundation. The presence of the malformed and/or displaced teeth with abnormal enamel was declared by half of the respondents indicating that MO might indeed be also associated with dental problems.

Introduction

Multiple osteochondromas (MO), previously known as hereditary multiple exostosis (HME), is a genetic dominant syndrome occurring at the frequency of 1:50,000 [1] that is caused by mutation of one of the two EXOSTOSIN (EXT) genes, EXT1 or EXT2 [1,2]. These two EXTs encode glycosyltransferases that are crucial for the polymerisation of heparan sulphate (HS). MO manifests itself during the first two decades of life by the development of benign bone tumours (osteochondromas) at multiple sites of the skeleton near growth plates (cartilage structures responsible for elongation of long bones). Osteochondromas predominately form around the knees and elbows, but all other enchondral bones can be affected. Growing tumours can press on muscles or nerves and thereby cause pain and discomfort. The associations of MO with general pain [3], autism [4], arthritis [5] and scarring [6] have also been reported. MO patients present additional complaints but the relevance of those in the relation to the syndrome requires validation.

Previously, using a zebrafish (Danio rerio) mutant called dackel (dak) which has a premature stop codon in ext2, we established a model for MO [7]. Whereas mice lacking Ext1 or Ext2 fail to gastrulate, ext2−/− fish complete gastrulation and show a strong skeletal phenotype. Although ext2 has a very broad expression in zebrafish larvae [8], ext2−/− fish have very specific and consistent phenotypes. We have shown that the cartilage phenotype in ext2−/− fish resembles that seen in osteochondromas providing evidence that lack of HS proteoglycans (HSPG) affects cartilage morphogenesis, without influencing early cartilage differentiation [7]. We also found that mutation in ext genes have a negative effect osteoblast differentiation. In addition we determined that ext2−/− cells behave autonomously, providing evidence that osteochondroma in humans form as a result of a loss of heterozygosity at an EXT locus.

HSPGs are present on membranes and/or in the extracellular matrix in most animal tissues. HSPGs were shown to have very specific spatio-temporal localisation in mice teeth [9]. Similar HSPGs pattern should also be present in human teeth suggesting that alteration of the level or the expression pattern of HS might play a role in dental defects. Indeed, abnormally high levels of HS are known to cause oral defects in patients with mucopolysaccharidoses (MPS), a group of diseases caused by mutations in genes
that degrade HS [10]. The presence of dental pathologies was reported by several MO patients, but up to date there is only one case report describing dental abnormalities in a patient with MO and severe vitamin D deficiencies [11]. In order to explore the potential consequences of reduced HS in MO patients we studied the tooth phenotype in dlk hetero- and homozygote mutants.

Zebrafish teeth are located on the most posterior (5th) ceratobranchial arch. The first developing teeth can be observed at 48 hours post fertilisation (hpf). This is the pair of 4V1 teeth that form at 48 hours post fertilisation (hpf). This is the pair of 4V1 teeth that were always found to be positioned at the end of the 5th arch (Figure 1B), whereas WT-teeth were located more centrally (Figure 1A). In all analysed cases (n>20), ext2-/- teeth were significantly shorter than control teeth (Figure 1C). Width-wise, ext2-/- tooth was also significantly larger than WT teeth (Figure 1D). Distal ossification was observed at 4 dpf in ext2-/- 5th pharyngeal arch and tooth (Figure S2). However, tooth attachment occurred on time (Figure 2B). At 6 dpf, in 90% of ext2-/- teeth (n=20), patches of non-ossified areas were found at the base of teeth (Figure 1B).

Results

ext2-/- larvae display a severe tooth phenotype

In addition to the previously described cartilage and bone defects in ext2-/- larvae [7], we now demonstrate that tooth number, development and morphology are also severely affected (Figure 1 and 2). In wild type zebrafish, by 6 dpf, three ossified teeth (3V1, 4V1 and 5V1) attached to each bilateral 5th pharyngeal arch can be visualised by Alizarin red stain (Figure 1A). All of these teeth have pointed tip and are fully ossified. During the first week of development, ext2-/- teeth were indistinguishable from WT, whereas in ext2-/- only one tooth (V1) was formed and attached to each 5th arch (Figure 1). Furthermore, ext2-/- teeth were always found to be positioned at the end of the 5th arch (Figure 1B), whereas WT-teeth were located more centrally (Figure 1A). In all analysed cases (n>20), ext2-/- teeth were significantly shorter than control teeth (Figure 1C). Width-wise, ext2-/- tooth was also significantly larger than WT teeth (Figure 1D). Distal ossification was observed at 4 dpf in ext2-/- 5th pharyngeal arch and tooth (Figure S2). However, tooth attachment occurred on time (Figure 2B). At 6 dpf, in 90% of ext2-/- teeth (n=20), patches of non-ossified areas were found at the base of teeth (Figure 1B).

