

Table S6. Lineage of the *rrn* inactivation strains. The following table traces the lineage of all *rrn* inactivation strains and indicates the length of each inactivated *rrn* operon that was transcribed, and in cases where the inactivated operon expressed a protein (*cat* or *lacZ* insertions), the length of the translated portion is also specified. A Δn strain indicates that n operons were inactivated. See S1.3 in Text S1 for further details.

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|---|---|
| $\Delta 0 = rrn^+ = TA563$ | source: $\Delta TX11$ from [106] |
| $\Delta 1 = \Delta Ac = TA566$ | |
| A transcription: 2147bp (with P_1P_2 -CAT fusion) ¹ . translation: cat^+: 657bp | source : P_1P_2 -CAT fusion BAG1 in [54] ² |
| $\Delta 2 = \Delta EAc = TA567$ | |
| E Completely deleted | source TX $\Delta 11$ [106]. See also fig. 1 in [91] |
| A transcription: 2147bp (with P_1P_2 -CAT fusion; see remarks in $\Delta 1$). translation: cat^+: 657bp | source : P_1P_2 -CAT fusion BAG1 in [54] |
| $\Delta 3 = \Delta EBAC = TA568$ | |
| E Completely deleted | source TX $\Delta 11$ [106]. See also fig. 1 in [91] |
| B transcription: 2287(PCR:16S-23S)+119(5S)³ = 2406 bp ⁴ | source: pMA101, Sall-Sall deletion [7] |
| A transcription: 2147bp (with P_1P_2 -CAT fusion; see remarks in $\Delta 1$). translation: cat^+: 657bp | source : P_1P_2 -CAT fusion BAG1 in [54] |
| $\Delta 4 = \Delta EBHAc = TA430$ | |
| E Completely deleted | source TX $\Delta 11$ [106]. See also fig. 1 in [91] |
| B transcription: 2287(PCR:16S-23S)+119(5S) = 2406 bp (see remarks in $\Delta 3$) | source: pMA101, Sall-Sall deletion [7] |
| H transcription: 1290(PCR:16S-23S) + 248(5S+distal tRNA) = 1538 bp ⁵ | source: pMA103, SacII-SacII deletion [7] |
| A transcription: 2147bp (with P_1P_2 -CAT fusion; see remarks in $\Delta 1$). translation: cat^+: 657bp | source : P_1P_2 -CAT fusion BAG1 in [54] |
| $\Delta 5 = \Delta EBHGzAc = TA476$ | |
| E Completely deleted | source TX $\Delta 11$ [106]. See also fig. 1 in [91] |
| B transcription: 2287(PCR:16S-23S)+119(5S) = 2406 bp (see remarks in $\Delta 3$) | source: pMA101, Sall-Sall deletion [7] |
| H transcription: 1290(PCR:16S-23S) + 248(5S+distal tRNA) = 1538 bp (see remark in $\Delta 4$) | source: pMA103, SacII-SacII deletion [7] |
| G transcription: 2928(16S-23S) + 3075(<i>lacZ</i>⁺)+212(5S) = 6215 ⁶ . translation: <i>lacZ</i>⁺ 3075 | source pNY30, smaI-HpaI deletion in [7] |
| A transcription: 2147bp (with P_1P_2 -CAT fusion; see remarks in $\Delta 1$). translation: cat^+: 657bp | source : P_1P_2 -CAT fusion BAG1 in [54] |
| $\Delta 6 = \Delta EBHGzADc = TA516$ | |
| E Completely deleted | source TX $\Delta 11$ [106]. See also fig. 1 in [91] |
| B transcription: 2287(PCR:16S-23S)+119(5S) = 2406 bp (see remarks in $\Delta 3$) | source: pMA101, Sall-Sall deletion [7] |
| H transcription: 1290(PCR:16S-23S) + 248(5S+distal tRNA) = 1538 bp (see remark in $\Delta 4$) | source: pMA103, SacII-SacII deletion [7] |
| G transcription: 2928 (16S-23S) + 3075(<i>lacZ</i>⁺)+212(5S) = 6215 (see remark in $\Delta 5$). translation: <i>lacZ</i>⁺ 3075 | source pNY30, smaI-HpaI deletion in [7] |
| A transcription: 1290 (<i>rrnH</i> PCR:16S-23S) + 213(5S) = 1503bp ⁷ | source: pNY34, SacII-SacII deletion [7] |
| D transcription: 2147bp (with P_1P_2 -CAT fusion). translation: cat^+: 657bp | source: W1485 ΔD in [19] ⁸ |

¹ Measured from figure 1 in [54].

² Lineage is traced as follows:

(i) BAG1 from [54] is designated CC164 in [19] and the A was inserted into the final strain.

(ii) A originated from the A1 strain of [54].

(iii) A1 was taken from [7] where it was designated W1485 ΔA . W1485 ΔA was formed by cutting *rrnA* at the first smaI restriction enzyme site (see figure 1 in [19]) and attaching to it a promoterless CAT gene from plasmid pKK232-8 [107]. pKK232-8 was also cut at smaI ([7] p. 4184, [107] p. 157). This is the first smaI site (see figure 1 in [19]) since according to [54] the CAT was inserted at position 612 bp from the start of 16S. According to figure 1 of [19] the first smaI is located at 605 bp position (assuming WT 16S-23S is 4719 bp [19]). This confirms that the scaling for this figure is correct.

(iv) pKK232-8 was designed by [107] and they used a CAT cassette based on [108].

(v) In [108] it is explained that CAT are derivatives of the Cm^r gene of the bacterial transposon Tn9 and contains the complete CAT polypeptide coding sequence. According to [108] this is 660 bp. This is confirmed in NCBI site for pKK232-8 (=657bp). Next we need to ascertain the length of the transcribed segment. The actual length of the CAT cassette in pKK232-8 is longer than 660 bp since (a) the coding region actually taken in [108] was 780bp and (b) pHH232-8 was cut at smaI which is upstream of the CAT cassette (figure 7 in [107]). Since the exact location of smaI was uncertain, the length of the transcribed gene was estimated from figure 1 in [54].

³ Measured from end of 23S

⁴ PCR length from [19] + 5S length taken from the NCBI site.

⁵ PCR length from [19] + 5S+tRNA length taken from the NCBI site.

⁶ Assuming a length of 4876 bp for the 16S-23S segment of *rrnG* (NCBI site) and from scaling of figure 1 in [19]. The first smaI was chosen, as described in figure 1 of [91]. *LacZ* length is from EcoCyc site, 5S length is from the NCBI site.

⁷ Assumed to be like *rrnH* PCR fragment since the 16S-23S sequence length of *rrnA* and *rrnH* are both 4892 bp and the same restriction enzymes were used; 5S length was taken from the NCBI site.

⁸ W1485 ΔD in [19] was obtained by the same procedure as W1485 ΔA described above: a promoterless CAT gene was inserted into the first smaI site at 612 bp; (see $\Delta 1$).