

**Table s1. SNPdetector Parameters Used to make Genotype and SNP calls.**

Parameter	Type of Variable	Underlying data	Description
<b>disqualify_by_stutter</b>	Binary	Ordinal	SNP is within a stutter region
<b>is_mismatch_cluster</b>	Binary	Ordinal	4 or more mismatches with phred quality >30
<b>good_homo_allele</b>	Binary	Ordinal	Quality check of adjacent bases
<b>strand_conflict</b>	Binary	Ordinal	Sample A has only single strand coverage and it has homozygous minor allele. However, in other samples (eg. B or C) with sequences from both orientations, the minor alleles detected in the same direction as sample A were found to be in conflict with the alleles detected in the other orientation. This information was used to evaluate potential sequencing error in those with single-strand coverage.
<b>pass_dirty_check</b>	Binary	Continuous	2° peak of het is distinct from “dirty” homo
<b>neighbor_spill</b>	Binary	Continuous	The 2° peak of the current position extends to more than 1 base.
<b>spill_ratio<sup>a</sup></b>	Continuous	Continuous	The max 2° peak height divided by the min 2° peak height for a putative “spill” region.
<b>max_regional_dirty_peaks<sup>b</sup></b>	Binary	Continuous	in the surrounding 20bp window, the maximum number of 2° peaks that exceed 80% of the 2° peak height of the current site.
<b>skip_hetero_analysis</b>	Binary	Ordinal	>90% of the reads have double peaks and longest region with no 2o peak is less than 50bp
<b>drop_a1_ratio<sup>c</sup></b>	Continuous	Continuous	The peak drop ratio of the first allele in a putative heterozygote compared to its homozygote counterpart.
<b>drop_a2_ratio</b>	Continuous	Continuous	Same as drop_a1_ratio except comparing the 2° peak to its homozygous read.
<b>hetero_has_peak_drop<sup>d</sup></b>	Categorical	Categorical	<b>0, no peak drop compared to corresponding homozygotes; 1, heterozygote has a peak drop compared to corresponding homozygotes; 2, fail to find a homozygote with comparable flanking peak profile.</b>
<b>flanking_region_score</b>	Continuous	Continuous	<b>Maximum of scores determined from flanking 4 bp and flanking 20 bp regions.</b>
<b>is_clean_hetero<sup>e</sup></b>	Binary	Categorical	<b>True if 5 tests in footnote e pass.</b>
<b>pass_poly_check<sup>f</sup></b>	Binary	Categorical	>0 if 2° peaks of a putative SNP site (homozygote or heterozygote) and its 4-bp flanking region have test in footnote e.
<b>High mismatch (short range)</b>	Binary	Ordinal	20 bp window <16 are identical to reference
<b>High mismatch (long range)</b>	Binary	Ordinal	In a 50 bp window, the distance between short-range High-mismatch segments is <5bp
<b>Low quality (short range)</b>	Binary	Ordinal	5 bp window, average quality below 20
<b>Low quality (long range)</b>	Binary	Ordinal	In a 50 bp window, the distance between short-range Low-quality segments is <5bp
<b>Region of 2° peaks</b>	Binary	Ordinal	10 bp window, 7 positions have 2° peak ratio ≥0.1

(short range)			
<b>Region of 2° peaks</b> (long range)	Binary	Ordinal	In a 50 bp window, the distance between short-range 2° peak is <5bp
<b>Region of high 2° pks</b> (short range)	Binary	Ordinal	10 bp window, 7 positions have 2° peak ratio ≥0.3
<b>Region of high 2° pks</b> (long range)	Binary	Ordinal	50 bp window
<b>Mismatch_cluster</b> (short range)	Binary	Ordinal	≥2 bp high quality (phred 30) mismatches in a 20 bp window.
<b>Mismatch_cluster</b> (long range)	Binary	Ordinal	≥4 bp high quality (phred 30) in a 40 bp window

a, Use of **spill\_ratio**. This ratio differentiates a spill from a SNP cluster. The latter tends to have similar secondary peak heights (e.g. spill ratio close to 1) while the former tends to have a large difference.