To further characterise ext2-/- tooth phenotype we examined tooth development using a set of molecular markers that were described previously in zebrafish [7,12,14,15]. The expression pattern of dlx2a, dlx2b, cx43 and osterix confirmed the presence of dental defects in the ext2-/- mutant (Figures 2 and 3). Transcripts of all tested dental markers were detected at 56 hpf in the 4V1 teeth in WT and in ext2-/- larvae. ext2-/- mutant expressed dlx2a, dlx2b and cx43 in one domain at 56 hpf, but the expression level was very weak (Table 1; Figure 3). By 72 hpf transcripts of dlx2, cx43 and osterix were detected in all WT and dak fish. However, while WT and dak siblings clearly expressed dlx2 and cx43 at position 4V1 and osterix at the positions 3V1 and 5V1, ext2-/- expressed all markers in one domain (Table 1; Figure 3). By 96 hpf, in all larvae, strong expression of cx43 and osterix were detected in two loci corresponding to 3V1 and 5V1 in WT and siblings, in one locus in ext2-/- fish (Table 2 and Figure 3). Transcript of dlx2b was detected in all WT and dak siblings but not in the homozygote mutant fish. The expression of dlx2a was not detected at 96 hpf in any of the fish. We have also analyzed the expression pattern of dental markers in the homozygote pinscher (pis) mutant. This strain has a mutation in the slc35b2 gene and there why lacks sulphation of various molecules including heparan sulphate [7]. slc35b2-/- larvae have a similar cartilage phenotype and forms two teeth out of which only one ossifies. We found all markers expressed at 56 and 72 hpf in one locus (Table 1, Figure 3). At later time-point, also weak expression in one locus was detected for cx43 and osterix.

As the tooth development depends on the interplay between epithelial and mesenchymal cells we have also investigated the expression pattern of the pitx2 gene that is known to be specifically expressed in the zebrafish pharyngeal epithelium [13]. In WT and dak mutants at 56 hpf signal was detected in one bilateral domain (Figure S3). The domain in ext2-/- has similar length to this from siblings, but the patches of pharyngeal epithelium which strongly expressed pitx2 had width restricted to one cell layer whereas in siblings minimum two cell width was observed (Figure S3).

Which pathways are impaired in HS-deficient teeth?

In order to find out which genetic pathway(s) might be affected in ext2-/- teeth we screened through known developmental mutants for those that affect tooth number. Out of 18 homozygous mutants, you too (yougl2a) and heart and soul (has/prkci) develop only a single tooth; acerebellar (acr/fgfb) and boxer (box/ext2s) had 1 to 3 teeth whereas pinscher (pis/slc35b2) had two teeth out of which only one ossifies (Table 2, Figure 4). In order to verify whether impairment of FGK, PKC and HH signalling indeed affects tooth formation, we have exposed fish to chemical inhibitors of different pathways: SU5402 (FGK), cycloamine (HH), G69876 (PKC), G69876 (PKC) and Bimi (PKC). With exception for Bimi that did not cause any visible changes, all other treatments affected tooth development.
formation. Fewer teeth were formed in WT, hetero- and homozygote 
mutant fish (Figure S4 for PKC, Figure S5 for FGF, data not shown for IHH). This finding indicated that FGF, IHH and PKC pathways may interact with HS during tooth development. Hence, we decided to test whether activation of those pathways could rescue the ext2+/2-tooth phenotype.

Treatments with purmorphamine (activator of IHH) or PMA (PKC activator) did not have any obvious effect on teeth in the ext2+/2 mutant and its siblings. An example of pharyngeal arches from fish treated with PKC activator is shown in Figure S4. For activation of FGF signalling, we implanted beads soaked in FGF8 into an area in which later forms teeth. Importantly, we have found that local exposure to FGF8 could stimulate tooth-bud development and increase ossification of the 5th arch in mutant larvae (Figure 5).

