

## **Automated construction of large kinetic models: mitochondrial respiratory complex III**

In the presented example the model of mitochondrial respiratory complex III, which was constructed based on the known biochemical mechanisms of electron and proton transport, simulated the experimentally measured triphasic reaction of cytochrome b<sub>H</sub> reduction by succinate in isolated succinate-cytochrome c reductase. These simulations validated the model and algorithms for its automated construction.

Multienzyme complexes, or multiple regulatory sites, which are of great importance in systems biology, require the construction of large number of differential equations. Even if such a complex has just several components, the combination of various states of each components result in huge number of states for the whole complex, and, respectively, equations to describe their evolution. An example of such multienzyme complex could be respiratory complexes in mitochondria.

The operation of respiratory complex III was simulated here using the basic principles of automated differential equations construction as described in (Selivanov et al, 2009) and schematically shown in Figure S1. The biochemical mechanism of electron transport and proton translocation for complex III, known as “Q-cycle” (Mitchell, 1975), was implemented in this model. The specific triphasic behavior of cytochrome b<sub>H</sub> reduction in the isolated complex is a direct consequence of Q-cycle mechanism (Tang and Trumpower, 1986). The origin of each phase of this reaction was analysed in details in a nice theoretical work (West, 1989). Here we simulated this reaction to show that the model reproduces this direct consequence of Q-cycle mechanism as a validation of our algorithms for model construction.

### **Simulation of particular experiments**

In experiments, evidently, could not be measured all the forms, which are accounted by the model, although the whole distribution of redox form defines the experimental measurement. To compare the result of calculation with experimental data, the program summarizes the calculated concentrations of redox form, so that the summarized value corresponds to that measured. For instance if reduced cytochrome b<sub>H</sub> is measured, the program summarizes all concentrations of all species for which b<sub>H</sub> reduced, e.g. for core it summarizes all forms 1xxx, etc. The obtained value of reduced b<sub>H</sub> could be compared with respective experimental data.

### **Implementation**

In a one of fundamental experimental works that reveal the main properties of complex III operation, it was shown that the reaction of cytochrome b<sub>H</sub> reduction by succinate in isolated succinate-cytochrome c reductase (complex III + complex II) has unusual triphasic behavior (Tang and Trumpower, 1986). In the original experimental study it was shown that such behavior is a direct consequence of Q-cycle

mechanism. Later, using fast equilibrium simulations, West (1989) had elucidated the mechanics of reduction, oxidation and subsequent ultimate reduction of  $b_H$ . Here we also simulated this triphasic reaction in order to check, whether our automatically constructed model correctly reflects the basic biochemical mechanism, which underlies such unusual behavior. Moreover, reproducing the experimentally observed behavior, we show a way of defining the proper parameters of system.

### **Model simulation of triphasic cytochrome $b_H$ reduction by succinate in isolated succinate-cytochrome c reductase.**

Figure S2 shows a model simulation of cytochromes  $b_H$  and  $c_1$  and ubiquinone reduction by succinate in the isolated system of complex III and complex II (there is no acceptors for  $c_1$ ), calculated with initial set of parameters. This initial set of parameters allows to reproduce the triphasic behavior of  $b_H$  reduction.

As it was proved experimentally (Tang and Trumpower, 1986) and theoretically (West, 1989), first phase of  $b_H$  reduction proceeds in accordance with the conventional way of net electron flow in complex III: ubiquinol produced by succinate dehydrogenase and bound at  $Q_o$  site delivers its first electron to  $Fe^{3+}$  of Rieske center, which then reduces  $c_1$ . The second electron of ubiquinol passes to heme  $b_L$  and then reduces  $b_H$ . In this way, in accordance with Q-cycle mechanism, reduction of  $c_1$  stoichiometrically linked with reduction of  $b_H$ .

In our simulation the initial rate of  $b_H$  reduction coincides with that for  $c_1$ . This indicates that the model correctly describes the separation of two electrons of  $QH_2$  in accordance with Q-cycle mechanism. Later the reduction of  $b_H$  gets slower because reduced  $b_H$  again oxidized at  $Q_i$ .

Since the experimental system lacks a respective electron acceptor, cytochrome  $c_1$  ultimately becomes completely reduced. As Q-cycle mechanisms assumes, reduction of  $c_1$  and thus block in electron flow coming from ubiquinol bound at  $Q_o$  site in the same time blocks the delivery of second electron to cytochrome b. Thus,  $Q_o$  site becomes blocked, which terminates the normal way of  $b_H$  reduction and first phase. Since  $b_H$  continues reducing ubiquinone at  $Q_i$  site, the levels of  $b_H$  reduction decrease, which constitutes the second phase of considered triphase reaction.

During the first phase, ubiquinol, produced by succinate dehydrogenase, is oxidized at  $Q_o$  site and therefore does not accumulate. After the  $Q_o$  site gets blocked at the end of first phase, succinate dehydrogenase reaction results in the accumulation of ubiquinol. This accumulation induces third phase of  $b_H$  reduction, which proceeds through  $Q_i$  site in the direction reverse to the normal net electron flow in Q-cycle.

The described sequence of events was revealed in the original paper (Tang and Trumpower, 1986). However, since such triphasic behavior strictly based of Q-cycle mechanism, the objective of presented

example was to show that our automated procedure correctly construct equations that correspond to the generally accepted mechanism of complex III operation. Moreover, in addition to the description of experimentally observed behavior, model simulation allowed revealing the details which were not measured, such as dynamics of ubiquinol production.

### **The analysis of characteristics of complex III operation**

Figure S3 shows how the change of these values would affect the shape of trace of  $b_H$  reduction. If the dissociation constants change so that to favor the forward direction of Q-cycle, i.e. tenfold increase rate of binding of  $QH_2$  at  $Q_o$  and  $Q$  at  $Q_i$  with respect to the rate of dissociation, and increase the rate of dissociation of  $Q$  at  $Q_o$  and  $QH_2$  at  $Q_i$  with respect to the rate of binding, the third phase of  $b_H$  reduction slows down as curve 1 indicates (compared to curve 0, which is calculated for initial set of parameters, as in Figure S2). If with the set taken for curve 1 only  $K_d$  for  $QH_2$  dissociation from  $Q_i$  (reaction 9) decreases favoring the direction reverse to Q-cycle, the third phase of  $b_H$  reduction accelerates (curve 2). These simulations show that fitting a particular experimental curve could be very useful in the determination of  $K_d$ , which essentially affect the third phase of  $b_H$  reduction.

In the literature we found a large variability of  $\Delta E_m$  values for first and second electron transition from  $b_H$  to  $Q$  at  $Q_i$ . This value varies even qualitatively, being negative (Rich, 1984) or positive (Covian et al, 2007) for the first electron, and positive (Rich, 1984) or negative (Covian et al, 2007) for the second electron. For the initial simulation we have taken  $\Delta E_m=0$  for both cases, but variation of these values in simulations would help to define these parameters for a particular experiment.

Figure S4 shows the simulations of  $b_H$  reduction for the different combinations of  $\Delta E_m$  for the first and second electron transitions from  $b_H$  to  $Q$  at  $Q_i$ .

If the values reported in (Rich, 1984) are taken for simulation, the phase 3 practically disappears. In contrary, if the values given in (Covian et al, 2007) are accepted, the third phase begins essentially before the completion of the second phase. It seems that the form of traces of  $b_H$  reduction is extremely sensitive to these parameters and fitting of a particular experiment would help to define them.

Thus, fitting particular experiments allows defining the parameters of the model and in this way to understand better the operation of respiratory chain in a specific experimental conditions.

### **References**

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