b, Use of **max\_regional\_dirty\_peaks**. This information was used to determine if the background noise of two traces are comparable in the “vertical” scan.

c, Use of **drop\_a1\_ratio**. A putative heterozygote site is compared to each of the homozygous read of the same orientation: to determine a) whether the left and the right flanking primary peaks in the two reads are comparable. A -1.0 value is assigned to those with incomparable homozygous flanking peaks; b) else (e.g. the flanking peaks are comparable), normalize the primary peak ratio of the homo/hetero at the SNP site to the average of homo/hetero at the left and the right flanking sites. The average ratio of all pair wise het-to-homo comparison (excluding the -1.0 cases) will be stored.

d, Use of **hetero\_has\_peak\_drop**. This value is initially set by the value of **drop\_a1\_ratio** of 0.55 (almost 50% reduction of a primary peak) subject to the following revisions:

- The forward and the reverse read have the same genotype and the reduction of the primary peak + the rise of the secondary peak ratio is approximately 1. This shows that the reduction of the primary peak can be explained by the addition of the secondary peak.
- When the secondary peak ratio of a putative heterozygote is less 20% of a dirty homozygote, the **peak\_drop\_ratio** is reset to 0.
- If a heterozygote has clean flanking region and its reduction of the primary peak can be explained by the addition of the secondary peak, then the flag is set to 1.

A non-clean heterozygote is used for SNP call only when its **hetero\_has\_peak\_drop** flag was set to 1.

e, Determination of **is\_clean\_hetero**.

- i. The putative heterozygote does not fit into a “spill” profile, i.e. a neighboring homozygote followed by at least 2 secondary peaks (with diminishing secondary peak area ratio) in its neighbor. This profile is evaluated with a sliding window method.
- ii. The heterozygote does not have any indel on its immediate left or right side.
- iii. The secondary peak represents a residue different from those of the primary peaks of its left and right neighbors.
- iv. There are no drastic peak height differences between the primary peak of the putative heterozygote site and its left/right neighbors. Specifically, the primary peak height should be ≥1/6 of its neighbor and ≤2 of its neighbor. If both the left and the right neighbor fail to meet this criterion, then the site fails in this test. The ≥1/6 test ensures that the site does not look like a deep valley (normally indicates a potential sequencing error). The ≤2 test will exclude a site if the primary peak appears to be twice as high as its neighbor because a heterozygote is expected to have its primary peak reduced compared to a homozygote. The reduced primary peak usually has lower peak height than its neighbors.
- v. The flanking region, excluding those that may appear to be putative heterozygote (secondary peak ratio ≥0.70), contains no site of secondary peak ratio ≥5%. If the secondary peak ratio of a putative heterozygote is below 60, then the test requires absence of secondary peak in the flanking region.

“#” in output indicates that a putative heterozygote has no noisy background (i.e. is **clean\_hetero**) nor apparent abnormalities in both its primary and secondary peaks compared to its immediate neighbors.

f, Calculation of **pass\_poly\_check**:. Define  $P = (\text{secondary\_peak\_area} / \text{primary\_peak\_area}) * 100$  (i.e. percent of primary peak area occupied by secondary peak). To evaluate noise at the flanking regions of a putative heterozygote or a homozygote, the program checks the secondary peak of each base in the flanking region. If each base in the flanking region passes the test of ( $P \leq 0, \leq 10, \leq 20$ ), then the flanking region is considered to have no, little, limited noise. To avoid penalizing secondary peak of a potential heterozygote in the flanking region, a site with a secondary peak ratio  $\geq 0.70$  is skipped. At the SNP site, the same test is applied to measure the noise level at a homozygous genotype. For a heterozygous genotype, the high, med and low is rewarded to those with  $P \geq 80, \geq 50$  and  $\geq 35$  respectively. \$ in output indicates pass\_poly\_check is greater than 0.