Tooth phenotype in ext2+/− adult fish

Since MO patients are heterozygous for mutations in Ext1 or Ext2, we were interested to know if ext2+/− mutant fish also display tooth defects. No defects were observed in dak (+/−) larvae up to 6 dpf (data not shown). A number of tooth pathologies were found in adult fish, and those changes were five times more frequent in heterozygote mutant fish than in the wild type siblings (Figures 6 and 7). As zebrafish teeth are being replaced throughout life, only teeth that were attached to the pharyngeal arch were analyzed.

There was no significant difference in the number of teeth number found in WT and mutant fish. However, we have observed changes in the distribution of the teeth (Figure 6). Normally, adult teeth are organised into three rows of teeth: five on the ventral side (1–5V), four in the middle (1–4MD) and two teeth on the dorsal side (1–2D) [16]. In the ext2+/− mutant, we have found a significant reduction in the number of dorsal teeth, whereas mediodorsal row was unaffected (Figure 6A–C). Interestingly, we have also observed the occurrence of a super-numeral tooth in the ventral row of the ext2+/− fish (Figure 6C, E, E′) and abnormal gaps between teeth (Figure 6F, F′). Incomplete enamel and split crowns were occasionally found in the ext2+/− fish, but not in their WT-siblings (Figure 7B–F). Cross section of teeth from adult fish did not show any notable difference in the morphology of ext2+/− teeth at the microscopic level (Figure S6).

Does MO affect human teeth?

In order to verify our findings on dental defects in zebrafish model, we have designed a questionnaire (Text S1) in which MO patients and their families were asked to provide information related to their condition and oral health. We have received 23 responses from 22 MO patients and one from a family member that was not diagnosed with MO. 32% of MO patients were genetically tested, out of whom only 3 knew their mutation status and shared this with us (one person had mutation in EXT1, one in

Figure 1. ext2+/− mutant displays severe tooth phenotype. Ventral view of alizarin-red-stained craniofacial skeleton and teeth at 6 dpf (A, B) and dissected and flat mounted 5th pharyngeal arches with teeth (A′, B′) reveals the presence on each pharyngeal arch of 3 teeth in siblings (A, A′) and only one misshapen tooth in ext2+/− larvae (B, B′). Note that the rod shaped branchial arch 5 to which the teeth attach is also ossified. Arrows point incomplete ossification of the mutant tooth. Tooth phenotype consisting of one misshapen tooth was observed in all (n>500) analysed ext2+/− embryos whereas heterozygote fish were indistinguishable from WT. Tooth lengths varies between 3V1, 4V1 and 5V1 in siblings (P<0.003). Each of those teeth was significantly longer then dak-tooth (P<0.0001) (C). Tooth widths of 3V1 and 5V1 were similar between siblings, and both were significantly narrower than 4V1 (D). ext2+/−-tooth was significantly broader than any of the siblings teeth (3V1, P<0.0001; 4V1, P = 0.023 and 5V1, P = 0.0001) (D). White boxes, siblings; grey boxes, homozygote mutant. Scale bar = 0.1 mm.

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the first to develop and attach – the 4V1 tooth. Note that single tooth germs of 3V1 and 5V1, but lost in the 4V1. Mineralised dentine outlines underlines the pharyngeal arches in sibling (A, A)

Discussion

MO is known as a genetic syndrome manifesting by the formation of multiple osteochondromas caused by the inactivation of EXT1 or EXT2 [17]. All other problems associated with MO, such as bone bowing, short stature or pain are considered as secondary defects caused by growing tumours. However, knowing the importance of HS one could expect that general skeletal, neurological, vascular and other changes would be common in MO. We became interested in the dental issues based upon discussions with members of the MHE Research Foundation (http://mheresearchfoundation.org/HOME.html) and decided to take advantage of the zebrafish model to analyse how reduced HS affects tooth health and development. We used homozygous mutant fish that have a very strong phenotype. Although in this setting, even more subtle changes are easy to be noticed, the results most likely overemphasise developmental changes caused by the absence of heparan sulphate that will not be observed in patients. This is why, we also analysed adult heterozygous fish. Even though results obtained from zebrafish studies are not directly applicable to patient data, they do give an indication of areas of interest for further study.

Justification of the zebrafish model for dental studies

Although zebrafish teeth differ from human teeth, they do have a similar organisation (Figure S7). In both cases tooth crowns are made of dentin that is covered with a protective layer of enameloid (fish) or enamel (human); dental pulp that occupies central part of tooth consists of odontoblasts [13,18]. The pulp cavity contains blood vessels and nerves, but in case of zebrafish this is only true for adult and not for larval teeth [19]. Furthermore, in contrast to human teeth that have roots, the presence of cementum was not described in zebrafish. This might be due to the fact that zebrafish are polyphyodont and hence they do not have permanent teeth as mammals do.

Why does only one tooth forms in dak homozygote mutants and what is its identity?

Comparing the expression patterns of various dental markers in the ext2−/− mutant that has one tooth, slc35b2−/− mutant that has 2 teeth and WT with three teeth, 5V1 tooth as a possible identity of the single ext2−/− tooth can be ruled out as no expression of dental markers was seen in any of the mutants. However, our data are no conclusive for other tooth identities. It might be that the tooth is 4V1 that is delayed or 3V1 that develops faster. Alternatively, ext2−/− tooth might result from V1 3 and 4 being fused together. In mammals, tooth development depends on the interplay between epithelium that originates from ectoderm and neural-crest-derived mesenchyme. Despite the disagreement whether zebrafish epithelium originates for ectoderm or endoderm, tooth development in ext2−/− might be caused by partial loss of contact between epithelium and mesenchyme. In fact, we have found that pitx2 expression domains in the pharyngeal epithelium do not undergo thickening in ext2−/− (Figure S5). In contrast to the rod-like shape of the pharyngeal cartilages in WT, ext2−/− pharyngeal are shorter and thicker [7,20]. Moreover, abnormal accumulation of cells undergoing apoptosis was noted at the lateral end of the ext2−/− pharyngeal (Figure S6). Hence, it is possible that 5th arch is not long enough to interact with the lateral part of epithelium i.e. the region where 5V1, and possibly 4V1, should be formed. This hypothesis could be strengthened by the fact that ext2−/− tooth is formed at the very distal end whereas the WT-teeth are always located to the middle of the pharyngeal arch. Furthermore, in the slc35b−/− mutant, a similar tooth loss coincides with shortening of the pharyngeal arch (Figure 4). Thus, defects in craniofacial morphogenesis may explain some aspects of the tooth phenotype.

The partial gaps in the ossification of the ext2−/− tooth at 6 dpf could indicate signs of natural events prior to tooth replacement. However, since ext2−/− tooth attaches on time and patches of non-ossified areas were also found in ext2−/− teeth already at 4 dpf, hence it is more likely that incomplete ossification reflect pathological changes in ext2−/− teeth. Given the osteoblast differentiation is reduced in ext2−/− fish [7], it is possible that also similar defects affect odontoblast and this incomplete ossification results from diminished odontoblast numbers.

Why tooth phenotype is observed only in every fifth heterozygote dak mutant?

In adult zebrafish, dental changes were observed only in 20% of fish that are known to carry a mutation in the ext2 gene. Which
raises the question why the remaining 80% of fish heterozygous for a mutation in the ext2 gene remain asymptomatic? Considering the huge variability of the bone phenotype in MO, involvement of other gene(s) cannot be excluded. However, the occurrence of the loss of heterozygosity (LOH) in tumours was confirmed in a number of solitary and multiple osteochondromas [21–23], which suggests that dental defects might take place as a result of a second hit mutation. Hence, if dental defects arise from a LOH, they would be sporadic and heterogeneous in nature. Moreover, the dental changes that we found in the ext2 heterozygous mutant fish were not specific. Similar changes could be observed in wild type fish, although at lower frequency. This is why they may have been previously overlooked.

Validation of the finding from the zebrafish model for MO

Mouse models have also been established for MO and used for studies on the formation of tumours [24–28] and synovial joints [29]. Although none of the authors characterised teeth of the Ext1 or Ext2-mutant mice, dental defect were described in Ndst1-null mice [30]. NDST1 is an enzyme that acts downstream of EXTs in the biosynthesis of heparan sulphate.

Online surveys might be biased towards a group having an active interest in given topic. Although, two-thirds of our respondents reported the presence of dental problems, the results of our dental survey still should be used as a guideline only. As survey was returned by one non-MO respondent only, we used general national statistic as an indicator of the frequencies of dental defects in normal population. According to a survey carried in the UK in 1998, adult dentate has on average 24.8 teeth with 15.3% of sound and untreated teeth (Office for National Statistics, UK). Although the number of teeth in adult human population varies depending on age, gender and country, in comparison to the UK population, our MO patient do not present any deviation from the norm, neither in the total number of teeth nor the number of untreated and good teeth. This is in line with our observation from zebrafish heterozygote adults carrying mutation in the ext2 gene that showed no significant difference in the tooth number as compared to WT siblings. The presence of misplaced and/or malformed teeth found from this zebrafish study was also indicated by 45% of the MO patients. Every third MO patient stated that was told by a dentist that they have abnormal (“too thin”) enamel. Unfortunately, technical limitations did not allow us to test enamel thickness in our zebrafish model.

Figure 3. The expression pattern of the dental markers at three stages of development. A panel of markers was used to dissect the identity of the ext2 mutant fish (dak). Progress of tooth development was monitored at 56, 72 and 96 hpf using dlx2a, dlx2b and cx43 markers. slc35b2-/- (pic) mutant was included as it has 2 teeth located to the end of pharyngeal arch. Ossification of teeth in slc35b2-/- is also delayed and progresses only in one tooth. ext2-/- and slc35b2-/- mutants were sorted by phenotype. A, ventral view and B, dorsal view. Scale bar = 0.1 mm.

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In addition to tooth defects observed in 100% of homozygote mutant larvae and in 20% of heterozygote adult fish, the dental questionnaire brought a high percentage of MO patients having problems with bleeding gums and gingivitis to light. However, as the occurrence of bleeding gums and/or gum inflammation are very frequent in normal population and the number of respondents were low (n = 23) we can not conclude if there are any significant differences between those two groups. It should be noted, that dental defects observed in MO patients do not look specific, and hence a special screening programme would be needed in order to tell whether tooth lesions, plaques on teeth and gingivitis are due to the improper dental hygiene or indeed due to MO.

How many people might be affected?
The frequency of MO occurrence was estimated at 1:50000. However since a) some of the osteochondromas are asymptomatic and b) genotyping or full body x-rays or MRI are not routinely

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Results from this study (a), were juxtaposed to information available from: Ablooglu et al, 2007 [14], (b); Jackman et al, 2004 [15], (c); and Borday-Birraux et al.,2006 [12], (d). 4V1 is the first tooth formed. The development of 4V1 is closely followed by the formation of neighbouring teeth. The development of 3V1 on the medial side of 4V1 slightly precedes the formation of 5V1 on the lateral side of 4V1. The expression pattern of dix2a, dix2b, connexin 43 and osterix does not clarify the identity of ext2+/− tooth. However, comparison of the expression patterns in ext2+/− and slc35b2−/− indicate loss of 5V1 in both mutants. Week expression is indicated by brackets; -, gene expression was not detected; na, not analysed.

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### Table 1. Summary of the expression pattern of dental markers during early tooth development.

<table>
<thead>
<tr>
<th></th>
<th>WT 56 hpf</th>
<th>WT 72 hpf</th>
<th>WT 96 hpf</th>
<th>dak 56 hpf</th>
<th>dak 72 hpf</th>
<th>dak 96 hpf</th>
<th>pic 56 hpf</th>
<th>pic 72 hpf</th>
<th>pic 96 hpf</th>
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<tr>
<td>cx43</td>
<td>3V1 kab</td>
<td></td>
<td></td>
<td>(V1)a</td>
<td>V1a</td>
<td>V1a</td>
<td>(V1)a</td>
<td>(V1)a</td>
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<td>dix2a</td>
<td>4V1 kab</td>
<td>4V1 kab</td>
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<td>(V1)a</td>
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<td>V1a</td>
<td>(V1)a</td>
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<tr>
<td>dix2b</td>
<td>4V1 kab</td>
<td>4V1 kab</td>
<td></td>
<td>(V1)a</td>
<td>V1a</td>
<td>V1a</td>
<td>(V1)a</td>
<td>(V1)a</td>
<td>V1a</td>
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<tr>
<td>osterix</td>
<td>3V1a</td>
<td>3V1a</td>
<td></td>
<td>(V1)a</td>
<td>V1a</td>
<td>V1a</td>
<td>na</td>
<td>na</td>
<td>(V1)a</td>
</tr>
</tbody>
</table>

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### Table 2. Summary of the tooth phenotypes found in various developmental mutants.

<table>
<thead>
<tr>
<th>mutant name</th>
<th>affected gene</th>
<th>pathway</th>
<th>number of teeth</th>
<th>comments</th>
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</thead>
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<tr>
<td>acerebellar (ace)</td>
<td>fgf8a</td>
<td>FGF</td>
<td>1–3</td>
<td>fish with more severe phenotype have single tooth</td>
</tr>
<tr>
<td>daedalus (dae)</td>
<td>fgf10a</td>
<td>FGF</td>
<td>3</td>
<td></td>
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<tr>
<td>lia (lia)</td>
<td>fgf3</td>
<td>FGF</td>
<td>3</td>
<td>normal shape</td>
</tr>
<tr>
<td>dreumes (dre)</td>
<td>sufu</td>
<td>IHH</td>
<td>3</td>
<td></td>
</tr>
<tr>
<td>detour (dtr)</td>
<td>gli1</td>
<td>IHH</td>
<td>3</td>
<td></td>
</tr>
<tr>
<td>you too (yot)</td>
<td>gli2a</td>
<td>IHH</td>
<td>1</td>
<td>broad and short tooth</td>
</tr>
<tr>
<td>u-shaped somites (you)</td>
<td>scube2</td>
<td>IHH</td>
<td>3</td>
<td></td>
</tr>
<tr>
<td>hi1002</td>
<td>casnk1</td>
<td>WNT/IHH</td>
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<td></td>
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<td>wnt5b</td>
<td>WNT</td>
<td>3</td>
<td></td>
</tr>
<tr>
<td>silberblick (sib)</td>
<td>wnt11</td>
<td>WNT</td>
<td>3</td>
<td></td>
</tr>
<tr>
<td>heart and soul (has)</td>
<td>prkci</td>
<td>PKC</td>
<td>1</td>
<td>normal shape</td>
</tr>
<tr>
<td>white tail (mis)</td>
<td>mib</td>
<td>NOTCH</td>
<td>3</td>
<td>normal shape</td>
</tr>
<tr>
<td>boxer (box)</td>
<td>ext13</td>
<td>3V and especially 5V are delayed</td>
<td></td>
<td></td>
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<tr>
<td>dockel (dak)</td>
<td>ext2</td>
<td>thick and short</td>
<td></td>
<td></td>
</tr>
<tr>
<td>hi307</td>
<td>b3gal3</td>
<td>3</td>
<td></td>
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<tr>
<td>pinicher (pic)</td>
<td>papst1</td>
<td>only 1 tooth ossifies</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

Mutant were raised till 6 dpf, fixed and stained with alizarin red. Dissected and flat mounted pharyngeal arches were analysed for number of the attached teeth, number of ossified teeth and abnormalities in tooth shape.

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done even on patients who are diagnosed with solitary osteochondroma than it is possible that many people are undiagnosed or misdiagnosed and the overall number of people having MO might be substantially higher. Recently Yasuda and colleagues [30] described dental defect in mice with null mutation in the Ndst1 gene. This implies that putative patients with a mutation in other genes involved in the biosynthesis of HS might increase the number of people with dental defects due to HS insufficiency. However, as patients with mucopolysaccharidosis who accumulate abnormally high levels of HS also have dental defects, it seems that any imbalance in HS might result in dental defects.

Here, we demonstrated that mutation in ext2 causes dental defect in zebrafish. Furthermore, we validated this observation in a pilot study on a group of MO patients. Our findings strongly suggest that MO is associated with dental problems. Problems with teeth might severely affect not only the self-esteem but also health of patients, hence it is important that MO patients receive appropriate dental care. More detailed studies on a larger group of patients should be performed for better understanding the clinical spectrum of this novel aspect of MO.

Materials and Methods

Animals and histology

Unless stated otherwise, all methods were based upon standardized protocols [31]. Zebrafish (Danio rerio H.) TL strain was used as wild type (WT). Homozygote acerebellar (ace, fgf8at282c), boxer (box, ext2tw24), dackel (dak, ext2to273b), detour (dtr, gli1tm276b), dreamies (drie, saga3146d), lia (fgf3, liaH0006-01), heart and soul (has, prkci m129), kanyek (kay, gpcV348), pinscher (pic, slc35b214mx), pipetail (ppt, wnt5bt265), hi307(b3gat3hi307), hi954(ux-s1hi954), hi1002(csk1a1hi1002), silberblick (slb, wnt11tx226), u-shaped somites (you, scubety97), you too (yot, gli2a ty119) and white tail (mibta52b) mutants were obtained in natural crosses and staged according to Kimmel et al. [32] Cartilages and bones were stained with Alcian blue and alizarin red respectively as described in [7].

Ethical statement

Patients data were obtained and handled according to ethical guidelines as described in the Code for Proper Secondary Use of Human Tissue in the Netherlands of the Dutch Federation of Medical Scientific Societies (www.federa.org). The institutional Review Board at the Department of Pathology at Leiden University Medical Centre imposes the use of the guidelines to any study performed with the use of human material. As the material was used in an anonymous, coded form informed consent was obtained. The study was approved by the Ethical Review Board of the University Medical Centre Leiden.

Figure 4. Tooth phenotype in zebrafish mutants affected in FGF, IHH and PKC pathways. Pharyngeal arch together with teeth from 6-days-old WT and dackel (dak/ext2), detour (dtr/gli1), heart and soul (has/prkci), pinscher (pic/sl35b2) and you too (yot/gli2a) homozygote mutants were stained with alizarin red, dissected and flat mounted. The average length of the 5th arch in WT larvae is 150 μm. doi:10.1371/journal.pone.0029734.g004

Figure 5. FGF8 stimulates growth of additional tooth-bud-like structures in ext2/- mutant. Beads were implanted at 36–39 hpf on one side of the body into an area in between the heart, ear and pectoral fin, where the teeth start to form. At 5 dpf, fish were fixed and stained with Alizarin red. Tooth-buds-like structures were formed on the pharyngeal arch neighboured by FGF-coated bead (arrowhead). Opposite arch was not affected. The tooth-bud-like structures were observed on each side of ext2/--tooth. Asterisk indicate position of the bead. doi:10.1371/journal.pone.0029734.g005
was not required. Zebrafish work was conducted in the Netherlands accordingly to Dutch law (article 9, Experiments on Animals Act) and did not require any licence for early stages. Studies performed with zebrafish in the UK were performed with Home Office approval under license PPL 40/2919.

**In situ hybridization**

Whole mount *in situ* hybridisation was performed as described in [33]. Antisense probes were amplified with M13 primers using *dlx2a*, *dlx2b*, *pitx2* [34], *cx43* [14] and *osterix* [35] plasmids as templates. Goat anti-DIG fab fragments (Roche) and NBT/BCIP substrate (Sigma) were used to develop the *in situ*. Prior to photo documentation embryos were cleared and preserved in 75% glycerol.

**Measurements**

Teeth were analysed in homozygote mutants and WT at 6 dpf. Measurements of the tooth length and width were taken on pictures of the dissected and flat mounted pharyngeal arches. Teeth were also dissected out along with whole pharyngeal arches from one year old fish. As zebrafish teeth undergo replacement throughout life, only attached teeth were analysed in this study. Teeth organisation was examined in adult heterozygote mutants and their siblings and compared to the pattern described previously [16]. Teeth from adult fish were also screened for other abnormalities such as incomplete enamel formation and crown splits. Teeth from WT and *ext2*+/2 fish were cross-sectioned and subjected to histological analysis. Groups of minimum ten fish were used for each measurement.

**Chemical treatment**

Drugs were added into Petri dish with 50 manually dechorionated embryos in 25 ml E3. FGF inhibitor (SU5402), IHH inhibitor (cyclopamine), IHH activator (purmorphamine), PKC inhibitors (Go6976, Go6983 and Bimü) and PKC activator (PMA)
were all purchased from Merck/Calbiochem. A 10 mM stock solution of cyclopamine in Ethanol was added into fish water to give final concentration corresponding to 10–100 μM. Stock solution of 40 μM SU5402 in DMSO was diluted in fish water up to final concentration of 40–200 nM. 10 mM stocks of Go6976, Go6983 and Bimi in DMSO were added into fish water and used at final 10–100 μM concentration. 1 mM PMA stock in DMSO was diluted in fish water and tested at concentrations: 0.02–1 μM.

Tested compounds and corresponding solvents (controls) were added to E3 at 24, 36 and 50 hpf and either kept for 24 h and washed off with fresh E3 or left unmoved up to 6 dpf, when fish were fixed and subjected to Alcian blue and alizarin red staining.

Bead implants

Heparin beads (Sigma) were coated with mouse recombinant FGF-8b (R&D systems) as described [36,37]. BSA-coated beads were used for control. Beads were implanted at 36–39 hpf into an area in between the heart, ear and pectoral fin, where the teeth start to form. Fish were raised until 96 hpf for molecular analysis (mRNA in situ hybridisation) and until 5 dpf for histological examination (alizarin red stain). Mice and zebrafish FGF8 share a high level of conservation (Figure S9).

Dental questionnaire

Dental questionnaire was design in a way that it could be answer without the need of consulting a dentist or another medical professional. The questionnaire was intended as an online survey for patients and their families (socio-economically matched control). The survey was placed on the EuroBoNet homepage (http://www.eurobonet.eu/news/News.php) and at the MHE Research Foundation website (http://mheresearchfoundation.org/HOME.html) located on an encrypted secure webpages. In addition, three patient support groups were directly invited to participate in the survey. Responses were collected, coded by an independent body in order to keep anonymity and analysed. The complete questionnaire is given in the Text S1.

Supporting Information

Figure S1 Simplified representation of the tooth development in the zebrafish larvae. During the first week of life, zebrafish develops pharyngeal teeth at three positions only, 3V, 4V and 5V. Tooth 4V1 is the first tooth to differentiate (48 hpf), attach into the pharyngeal arch (80 hpf) and undergo replacement by 4V2 at 12 days post fertilisation. Teeth, 3V and 5V start to differentiate at 56 hpf and become attached at 144 hpf [12,13]. (TIF)

Figure S2 Tooth ossification is delayed in ext2+/− mutant. Is indicated by Alizarin red stain at 96 hpf, single tooth is ossified in both ext2+/− mutant and its siblings. Interestingly, the ossification of the pharyngeal arches starts in the mid part in siblings (A, A’) and at the end of arch in ext2+/− mutant (B, B’).
Moreover, weaker intensity of the Alizarin red in ext2<sup>+/−</sup> suggests general delay in ossification. A’ and B’, line outline of the branchial arch 5 and attached teeth. Scale bar = 0.1 mm. (TIF)

Figure S3 The expression pattern of pitx2 indicates slight defect in thickening of the pharyngeal epithelium in the ext<sup>2</sup>−/− mutant. pitx2-expressing bilateral domain in the ext<sup>2</sup>−/− are of similar length but more narrower than the one from siblings. A, siblings and B, ext<sup>2</sup>−/− at 56 hpf. A’ and B’, magnification of the pharyngeal area. Scale bar = 0.1 mm. (TIF)

Figure S4 Inhibition of PKC affects tooth formation. Similarly to has (PKC) mutant, one tooth-phenotype was also observed in fish treated with PKC inhibitor. PMA – activator of PKC does not stimulate formation of additional teeth in WT, nor rescues tooth phenotype in the dak homozygote mutant. Cartilaginous skeletons were stained with Alcian blue at 6 dpf. Pharyngeal arches were dissected out and flat mounted. (TIF)

Figure S5 Inhibition of FGF by SU5402 tooth formation in WT and dak mutant. Embryos were treated from 50 hpf till 6 dpf. The ext<sup>2</sup>−/− fish treated with SU5402 does not form teeth hence picture was not included. Alizarin-red-stained pharyngeal arches were dissected and flat mounted. Arrow points a single bilateral tooth formed in dak siblings. (TIF)

Figure S6 Tooth morphology in adult dak heterozygote mutant. Cross section of teeth from adult fish did not reveal any obvious morphological differences between WT and ext<sup>2</sup>−/−. 4 μm sections of teeth were stained with hematoxalin and cosin. (TIF)

References


Figure S7 Schematic representation of adult human and zebrafish teeth. Organization of the zebrafish tooth was adapted from work by Neues and colleagues [18]. (TIF)

Figure S8 Accumulation of cells undergoing cell death at the end of the pharyngeal arch in the ext<sup>2</sup>−/− mutant. TUNEL staining was performed in fish at 6 dpf. Pharyngeal arches were dissected and flat mounted. (TIF)

Figure S9 Alignment of the zebrafish and mouse FGF8 shows high level of conservation between proteins. Accession numbers: mouse FGFb, P37247-2; zebrafish FGF8a, Q5PRC3; and zebrafish FGF8b, B3DJ36. (TIF)

Text S1 Dental questionnaire for MO patients and their families. (PDF)

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Author Contributions

Conceived and designed the experiments: MIW RJPvM PCWH. Performed the experiments: MIW ZZ. Analyzed the data: MIW ZZ RJPvM PCWH. Contributed reagents/materials/analysis tools: HHR PCWH. Wrote the paper: MIW ZZ RJPvM HHR PCWH.

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